



Perspective Tellurium-Modified Nucleosides, Nucleotides, and Nucleic Acids with Potential Applications

Cen Chen ^{1,2} and Zhen Huang ^{1,3,*}

- ¹ SeNA Research Institute and Szostak-CDHT Large Nucleic Acids Institute, Chengdu 610041, China
- ² Firebird Biomolecular Sciences LLC, Alachua, FL 32615, USA
- ³ College of Life Sciences, Sichuan University, Chengdu 610064, China

Correspondence: huang@senaresearch.org

Abstract: Tellurium was successfully incorporated into proteins and applied to protein structure determination through X-ray crystallography. However, studies on tellurium modification of DNA and RNA are limited. This review highlights the recent development of Te-modified nucleosides, nucleotides, and nucleic acids, and summarizes the main synthetic approaches for the preparation of 5-PhTe, 2'-MeTe, and 2'-PhTe modifications. Those modifications are compatible with solid-phase synthesis and stable during Te-oligonucleotide purification. Moreover, the ideal electronic and atomic properties of tellurium for generating clear isomorphous signals give Te-modified DNA and RNA great potential applications in 3D crystal structure determination through X-ray diffraction. STM study also shows that Te-modified DNA has strong topographic and current peaks, which immediately suggests potential applications in nucleic acid direct imaging, nanomaterials, molecular electronics, and diagnostics. Theoretical studies indicate the potential application of Te-modified nucleosides in cancer therapy.

Keywords: tellurium; crystallography; nucleic acids



Citation: Chen, C.; Huang, Z. Tellurium-Modified Nucleosides, Nucleotides, and Nucleic Acids with Potential Applications. *Molecules* 2022, 27, 8379. https://doi.org/ 10.3390/molecules27238379

Academic Editors: Xinjing Tang and Changmai Chen

Received: 20 October 2022 Accepted: 23 November 2022 Published: 1 December 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/).

1. Introduction

Beyond all doubt, nucleic acids are the most important biomolecules for all known forms of life, which store all genetic information and pass it from one generation to the next. Extensive studies of nucleic acid have revealed its structure, functions, and properties [1–4]. The unique properties of nucleic acid promote its application in lots of areas such as diagnostics [5–12], therapeutics [13–18], crystallography [19–22], catalysis [23–27], material science [28–32], as well as vaccinology [33–38]. It is worth mentioning that nucleic acid as a therapeutic has greatly developed in recent decades and 14 oligonucleotide drugs have been approved since 1998, including siRNA, antisense, aptamer, etc. [39].

Notably, modified nucleic acid has played a vital role in almost all the above applications, because of the narrow chemical diversity and poor physiological stability of nucleic acids which restrict their utilization. Various strategies of modification have been employed to develop novel DNA and RNA analogs to overcome those limitations, such as changing the structure of the backbones [40–48], sugars [49–55], nucleobases [56–65], introducing different functional groups to endow the nucleic acid with specific function [66–73], or introducing special elements that do not exist in nucleic acids such as fluorine [74–77], bromine [78–80], mercury [81], and other heavy atoms [82].

As the same group element of tellurium, selenium modifications have been widely applied for protein structure determination, by replacing the sulfur in methionine, in which the selenium can be used as an ideal scattering center for multiwavelength anomalous dispersion (MAD) [83–86]. The selenium-derivatized nucleic acid (SeNA) has also achieved great success in 3D crystal structure determination and selenium has been introduced to different positions of the ribose, the phosphate backbone, as well as the nucleobases

(Scheme 1) [87–98]. These works have been well reviewed here [99,100]. It is worth mentioning that the incorporation of 2'-selenium modified nucleoside into DNA oligo not only solved the phase problem but also greatly facilitated crystallization, especially because, compared with protein crystallization, there are greater challenges in nucleic acid crystallization due to the negatively charged repetitive phosphate groups. Besides crystallography studies, selenium-modified nucleic acids have also been found to improve the specificity and sensitivity of DNA polymerization [12,47,101] However, compared with selenium, the application of tellurium in nucleic acid is still quite limited.



Scheme 1. Atom-specific selenium substitution of oxygen atoms in nucleic acids.

2. Discussion

Tellurium is a metalloid located in group VI, also known as chalcogens, of the periodic table following sulfur and selenium. Its chemical or physical properties, reactivities, structures, and functions have been well studied and show multiple potential applications in different areas [102-107]. Tellurium has a larger radius (1.35 Å) and a weaker electronegativity (2.0) in comparison to sulfur (1.04 A, 2.58) and selenium (1.17A, 2.55). The larger electrovalent and coordination sphere radius provide tellurium with strong metallic properties and result in weak covalent bonds with carbon and hydrogen [108]. Elemental tellurium has a lower abundance (1 ppb) than gold, platinum, or "rare-earth" elements on Earth [109]. Naturally occurring tellurium contains a series of isotopes, including 120 Te (natural abundance 0.09%), ¹²²Te (2.55%), ¹²³Te (0.89%), ¹²⁴Te (4.74%), ¹²⁵Te (7.07%), ¹²⁶Te (18.84%), ¹²⁸Te (31.74%), and ¹³⁰Te (34.08%) [110], which result in a unique isotope pattern in mass spectrometry for Te-containing compounds. Meanwhile, the diamagnetic nucleus ¹²⁵Te (spin 1/2) enables Te-NMR studies and has wide chemical shifts ranging from -1400 ppm to 3400 ppm, which facilitates the identification of tellurium compounds with different chemical environments. Moreover, it also has excellent sensitivity due to the high natural abundance (7.07%) compared with ^{13}C (1.1%) [111].

Although it belongs to the chalcogen family, the structures and chemical properties of tellurium compounds frequently differ from its family members [86,112]. For example, with the same range of oxidation states from -2 to +6 as sulfur and selenium, the higher oxidation states of tellurium are more stable due to the lower ionization energies [113,114]. In addition, the σ - and π - bond energies of tellurium are also significantly lower than their chalcogen analogs, [112,115–117] which contributes to its higher lability such as photochemical sensitivity [118] and lower tendency to catenate [119–122] compared to sulfur and selenium. Another intriguing feature of the chemistry of tellurium compounds is its proclivity to engage in hypervalent interactions, which was rationalized in terms of three-center–four-electron (3c–4e) bonding, charge-transfer interactions, hyperconjugation, and secondary bonding interactions (SBIs) [123–129].

Tellurium was successfully incorporated into protein in a tellurium-tolerant fungus in 1989, which was achieved by growing the Te-resistant fungi on a sulfur-free medium, and an extraordinarily high level of tellurium was detected [130]. Later, telluromethionine was reported to be selectively incorporated into dihydrofolate reductase [131]. Further

studies optimized the bioincorporation technique of TeMet into protein and provide a promising approach for the X-ray structure study of protein [132]. The absorption edge of tellurium is about 0.3 Å, which indicates that it is not as suitable as selenium (0.9795 Å) as a scattering center in a MAD experiment. But the ideal electronic and atomic properties of tellurium for generating clear isomorphous signals make it a suitable heavy-atom for isomorphous replacement without the need for synchrotron radiation [108]. Interestingly, besides being covalently incorporated into protein to solve the phasing problem, the tellurium-centered Anderson–Evans polyoxotungstate (TEW) was used as a universal additive to enable or improve the crystallization of proteins to achieve high-quality crystals through electrostatic interactions [133]. Moreover, it was found that the incorporation of tellurium into phycocyanin (PC) and allophycocyanin (APC) enhanced their antioxidant activities [134].

Te Modifications in Nucleoside, Nucleotide, and Nucleic Acids

The incorporation of Te into nucleic acid has also been achieved in the past decades. The first Te-modified nucleoside was reported [135] by Huang et al. in 2008 and then successfully incorporated into DNA oligo through solid-phase synthesis [136] (Scheme 2). The tellurium functionality was protected by alkylation with either the phenyl or methyl group and was introduced into the 2'-position of both uridine and ribo-thymidine with good yield.



Scheme 2. Synthesis of the Te-DNA. (**a**) PhTeTePh, NaBH4; (**b**) 2-cyanoethyl, N,N-diisopropyl chlorophosphoramidite; (**c**) solid-phase synthesis.

Interestingly, unlike the MeSe functionality [19,93,137–140], the 1',2'- and 2',3'-eliminations were observed during Te functionalization when using sodium borohydride as reducing reagent at room temperature (Scheme 3), which provides a new method for the synthesis of the 2',3'-didehydro-2',3'-didoxynucleotides (d4Ns).

To get the desired product X, a stronger reducing reagent and lower temperature (0 °C) were applied together with crown ether (12-*crown*-4) to chelate the lithium ions to enhance the MeTe reactivity. The desired product was obtained in 47% yield with the 1',2'-elimination products as the major byproduct. Both PhTe- and MeTe-modified

nucleosides were incorporated into DNA oligos by solid-phase synthesis following a standard protocol [141] and quantitative coupling yield was achieved. A few of Te-DNAs have been oxidized to tellurides during the solid-phase synthesis which can be reduced by treating with diborane after the deprotection step (Scheme 4). It was found that both methyltelluride and phenyltelluride functionalities were stable with the treatment of mild acid and base during the deprotection and purification. Interestingly, under heating (50 °C) in the presence of B_2H_6 or I_2 , 2'-TePh DNA undergoes 2',3'-elimination at the modification site and generates the fragmented product. However, the 1',2'-elimination was observed for 2'-TeMe DNA and creates the abasic product (Scheme 4). The decrease in the melting temperature was observed during the UV melting study which was probably caused by the perturbation introduced by the bulky Te functionality (Figure 1).



Scheme 3. Elimination reactions resulting from the 2'-TeMe functionality.



Scheme 4. Redox and fragmentation of Te-DNA oligonucleotides.



Figure 1. UV melting curves of DNA duplexes (5'-C(TeXU)TCTTGTCCG-3' and 3'-CGGACAAGAAG-5'; X = Me or Ph).

In 2011, 5-PhTe modified nucleoside was successfully synthesized by applying the lithium–halogen exchange reaction [142] on a protected 5-iodo-2'-deoxyuridine and achieved medium yield (64%) [143] (Scheme 5). The key steps of the reaction are the deprotonation of the NH and the treatment with n-BuLi followed by the addition of Ph₂Te₂. An elevated concentration of the reactant (0.15–0.18 M) is necessary to avoid the generation of a 6-PhTe isomer, which is inseparable. The synthesis of the corresponding phosphoramidite followed the standard protocol and applied to solid-phase synthesis. The results show that the PhTe functionality is well compatible with the solid-phase synthesis condition, deprotection, and purification. The UV-thermal denaturation studies show similar stability between the Te-derivatized duplex and the corresponding native which suggested that the bulky PhTe moiety is well accommodated and does not significantly change the duplex structure (Figure 2).



Scheme 5. Synthesis of 5-phenyltelluro-2'-deoxyuridine.



Figure 2. Crystal photo of the Te-DNA octamer [5'-G(2'-SeMe-dU)G(TeT)ACAC-3]₂.

The Te-DNA crystal structure was also obtained by the same author by using 2'-Se modification strategy [137–139,144]. The results reveal that Te-DNA has virtually identical global and local structures as the corresponding native DNA (Figure 3). This result further confirms that the Te-functionality does not cause significant perturbation. The Te-DNA was quite stable under a high temperature (90 °C) but it was found to be sensitive to X-ray irradiation. Partial cleavage of the Te–C bond was detected through MALDI-TOF-MS after X-ray irradiation. Due to the metallic property of the tellurium atom, STM imaging studies show stronger topographic and current peaks for the Te-modified DNA duplex compared to the native one (Figure 4).



Figure 3. The global and local structures of the Te-DNA duplex. (**A**) The Te-dsDNA structure (in cyan; 1.50 Å resolution; PDB ID: 3FA1) is superimposed over the native one (in magenta; PDB ID: 1DNS). (**B**) The local structure of the TeT/A base pair (in cyan) is superimposed over the native T/A base pair (in magenta).

Photodynamic therapy (PDT) is a promising medical treatment using visible light irradiation in conjunction with a photosensitizer (PS), which is non-toxic in the dark, to selectively treat the targeted issue [144,145]. A study [146] investigated the photophysical properties of 2-/4-position Te-substituted thymidine, indicating its potential application as a UVA chemotherapeutic agent. The lowest triplet states were found to lie above the energy required to produce cytotoxic excited oxygen molecule and the absorption energies are short enough to penetrate the issue, via density functional theory (DFT) and time-dependent



density functional theory (TDDFT) calculation. Further study [147] of the Te-subsituted deoxyguanosine revealed its ability to act as photosensitizer in cancer therapy.

Figure 4. The STM images of the Te-modified DNA duplex (5'-ATGG(TeT)-GCTC-3' and 5'-(GAGCACCAT)6-3') and its counterpart native duplex on HOPG. The arrows indicate the edges or current peaks of the measured molecules. (**A**) Topographic image of Te-duplex; (**B**) Current image of Te-duplex; (**C**) Topographic image of the native duplex; (**D**) Current image of the native duplex.

3. Conclusions

The Te-modified DNA and RNA are a promising strategy for investigating the structure and function of nucleic acid. However, studies in this area are still quite limited and only a few papers on the subject have been published in the last 10 years. The 2'- and 5-position tellurium modified nucleoside has been successfully synthesized, and both are compatible with solid-phase synthesis, deprotection, and purification. The particular redox properties and selective elimination of the 2'-Te modified DNA oligo could be useful in studying DNA fragmentation and nucleobase damage. The location of the Te functionality modification and the size of the protecting group directly affect the melting temperature of the duplex, which could be used as a useful strategy for detecting DNA and RNA polymerization and catalysis. Furthermore, due to the metallic property of the tellurium atom, the Te-modified DNA duplex becomes visible under STM, which suggests a promising strategy for the direct imaging of DNA without structural perturbation. This will further help us conduct mechanism and function studies or even produce novel nano-electronic materials.

Author Contributions: Writing—original draft preparation, C.C.; writing—review and editing, Z.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Watson, J.D.; Crick, F.H. Molecular structure of nucleic acids. Nature 1953, 171, 737–738. [CrossRef]
- Blount, K.F.; Uhlenbeck, O.C. The structure-function dilemma of the hammerhead ribozyme. *Annu. Rev. Biophys. Biomol. Struct.* 2005, 34, 415–440. [CrossRef] [PubMed]
- 3. Eddy, S.R. Non-coding RNA genes and the modern RNA world. Nat. Rev. Genet. 2001, 2, 919–929. [CrossRef] [PubMed]
- 4. Davidson, J.N. The Biochemistry of the Nucleic Acids; Academic Press: Cambridge, MA, USA, 2012.
- van Kasteren, P.B.; van Der Veer, B.; van den Brink, S.; Wijsman, L.; de Jonge, J.; van den Brandt, A.; Molenkamp, R.; Reusken, C.B.; Meijer, A. Comparison of seven commercial RT-PCR diagnostic kits for COVID-19. *J. Clin. Virol.* 2020, 128, 104412. [CrossRef] [PubMed]
- 6. Pokhrel, P.; Hu, C.; Mao, H. Detecting the coronavirus (COVID-19). ACS Sens. 2020, 5, 2283–2296. [CrossRef]
- Scheler, O.; Glynn, B.; Kurg, A. Nucleic acid detection technologies and marker molecules in bacterial diagnostics. *Expert Rev. Mol. Diagn.* 2014, 14, 489–500. [CrossRef]
- 8. Hartman, M.R.; Ruiz, R.C.; Hamada, S.; Xu, C.; Yancey, K.G.; Yu, Y.; Han, W.; Luo, D. Point-of-care nucleic acid detection using nanotechnology. *Nanoscale* 2013, *5*, 10141–10154. [CrossRef]
- 9. O'Connor, L.; Glynn, B. Recent advances in the development of nucleic acid diagnostics. *Expert Rev. Med. Devices* 2010, 7, 529–539. [CrossRef]
- 10. Piepenburg, O.; Williams, C.H.; Stemple, D.L.; Armes, N.A. DNA detection using recombination proteins. *PLoS Biol.* **2006**, *4*, e204. [CrossRef]
- El Wahed, A.A.; Patel, P.; Maier, M.; Pietsch, C.; Rüster, D.; Böhlken-Fascher, S.; Kissenkötter, J.; Behrmann, O.; Frimpong, M.; Diagne, M.M. Suitcase Lab for rapid detection of SARS-CoV-2 based on recombinase polymerase amplification assay. *Anal. Chem.* 2021, 93, 2627–2634. [CrossRef]
- Luo, G.; Zhang, J.; Zhang, S.; Hu, B.; Hu, L.; Huang, Z. High-quality RT-PCR with chemically modified RNA controls. *Talanta* 2021, 224, 121850. [CrossRef] [PubMed]
- 13. Jordheim, L.P.; Durantel, D.; Zoulim, F.; Dumontet, C. Advances in the development of nucleoside and nucleotide analogues for cancer and viral diseases. *Nat. Rev. Drug Discov.* **2013**, *12*, 447–464. [CrossRef] [PubMed]
- 14. Burnett, J.C.; Rossi, J.J. RNA-based therapeutics: Current progress and future prospects. Chem. Biol. 2012, 19, 60–71. [CrossRef]
- 15. Shen, X.; Corey, D.R. Chemistry, mechanism and clinical status of antisense oligonucleotides and duplex RNAs. *Nucleic Acids Res.* **2018**, *46*, 1584–1600. [CrossRef]
- 16. Stein, C.A.; Castanotto, D. FDA-approved oligonucleotide therapies in 2017. Mol. Ther. 2017, 25, 1069–1075. [CrossRef]
- 17. Deleavey, G.F.; Damha, M.J. Designing chemically modified oligonucleotides for targeted gene silencing. *Chem. Biol.* **2012**, *19*, 937–954. [CrossRef]
- Crooke, S.T.; Vickers, T.A.; Liang, X.-H. Phosphorothioate modified oligonucleotide–protein interactions. *Nucleic Acids Res.* 2020, 48, 5235–5253. [CrossRef]
- 19. Du, Q.; Carrasco, N.; Teplova, M.; Wilds, C.J.; Egli, M.; Huang, Z. Internal Derivatization of Oligonucleotides with Selenium for X-ray Crystallography Using MAD. *J. Am. Chem. Soc.* **2002**, *124*, 24–25. [CrossRef] [PubMed]
- 20. Campbell, N.H.; Parkinson, G.N. Crystallographic studies of quadruplex nucleic acids. Methods 2007, 43, 252–263. [CrossRef]
- 21. Egli, M.; Pallan, P.S. Crystallographic studies of chemically modified nucleic acids: A backward glance. *Chem. Biodivers.* **2010**, *7*, 60–89. [CrossRef]
- 22. Ennifar, E. Nucleic Acid Crystallography; Springer: New York, NY, USA, 2016.
- Park, S.; Okamura, I.; Sakashita, S.; Yum, J.H.; Acharya, C.; Gao, L.; Sugiyama, H. Development of DNA Metalloenzymes Using a Rational Design Approach and Application in the Asymmetric Diels–Alder Reaction. ACS Catal. 2015, 5, 4708–4712. [CrossRef]
- 24. Roelfes, G.; Feringa, B.L. DNA-Based Asymmetric Catalysis. Angew. Chem. Int. Ed. 2005, 44, 3230–3232. [CrossRef] [PubMed]
- Duchemin, N.; Skiredj, A.; Mansot, J.; Leblanc, K.; Vasseur, J.J.; Beniddir, M.A.; Evanno, L.; Poupon, E.; Smietana, M.; Arseniyadis, S. DNA-Templated [2+2] Photocycloaddition: A Straightforward Entry into the Aplysinopsin Family of Natural Products. *Angew. Chem. Int. Ed.* 2018, *57*, 11786–11791. [CrossRef] [PubMed]
- Amirbekyan, K.; Duchemin, N.; Benedetti, E.; Joseph, R.; Colon, A.; Markarian, S.A.; Bethge, L.; Vonhoff, S.; Klussmann, S.; Cossy, J. Design, synthesis, and binding affinity evaluation of hoechst 33,258 derivatives for the development of sequence-specific DNA-based asymmetric catalysts. ACS Catal. 2016, 6, 3096–3105. [CrossRef]
- 27. Ward, W.L.; Plakos, K.; DeRose, V.J. Nucleic acid catalysis: Metals, nucleobases, and other cofactors. *Chem. Rev.* 2014, 114, 4318–4342. [CrossRef]
- 28. Niemeyer, C.M. Nanoparticles, proteins, and nucleic acids: Biotechnology meets materials science. *Angew. Chem. Int. Ed.* 2001, 40, 4128–4158. [CrossRef]
- 29. Baillet, J.; Desvergnes, V.; Hamoud, A.; Latxague, L.; Barthélémy, P. Lipid and nucleic acid chemistries: Combining the best of both worlds to construct advanced biomaterials. *Adv. Mater.* **2018**, *30*, 1705078. [CrossRef]
- Ge, Z.; Gu, H.; Li, Q.; Fan, C. Concept and development of framework nucleic acids. J. Am. Chem. Soc. 2018, 140, 17808–17819. [CrossRef]
- 31. Dong, Y.; Yao, C.; Zhu, Y.; Yang, L.; Luo, D.; Yang, D. DNA functional materials assembled from branched DNA: Design, synthesis, and applications. *Chem. Rev.* **2020**, *120*, *9*420–9481. [CrossRef]
- 32. Cutler, J.I.; Auyeung, E.; Mirkin, C.A. Spherical nucleic acids. J. Am. Chem. Soc. 2012, 134, 1376–1391. [CrossRef]

- 33. Restifo, N.; Ying, H.; Hwang, L.; Leitner, W. The promise of nucleic acid vaccines. Gene Ther. 2000, 7, 89–92. [CrossRef] [PubMed]
- Geall, A.J.; Mandl, C.W.; Ulmer, J.B. RNA: The new revolution in nucleic acid vaccines. *Semin. Immunol.* 2013, 25, 152–159. [CrossRef] [PubMed]
- Deering, R.P.; Kommareddy, S.; Ulmer, J.B.; Brito, L.A.; Geall, A.J. Nucleic acid vaccines: Prospects for non-viral delivery of mRNA vaccines. *Expert Opin. Drug Deliv.* 2014, 11, 885–899. [CrossRef] [PubMed]
- Pardi, N.; Hogan, M.J.; Weissman, D. Recent advances in mRNA vaccine technology. *Curr. Opin. Immunol.* 2020, 65, 14–20. [CrossRef] [PubMed]
- Chen, W.-H.; Strych, U.; Hotez, P.J.; Bottazzi, M.E. The SARS-CoV-2 vaccine pipeline: An overview. *Curr. Trop. Med. Rep.* 2020, 7, 61–64. [CrossRef] [PubMed]
- Smith, T.R.; Patel, A.; Ramos, S.; Elwood, D.; Zhu, X.; Yan, J.; Gary, E.N.; Walker, S.N.; Schultheis, K.; Purwar, M. Immunogenicity of a DNA vaccine candidate for COVID-19. *Nat. Commun.* 2020, *11*, 2601. [CrossRef]
- Xiong, H.; Veedu, R.N.; Diermeier, S.D. Recent advances in oligonucleotide therapeutics in oncology. *Int. J. Mol. Sci.* 2021, 22, 3295.
 [CrossRef]
- Singh, J.; Ripp, A.; Haas, T.M.; Qiu, D.; Keller, M.; Wender, P.A.; Siegel, J.S.; Baldridge, K.K.; Jessen, H.J. Synthesis of modified nucleoside oligophosphates simplified: Fast, pure, and protecting group free. J. Am. Chem. Soc. 2019, 141, 15013–15017. [CrossRef]
- 41. Knouse, K.W.; Justine, N.; Schmidt, M.A.; Zheng, B.; Vantourout, J.C.; Kingston, C.; Mercer, S.E.; Mcdonald, I.M.; Olson, R.E.; Zhu, Y. Unlocking P (V): Reagents for chiral phosphorothioate synthesis. *Science* **2018**, *361*, 1234–1238. [CrossRef]
- Sharma, V.K.; Singh, S.K.; Krishnamurthy, P.M.; Alterman, J.F.; Haraszti, R.A.; Khvorova, A.; Prasad, A.K.; Watts, J.K. Synthesis and biological properties of triazole-linked locked nucleic acid. *Chem. Commun.* 2017, 53, 8906–8909. [CrossRef]
- 43. Paul, S.; Caruthers, M.H. Synthesis of phosphorodiamidate morpholino oligonucleotides and their chimeras using phosphoramidite chemistry. *J. Am. Chem. Soc.* **2016**, *138*, 15663–15672. [CrossRef] [PubMed]
- 44. Li, P.; Sergueeva, Z.A.; Dobrikov, M.; Shaw, B.R. Nucleoside and oligonucleoside boranophosphates: Chemistry and properties. *Chem. Rev.* 2007, 107, 4746–4796. [CrossRef]
- 45. Eckstein, F. Phosphorothioate oligodeoxynucleotides: What is their origin and what is unique about them? *Antisense Nucleic Acid Drug Dev.* **2000**, *10*, 117–121. [CrossRef]
- 46. Eckstein, F. Phosphorothioate analogs of nucleotides. Acc. Chem. Res. 1979, 12, 204–210. [CrossRef]
- Hu, B.; Wang, Y.; Sun, S.; Yan, W.; Zhang, C.; Luo, D.; Deng, H.; Hu, L.R.; Huang, Z. Synthesis of Selenium-Triphosphates (dNTPαSe) for More Specific DNA Polymerization. *Angew. Chem.* 2019, 131, 7917–7921. [CrossRef]
- Liu, C.; Cozens, C.; Jaziri, F.; Rozenski, J.; Marechal, A.; Dumbre, S.; Pezo, V.; Marlière, P.; Pinheiro, V.B.; Groaz, E. Phosphonomethyl oligonucleotides as backbone-modified artificial genetic polymers. *J. Am. Chem. Soc.* 2018, 140, 6690–6699. [CrossRef]
- 49. Sawamoto, H.; Arai, Y.; Yamakoshi, S.; Obika, S.; Kawanishi, E. Synthetic method for 2'-amino-LNA bearing any of the four nucleobases via a transglycosylation reaction. *Org. Lett.* **2018**, *20*, 1928–1931. [CrossRef]
- Mitsuoka, Y.; Yamamoto, T.; Kugimiya, A.; Waki, R.; Wada, F.; Tahara, S.; Sawamura, M.; Noda, M.; Fujimura, Y.; Kato, Y. Triazoleand Tetrazole-Bridged Nucleic Acids: Synthesis, Duplex Stability, Nuclease Resistance, and in Vitro and in Vivo Antisense Potency. J. Org. Chem. 2017, 82, 12–24. [CrossRef]
- 51. Rozners, E. Recent advances in chemical modification of peptide nucleic acids. J. Nucleic Acids 2012, 2012, 518162. [CrossRef]
- Kaur, H.; Babu, B.R.; Maiti, S. Perspectives on chemistry and therapeutic applications of Locked Nucleic Acid (LNA). *Chem. Rev.* 2007, 107, 4672–4697. [CrossRef]
- 53. Ichikawa, E.; Kato, K. Sugar-modified nucleosides in past 10 years, a review. *Curr. Med. Chem.* 2001, *8*, 385–423. [CrossRef] [PubMed]
- Summerton, J.; Weller, D. Morpholino antisense oligomers: Design, preparation, and properties. *Antisense Nucleic Acid Drug Dev.* 1997, 7, 187–195. [CrossRef] [PubMed]
- Campbell, M.A.; Wengel, J. Locked vs. unlocked nucleic acids (LNA vs. UNA): Contrasting structures work towards common therapeutic goals. *Chem. Soc. Rev.* 2011, 40, 5680–5689. [CrossRef]
- 56. Oh, J.; Shin, J.; Unarta, I.C.; Wang, W.; Feldman, A.W.; Karadeema, R.J.; Xu, L.; Xu, J.; Chong, J.; Krishnamurthy, R. Transcriptional processing of an unnatural base pair by eukaryotic RNA polymerase II. *Nat. Chem. Biol.* **2021**, *17*, 906–914. [CrossRef] [PubMed]
- Fischer, E.C.; Hashimoto, K.; Zhang, Y.; Feldman, A.W.; Dien, V.T.; Karadeema, R.J.; Adhikary, R.; Ledbetter, M.P.; Krishnamurthy, R.; Romesberg, F.E. New codons for efficient production of unnatural proteins in a semisynthetic organism. *Nat. Chem. Biol.* 2020, 16, 570–576. [CrossRef]
- Zhou, A.X.-Z.; Sheng, K.; Feldman, A.W.; Romesberg, F.E. Progress toward eukaryotic semisynthetic organisms: Translation of unnatural codons. J. Am. Chem. Soc. 2019, 141, 20166–20170. [CrossRef] [PubMed]
- Feldman, A.W.; Romesberg, F.E. Expansion of the genetic alphabet: A chemist's approach to synthetic biology. Acc. Chem. Res. 2018, 51, 394–403. [CrossRef]
- 60. Zhang, Y.; Ptacin, J.L.; Fischer, E.C.; Aerni, H.R.; Caffaro, C.E.; San Jose, K.; Feldman, A.W.; Turner, C.R.; Romesberg, F.E. A semi-synthetic organism that stores and retrieves increased genetic information. *Nature* **2017**, *551*, 644–647. [CrossRef]
- 61. Hoshika, S.; Leal, N.A.; Kim, M.-J.; Kim, M.-S.; Karalkar, N.B.; Kim, H.-J.; Bates, A.M.; Watkins, N.E.; SantaLucia, H.A.; Meyer, A.J. Hachimoji DNA and RNA: A genetic system with eight building blocks. *Science* **2019**, *363*, 884–887. [CrossRef]

- 62. Biondi, E.; Benner, S.A. Artificially expanded genetic information systems for new aptamer technologies. *Biomedicines* **2018**, *6*, 53. [CrossRef]
- 63. Hoshika, S.; Chen, F.; Leal, N.A.; Benner, S.A. Artificial Genetic Systems: Self-Avoiding DNA in PCR and Multiplexed PCR. *Angew. Chem.* **2010**, 122, 5686–5689. [CrossRef]
- 64. Yang, Z.; Le, J.T.; Hutter, D.; Bradley, K.M.; Overton, B.R.; McLendon, C.; Benner, S.A. Eliminating primer dimers and improving SNP detection using self-avoiding molecular recognition systems. *Biol. Methods Protoc.* **2020**, *5*, bpaa004. [CrossRef] [PubMed]
- 65. Xu, W.; Chan, K.M.; Kool, E.T. Fluorescent nucleobases as tools for studying DNA and RNA. *Nat. Chem.* **2017**, *9*, 1043–1055. [CrossRef]
- 66. Tsao, Y.-Y.T.; Wooley, K.L. Synthetic, functional thymidine-derived polydeoxyribonucleotide analogues from a six-membered cyclic phosphoester. J. Am. Chem. Soc. 2017, 139, 5467–5473. [CrossRef]
- 67. Song, J.; Yi, C. Chemical modifications to RNA: A new layer of gene expression regulation. *ACS Chem. Biol.* **2017**, *12*, 316–325. [CrossRef] [PubMed]
- Tolle, F.; Brändle, G.M.; Matzner, D.; Mayer, G. A versatile approach towards nucleobase-modified aptamers. *Angew. Chem. Int. Ed.* 2015, 54, 10971–10974. [CrossRef]
- 69. Gottfried, A.; Weinhold, E. Sequence-specific covalent labelling of DNA. Biochem. Soc. Trans. 2011, 39, 623–628. [CrossRef]
- 70. Schulz, D.; Holstein, J.M.; Rentmeister, A. A chemo-enzymatic approach for site-specific modification of the RNA cap. *Angew. Chem. Int. Ed.* **2013**, *52*, 7874–7878. [CrossRef]
- 71. Weisbrod, S.H.; Marx, A. Novel strategies for the site-specific covalent labelling of nucleic acids. *Chem. Commun.* 2008, 44, 5675–5685. [CrossRef]
- Rashid, J.I.A.; Yusof, N.A. The strategies of DNA immobilization and hybridization detection mechanism in the construction of electrochemical DNA sensor: A review. Sens. Bio-Sens. Res. 2017, 16, 19–31. [CrossRef]
- Farzan, V.M.; Aparin, I.O.; Veselova, O.A.; Podkolzin, A.T.; Shipulin, G.A.; Korshun, V.A.; Zatsepin, T.S. Cy5/BHQ dye–quencher pairs in fluorogenic qPCR probes: Effects of charge and hydrophobicity. *Anal. Methods* 2016, 8, 5826–5831. [CrossRef]
- Malek-Adamian, E.; Guenther, D.C.; Matsuda, S.; Martínez-Montero, S.I.; Zlatev, I.; Harp, J.; Burai Patrascu, M.; Foster, D.J.; Fakhoury, J.; Perkins, L. 4'-C-Methoxy-2'-deoxy-2'-fluoro modified ribonucleotides improve metabolic stability and elicit efficient RNAi-mediated gene silencing. *J. Am. Chem. Soc.* 2017, 139, 14542–14555. [CrossRef] [PubMed]
- 75. Abou Assi, H.; El-Khoury, R.; González, C.; Damha, M.J. 2'-Fluoroarabinonucleic acid modification traps G-quadruplex and i-motif structures in human telomeric DNA. *Nucleic Acids Res.* **2017**, *45*, 11535–11546. [CrossRef] [PubMed]
- Martínez-Montero, S.; Deleavey, G.F.; Dierker-Viik, A.; Lindovska, P.; Ilina, T.; Portella, G.; Orozco, M.; Parniak, M.A.; González, C.; Damha, M.J. Synthesis and properties of 2'-deoxy-2', 4'-difluoroarabinose-modified nucleic acids. *J. Org. Chem.* 2015, 80, 3083–3091. [CrossRef]
- Martinez-Montero, S.; Deleavey, G.F.; Martín-Pintado, N.; Fakhoury, J.F.; Gonzalez, C.; Damha, M.J. Locked 2'-deoxy-2', 4'difluororibo modified nucleic acids: Thermal stability, structural studies, and siRNA activity. ACS Chem. Biol. 2015, 10, 2016–2023. [CrossRef]
- 78. Jones, A.; Woodhouse, D. Bromination of nucleic acids and their derivatives. Nature 1959, 183, 1603–1605. [CrossRef]
- 79. Ennifar, E.; Carpentier, P.; Ferrer, J.-L.; Walter, P.; Dumas, P. X-ray-induced debromination of nucleic acids at the Br K absorption edge and implications for MAD phasing. *Acta Crystallogr. Sect. D Biol. Crystallogr.* **2002**, *58*, 1262–1268. [CrossRef]
- 80. Egli, M. Nucleic acid crystallography: Current progress. Curr. Opin. Chem. Biol. 2004, 8, 580–591. [CrossRef]
- 81. Ukale, D.; Lönnberg, T. Organomercury Nucleic Acids: Past, Present and Future. ChemBioChem 2021, 22, 1733. [CrossRef]
- 82. Zhang, W.; Szostak, J.W.; Huang, Z. Nucleic acid crystallization and X-ray crystallography facilitated by single selenium atom. *Front. Chem. Sci. Eng.* **2016**, *10*, 196–202. [CrossRef]
- 83. Hendrickson, W.A. Determination of Macromolecular Structures from Anomalous Diffraction of. *Science* **1991**, 254, 5028. [CrossRef] [PubMed]
- 84. Hendrickson, W.A. Synchrotron crystallography. Trends Biochem. Sci. 2000, 25, 637–643. [CrossRef] [PubMed]
- 85. Hendrickson, W.A.; Horton, J.R.; LeMaster, D.M. Selenomethionyl proteins produced for analysis by multiwavelength anomalous diffraction (MAD): A vehicle for direct determination of three-dimensional structure. *EMBO J.* **1990**, *9*, 1665. [CrossRef] [PubMed]
- Chen, C. Synthesis of Selenium and Tellurium Modified Nucleic Acids For DNA Crystallization, Structure and Function Studies. Ph.D. Thesis, Georgia State University, Atlanta, GA, USA, 2020.
- 87. Sheng, J.; Huang, Z. Selenium Derivatization of Nucleic Acids for Phase and Structure Determination in Nucleic Acid X-ray Crystallography. *Int. J. Mol. Sci.* 2008, *9*, 258–271. [CrossRef] [PubMed]
- Sheng, J.; Huang, Z. Selenium Derivatization of Nucleic Acids for X-Ray Crystal-Structure and Function Studies. *Chem. Biodivers.* 2010, 7, 753–785. [CrossRef] [PubMed]
- Salon, J.; Sheng, J.; Jiang, J.; Chen, G.; Caton-Williams, J.; Huang, Z. Oxygen replacement with selenium at the thymidine 4-position for the Se base pairing and crystal structure studies. *J. Am. Chem. Soc.* 2007, 129, 4862–4863. [CrossRef]
- 90. Sun, H.; Sheng, J.; Hassan, A.E.; Jiang, S.; Gan, J.; Huang, Z. Novel RNA base pair with higher specificity using single selenium atom. *Nucleic Acids Res.* 2012, 40, 5171–5179. [CrossRef]
- Salon, J.; Chen, G.; Portilla, Y.; Germann, M.W.; Huang, Z. Synthesis of a 2'-Se-uridine Phosphoramidite and Its Incorporation into Oligonucleotides for Structural Study. Org. Lett. 2005, 7, 5645–5648. [CrossRef]

- 92. Salon, J.; Jiang, J.; Sheng, J.; Gerlits, O.O.; Huang, Z. Derivatization of DNAs with selenium at 6-position of guanine for function and crystal structure studies. *Nucleic Acids Res.* 2008, *36*, 7009–7018. [CrossRef]
- 93. Salon, J.; Gan, J.; Abdur, R.; Liu, H.; Huang, Z. Synthesis of 6-Se-guanosine RNAs for structural study. *Org. Lett.* **2013**, *15*, 3934–3937. [CrossRef]
- 94. Tram, K.; Wang, X.; Yan, H. Facile synthesis of oligonucleotide phosphoroselenoates. *Org. Lett.* **2007**, *9*, 5103–5106. [CrossRef] [PubMed]
- Abdur, R.; Gerlits, O.O.; Gan, J.; Jiang, J.; Salon, J.; Kovalevsky, A.Y.; Chumanevich, A.A.; Weber, I.T.; Huang, Z. Novel complex MAD phasing and RNase H structural insights using selenium oligonucleotides. *Acta Crystallogr. Sect. D Biol. Crystallogr.* 2014, 70, 354–361. [CrossRef] [PubMed]
- 96. Caton-Williams, J.; Huang, Z. Synthesis and DNA-polymerase incorporation of colored 4-selenothymidine triphosphate for polymerase recognition and DNA visualization. *Angew. Chem. Int. Ed.* **2008**, *47*, 1723–1725. [CrossRef] [PubMed]
- 97. Hassan, A.E.; Sheng, J.; Zhang, W.; Huang, Z. High fidelity of base pairing by 2-selenothymidine in DNA. J. Am. Chem. Soc. 2010, 132, 2120–2121. [CrossRef]
- Chen, C.; Fang, Z.; Huang, Z. 2'-β-Selenium Atom on Thymidine to Control β-Form DNA Conformation and Large Crystal Formation. *Cryst. Growth Des.* 2022, 22, 3601–3604. [CrossRef]
- 99. Lin, L.; Sheng, J.; Huang, Z. Nucleic acid X-ray crystallography via direct selenium derivatization. *Chem. Soc. Rev.* 2011, 40, 4591–4602. [CrossRef]
- 100. Caton-Williams, J.; Huang, Z. Biochemistry of selenium-derivatized naturally occurring and unnatural nucleic acids. *Chem. Biodivers.* **2008**, *5*, 396–407. [CrossRef]
- Adenis, C.; Langer, V.; Lindqvist, O. Reinvestigation of the structure of tellurium. *Acta Crystallogr. Sect. C Cryst. Struct. Commun.* 1989, 45, 941–942. [CrossRef]
- 102. Ba, L.A.; Döring, M.; Jamier, V.; Jacob, C. Tellurium: An element with great biological potency and potential. *Org. Biomol. Chem.* **2010**, *8*, 4203–4216. [CrossRef]
- 103. Petragnani, N.; Stefani, H.A. Tellurium in Organic Synthesis, 2nd ed.; Academic Press: Cambridge, MA, USA, 2010.
- 104. Caldwell, R.S.; Fan, H. Optical properties of tellurium and selenium. Phys. Rev. 1959, 114, 664. [CrossRef]
- 105. Shi, Z.; Cao, R.; Khan, K.; Tareen, A.K.; Liu, X.; Liang, W.; Zhang, Y.; Ma, C.; Guo, Z.; Luo, X. Two-dimensional tellurium: Progress, challenges, and prospects. *Nano-Micro Lett.* 2020, 12, 99. [CrossRef] [PubMed]
- 106. Brumaghim, J.L. *Biochalcogen Chemistry: The Biological Chemistry of Sulfur, Selenium, and Tellurium;* American Chemical Society: Washington, DC, USA, 2013.
- 107. Lippolis, V.; Santi, C. Selenium & tellurium chemistry at the beginning of the 3rd millennium: A celebration of ICCST. *New J. Chem.* **2019**, *43*, 11032–11033.
- 108. Moroder, L. Isosteric replacement of sulfur with other chalcogens in peptides and proteins. *J. Peptide Sci.* 2005, *11*, 187–214. [CrossRef] [PubMed]
- 109. Ibers, J. Tellurium in a twist. Nat. Chem. 2009, 1, 508. [CrossRef]
- 110. Rosman, K.J.R.; Taylor, P.D.P. Isotopic compositions of the elements 1997. Pure Appl. Chem. 1998, 70, 217–235. [CrossRef]
- 111. Saito, S.; Zhang, J.; Tanida, K.; Takahashi, S.; Koizumi, T. A Systematic ¹²⁵Te NMR Study of Organotellurium Compounds: The Effect of Oxidation States and Substituents. *Tetrahedron* 1999, 55, 2545–2552. [CrossRef]
- 112. Li, G.M.; Zingaro, R.A.; Segi, M.; Reibenspies, J.H.; Nakajima, T. Synthesis and structure of telluroamides and selenoamides. The first crystallographic study of telluroamides. *Organometallics* **1997**, *16*, 756–762. [CrossRef]
- Beckmann, J.; Hesse, M.; Poleschner, H.; Seppelt, K. Formation of Mixed-Valent Aryltellurenyl Halides RX2TeTeR. Angew. Chem. Int. Ed. 2007, 46, 8277–8280. [CrossRef]
- 114. Miyasato, M.; Minoura, M.; Akiba, K.Y. Cleavage of Tellurium–Carbon Bonds of Hexavalent Organotellurium Compounds by Potassium Graphite. *Angew. Chem. Int. Ed.* 2001, *40*, 2674–2676. [CrossRef]
- 115. Housecroft, C.; Sharpe, A. Inorganic Chemistry, Pearson Education Limited, 4th ed.; Pearson: London, UK, 2012.
- 116. Mutoh, Y.; Murai, T.; Yamago, S. Telluration of seleno-and chloroiminium salts leading to various telluroamides, and their structure and NMR properties. *J. Organomet. Chem.* **2007**, *692*, 129–135. [CrossRef]
- 117. Kuhn, N.; Henkel, G.; Kratz, T. Beiträge zur Chemie des Imidazols, III. 2-Telluroimidazoline—Stabile Tellurocarbonyl-Verbindungen. *Chem. Ber.* **1993**, *126*, 2047–2049. [CrossRef]
- Webber, D.H.; Brutchey, R.L. Photolytic preparation of tellurium nanorods. *Chem. Commun.* 2009, 38, 5701–5703. [CrossRef]
 [PubMed]
- Graf, C.; Assoud, A.; Mayasree, O.; Kleinke, H. Solid state polyselenides and polytellurides: A large variety of Se–Se and Te–Te interactions. *Molecules* 2009, 14, 3115–3131. [CrossRef] [PubMed]
- 120. Sheldrick, W.S.; Wachhold, M. Discrete Crown-Shaped Te₈ Rings in Cs₃ Te₂₂. *Angew. Chem. Int. Ed. Engl.* **1995**, *34*, 450–451. [CrossRef]
- Deiseroth, H.J.; Wagener, M.; Neumann, E. (AgI) 2Te₆ and (AgI) 2Se₆: New Composite Materials with Cyclic Te₆ and Se₆ Molecules Stabilized in the "Solid Solvent" AgI. *Eur. J. Inorg. Chem.* 2004, 2004, 4755–4758. [CrossRef]
- 122. Günther, A.; Isaeva, A.; Baranov, A.I.; Ruck, M. Neutral Tellurium Rings in the Coordination Polymers [Ru (Te₉)](InCl₄)₂, [Ru (Te₈)] Cl₂, and [Rh (Te₆)] Cl₃. *Chem. A Eur. J.* **2011**, *17*, 6382–6388. [CrossRef]

- 123. Jeske, J.; du Mont, W.W.; Jones, P.G. Synthesis of a Triiodide-Like Pentamesityl-tritellurium Cation by Addition of Dimesityltelluride to the Remarkably Electrophilic Trimesitylditelluronium Ion. *Angew. Chem. Int. Ed. Engl.* 1997, 36, 2219–2221. [CrossRef]
- 124. Hayashi, S.; Nakanishi, W. Handbook of Chalcogen Chemistry: New Perspectives in Sulfur, Selenium and Tellurium, 2nd ed.; Devillanova, F.A., du Mont, W.-W., Eds.; RSC Publishing: London, UK, 2013; Volume 2.
- 125. Lin, T.P.; Gabbaï, F.P. Telluronium Ions as σ-Acceptor Ligands. Angew. Chem. Int. Ed. 2013, 52, 3864–3868. [CrossRef]
- 126. Zhao, H.; Gabbaï, F.P. A bidentate Lewis acid with a telluronium ion as an anion-binding site. *Nat. Chem.* **2010**, *2*, 984–990. [CrossRef]
- Knight, F.R.; Arachchige, K.S.A.; Randall, R.A.; Bühl, M.; Slawin, A.M.; Woollins, J.D. Exploring hypervalency and three-centre, four-electron bonding interactions: Reactions of acenaphthene chalcogen donors and dihalogen acceptors. *Dalton Trans.* 2012, 41, 3154–3165. [CrossRef]
- 128. Bühl, M.; Knight, F.R.; Křístková, A.; Malkin Ondík, I.; Malkina, O.L.; Randall, R.A.; Slawin, A.M.; Woollins, J.D. Weak Te, Te Interactions through the Looking Glass of NMR Spin–Spin Coupling. *Angew. Chem.* **2013**, *125*, 2555–2558. [CrossRef]
- Knight, F.R.; Diamond, L.; Athukorala Arachchige, K.S.; Sanz Camacho, P.; Randall, R.A.M.; Ashbrook, S.E.; Bühl, M.; Slawin, A.M.Z.; Woollins, J.D. Conformational dependence of through-space tellurium-tellurium spin-spin coupling in peri-substituted bis (tellurides). *Chem. A Eur. J.* 2014, 21, 3613–3627. [CrossRef]
- Ramadan, S.E.; Razak, A.; Ragab, A.M.; El-Meleigy, M. Incorporation of tellurium into amino acids and proteins in a telluriumtolerant fungi. *Biol. Trace Elem. Res.* 1989, 20, 225–232. [CrossRef] [PubMed]
- Boles, J.O.; Lewinski, K.; Kunkle, M.; Odom, J.D.; Dunlap, R.B.; Lebioda, L.; Hatada, M. Bio-incorporation of telluromethionine into buried residues of dihydrofolate reductase. *Nat. Struct. Biol.* **1994**, *1*, 283–284. [CrossRef] [PubMed]
- Budisa, N.; Karnbrock, W.; Steinbacher, S.; Humm, A.; Prade, L.; Neuefeind, T.; Moroder, L.; Huber, R. Bioincorporation of Telluromethionine into Proteins: A Promising New Approach for X-ray Structure Analysis of Proteins. *J. Mol. Biol.* 1997, 270, 616–623. [CrossRef]
- 133. Bijelic, A.; Rompel, A. Ten good reasons for the use of the tellurium-centered Anderson–Evans polyoxotungstate in protein crystallography. *Acc. Chem. Res.* 2017, *50*, 1441–1448. [CrossRef]
- 134. Yang, F.; Wong, K.-H.; Yang, Y.; Li, X.; Jiang, J.; Zheng, W.; Wu, H.; Chen, T. Purification and in vitro antioxidant activities of tellurium-containing phycobiliproteins from tellurium-enriched Spirulina platensis. *Drug Des. Dev. Ther.* **2014**, *8*, 1789.
- 135. Sheng, J.; Hassan, A.E.A.; Huang, Z. New Telluride-Mediated Elimination for Novel Synthesis of 2',3'-Didehydro-2'.3'dideoxynucleosides. J. Org. Chem. 2008, 73, 3725–3729. [CrossRef]
- Sheng, J.; Hassan, A.E.A.; Huang, Z. Synthesis of the First Tellurium-Derivatized Oligonucleotides for Structural and Functional Studies. *Chem. Eur. J.* 2009, 15, 10210–10216. [CrossRef]
- Jiang, J.; Sheng, J.; Carrasco, N.; Huang, Z. Selenium derivatization of nucleic acids for crystallography. *Nucleic Acids Res.* 2006, 35, 477–485. [CrossRef]
- Salon, J.; Sheng, J.; Gan, J.; Huang, Z. Synthesis and crystal structure of 2'-Se-modified guanosine containing DNA. J. Org. Chem. 2010, 75, 637–641. [CrossRef] [PubMed]
- Sheng, J.; Salon, J.; Gan, J.; Huang, Z. Synthesis and crystal structure study of 2'-Se-adenosine-derivatized DNA. *Sci. China Chem.* 2010, 53, 78–85. [CrossRef]
- 140. Yano, S.; Hirohara, S.; Obata, M.; Hagiya, Y.; Ogura, S.-I.; Ikeda, A.; Kataoka, H.; Tanaka, M.; Joh, T. Current states and future views in photodynamic therapy. *J. Photochem. Photobiol. C Photochem. Rev.* **2011**, *12*, 46–67. [CrossRef]
- Carrasco, N.; Buzin, Y.; Tyson, E.; Halpert, E.; Huang, Z. Selenium derivatization and crystallization of DNA and RNA oligonucleotides for X-ray crystallography using multiple anomalous dispersion. *Nucleic Acids Res.* 2004, 32, 1638–1646. [CrossRef] [PubMed]
- Karino, N.; Ueno, Y.; Matsuda, A. Synthesis and properties of oligonucleotides containing 5-formyl-2'-deoxycytidine: In vitro DNA polymerase reactions on DNA templates containing 5-formyl-2'-deoxycytidine. *Nucleic Acids Res.* 2001, 29, 2456–2463. [CrossRef]
- 143. Sheng, J.; Hassan, A.E.A.; Zhang, W.; Zhou, J.; Xu, B.; Soares, A.S.; Huang, Z. Synthesis, structure and imaging of oligodeoxyribonucleotides with tellurium-nucleobase derivatization. *Nucleic Acids Res.* **2011**, *39*, 3962–3971. [CrossRef] [PubMed]
- 144. Jain, S.; Zon, G.; Sundaralingam, M. Base only binding of spermine in the deep groove of the A-DNA octamer d (GTGTACAC). *Biochemistry* **1989**, *28*, 2360–2364. [CrossRef]
- 145. Brown, S.B.; Brown, E.A.; Walker, I. The present and future role of photodynamic therapy in cancer treatment. *Lancet Oncol.* 2004, *5*, 497–508. [CrossRef]
- 146. Pirillo, J.; De Simone, B.C.; Russo, N. Photophysical properties prediction of selenium-and tellurium-substituted thymidine as potential UVA chemotherapeutic agents. *Theor. Chem. Acc.* **2016**, *135*, 1–5. [CrossRef]
- Pirillo, J.; Mazzone, G.; Russo, N.; Bertini, L. Photophysical properties of S, Se and Te-substituted deoxyguanosines: Insight into their ability to act as chemotherapeutic agents. J. Chem. Inf. Model. 2017, 57, 234–242. [CrossRef]