

Production Review of Accelerator-Based Medical Isotopes

Yiwei Wang¹, Daiyuan Chen¹, Ricardo dos Santos Augusto², Jixin Liang³, Zhi Qin⁴, Juntao Liu^{1,5,*}  and Zhiyi Liu^{1,5,*}

¹ School of Nuclear Science and Technology, Lanzhou University, Lanzhou 730000, China

² Brookhaven National Laboratory, United States Department of Energy Upton, New York, NY 11973-5000, USA

³ Department of Nuclear Technology and Application, China Institute of Atomic Energy, Beijing 102413, China

⁴ Institute of Modern Physics, Chinese Academy of Sciences, Lanzhou 730000, China

⁵ Frontiers Science Center for Rare Isotopes, Lanzhou University, Lanzhou 730000, China

* Correspondence: ljt@lzu.edu.cn (J.L.); zhiyi@lzu.edu.cn (Z.L.)

Abstract: The production of reactor-based medical isotopes is fragile, which has meant supply shortages from time to time. This paper reviews alternative production methods in the form of cyclotrons, linear accelerators and neutron generators. Finally, the status of the production of medical isotopes in China is described.

Keywords: medical isotope production; accelerator; nuclear medicine; review

1. Introduction

1.1. Definition of Medical Isotopes

Medical isotopes are radioisotopes that emit positrons or gamma rays for medical diagnosis or particulate radiation, such as alpha or beta particles for medical therapy [1].

1.2. Medical Use

The application process for medical isotopes is depicted in Figure 1 and can be summarized in four steps:

- (1a) In a reactor, irradiate a suitable target with neutrons to induce a nuclear reaction;
- (1b) In an accelerator, irradiate a suitable target with protons, alpha, or deuteron particles to induce a nuclear reaction;
- (2) Separate radioisotopes from the irradiated targets;
- (3) Combine the ligands with radioisotopes to prepare radiopharmaceuticals;
- (4) Employ the radiopharmaceuticals in nuclear medicine.

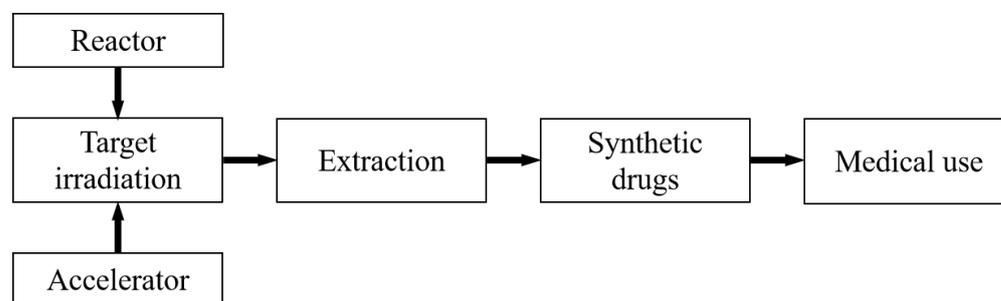


Figure 1. Process for the application of medical isotopes.

Depending on the physical characteristics of the isotopes applied, radiopharmaceuticals have different medical uses in diagnosis, therapy, or both (theranostics) [2], leading to a steady increase in the use of medical isotopes in nuclear medicine over time [3,4].



Citation: Wang, Y.; Chen, D.; dos Santos Augusto, R.; Liang, J.; Qin, Z.; Liu, J.; Liu, Z. Production Review of Accelerator-Based Medical Isotopes. *Molecules* **2022**, *27*, 5294. <https://doi.org/10.3390/molecules27165294>

Academic Editor: Kazuma Ogawa

Received: 26 June 2022

Accepted: 16 August 2022

Published: 19 August 2022

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1.2.1. Radiopharmaceuticals for Diagnosis

Radiopharmaceuticals are generally injected intravenously or, in some cases, taken orally [5,6]. They are transported in the blood throughout the body and, due to their high affinities with specific organs, can target different diseases, especially tumors. The γ rays emitted by radiopharmaceuticals are used for imaging. Currently, there are two main imaging applications for diagnosis in nuclear medicine: Single Photon Emission Computed Tomography (SPECT) [7–9] and Positron Emission Tomography (PET) [10–12]. The distribution of radiotracers in vivo can be detected using SPECT and PET cameras.

The main advantage of nuclear medicine diagnosis lies in its ability to find lesions earlier since diseased tissues usually first denote functional changes before later evolving into shape and structural changes [13]. Another major feature of nuclear medicine diagnosis is its ability to specifically show the locations and sizes of tumors, especially when combined with Computed Tomography (CT) or Magnetic Resonance Imaging (MRI) [14,15].

1.2.2. Radiopharmaceuticals for Therapy

Therapeutic radiopharmaceuticals accumulate in diseased tissue after entering the human body. Then, their cumulative radioactive emissions can produce biological effects (e.g., killing tumor cells), which makes radiopharmaceuticals particularly suitable for cancer treatment [16]. The applications of radiopharmaceuticals for therapy include α therapy, β therapy, and Auger therapy. This review focuses on α therapy and β therapy.

1.2.3. Radiopharmaceuticals for Theranostics

In theranostics, radiopharmaceuticals can be used to perform diagnostic imaging and medical treatment [17–19]. Imaging diagnosis is used to determine an optimal treatment modality and can help monitor and evaluate the medical treatment progress [18,20,21]. Currently, radiopharmaceuticals for theranostics use either the same radiopharmaceutical, which emits γ rays for diagnosis and α or β particles for treatment [22,23], or two different radiopharmaceuticals (one for diagnosis and the other for treatment) [24].

Radiopharmaceuticals for theranostics have developed rapidly in recent years with great progress in treating neuroendocrine tumors, thyroid cancer [20,21,25,26], prostate cancer, breast cancer [27,28], and other diseases.

1.3. The Status of Medical Isotope Production

Radioisotopes are divided into natural and artificial radioisotopes. Currently, there are about 200 radioisotopes in use, most of which are produced artificially [29].

With the widespread usage of radiopharmaceuticals, the stable production and supply of medical isotopes is becoming increasingly important.

Medical isotopes are generally produced via either reactors or accelerators. Typically, reactor-based medical isotopes are neutron-rich isotopes commonly characterized by a long half-life, while accelerator-based medical isotopes tend to offer a shorter half-life and usually emit positrons or γ rays [30]. Reactor irradiation is currently the most commonly used method to produce medical isotopes due to their high yield, low cost, and ease of target preparation. However, this supply is sustained by reactors that were built in the 1950–60s (Table 1). The majority of these reactors will gradually shut down before 2030.

Table 1. Information on the world's major reactors producing medical isotopes [31–34].

Country	Reactor	Power [MW]	Year of First Criticality	Estimated Retirement Time
Belgium	BR-2	100	1961	2026
Netherlands	HFR	45	1961	2024
Czech Republic	LVR-15	10	1957	2028
Poland	MARIA	20	1974	2030
South Africa	SAFARI-1	20	1965	2030
Russia	WWR-TS	15	1964	2025
United States	HFIR	100	1965	2035
Australia	OPAL	20	2006	2057
Germany	FRM-II	20	2004	2054

Moreover, due to their age, and as part of the decommissioning process, reactors can be expected to have longer periods of down time due to maintenance or unplanned shutdown events for safety or technical reasons [35], increasing the risk of supply interruptions or persistent shortages. Additionally, most irradiated targets for ^{99}Mo production in a reactor context use highly enriched uranium (HEU) targets that generate considerable amounts of highly radioactive waste and increase the risk of nuclear proliferation [36,37]. These factors strengthen the argument that medical isotopes produced via reactors should be replaced by accelerator-based production [38,39].

The growing interest and recent improvements in accelerator technologies have already led some medical isotopes produced via reactors to be replaced or partly replaced by accelerator-produced isotopes. There are many advantages to using medical isotopes produced by accelerators:

- (1) Supervision is easier, and safety is improved [40];
- (2) The maintenance and decommissioning costs are lower [29];
- (3) The amount of radioactive waste produced is less than 10% of the amount produced by a reactor, and the radiation levels are lower [41];
- (4) It has no risk of nuclear proliferation [42].

As shown in Figure 2, the number of cyclotrons producing radioisotopes is increasing, while the number of reactors is slowly decreasing.

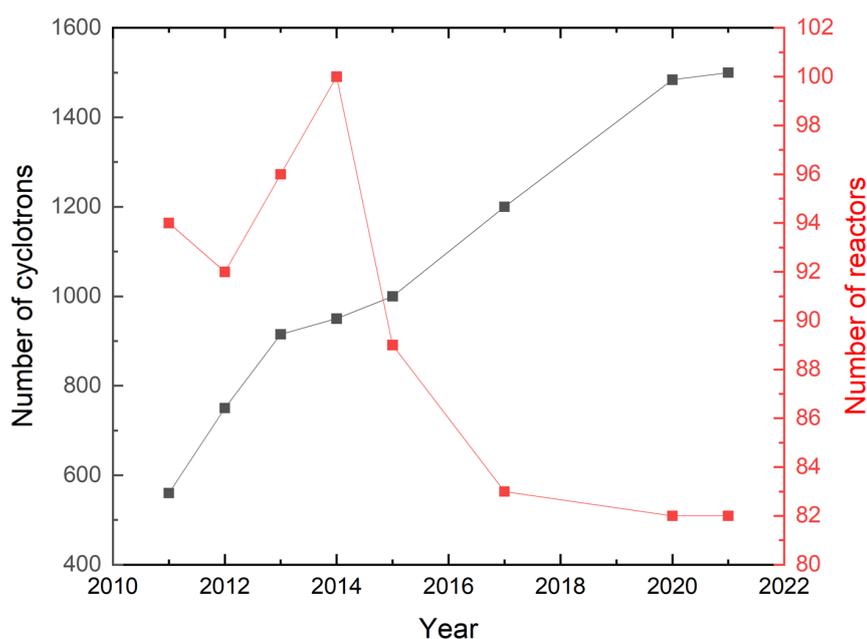


Figure 2. A comparison of the number of cyclotrons in the world and the number of reactors reported by the IAEA [43–57].

2. Medical Isotopes

This section reviews the medical isotopes produced by cyclotrons, linear accelerators, and neutron generators and lists some of the most commonly used medical isotopes, as well as their characteristics, applications, and production methods.

2.1. Medical Isotopes Produced by Cyclotrons (1–5: PET Radioisotopes, 6–7: SPECT Radioisotopes, 8–10: Therapeutic Radioisotopes)

A cyclotron is a particle accelerator that accelerates charged particles and uses an electromagnetic field to get the particles to follow a spiral path to ever-increasing energies until achieving the energy necessary to produce medical isotopes via nuclear interactions [58]. Compared with linear accelerators, the beams from cyclotrons have characteristically lower beam intensity, but their energy can be higher [59]. Cyclotrons are classified according to

the energy of the particles they produce. As shown in Table 2, different types of cyclotrons can produce medical isotopes for a wide range of applications.

Table 2. Classification of medical cyclotrons [60].

Type	The Energy of Particles [MeV]	Application
Small medical cyclotron	<20	Short-lived radioisotopes for PET
Medium-energy cyclotron	20–35	Production of SPECT and some PET radioisotopes
High-energy cyclotron	>35	Production of radioisotopes for therapy

2.1.1. ^{18}F

^{18}F ($T_{1/2} = 109.8$ min) decays and emits positrons with an average energy of 0.25 MeV; hence, the distance traveled until reaching positron annihilation in tissues is short. ^{18}F is the most commonly used PET radioisotope. At present, the Food and Drug Administration (FDA) has approved ^{18}F radiopharmaceuticals for use in the diagnosis of a variety of diseases, such as Alzheimer's disease, infections, and many types of cancer, as well as to evaluate treatment outcomes [61,62]. According to clinical data, [^{18}F]FDG can distinguish between Parkinson's Disease (PD), MSA with predominant Parkinsonism (MSA-P), and MSA with predominant cerebellar features (MSA-C) [63,64]. PET diagnosis is expensive and can cost over \$1000, while doctors can make an early and accurate diagnosis. For that reason, the annual number of PET scans has steadily increased for many years [65]. Most ^{18}F is produced via cyclotrons by exploiting two nuclear reactions:

- (1) $^{18}\text{O}(\text{p}, \text{n})^{18}\text{F}$: This reaction requires enriched (and more expensive) ^{18}O target materials to produce ^{18}F in a high yield [66]. Technology developments led to improvements in the target system and the production of ^{18}F up to 34 GBq, as well as specific activities of 350–600 GBq/mmol 30 min after the end of bombardment [67]. Subsequently, it was found that with the irradiation of 11 MeV protons, the yield of ^{18}F further increased directly with the proton current. However, the impurities also increased such that for a proton current of 20 μA , the yield of ^{56}Co (4.86 MBq) and $^{110\text{m}}\text{Ag}$ (1.51 MBq) doubled [68]. Many developing countries do not have medical isotope production facilities. If these countries desire to become self-sufficient in the production of medical isotopes, they could start by installing low-energy cyclotrons to produce ^{18}F [69].
- (2) $^{20}\text{Ne}(\text{d}, \alpha)^{18}\text{F}$: This is the first production method used to produce ^{18}F . This reaction is characterized by lower yields and low specific activity, so it is gradually being replaced. However, with production improvements, this method could again become an attractive alternative [70].

2.1.2. ^{68}Ga

^{68}Ga ($T_{1/2} = 68$ min) is a metal PET radioisotope. Currently, there are about 100 ongoing clinical tests with ^{68}Ga [61], indicating the rapid development of ^{68}Ga -labelled radiotracers. Radiopharmaceuticals labeled with ^{68}Ga are used for the diagnosis of neuroendocrine tumors and are highly accurate when used in patients with suspected but yet not localized neuroendocrine tumors [71]. In addition, ^{68}Ga and ^{177}Lu ($T_{1/2} = 6.7$ d) have a similar coordination chemistry, rendering them some of the most promising radiopharmaceuticals for theranostics. For neuroendocrine tumors, both [^{68}Ga]Ga-DOTA-TATE and [^{177}Lu]Lu-DOTA-TATE have been approved by the FDA for clinical PET diagnosis and medical treatment [72–74]. [^{68}Ga]Ga-PSMA-11 is the first radiopharmaceutical approved by the FDA for PET imaging of PSMA-positive prostate cancer, and [^{177}Lu]Lu-PSMA-617 has also been used for PSMA-targeted therapy [74–77].

^{68}Ga is generally available using a $^{68}\text{Ge}/^{68}\text{Ga}$ generator and represents a relatively simple and convenient method [78] that can yield up to 1.85 GBq [79]. With the development of

technology, the commercial “ionic” generators have made ^{68}Ga clinically successful [80,81]. ^{68}Ga obtained by generators cannot meet the growing demands, however, so the use of accelerators to obtain ^{68}Ga has aroused scientific interest. Moreover, higher yields of ^{68}Ga can be obtained with the $^{68}\text{Zn}(p, n)^{68}\text{Ga}$ reaction using a small cyclotron [82,83]. The yield when using a solid target was reported as $5.032\text{ GBq}/\mu\text{A}\cdot\text{h}$ [83]. After 6 h, impurities such as ^{66}Ga and ^{67}Ga only accounted for 0.51% of the total activity [84]. Compared with using a generator, this production method does not require radioactive waste treatment. Although the solid target system is complex, and the separation steps are lengthy, an automated process was developed to separate the solid target and is simpler to operate than alternative methods [85]. This nuclear reaction can also take place in a liquid target, with radiochemical and radionuclidic purities both above 99.9%. However, the yield using a liquid target was found to be significantly lower (192.5 ± 11.0) $\text{MBq}/\mu\text{A}\cdot\text{h}$ [86]. This production method using the liquid target as an alternative method still needs further optimization to improve the yield.

2.1.3. ^{64}Cu

Upon decay, ^{64}Cu ($T_{1/2} = 12.7$ h) emits positrons and electrons that can be utilized for PET diagnosis and have potential applications in β therapy, thus making ^{64}Cu useful as a radiopharmaceutical for theranostics. Furthermore, ^{64}Cu and ^{67}Cu ($T_{1/2} = 61.76$ h) can be radiopharmaceuticals for theranostics in order to conduct pre-targeted radioimmunotherapy [87]. Presently, the FDA has approved [^{64}Cu]Cu-DOTA-TATE to localize somatostatin receptor-positive neuroendocrine tumors in adult patients. In clinical experiments, [^{64}Cu]Cu-DOTA-TATE has excellent imaging quality and higher detection rates for lesions [88].

^{64}Cu can be produced by small medical cyclotrons via $^{64}\text{Ni}(p, n)^{64}\text{Cu}$ reaction with high specific activity. This production method requires an enriched ^{64}Ni (at least 96%) target to obtain a high yield of $5.89\text{ GBq}/\mu\text{A}\cdot\text{h}$ and ^{64}Cu with radionuclidic purity higher than 99% [89]. The disadvantage is that the ^{64}Ni target material has a low isotopic abundance (0.926%) in nature [90], meaning that the target material is expensive and must be recycled to improve its cost-effectiveness [91,92]. Alternative methods of ^{64}Cu production can also be deuteron-zinc reactions such as $^{\text{nat}}\text{Zn}(d, x)^{64}\text{Cu}$, and $^{66}\text{Zn}(d, \alpha)^{64}\text{Cu}$. Although they have lower costs, their yields are lower, and high-energy deuterons are required [93]. These factors limit actual production through such reactions.

The $^{64}\text{Ni}(p, n)^{64}\text{Cu}$ reaction is the preferred choice for ^{64}Cu production in clinical applications. During the past decade, more than 20 countries, including the United States, Japan, Finland, and China, have developed $^{64}\text{Ni}(p, n)^{64}\text{Cu}$ methods for ^{64}Cu production [89,91,94], some of which are shown in Table 3.

Table 3. Facilities that have reported the production of ^{64}Cu [91,94–99].

Facility/Location	Nuclear Reaction	Irradiation Parameters	Yield
Fukui Medical University	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	12 MeV, (50 ± 3) μA	2–24 GBq in 2 h
The University of Sherbrooke PET Imaging Centre	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	15 MeV, 18 μA	3.9 GBq in 4 h
IBA	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	10 MeV, 12 μA	5123 MBq in 3 h
Paul Scherrer Institute	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	11 MeV, 40–50 μA	Max 8.2 GBq in 4–5 h
Turku PET Centre	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	15.7 MeV, < 100 μA	Max 9.4GBq after purification
Sumitomo HM-20 cyclotron	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	12.5 MeV, 20 μA	7.4 GBq in 5–7 h
NIRS AVF-930 cyclotron	$^{64}\text{Ni}(p, n)^{64}\text{Cu}$	24 MeV HH^+ , 10 $\text{e}\mu\text{A}$	5.2–13GBq in 1–3 h

2.1.4. ^{89}Zr

^{89}Zr ($T_{1/2} = 78.4$ h) is a positron emitter and a new metal PET radioisotope ideal for immunoimaging [100]. To date, ^{89}Zr -atezolizumab has been studied in renal cell carcinoma (RCC), but some obstacles were encountered, so further research is needed [101]. ^{89}Zr is produced by cyclotrons involving the following nuclear reactions:

- (1) $^{89}\text{Y} (p, n) ^{89}\text{Zr}$: This reaction only requires low-energy protons (5–15 MeV) and targets with natural abundance ^{89}Y (100%), which reduces the costs significantly. The number of interference nuclear reactions is limited; hence, one can obtain a high specific activity of ^{89}Zr [102–104]. The yield of this (p, n) reaction can be as high as 44 MBq/ $\mu\text{A}\cdot\text{h}$ under irradiation of 14 MeV protons [105]. Various methods for the isolation and purification of ^{89}Zr have been proposed, including solvent extraction, anion exchange chromatography, and weak cation exchange chromatography, which can obtain ^{89}Zr with high specific activity and radionuclidic purity [106]. The proton energy from small medical cyclotrons installed in hospitals can meet the requirements for bombarding the ^{89}Y target, which is the main reason why many hospitals have developed ^{89}Zr production processes.
- (2) $^{89}\text{Y} (d, 2n) ^{89}\text{Zr}$: This reaction uses low-energy deuterons (also 5–15 MeV) and has the same advantages as the aforementioned production method [102–104], as well as offering a higher yield of 58 MBq/ $\mu\text{A}\cdot\text{h}$. However, one must still factor in the availability of the beam of particles and the costs of these two production methods [105]. Thus, more research is needed.
- (3) $^{\text{nat}}\text{Sr} (\alpha, xn) ^{89}\text{Zr}$: Besides requiring α beams, if $^{\text{nat}}\text{Sr}$ targets are used, abundant quantities of impurities such as ^{88}Zr and ^{86}Zr can easily be produced. For the moment, this production method is only theoretically feasible [107].

2.1.5. ^{124}I

^{124}I ($T_{1/2} = 4.176$ d) is a PET nuclide that can provide a higher quality diagnostic image [108]. Currently, ^{124}I is used for the clinical diagnosis of thyroid cancer [109] and neuroblastoma [110]. ^{124}I and ^{131}I can also be combined as radiopharmaceuticals for theranostics to treat thyroid cancer [20].

^{124}I is produced via cyclotrons through two different production methods:

- (1) $^{124}\text{Te} (p, n) ^{124}\text{I}$: This is the main production method currently employed. Although this method offers a relatively low production rate, it can achieve high currents and use enriched targets to improve the overall yield [108]. The average yield of this reaction is 16 MBq/ $\mu\text{A}\cdot\text{h}$, and at the end of bombardment, the impurity content of ^{123}I and ^{125}I only reaches about 1% [111]. Dry distillation is used to extract ^{124}I [112]. On the downside, the enriched ^{124}Te target material costs about 10000\$/g, which is relatively expensive [113].
- (2) $^{124}\text{Te} (d, 2n) ^{124}\text{I}$: Has a high production yield of 17.5 MBq/ $\mu\text{A}\cdot\text{h}$, however, this reaction requires a beam of deuterons, which may be difficult to obtain and can result in impurities such as ^{125}I (reaching about 1.7%) [111,114].

2.1.6. $^{99\text{m}}\text{Mo}/^{99\text{m}}\text{Tc}$

$^{99\text{m}}\text{Tc}$ ($T_{1/2} = 6.02$ h) emits single γ rays with 0.141 MeV and is mostly used in SPECT; for the diagnosis of stroke; and to examine bone, myocardium, kidneys, thyroid, salivary glands, and other organs [61,62]. The proportion of nuclear medicine diagnosis applying $^{99\text{m}}\text{Tc}$ accounts for approximately 80% of all nuclear medicine procedures, representing around 40 million examinations worldwide every year [115]. $^{99\text{m}}\text{Tc}$ is mainly produced using a $^{99}\text{Mo}/^{99\text{m}}\text{Tc}$ generator. Currently, $^{99\text{m}}\text{Tc}$ can be produced by cyclotrons through the following reactions:

- (1) $^{100}\text{Mo} (p, 2n) ^{99\text{m}}\text{Tc}$ [116,117]: This is the main production method and is optimal with a proton energy range of 19–24 MeV and a highly enriched ^{100}Mo target, such that ^{98}Tc , ^{97}Tc , and other impurities can be reduced to a minimum. According to

the experimental data, with a proton beam energy of 24 MeV, the yield of ^{99m}Tc is about 592 GBq/mA·h [118]. A target irradiated with a 24 MeV proton beam at 500 μA for 12 h yielded 2.59 TBq of ^{99m}Tc [119]. GE PETtrace880 machines have obtained approximately 174 GBq after 6 h [116]. To date, TRIUMF and its partners have successfully verified the feasibility of using a 24 MeV cyclotron to produce ^{99m}Tc to supply the needs of all applications in Vancouver by developing a complete process based on 16, 19, and 24 MeV cyclotron production and applied the results to relevant patents [120]. Automated modules to separate ^{99m}Tc from irradiated targets of ^{100}Mo are under development [121]. However, the shipped distance should be considered based on the direct product and its half-life [122];

- (2) $^{96}\text{Zr} (\alpha, n) ^{99}\text{Mo} \rightarrow ^{99m}\text{Tc}$ [123,124]: This production method can produce ^{99m}Tc with high specific activity. However, it has a low yield, and a beam with a high current is difficult to obtain, which limits the applicability of this production method.

2.1.7. ^{123}I

^{123}I ($T_{1/2} = 13.2$ h) is a γ -ray emitter that can be utilized for SPECT diagnosis. It has especially been used for the diagnosis of Parkinson's disease, primary and metastatic pheochromocytoma, and neuroblastoma. The sensitivity and specificity of this technology are greater than 90% [125]. It also can be used for diagnosis of the thyroid, brain, and myocardium.

Presently, there are three common production routes yielding ^{123}I :

(1–2) $^{124}\text{Xe} (p, 2n) ^{123}\text{Cs} \rightarrow ^{123}\text{Xe} \rightarrow ^{123}\text{I}$ and $^{124}\text{Xe} (p, pn) ^{123}\text{Xe} \rightarrow ^{123}\text{I}$: These nuclear reactions require a medium-energy cyclotron and can obtain with a high radionuclidic purity. The yield of these reactions simulated by MCNP was 757 MBq/ $\mu\text{A}\cdot\text{h}$. Compared with the experimental data, the maximum fluctuation was about 185 MBq/ $\mu\text{A}\cdot\text{h}$ [126,127]. However, due to the use of enriched ^{124}Xe targets, these methods are costly [128,129].

(3) $^{123}\text{Te} (p, n) ^{123}\text{I}$: This production method can apply a low-energy cyclotron. When enriched targets of ^{123}Te (enrichment of 99.3%) were used, an ultrapure nuclide was obtained, and the yield increased from nearly 18.5 to 37GBq 30 h after EOB (end of the bombardment) [130–132]. This production method is also costly because of the enriched target of ^{123}Te . This alternative production method was proven feasible to produce ^{123}I .

2.1.8. ^{225}Ac

^{225}Ac ($T_{1/2} = 9.92$ d) has a unique decay chain that can emit four α rays, causing it to be more effective in destroying tumor cells than other isotopes. Presently, the first use of [^{225}Ac]Ac-PSMA-I&T in a clinical context was successful in treating advanced metastatic castration-resistant prostate cancer [133–135]. Additionally, the research of [^{225}Ac]Ac-DOTAGA-SP for the treatment of malignant gliomas is ongoing [136].

^{225}Ac can be produced with medium-energy protons via the $^{226}\text{Ra} (p, 2n) ^{225}\text{Ac}$ reaction. The yield was only about 2.4 MBq after EOB [137], moreover, its radioactive inventory is difficult to handle [137–139]. Production of ^{225}Ac applying high-energy protons (60–140 MeV) through bombarding a ^{232}Th target can produce a high yield of 96 GBq, but this yield requires high intensity and energy [140], which are not readily available. Currently, the U.S. Department of Energy Isotope Program produces ^{225}Ac using a spallation-induced reaction with high-energy protons on natural thorium.

2.1.9. ^{211}At

^{211}At ($T_{1/2} = 7.2$ h) emits α particles that can be utilized in α therapy [141]. Currently, ^{211}At in the form of [^{211}At]At-PA and [^{211}At]At-ch81C6 has been studied in glioma and recurrent brain tumors [142,143]. Gothenburg (Sweden) [144] is undergoing a clinical research using [^{211}At]At-MX35(Fab) $_2$ to treat ovarian cancer patients, which is an alpha-emitting radionuclide with great clinical potential [145].

^{211}At is commonly produced by a medium-energy cyclotron bombarding a ^{209}Bi target with α particles, causing a $^{209}\text{Bi} (\alpha, 2n) ^{211}\text{At}$ reaction to take place [146,147]. Purifying the

^{211}At from the target material was either done by a wet extraction or a dry distillation. The National Institutes of Health (Bethesda, USA) produced a maximum of 1.71 GBq in one hour, while Sichuan University in China produced a maximum of 200 MBq in 2 h [148]. However, due to the product of toxic impurities such as ^{210}Po , the energy of the α beam needs to be monitored [148,149].

2.1.10. ^{67}Cu

^{67}Cu ($T_{1/2} = 61.76$ h) emits γ rays for SPECT diagnosis and β particles that can be used for medical treatment. Thus, ^{67}Cu can be used individually or with ^{64}Cu as a radiopharmaceutical for theranostics. Presently, ^{67}Cu is used for the nuclear medicinal diagnosis of neuroendocrine tumors and lymphomas [150,151] and the medical treatment of lymphoma and colon cancer [152].

^{67}Cu is generally produced via the $^{68}\text{Zn}(p, 2p)^{67}\text{Cu}$ reaction. This reaction has high recovery and needs both a medium-energy cyclotron and a highly enriched target [153–155]. Due to the need for high-energy protons, there are only a few laboratories in the world that can produce ^{67}Cu [156]. The yield of the integral physical thick target was calculated and is shown in Figure 3.

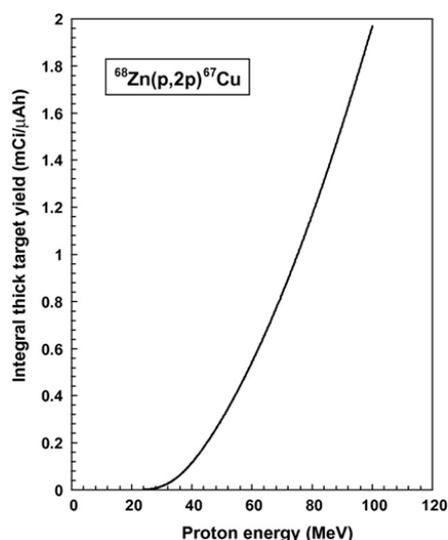


Figure 3. Integral physical thick target yields for the $^{68}\text{Zn}(p, 2p)^{67}\text{Cu}$ reaction [157].

In addition to the medical isotopes mentioned above, ^{11}C [158], ^{13}N [159], ^{15}O [160], ^{86}Y [161], ^{44}Sc [162,163], ^{201}Tl [164], ^{47}Sc [165,166], ^{32}P [167], ^{67}Ga [168], and other medical isotopes produced by cyclotrons have also been reported.

Cyclotrons are the main accelerator-based drivers of medical isotope production. Their output is constantly improving due to advancements in targets [169,170], research on new nuclear reactions [171–173], and accelerator technology developments [174–176], leading not only to increased yields but also to a reduction in radioactive impurities. Most medical isotopes currently produced by reactors can also alternatively be produced by cyclotrons, and the constant improvements to the medical-isotope-producing abilities of cyclotrons have contributed to the stable supply of medical isotopes.

2.2. Medical Isotopes Produced by Linacs

The charged particles accelerated by a linac pass through the focusing magnetic field and the linear acceleration field once without deflection [58]. Once ejected, these particles irradiate their targets to produce medical isotopes. Linac beams are characterized by high beam intensity and lower energy [59].

In terms of linacs currently used to produce medical isotopes, proton linacs can be relatively easily employed in medical isotope production. For example, proton linacs

that produce PET nuclides can reduce the weight of cyclotron magnets, and some high-energy and high-fluxes proton linacs can produce therapeutic nuclides [177–179]. While feasibility reports on the ability of electron linacs to produce medical isotopes are common, the pulsed beams and the cross-sections of linacs can create challenges when used in practice [41,180–182]. There are other linacs that accelerate other charged particles; however, these linacs will not be described here.

2.2.1. ^{18}F

PET radioisotopes can be produced with proton linacs. The first compact proton linear accelerator in the United States for the generation of medical isotopes produces ^{18}F for a local hospital [183]. Additionally, Hitachi, Ltd. and AccSys Technology, Inc. (Hitachi's subsidiary company) also developed a proton linac to produce PET nuclides. After bombardment for one hour, 23.5 GBq ^{18}F was produced, indicating that batch production of ^{18}F could be achieved [177].

^{18}F ($T_{1/2} = 109.8$ min) can also be produced by electron linacs through a photonuclear reaction $^{19}\text{F}(\gamma, n)^{18}\text{F}$, as well as other commonly used PET radioisotopes such as ^{11}C ($T_{1/2} = 20.38$ min), ^{13}N ($T_{1/2} = 9.96$ min), and ^{15}O ($T_{1/2} = 122$ s). When using a photonuclear reaction to produce these PET radioisotopes, the yields are generally lower since the cross-section is 1–2 orders of magnitude lower than that under a proton reaction. However, photonuclear reactions can use a natural target of ^{19}F , thus providing lower costs compared to proton reactions [177]. Many feasibility reports on producing PET nuclides via photonuclear reactions have been published, but actual production still needs further study.

2.2.2. ^{99}Mo

^{99}Mo ($T_{1/2} = 66$ h) decays into $^{99\text{m}}\text{Tc}$ ($T_{1/2} = 6.02$ h). An electron linac can be utilized to produce ^{99}Mo via the photonuclear reaction $^{100}\text{Mo}(\gamma, n)^{99}\text{Mo}$ [184–186]. It was reported that the yield of ^{99}Mo obtained after 6.5 days of continuous bombardment of a 6 g high-purity ^{100}Mo target with 36 MeV electrons was 458.8 GBq (average beam power of ~8 kW) [187]. The cost of this production method can be reduced by using a natural target and, although this method will produce the isotopes of Mo, isotopes of Tc will not be produced, making it easy to separate ^{99}Mo via chemical difference or evaporation temperature difference [188]. NorthStar and its partners have studied this production method and listed it as the main ^{99}Mo supply option in their long-term plans [187]. Canadian Light Source (CLS) and TRIUMF also conducted feasibility research on this production method and plan to put it into production [189,190].

In addition to the medical isotopes mentioned above, the production of ^{67}Cu [191–193], ^{64}Cu [194], ^{225}Ac [195,196], ^{68}Ga [197], ^{111}In [181], ^{177}Lu [198], ^{47}Sc [199], and other medical isotopes through linacs have been reported.

Overall, linacs have some disadvantages in terms of their design and yields [41,182,200,201]. As a backup method for the production of medical isotopes, linacs still require further research.

2.3. Medical Isotopes Produced by Neutron Generators

A neutron generator is an accelerator-based neutron source device that is capable of delivering neutrons through nuclear fusion reactions. These neutrons will, in turn, irradiate the target to produce medical isotopes. The nuclear fusion reactions commonly used to produce neutrons are shown in Table 4.

Table 4. Fusion reactions that produce neutrons [202–206].

Reaction	Energy [MeV]	The Suitable Reaction of Isotope Production
D-D reaction	2–3	(n, γ)
D-T reaction	14–15	(n, 2n) (n, p)
D- ^7Li reaction	10&13	(n, 2n) (n, p)

2.3.1. $^{99}\text{Mo}/^{99\text{m}}\text{Tc}$

$^{99\text{m}}\text{Tc}$ ($T_{1/2} = 6.02\text{h}$) can be produced by neutron generators [207,208]. After neutron moderation, neutrons with a specific energy can be obtained and then used to produce $^{99\text{m}}\text{Tc}$ via the nuclear reaction of $^{235}\text{U} (n, f) ^{99}\text{Mo} \rightarrow ^{99\text{m}}\text{Tc}$. The advantages of this production method include both ease of supervision and overall safety, but the yield will be 1–2 orders of magnitude lower than that produced by a reactor [209]. SHINE and Phoenix Laboratory used a DT neutron generator to bombard UO_2SO_4 to produce ^{99}Mo . After irradiation of a 5 L UO_2SO_4 solution for about 20 h, the yield of ^{99}Mo was 51.8 GBq [210]. The disadvantage of this production method is that a long-term, stable, and high-intensity beam is difficult to achieve [211].

In addition, ^{99}Mo can be produced via the nuclear reactions of $^{98}\text{Mo} (n, \gamma) ^{99}\text{Mo}$ and $^{100}\text{Mo} (n, 2n) ^{99}\text{Mo}$, both of which use Mo targets instead of U targets. Additionally, sufficient activity of ^{99}Mo can be produced in principle [207,208,212]. The yields of these two nuclear reactions can be increased by improving the fluxes of neutrons and the irradiation time and/or using highly enriched targets, in addition to other methods [213]. However, ^{99}Mo from an irradiated $^{98}\text{Mo}/^{100}\text{Mo}$ target is a carrier-added product with a low specific activity. The biggest challenge for this method is how to develop a new type of $^{99}\text{Mo}/^{99\text{m}}\text{Tc}$ generator that meets medical requirements.

2.3.2. ^{67}Cu

^{67}Cu ($T_{1/2} = 61.76\text{h}$) is generally produced by cyclotrons. Kin proposed using neutrons to produce ^{67}Cu [212]. Presently, using neutron generators via the D-T reaction in the form of $^{67}\text{Zn} (n, p) ^{67}\text{Cu}$ can produce ^{67}Cu . Due to the developments of neutron generators, ^{67}Cu can be produced in the hospital without the need to transport the isotope over long distances. This production method does not produce a large number of impurities [156,214], and the activity can reach hundreds to thousands of MBq [212]. However, when dealing with radioactive isotopes with GBq, the radiation facility will result in higher costs [212].

In addition to the medical isotopes mentioned above, ^{89}Sr [215–217], ^{64}Cu [218], ^{47}Sc [219], ^{132}Xe [220], ^{225}Ac [212], and other medical isotopes produced by neutron generators have also been reported.

As a neutron source, a neutron generator is essential to produce neutron-rich medical isotopes. Although such generators have the advantages of low cost and target reusability [212,221], providing continuously high fluxes of neutrons and engaging in separation-extraction of the medical isotopes remain challenging topics [221]. Despite these challenges, generators are presently regarded as a viable alternative to the reactor-production method.

3. The Status of Medical Isotope Production via Accelerators in China

3.1. Available Accelerators for Medical Isotope Production in China

Currently, there are about 160 PET small medical cyclotrons for the routine production of ^{11}C , ^{18}F , and other medical isotopes to meet clinical demands in China [222]. Additionally, there are several medium- and high-energy accelerators used for medical isotope production in China.

The Chinese Institute of Atomic Energy (CIAE) and Shanghai Ansheng Kexing Company each have a C-30 cyclotron with adjustable proton energy of 15.5–30 MeV and beam currents up to 350 μA . These can be used to produce medical isotopes such as ^{11}C , ^{18}F , ^{64}Cu , ^{68}Ge , ^{89}Zr , ^{123}I , ^{124}I , and ^{201}Tl . CIAE has a 100 MeV proton cyclotron (C-100) with a beam current up to 200 μA capable of producing ^{67}Cu , ^{225}Ac , and other medical isotopes of interest.

The Sichuan University owns a cyclotron capable of delivering beams of protons, as well as alpha and deuteron particles (p –26 MeV and α –30 MeV).

The Chinese Academy of Sciences Institute of Modern Physics built a 25 MeV superconducting proton linear accelerator with an intensity in the order of milliamperes. At present, the linac can accelerate various beams such as proton beams, $^3\text{He}^{2+}$ beams, and $^4\text{He}^{2+}$ beams. The energy of $^3\text{He}^{2+}$ beams can reach 36 MeV at an intensity of 200 μA , while the

energy of $^4\text{He}^{2+}$ beams can reach 32 MeV with a current of 100 μA . The accelerator can meet the needs of medical isotope production and produce various radioisotopes such as $^{99}\text{Mo}/^{99\text{m}}\text{Tc}$, $^{117\text{m}}\text{Sn}$, ^{211}At , ^{55}Fe , ^{73}As , ^{225}Ac , ^{109}Cd , ^{88}Y , and ^{75}Se .

Lanzhou University has been instrumental in the development of advanced ion source selection, ion beam extraction, and acceleration system design, as well as target system design. Additionally, the university independently built a series of neutron generators based on D-D and D-T reactions [223].

3.2. The Status of Medical Isotope Production via Accelerators

There is a solid research foundation for accelerator-based medical isotope production in China. In the 1980s, Sichuan University and others successfully developed production technology for medical isotopes such as ^{211}At , ^{123}I , ^{111}In , and ^{201}Tl by relying on domestic cyclotrons and a CS-30 cyclotron [224]. Since the 1990s, CIAE has produced medical isotopes such as ^{18}F , ^{111}In , and ^{201}Tl using a C-30 cyclotron.

In the last two decades, with the popularization and rapid development of domestic nuclear medicine, the amount of PET equipment increased to 427 by 2019. Today, 117 hospitals equipped with small medical cyclotrons routinely produce ^{18}F to meet clinical needs, with an annual consumption of more than 1850 TBq. Additionally, some emerging isotopes such as ^{64}Cu , ^{89}Zr , and $^{123/124}\text{I}$ have been rapidly developed for medical applications. In 2007, CIAE cooperated with Atom Hitech to carry out research on ^{123}I production using enriched ^{124}Xe gas at 111 GBq for each batch with a C-30 cyclotron. In 2012, Atom Hitech produced carrier-free ^{64}Cu with enriched ^{64}Ni at 37–74 GBq for each batch based on a C-30 cyclotron. In 2016, Sichuan University bombarded an ^{89}Y target with 13 MeV protons and obtained ^{89}Zr with a radionuclidic purity of more than 99% [16]. However, due to the limited availability of high-energy particle accelerators for the production of therapeutic nuclides such as ^{67}Cu , ^{225}Ac , and ^{223}Ra , China is significantly lagging behind the advanced international levels of development. In 2021, for the first time, CIAE obtained around 22.2 MBq of ^{225}Ac with radionuclidic purity greater than 99% using a C-100 cyclotron.

4. Summary

Presently, cyclotrons remain the primary facilities for accelerator-based medical isotope production, although linacs and neutron generators are rapidly becoming a viable alternative.

Cyclotrons with adjustable energy ranges or medium energy can produce various kinds of medical isotopes and can cover most radiopharmaceutical production needs in a region [59]. Yield and purity improvements in medical isotopes and the overall cost of cyclotron production have led researchers to explore further possibilities, including proton linacs, which have significant advantages in providing proton beams in the order of tens to hundreds of MeV [179]. These linacs can be developed in research institutes or laboratories conducting scientific experiments and physical research at the same time. For electron linacs, the cross-section of photonuclear interactions is relatively low, which restricts their practical applications. Other factors, such as impurity products and economic costs, also play major roles when evaluating production techniques and methodologies. Attempts to produce medical isotopes through neutron generators are promising and could theoretically yield the medical isotopes that are currently produced by reactors. However, improving the neutron flux rate remains a major consideration.

As medical isotopes produced by reactors often face supply shortages, interest in the use of accelerator-based techniques to produce medical isotopes will increase. We hope to develop an accelerator with the right energy, right beam types, right location, and good shielding facilities, which will play an important role in the supply of medical isotopes.

Author Contributions: Conceptualization, Y.W. and D.C.; methodology, D.C.; investigation, Y.W., D.C., R.d.S.A., J.L. (Jixin Liang) and Z.L.; resources, Z.L., R.d.S.A., J.L. (Jixin Liang), Z.Q. and J.L. (Juntao Liu); writing—original draft preparation, Y.W.; writing—review and editing, R.d.S.A. and J.L. (Jixin Liang); supervision, Z.Q. and J.L. (Juntao Liu); project administration, Z.L.; funding acquisition, J.L. (Juntao Liu). All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National natural Science Foundation of China, grant number 11975115; Special Projects of the Central Government in Guidance of Local Science and Technology Development (Research and development of three-dimensional prospecting technology based on Cosmic-ray muons), grant number YDZX20216200001297; the Research and Development of Medical Isotopes based on High-current Superconducting Linear Accelerator Project, the Fundamental Research Funds for the Central Universities, grant number lzujbky-2019-54; the Science and Technology Planning Project of Gansu, grant number 20JR10RA645; Lanzhou University Talent Cooperation Research Funds sponsored by Lanzhou City, grant number 561121203 and Gansu provincial science and technology plan projects for talents, grant number 054000029.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: No new data were created or analyzed in this study. Data sharing is not applicable to this article.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Radioisotopes in Medicine [EB/OL]. Available online: <https://world-nuclear.org/information-library/non-power-nuclear-applications/radioisotopes-research/radioisotopes-in-medicine.aspx> (accessed on 1 June 2022).
2. AMA Manual of Style Committee. *AMA manual of style: A guide for authors and editors, 10 th ed.*; Oxford University Press: New York, NY, USA, 2007; ISBN 978-0-19-517633-9.
3. Reuzé, S.; Schernberg, A.; Orhac, F.; Sun, R.; Chargari, C.; Derclé, L.; Deutsch, E.; Buvat, I.; Robert, C. Radiomics in nuclear medicine applied to radiation therapy: Methods, pitfalls, and challenges. *Int. J. Radiat. Oncol. Biol. Phys.* **2018**, *102*, 1117–1142. [[CrossRef](#)] [[PubMed](#)]
4. Langbein, T.; Weber, W.A.; Eiber, M. Future of theranostics: An outlook on precision oncology in nuclear medicine. *J. Nucl. Med.* **2019**, *60*, 13S–19S. [[CrossRef](#)] [[PubMed](#)]
5. Kar, N.R. Production and applications of radiopharmaceuticals: A review. *Int. J. Pharm. Investig.* **2019**, *9*, 36–42. [[CrossRef](#)]
6. Vermeulen, K.; Vandamme, M.; Bormans, G.; Cleeren, F. Design and challenges of radiopharmaceuticals. In *Seminars in Nuclear Medicine*; WB Saunders: Philadelphia, PA, USA, 2019; Volume 49, pp. 339–356.
7. Holly, T.A.; Abbott, B.G.; Al-Mallah, M.; Calnon, D.A.; Cohen, M.C.; DiFilippo, F.P.; Ficaro, E.P.; Freeman, M.R.; Hendel, R.C.; Jain, D.; et al. Single photon-emission computed tomography. *J. Nucl. Cardiol.* **2010**, *17*, 941–973. [[CrossRef](#)] [[PubMed](#)]
8. Jaszczak, R.J.; Coleman, R.E.; Lim, C.B. SPECT: Single photon emission computed tomography. *IEEE Trans. Nucl. Sci.* **1980**, *27*, 1137–1153. [[CrossRef](#)]
9. Jaszczak, R.J.; Coleman, R.E. Single photon emission computed tomography (SPECT). Principles and instrumentation. *Investig. Radiol.* **1985**, *20*, 897–910. [[CrossRef](#)]
10. Valk, P.E.; Delbeke, D.; Bailey, D.L.; Townsend, D.W.; Maisey, M.N. *Positron Emission Tomography*; Springer: London, UK, 2005.
11. Kubota, K. From tumor biology to clinical PET: A review of positron emission tomography (PET) in oncology. *Ann. Nucl. Med.* **2001**, *15*, 471–486. [[CrossRef](#)]
12. Wagner, H.N., Jr. A brief history of positron emission tomography (PET). In *Seminars in Nuclear Medicine*; WB Saunders: Philadelphia, PA, USA, 1998; Volume 28, pp. 213–220.
13. Wheat, J.M.; Currie, G.M.; Davidson, R.; Kiat, H. An introduction to nuclear medicine. *Radiographer* **2011**, *58*, 38–45. [[CrossRef](#)]
14. Mariani, G.; Bruselli, L.; Kuwert, T.; Kim, E.E.; Flotats, A.; Israel, O.; Dondi, M.; Watanabe, N. A review on the clinical uses of SPECT/CT. *Eur. J. Nucl. Med. Mol. Imaging* **2010**, *37*, 1959–1985. [[CrossRef](#)]
15. Palmedo, H.; Bucerius, J.; Joe, A.; Strunk, H.; Hortling, N.; Meyka, S.; Roedel, R.; Wolff, M.; Wardelmann, E.; Biersack, H.J.; et al. Integrated PET/CT in differentiated thyroid cancer: Diagnostic accuracy and impact on patient management. *J. Nucl. Med.* **2006**, *47*, 616–624.
16. Liqun, H.; Shufang, L.; Ge, S.; Huan, L.; Jianguo, L.; Quan, A.; Zhongwen, W. Current Applications and Prospects of Radionuclide for Therapy. *J. Isot.* **2021**, *34*, 412.
17. Rösch, F.; Baum, R.P. Generator-based PET radiopharmaceuticals for molecular imaging of tumours: On the way to THERANOSTICS. *Dalton Trans.* **2011**, *40*, 6104–6111. [[CrossRef](#)]
18. Notni, J.; Wester, H.J. Re-thinking the role of radiometal isotopes: Towards a future concept for theranostic radiopharmaceuticals. *J. Label. Compd. Radiopharm.* **2018**, *61*, 141–153. [[CrossRef](#)]

19. Qaim, S.M.; Scholten, B.; Neumaier, B. New developments in the production of theranostic pairs of radionuclides. *J. Radioanal. Nucl. Chem.* **2018**, *318*, 1493–1509. [[CrossRef](#)]
20. Nagarajah, J.; Janssen, M.; Hetkamp, P.; Jentzen, W. Iodine symporter targeting with ¹²⁴I/¹³¹I theranostics. *J. Nucl. Med.* **2017**, *58* (Suppl. S2), 34S–38S. [[CrossRef](#)]
21. Eberlein, U.; Cremonesi, M.; Lassmann, M. Individualized dosimetry for theranostics: Necessary, nice to have, or counterproductive? *J. Nucl. Med.* **2017**, *58* (Suppl. S2), 97S–103S. [[CrossRef](#)]
22. Braccini, S.; Belver-Aguilar, C.; Carzaniga, T.; Dellepiane, G.; Häffner, P.; Scampoli, P. Novel irradiation methods for theranostic radioisotope production with solid targets at the Bern medical cyclotron. In Proceedings of the International Conference on Cyclotrons and their Applications (CYC), Cape Town, South Africa, 22–27 September 2019; pp. 22–27.
23. Brandt, M.; Cardinale, J.; Aulsebrook, M.L.; Gasser, G.; Mindt, T.L. An overview of PET radiochemistry, part 2: Radiometals. *J. Nucl. Med.* **2018**, *59*, 1500–1506. [[CrossRef](#)]
24. Poschenrieder, A.; Schottelius, M.; Schwaiger, M.; Kessler, H.; Wester, H.-J. The influence of different metal-chelate conjugates of pentixafor on the CXCR4 affinity. *EJNMMI Res.* **2016**, *6*, 36. [[CrossRef](#)]
25. Ahn, B.C. Personalized medicine based on theranostic radioiodine molecular imaging for differentiated thyroid cancer. *BioMed Res. Int.* **2016**, *2016*, 1680464. [[CrossRef](#)]
26. Miller, C.; Rousseau, J.; Ramogida, C.F.; Celler, A.; Rahmim, A.; Uribe, C.F. Implications of physics, chemistry and biology for dosimetry calculations using theranostic pairs. *Theranostics* **2022**, *12*, 232. [[CrossRef](#)]
27. Ehlerding, E.B.; Ferreira, C.A.; Aluicio-Sarduy, E.; Jiang, D.; Lee, H.J.; Theuer, C.P.; Engle, J.W.; Cai, W. ^{86/90}Y-based theranostics targeting angiogenesis in a murine breast cancer model. *Mol. Pharm.* **2018**, *15*, 2606–2613. [[CrossRef](#)]
28. Ferreira, C.A.; Ehlerding, E.B.; Rosenkrans, Z.T.; Jiang, D.; Sun, T.; Aluicio-Sarduy, E.; Engle, J.W.; Ni, D.; Cai, W. ^{86/90}Y-Labeled monoclonal antibody targeting tissue factor for pancreatic cancer theranostics. *Mol. Pharm.* **2020**, *17*, 1697–1705. [[CrossRef](#)]
29. Ming-qi, L.I.; Qi-min, D.; Zuo-yong, C.; Mao-liang, L.I. Production and application of medical radionuclide: Status and urgent problems to be resolved in China. *J. Isot.* **2013**, *26*, 186. (In Chinese)
30. Mushtaq, A. Reactors are indispensable for radioisotope production. *Ann. Nucl. Med.* **2010**, *24*, 759–760. [[CrossRef](#)]
31. Xoubi, N.; Primm, R.T., III. *Modeling of the High Flux Isotope Reactor Cycle 400*; ORNL/TM-2004/251; Oak Ridge National Laboratory: Oak Ridge, Tennessee, USA, 2005.
32. Ruth, T.J. The medical isotope crisis: How we got here and where we are going. *J. Nucl. Med. Technol.* **2014**, *42*, 245–248. [[CrossRef](#)]
33. Kolečka, M.; Lahodová, Z.; Šoltés, J.; Viererbl, L.; Ernest, J.; Vinš, M.; Stehno, J. Capabilities of the LVR-15 research reactor for production of medical and industrial radioisotopes. *J. Radioanal. Nucl. Chem.* **2015**, *305*, 51–59. [[CrossRef](#)]
34. OECD-NEA. *The Supply of Medical Radioisotopes: 2019 Medical Isotope Supply and Capacity Projection for the 2019–2024 Period*; OECD-NEA: Paris, France, 2019.
35. Gao, F.; Lin, L.; Liu, Y.; Ma, X. Production situation and technology prospect of medical isotopes. *J. Isot.* **2016**, *29*, 116–120. (In Chinese)
36. Kurenkov, N.V.; Shubin, Y.N. Radionuclides for nuclear medicine. *Медицинская Радиология И Радиационная Безопасность* **1996**, *41*, 54–63.
37. IAEA. *Nuclear Research Reactors in the World*; IAEA: New York, NY, USA, 1997; 120p, ISBN 92-0-100298-X.
38. Hoedl, S.A.; Updegraff, W.D. The production of medical isotopes without nuclear reactors or uranium enrichment. *Sci. Glob. Secur.* **2015**, *23*, 121–153. [[CrossRef](#)]
39. Van der Keur, H. Medical radioisotopes production without a nuclear reactor. 2010. Available online: http://www.laka.org/info/publicaties/2010-medical_isotopes.pdf (accessed on 20 June 2022).
40. Ziwei, L.; Yuncheng, H.; Xiaoyu, W.; Jiachen, Z.; Yongfeng, W.; Qunying, H. Production Status and Technical Prospects of Medical Radioisotope ⁹⁹Mo/^{99m}Tc. *Nucl. Phys. Rev.* **2019**, *36*, 170–183. (In Chinese)
41. Starovoitova, V.N.; Tchelidze, L.; Wells, D.P. Production of medical radioisotopes with linear accelerators. *Appl. Radiat. Isot.* **2014**, *85*, 39–44. [[CrossRef](#)] [[PubMed](#)]
42. Kaur, C.D.; Mishra, K.K.; Sahu, A.; Panik, R.; Kashyap, P.; Mishra, S.P.; Kumar, A. Theranostics: New era in nuclear medicine and radiopharmaceuticals. In *Medical Isotopes*; Naqvi, S.A.R., Imrani, M.B., Eds.; IntechOpen: London, UK, 2020.
43. Zhang, T.; Fan, M.; Wei, S.; Chen, S.; Yang, F. The present situation and the prospect of medical cyclotrons in China. *Sci. China Phys. Mech. Astron.* **2011**, *54*, 260–265. [[CrossRef](#)]
44. Sunderland, J.; Erdahl, C.; Bender, B.; Sensoy, L.; Watkins, G. Considerations, measurements and logistics associated with low-energy cyclotron decommissioning. In Proceedings of the AIP Conference Proceedings, Playa del Carmen, México, 26–29 August 2012; American Institute of Physics: New York, NY, USA, 2012; Volume 1509, pp. 16–20.
45. International Atomic Energy Agency. *Alternative Radionuclide Production with a Cyclotron*; IAEA Radioisotopes and Radiopharmaceuticals Reports No. 4; IAEA: Vienna, Austria, 2021.
46. Chernyaev, A.P.; Varzar, S.M. Particle accelerators in modern world. *Phys. At. Nucl.* **2014**, *77*, 1203–1215. [[CrossRef](#)]
47. Goethals, P.E.; Zimmermann, R.G. *Cyclotrons used in Nuclear Medicine World Market Report & Directory*; MEDraysintell: Louvain-la-Neuve, Belgium, 2015.
48. Available online: <https://www.machinedesign.com/learning-resources/whats-the-difference-between/article/21832184/what-are-the-differences-between-linear-accelerators-cyclotrons-and-synchrotrons> (accessed on 1 June 2022).

49. Available online: <https://www.iaea.org/newscenter/news/cyclotrons-what-are-they-and-where-can-you-find-them> (accessed on 10 July 2022).
50. Available online: <https://www.iaea.org/sites/default/files/gc/gc65-inf2.pdf> (accessed on 10 July 2022).
51. Available online: <https://www.iaea.org/sites/default/files/gc/gc64-inf2.pdf> (accessed on 10 July 2022).
52. Available online: https://www.iaea.org/sites/default/files/gc/gc61inf-4_en.pdf (accessed on 10 July 2022).
53. Available online: <https://www.iaea.org/sites/default/files/ntr2015.pdf> (accessed on 10 July 2022).
54. Available online: <https://www.iaea.org/sites/default/files/ntr2014.pdf> (accessed on 10 July 2022).
55. Available online: https://www-legacy.iaea.org/OurWork/ST/NE/Pess/assets/13-25751_rep_ntr_2013_web.pdf (accessed on 10 July 2022).
56. Available online: https://www-legacy.iaea.org/OurWork/ST/NE/Pess/assets/ntr2012_web.pdf (accessed on 10 July 2022).
57. Available online: <https://www-legacy.iaea.org/OurWork/ST/NE/Pess/assets/ntr2011.pdf> (accessed on 10 July 2022).
58. Chao, A.W.; Chou, W. (Eds.) *Reviews of Accelerator Science and Technology-Volume 3: Accelerators as Photon Sources*; World Scientific: Chiyoda City, Tokyo, Japan, 2011.
59. Leo, K.W.K.; Hashim, S. Accelerator Selection for Industry and Medical Applications. (This is a preprint article, it offers immediate access but has not been peer reviewed).
60. Synowiecki, M.A.; Perk, L.R.; Nijssen, J.F.W. Production of novel diagnostic radionuclides in small medical cyclotrons. *EJNMMI Radiopharm. Chem.* **2018**, *3*, 3. [[CrossRef](#)]
61. Zuoyuan, D.; Bin, W. *Securities Research Report-In-Depth Discussion Series-Nuclear Medicine*; Pacific Securities: Guangdong, China, 2019.
62. Yuan, Z. FDA approved radiopharmaceuticals. In *Foreign Medical Sciences*; Section of Radiation Medicine and Nuclear Medicine: Tianjin, China, 2000; Volume 24, pp. 161–163, ISSN 1001-098X.
63. Racette, B.A.; Antenor, J.A.; McGee-Minnich, L.; Moerlein, S.M.; Videen, T.O.; Kotagal, V.; Perlmutter, J.S. [¹⁸F] FDOPA PET and clinical features in parkinsonism due to manganese. *Mov. Disord.* **2005**, *20*, 492–496. [[CrossRef](#)]
64. Zhao, P.; Zhang, B.; Gao, S.; Li, X. Clinical features, MRI, and ¹⁸F-FDG-PET in differential diagnosis of Parkinson disease from multiple system atrophy. *Brain Behav.* **2020**, *10*, e01827. [[CrossRef](#)]
65. Rahmim, A.; Zaidi, H. PET versus SPECT: Strengths, limitations and challenges. *Nucl. Med. Commun.* **2008**, *29*, 193–207. [[CrossRef](#)]
66. Ruth, T.J.; Wolf, A.P. Absolute cross sections for the production of ¹⁸F via the ¹⁸O (p, n) ¹⁸F reaction. *Radiochim. Acta* **1979**, *26*, 21–24. [[CrossRef](#)]
67. Hess, E.; Blessing, G.; Coenen, H.H.; Qaim, S.M. Improved target system for production of high purity [¹⁸F] fluorine via the ¹⁸O (p, n) ¹⁸F reaction. *Appl. Radiat. Isot.* **2000**, *52*, 1431–1440. [[CrossRef](#)]
68. Kambali, I.; Parwanto; Suryanto, H.; Huda, N.; Listiawadi, F.D.; Astarina, H.; Ismuha, R.R.; Kardinah. Dependence of ¹⁸F Production Yield and Radioactive Impurities on Proton Irradiation Dose. *Phys. Res. Int.* **2017**, *2017*, 2124383. [[CrossRef](#)]
69. P Perini, E.A.; Skopchenko, M.; Hong, T.T.; Harianto, R.; Maître, A.; Rodríguez, M.R.R.; de Oliveira Santos, N.; Guo, Y.; Qin, X.; Zeituni, C.A.; et al. Pre-feasibility study for establishing radioisotope and radiopharmaceutical production facilities in developing countries. *Curr. Radiopharm.* **2019**, *12*, 187–200. [[CrossRef](#)]
70. Barnhart, T.E.; Nickles, R.J.; Roberts, A.D. Revisiting Low Energy Deuteron Production of [¹⁸F] Fluoride and Fluorine for PET. In Proceedings of the AIP Conference Proceedings, Denton, Texas, USA, 12–16 November 2002; American Institute of Physics: New York, NY, USA, 2003; Volume 680, pp. 1086–1089.
71. Haug, A.R.; Cindea-Drimus, R.; Auernhammer, C.J.; Reincke, M.; Wängler, B.; Uebleis, C.; Schmidt, G.P.; Göke, B.; Bartenstein, P.; Hacker, M. The role of ⁶⁸Ga-DOTATATE PET/CT in suspected neuroendocrine tumors. *J. Nucl. Med.* **2012**, *53*, 1686–1692. [[CrossRef](#)]
72. Kręcis, P.; Czarnecka, K.; Królicki, L.; Mikiciuk-Olasik, E.b.; Szymański, P. Radiolabeled peptides and antibodies in medicine. *Bioconjugate Chem.* **2020**, *32*, 25–42. [[CrossRef](#)]
73. Vaughn, B.A. *Chelation Approaches for the Theranostic Radioisotopes of Copper, Scandium and Lutetium*; State University of New York at Stony Brook: York, NE, USA, 2021.
74. Krebs, S.; O'Donoghue, J.A.; Biegel, E.; Beattie, B.J.; Reidy, D.; Lyashchenko, S.K.; Lewis, J.S.; Bodei, L.; Weber, W.A.; Pandit-Taskar, N. Comparison of ⁶⁸Ga-DOTA-JR11 PET/CT with dosimetric ¹⁷⁷Lu-satoreotide tetraxetan (¹⁷⁷Lu-DOTA-JR11) SPECT/CT in patients with metastatic neuroendocrine tumors undergoing peptide receptor radionuclide therapy. *Eur. J. Nucl. Med. Mol. Imaging* **2020**, *47*, 3047–3057. [[CrossRef](#)]
75. Maffey-Steffan, J.; Scarpa, L.; Sviridenka, A.; Nilica, B.; Mair, C.; Buxbaum, S.; Bektic, J.; von Guggenberg, E.; Uprimny, C.; Horninger, W.; et al. The ⁶⁸Ga/¹⁷⁷Lu-theragnostic concept in PSMA-targeting of metastatic castration-resistant prostate cancer: Impact of post-therapeutic whole-body scintigraphy in the follow-up. *Eur. J. Nucl. Med. Mol. Imaging* **2020**, *47*, 695–712. [[CrossRef](#)]
76. Scarpa, L.; Buxbaum, S.; Kendler, D.; Fink, K.; Bektic, J.; Gruber, L.; Decristoforo, C.; Uprimny, C.; Lukas, P.; Horninger, W.; et al. The ⁶⁸Ga/¹⁷⁷Lu theragnostic concept in PSMA targeting of castration-resistant prostate cancer: Correlation of SUVmax values and absorbed dose estimates. *Eur. J. Nucl. Med. Mol. Imaging* **2017**, *44*, 788–800. [[CrossRef](#)]
77. Sartor, O.; Herrmann, K. Prostate Cancer Treatment: ¹⁷⁷Lu-PSMA-617 Considerations, Concepts, and Limitations. *J. Nucl. Med.* **2022**, *63*, 823–829. [[CrossRef](#)]
78. Velikyan, I. ⁶⁸Ga-based radiopharmaceuticals: Production and application relationship. *Molecules* **2015**, *20*, 12913–12943. [[CrossRef](#)]

79. Lin, M.; Waligorski, G.J.; Lepera, C.G. Production of curie quantities of ^{68}Ga with a medical cyclotron via the ^{68}Zn (p, n) ^{68}Ga reaction. *Appl. Radiat. Isot.* **2018**, *133*, 1–3. [[CrossRef](#)]
80. Razbash, A.A.; Sevastianov, Yu.G.; Krasnov, N.N.; Leonov, A.I.; Pavlekin, V.E. Germanium-68 row of products. In Proceedings of the 5th International Conference on Isotopes, 5ICI, Brussels, Belgium, 25–29 April 2005; Medimond: Bologna, Italy; p. 147.
81. Rösch, F. Past, present and future of $^{68}\text{Ge}/^{68}\text{Ga}$ generators. *Appl. Radiat. Isot.* **2013**, *76*, 24–30. [[CrossRef](#)]
82. Engle, J.; Lopez-Rodriguez, V.; Gaspar-Carcamo, R.; Valdovinos, H.; Valle-Gonzalez, M.; Trejo-Ballado, F.; Severin, G.W.; Barnhart, T.; Nickles, R.; Avila-Rodriguez, M.A. Very high specific activity $^{66/68}\text{Ga}$ from zinc targets for PET. *Appl. Radiat. Isot.* **2012**, *70*, 1792–1796. [[CrossRef](#)]
83. Sadeghi, M.; Kakavand, T.; Rajabifar, S.; Mokhtari, L.; Rahimi-Nezhad, A. Cyclotron production of ^{68}Ga via proton-induced reaction on ^{68}Zn target. *Nukleonika* **2009**, *54*, 25–28.
84. Nelson, B.J.; Wilson, J.; Richter, S.; Duke, M.J.M.; Wuest, M.; Wuest, F. Taking cyclotron ^{68}Ga production to the next level: Expeditious solid target production of ^{68}Ga for preparation of radiotracers. *Nucl. Med. Biol.* **2020**, *80*, 24–31. [[CrossRef](#)]
85. Mardon, A.; Saleem, H.; Parish, G.; Syed, M.; Inayat, E.; Henry, J.; Heinen, L.; Amiscaray, D.E.; Dong, F.; Mak, E.; et al. *What in the World is Medical Isotope Production?* Golden Meteorite Press: Edmonton, Alberta, Canada, 2021.
86. Pandey, M.K.; Byrne, J.F.; Jiang, H.; Packard, A.B.; DeGrado, T.R. Cyclotron production of ^{68}Ga via the ^{68}Zn (p, n) ^{68}Ga reaction in aqueous solution. *Am. J. Nucl. Med. Mol. Imaging* **2014**, *4*, 303.
87. Keinänen, O.; Fung, K.; Brennan, J.M.; Zia, N.; Harris, M.; van Dam, E.; Biggin, C.; Hedt, A.; Stoner, J.; Donnelly, P.S.; et al. Harnessing $^{64}\text{Cu}/^{67}\text{Cu}$ for a theranostic approach to pretargeted radioimmunotherapy. *Proc. Natl. Acad. Sci. USA* **2020**, *117*, 28316–28327. [[CrossRef](#)]
88. Pfeifer, A.; Knigge, U.; Mortensen, J.; Oturai, P.; Berthelsen, A.K.; Loft, A.; Binderup, T.; Rasmussen, P.; Elema, D.; Klausen, T.L.; et al. Clinical PET of neuroendocrine tumors using ^{64}Cu -DOTATATE: First-in-humans study. *J. Nucl. Med.* **2012**, *53*, 1207–1215. [[CrossRef](#)]
89. Avila-Rodriguez, M.A.; Nye, J.A.; Nickles, R.J. Simultaneous production of high specific activity ^{64}Cu and ^{61}Co with 11.4 MeV protons on enriched ^{64}Ni nuclei. *Appl. Radiat. Isot.* **2007**, *65*, 1115–1120. [[CrossRef](#)]
90. Szelecsényi, F.; Kovács, Z.; Nagatsu, K.; Zhang, M.R.; Suzuki, K. Excitation function of (p, α) nuclear reaction on enriched ^{67}Zn : Possibility of production of ^{64}Cu at low energy cyclotron. *Radiochim. Acta* **2014**, *102*, 465–472. [[CrossRef](#)]
91. Obata, A.; Kasamatsu, S.; McCarthy, D.W.; Welch, M.J.; Saji, H.; Yonekura, Y.; Fujibayashi, Y. Production of therapeutic quantities of ^{64}Cu using a 12 MeV cyclotron. *Nucl. Med. Biol.* **2003**, *30*, 535–539. [[CrossRef](#)]
92. McCarthy, D.W.; Shefer, R.E.; Klinkowstein, R.E.; Bass, L.A.; Margeneau, W.H.; Cutler, C.S.; Anderson, C.J.; Welch, M.J. Efficient production of high specific activity ^{64}Cu using a biomedical cyclotron. *Nucl. Med. Biol.* **1997**, *24*, 35–43. [[CrossRef](#)]
93. Hilgers, K.; Stoll, T.; Skakun, Y.; Coenen, H.H.; Qaim, S.M. Cross-section measurements of the nuclear reactions natZn (d, x) ^{64}Cu , ^{66}Zn (d, α) ^{64}Cu and ^{68}Zn (p, α n) ^{64}Cu for production of ^{64}Cu and technical developments for small-scale production of ^{67}Cu via the ^{70}Zn (p, α) ^{67}Cu process. *Appl. Radiat. Isot.* **2003**, *59*, 343–351. [[CrossRef](#)]
94. Elomaa, V.V.; Jurttila, J.; Rajander, J.; Solin, O. Automation of ^{64}Cu production at Turku PET Centre. *Appl. Radiat. Isot.* **2014**, *89*, 74–78. [[CrossRef](#)]
95. Zeisler, S.K.; Pavan, R.A.; Orzechowski, J.; Langlois, R.; Rodrigue, S.; Van Lier, J.E. Production of ^{64}Cu on the Sherbrooke TR-PET cyclotron. *J. Radioanal. Nucl. Chem.* **2003**, *257*, 175–177. [[CrossRef](#)]
96. Thieme, S.; Walther, M.; Pietzsch, H.J.; Henniger, J.; Preusche, S.; Mäding, P.; Steinbach, J. Module-assisted preparation of ^{64}Cu with high specific activity. *Appl. Radiat. Isot.* **2012**, *70*, 602–608. [[CrossRef](#)]
97. Van der Meulen, N.P.; Hasler, R.; Blanc, A.; Farkas, R.; Benešová, M.; Talip, Z.; Müller, C.; Schibli, R. Implementation of a new separation method to produce qualitatively improved ^{64}Cu . *J. Label. Compd. Radiopharm.* **2019**, *62*, 460–470. [[CrossRef](#)]
98. Xie, Q.; Zhu, H.; Wang, F.; Meng, X.; Ren, Q.; Xia, C.; Yang, Z. Establishing reliable Cu-64 production process: From target plating to molecular specific tumor micro-PET imaging. *Molecules* **2017**, *22*, 641. [[CrossRef](#)]
99. Ohya, T.; Nagatsu, K.; Suzuki, H.; Fukada, M.; Minegishi, K.; Hanyu, M.; Fukumura, T.; Zhang, M.-R. Efficient preparation of high-quality ^{64}Cu for routine use. *Nucl. Med. Biol.* **2016**, *43*, 685–691. [[CrossRef](#)]
100. Verel, I.; Visser, G.W.; Boellaard, R.; Stigter-van Walsum, M.; Snow, G.B.; Van Dongen, G.A. ^{89}Zr immuno-PET: Comprehensive procedures for the production of ^{89}Zr -labeled monoclonal antibodies. *J. Nucl. Med.* **2003**, *44*, 1271–1281.
101. Vento, J.; Mulgaonkar, A.; Woolford, L.; Nham, K.; Christie, A.; Bagrodia, A.; de Leon, A.D.; Hannan, R.; Bowman, I.; McKay, R.M.; et al. PD-L1 detection using ^{89}Zr -atezolizumab immuno-PET in renal cell carcinoma tumorgrafts from a patient with favorable nivolumab response. *J. Immunother. Cancer* **2019**, *7*, 144. [[CrossRef](#)]
102. Taghilo, M.; Kakavand, T.; Rajabifar, S.; Sarabadani, P. Cyclotron production of ^{89}Zr : A potent radionuclide for positron emission tomography. *Int. J. Phys. Sci.* **2012**, *7*, 1321–1325. [[CrossRef](#)]
103. Ciarmatori, A.; Cicoria, G.; Pancaldi, D.; Infantino, A.; Boschi, S.; Fanti, S.; Marengo, M. Some experimental studies on ^{89}Zr production. *Radiochim. Acta* **2011**, *99*, 631–634. [[CrossRef](#)]
104. Sadeghi, M.; Enferadi, M.; Bakhtiari, M. Accelerator production of the positron emitter zirconium-89. *Ann. Nucl. Energy* **2012**, *41*, 97–103. [[CrossRef](#)]
105. Tang, Y.; Li, S.; Yang, Y.; Chen, W.; Wei, H.; Wang, G.; Yang, J.; Liao, J.; Luo, S.; Liu, N. A simple and convenient method for production of ^{89}Zr with high purity. *Appl. Radiat. Isot.* **2016**, *118*, 326–330. [[CrossRef](#)] [[PubMed](#)]

106. Deri, M.A.; Zeglis, B.M.; Francesconi, L.C.; Lewis, J.S. PET imaging with ^{89}Zr : From radiochemistry to the clinic. *Nucl. Med. Biol.* **2013**, *40*, 3–14. [CrossRef]
107. Kandil, S.A.; Spahn, I.; Scholten, B.; Saleh, Z.A.; Saad, S.M.M.; Coenen, H.H.; Qaim, S.M. Excitation functions of (α , xn) reactions on natRb and natSr from threshold up to 26 MeV: Possibility of production of ^{87}Y , ^{88}Y and ^{89}Zr . *Appl. Radiat. Isot.* **2007**, *65*, 561–568. [CrossRef]
108. Liqiang, L.; Feng, W.; Teli, L.; Hua, Z.; Zhi, Y. Production of Iodine-124 and Its Application in PET Molecular Imaging. *J. Isot.* **2018**, *31*, 188. (In Chinese)
109. Freudenberg, L.S.; Jentzen, W.; Stahl, A.; Bockisch, A.; Rosenbaum-Krumme, S.J. Clinical applications of ^{124}I -PET/CT in patients with differentiated thyroid cancer. *Eur. J. Nucl. Med. Mol. Imaging* **2011**, *38*, 48–56. [CrossRef]
110. Aboian, M.S.; Huang, S.-y.; Hernandez-Pampaloni, M.; Hawkins, R.A.; VanBrocklin, H.F.; Huh, Y.; Vo, K.T.; Gustafson, W.C.; Matthay, K.K.; Seo, Y. ^{124}I -MIBG PET/CT to monitor metastatic disease in children with relapsed neuroblastoma. *J. Nucl. Med.* **2021**, *62*, 43–47. [CrossRef]
111. Lewis, J.S. *Production, Use and Applications of ^{124}I . PowerPoint Slides*; Memorial–Sloan Kettering Cancer Center: New York, NY, USA, 2020.
112. Braghirolli AM, S.; Waissmann, W.; da Silva, J.B.; dos Santos, G.R. Production of iodine-124 and its applications in nuclear medicine. *Appl. Radiat. Isot.* **2014**, *90*, 138–148. [CrossRef]
113. Bzowski, P.; Borys, D.; Gorczewski, K.; Chmura, A.; Daszewska, K.; Gorczewska, I.; Kastelik-Hryniewiecka, A.; Szydło, M.; d’Amico, A.; Sokół, M. Efficiency of ^{124}I radioisotope production from natural and enriched tellurium dioxide using ^{124}Te (p, xn) ^{124}I reaction. *EJNMMI Phys.* **2022**, *9*, 41. [CrossRef]
114. Bastian, T.; Coenen, H.H.; Qaim, S.M. Excitation functions of ^{124}Te (d, xn) $^{124,125}\text{I}$ reactions from threshold up to 14 MeV: Comparative evaluation of nuclear routes for the production of ^{124}I . *Appl. Radiat. Isot.* **2001**, *55*, 303–308. [CrossRef]
115. Payolla, F.B.; Massabni, A.C.; Orvig, C. Radiopharmaceuticals for diagnosis in nuclear medicine: A short review. *Eclética Química* **2019**, *44*, 11–19.
116. Schaffer, P.; Bénard, F.; Bernstein, A.; Buckley, K.; Celler, A.; Cockburn, N.; Corsaut, J.; Dodd, M.; Economou, C.; Eriksson, T.; et al. Direct production of $^{99\text{m}}\text{Tc}$ via ^{100}Mo (p, 2n) on small medical cyclotrons. *Phys. Procedia* **2015**, *66*, 383–395. [CrossRef]
117. Takacs, S.; Hermanne, A.; Ditroi, F.; Tárkányi, F.; Aikawa, M. Reexamination of cross sections of the ^{100}Mo (p, 2n) $^{99\text{m}}\text{Tc}$ reaction. *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2015**, *347*, 26–38. [CrossRef]
118. Scholten, B.; Lambrecht, R.M.; Cogneau, M.; Ruiz, H.V.; Qaim, S.M. Excitation functions for the cyclotron production of $^{99\text{m}}\text{Tc}$ and ^{99}Mo . *Appl. Radiat. Isot.* **1999**, *51*, 69–80. [CrossRef]
119. Rodrigue, S.; van Lier, J.E.; van Lier, M.A.S.E. Cyclotron production of $^{99\text{m}}\text{Tc}$: An approach to the medical isotope crisis. *J. Nucl. Med.* **2010**, *51*, 13N.
120. Hoehr, C.; Bénard, F.; Buckley, K.; Crawford, J.; Gottberg, A.; Hanemaayer, V.; Kunz, P.; Ladouceur, K.; Radchenko, V.; Ramogida, C.; et al. Medical isotope production at TRIUMF—from imaging to treatment. *Phys. Procedia* **2017**, *90*, 200–208. [CrossRef]
121. Pillai, M.R.A.; Dash, A.; Knapp, F.F.R. Sustained availability of $^{99\text{m}}\text{Tc}$: Possible paths forward. *J. Nucl. Med.* **2013**, *54*, 313–323. [CrossRef]
122. Lebeda, O.; van Lier, E.J.; Štursa, J.; Ráliš, J.; Zyuzin, A. Assessment of radionuclidic impurities in cyclotron produced $^{99\text{m}}\text{Tc}$. *Nucl. Med. Biol.* **2012**, *39*, 1286–1291. [CrossRef]
123. Pupillo, G.; Esposito, J.; Gambaccini, M.; Haddad, F.; Michel, N. Experimental cross section evaluation for innovative ^{99}Mo production via the (α , n) reaction on ^{96}Zr target. *J. Radioanal. Nucl. Chem.* **2014**, *302*, 911–917. [CrossRef]
124. Hagiwara, M.; Yashima, H.; Sanami, T.; Yonai, S. Measurement of the excitation function of ^{96}Zr (α , n) ^{99}Mo for an alternative production source of medical radioisotopes. *J. Radioanal. Nucl. Chem.* **2018**, *318*, 569–573. [CrossRef]
125. Jacobson, A.F.; Deng, H.; Lombard, J.; Lessig, H.J.; Black, R.R. ^{123}I -meta-iodobenzylguanidine scintigraphy for the detection of neuroblastoma and pheochromocytoma: Results of a meta-analysis. *J. Clin. Endocrinol. Metab.* **2010**, *95*, 2596–2606. [CrossRef]
126. Eslami, M.; Kakavand, T.; Mirzaii, M. Simulation of Proton beam using the MCNPX code; A prediction for the production of ^{123}I via ^{124}Xe (p, x) ^{123}I reaction. In Proceedings of the DAE-BRNS symposium on nuclear physics, Mumbai, India, 2–6 December 2013; Volume 58, p. 860.
127. EXFOR. Experimental Nuclear Reaction Data. 2011. Available online: <http://www-nds.iaea.org/exfor> (accessed on 15 June 2022).
128. Kakavand, T.; Sadeghi, M.; Kamali Moghaddam, K.; Shokri Bonab, S.; Fateh, B. Computer simulation techniques to design Xenon-124 solid target for iodine-123 production. *Iran. J. Radiat. Res.* **2008**, *5*, 207–212.
129. Tárkányi, F.; Qaim, S.M.; Stöcklin, G.; Sajjad, M.; Lambrecht, R.M.; Schweickert, H. Excitation functions of (p, 2n) and (p, pn) reactions and differential and integral yields of ^{123}I in proton induced nuclear reactions on highly enriched ^{124}Xe . *Int. J. Radiat. Appl. Instrumentation. Part A Appl. Radiat. Isot.* **1991**, *42*, 221–228. [CrossRef]
130. Hupf, H.B.; Beaver, J.E.; Armbruster, J.M.; Pendola, J.P. Production of ultra-pure I-123 from the ^{123}Te (p, n) ^{123}I reaction. *AIP Conf Proc* **2001**, *576*, 845–848.
131. Mertens, J. New Development in Radio-Iodinated Radiopharmaceuticals for SPECT and Radionuclide Therapy. In Proceedings of the IAEA-CN-130 International Symposium on Trends in Radiopharmaceuticals ISTR-2005, Vienna, Austria, 14–18 November 2005; IAEA: New York, NY, USA; pp. 101–103.

132. Scholten, B.; Qaim, S.M.; Stöcklin, G. Excitation functions of proton induced nuclear reactions on natural tellurium and enriched ^{123}Te : Production of ^{123}I via the $^{123}\text{Te}(p, n)^{123}\text{I}$ -process at a low-energy cyclotron. *Int. J. Radiat. Appl. Instrumentation. Part A Appl. Radiat. Isot.* **1989**, *40*, 127–132. [[CrossRef](#)]
133. Kratochwil, C.; Haberkorn, U.; Giesel, F.L. ^{225}Ac -PSMA-617 for therapy of prostate cancer. In *Seminars in Nuclear Medicine*; WB Saunders: Philadelphia, PA, USA, 2020; Volume 50, pp. 133–140.
134. Kratochwil, C.; Bruchertseifer, F.; Giesel, F.L.; Weis, M.; Verburg, F.A.; Mottaghy, F.; Kopka, K.; Apostolidis, C.; Haberkorn, U.; Morgenstern, A. ^{225}Ac -PSMA-617 for PSMA-targeted α -radiation therapy of metastatic castration-resistant prostate cancer. *J. Nucl. Med.* **2016**, *57*, 1941–1944. [[CrossRef](#)]
135. Zacherl, M.J.; Gildehaus, F.J.; Mittlmeier, L.; Böning, G.; Gosewisch, A.; Wenter, V.; Unterrainer, M.; Schmidt-Hegemann, N.; Belka, C.; Kretschmer, A. First clinical results for PSMA-targeted α -therapy using ^{225}Ac -PSMA-I&T in advanced-mCRPC patients. *J. Nucl. Med.* **2021**, *62*, 669–674.
136. Królicki, L.; Kunikowska, J.; Bruchertseifer, F.; Koziara, H.; Królicki, B.; Jakuciński, M.; Pawlak, D.; Rola, R.; Morgenstern, A.; Rosiak, E.; et al. ^{225}Ac - and ^{213}Bi -substance P analogues for glioma therapy. In *Seminars in Nuclear Medicine*; WB Saunders: Philadelphia, PA, USA, 2020; Volume 50, pp. 141–151.
137. Nagatsu, K.; Suzuki, H.; Fukada, M.; Ito, T.; Ichinose, J.; Honda, Y.; Minegishi, K.; Higashi, T.; Zhang, M.-R. Cyclotron production of ^{225}Ac from an electroplated ^{226}Ra target. *Eur. J. Nucl. Med. Mol. Imaging* **2021**, *49*, 279–289. [[CrossRef](#)]
138. Lee, K.C. ^{225}Ac production at KIRAMS. In Proceedings of the IAEA Workshop on the Supply of ^{225}Ac , Vienna, Austria, 9–10 October 2018, (unpublished).
139. Bruchertseifer, F.; Kellerbauer, A.; Malmbeck, R.; Morgenstern, A. Targeted alpha therapy with bismuth-213 and actinium-225: Meeting future demand. *J. Label. Compd. Radiopharm.* **2019**, *62*, 794–802. [[CrossRef](#)]
140. Ermolaev, S.; Zhuikov, B.; Kokhanyuk, V.; Matushko, V.; Kalmykov, S.N.; Aliev, R.A.; Tananaev, I.G.; Myasoedov, B.F. Production of actinium, thorium and radium isotopes from natural thorium irradiated with protons up to 141 MeV. *Radiochim. Acta* **2012**, *100*, 223–229. [[CrossRef](#)]
141. Chen, D.; Liu, W.; Huang, Q.; Cao, S.; Tian, W.; Yin, X.; Tan, C.; Wang, J.; Chu, J.; Jia, Z.; et al. Accelerator Production of the Medical Isotope ^{211}At and Monoclonal Antibody Labeling. *Acta Chim. Sin.* **2021**, *79*, 1376–1384. [[CrossRef](#)]
142. Watabe, T.; Kaneda-Nakashima, K.; Shirakami, Y.; Liu, Y.; Ooe, K.; Teramoto, T.; Toyoshima, A.; Shimosegawa, E.; Nakano, T.; Kanai, Y.; et al. Targeted alpha therapy using astatine (^{211}At)-labeled phenylalanine: A preclinical study in glioma bearing mice. *Oncotarget* **2020**, *11*, 1388. [[CrossRef](#)]
143. Zalutsky, M.R.; Reardon, D.A.; Akabani, G.; Coleman, R.E.; Friedman, A.H.; Friedman, H.S.; McLendon, R.E.; Wong, T.Z.; Bigner, D.D. Clinical experience with α -particle-emitting ^{211}At : Treatment of recurrent brain tumor patients with ^{211}At -labeled chimeric antitenascin monoclonal antibody 81C6. *J. Nucl. Med.* **2008**, *49*, 30–38. [[CrossRef](#)] [[PubMed](#)]
144. Lindegren, S.; Albertsson, P.; Bäck, T.; Jensen, H.; Palm, S.; Aneheim, E. Realizing clinical trials with astatine-211: The chemistry infrastructure. *Cancer Biother. Radiopharm.* **2020**, *35*, 425–436. [[CrossRef](#)]
145. Cederkrantz, E.; Andersson, H.; Bernhardt, P.; Bäck, T.; Hultborn, R.; Jacobsson, L.; Jensen, H.; Lindegren, S.; Ljungberg, M.; Magnander, T.; et al. Absorbed doses and risk estimates of ^{211}At -MX35 F (ab')₂ in intraperitoneal therapy of ovarian cancer patients. *Int. J. Radiat. Oncol. Biol. Phys.* **2015**, *93*, 569–576. [[CrossRef](#)]
146. Washiyama, K.; Oda, T.; Sasaki, S.; Aoki, M.; Gomez, F.L.G.; Taniguchi, M.; Nishijima, K.-i.; Takahashi, K. At-211 production using the CYPRIIS MP-30. *J. Med. Imaging Radiat. Sci.* **2019**, *50*, S42. [[CrossRef](#)]
147. Alfaraano, A.; Abbas, K.; Holzwarth, U.; Bonardi, M.; Groppi, F.; Alfassi, Z.; Menapace, E.; Gibson, P. Thick target yield measurement of ^{211}At through the nuclear reaction $^{209}\text{Bi}(\alpha, 2n)$. In *Journal of Physics: Conference Series*; IOP Publishing: Bristol, UK, 2006; Volume 41, p. 009.
148. Feng, Y.; Zalutsky, M.R. Production, purification and availability of ^{211}At : Near term steps towards global access. *Nucl. Med. Biol.* **2021**, *100*, 12–23. [[CrossRef](#)]
149. Guérard, F.; Gestin, J.F.; Brechbiel, M.W. Production of [^{211}At]-astatinated radiopharmaceuticals and applications in targeted α -particle therapy. *Cancer Biother. Radiopharm.* **2013**, *28*, 1–20. [[CrossRef](#)]
150. Cullinane, C.; Jeffery, C.M.; Roselt, P.D.; van Dam, E.M.; Jackson, S.; Kuan, K.; Jackson, P.; Binns, D.; van Zuylekom, J.; Harris, M.; et al. Peptide receptor radionuclide therapy with ^{67}Cu -CuSarTATE is highly efficacious against a somatostatin-positive neuroendocrine tumor model. *J. Nucl. Med.* **2020**, *61*, 1800–1805. [[CrossRef](#)]
151. DeNardo, S.J.; DeNardo, G.L.; Kukis, D.L.; Shen, S.; Kroger, L.A.; DeNardo, D.A.; Goldstein, D.S.; Mirick, G.R.; Salako, Q.; Mausner, L.F.; et al. ^{67}Cu -21T-BAT-Lym-1 pharmacokinetics, radiation dosimetry, toxicity and tumor regression in patients with lymphoma. *J. Nucl. Med.* **1999**, *40*, 302–310.
152. Pupillo, G.; Sounalet, T.; Michel, N.; Mou, L.; Esposito, J.; Haddad, F. New production cross sections for the theranostic radionuclide ^{67}Cu . *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2018**, *415*, 41–47. [[CrossRef](#)]
153. Katabuchi, T.; Watanabe, S.; Ishioka, N.S.; Iida, Y.; Hanaoka, H.; Endo, K.; Matsuhashi, S. Production of ^{67}Cu via the $^{68}\text{Zn}(p, 2p)^{67}\text{Cu}$ reaction and recovery of ^{68}Zn target. *J. Radioanal. Nucl. Chem.* **2008**, *277*, 467–470. [[CrossRef](#)]
154. Mou, L.; Martini, P.; Pupillo, G.; Cieszykowska, I.; Cutler, C.S.; Mikołajczak, R. ^{67}Cu production capabilities: A mini review. *Molecules* **2022**, *27*, 1501. [[CrossRef](#)] [[PubMed](#)]
155. Hovhannisyanyan, G.H.; Stepanyan, A.V.; Saryan, E.R.; Amirakyan, L.A. Methods of Production the Isotope ^{67}Cu . *J. Contemp. Phys. (Armen. Acad. Sci.)* **2020**, *55*, 183–190. [[CrossRef](#)]

156. Kin, T.; Nagai, Y.; Iwamoto, N.; Minato, F.; Iwamoto, O.; Hatsukawa, Y.; Segawa, M.; Harada, H.; Konno, C.; Ochiai, K.; et al. New production routes for medical isotopes ^{64}Cu and ^{67}Cu using accelerator neutrons. *J. Phys. Soc. Jpn.* **2013**, *82*, 034201. [[CrossRef](#)]
157. Szelecsényi, F.; Steyn, G.F.; Dolley, S.G.; Kovács, Z.; Vermeulen, C.; Van der Walt, T.N. Investigation of the ^{68}Zn (p, 2p) ^{67}Cu nuclear reaction: New measurements up to 40 MeV and compilation up to 100 MeV. *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2009**, *267*, 1877–1881. [[CrossRef](#)]
158. Pandey, M.K.; DeGrado, T.R. Cyclotron production of PET radiometals in liquid targets: Aspects and prospects. *Curr. Radiopharm.* **2021**, *14*, 325–339. [[CrossRef](#)]
159. Deng, X.; Rong, J.; Wang, L.; Vasdev, N.; Zhang, L.; Josephson, L.; Liang, S.H. Chemistry for positron emission tomography: Recent advances in ^{11}C -, ^{18}F -, ^{13}N -, and ^{15}O -labeling reactions. *Angew. Chem. Int. Ed.* **2019**, *58*, 2580–2605. [[CrossRef](#)]
160. McQuade, P.; Rowland, D.J.; Lewis, J.S.; Welch, M.J. Positron-emitting isotopes produced on biomedical cyclotrons. *Curr. Med. Chem.* **2005**, *12*, 807–818. [[CrossRef](#)]
161. Schmitz, J. The production of [^{124}I] iodine and [^{86}Y] yttrium. *Eur. J. Nucl. Med. Mol. Imaging* **2011**, *38*, 4–9. [[CrossRef](#)]
162. van der Meulen, N.P.; Bunka, M.; Domnanich, K.A.; Müller, C.; Haller, S.; Vermeulen, C.; Türler, A.; Schibli, R. Cyclotron production of ^{44}Sc : From bench to bedside. *Nucl. Med. Biol.* **2015**, *42*, 745–751. [[CrossRef](#)]
163. van der Meulen, N.P.; Hasler, R.; Talip, Z.; Grundler, P.V.; Favaretto, C.; Umbricht, C.A.; Müller, C.; Dellepiane, G.; Carzaniga, T.S.; Braccini, S. Developments toward the implementation of ^{44}Sc production at a medical cyclotron. *Molecules* **2020**, *25*, 4706. [[CrossRef](#)]
164. Lagunas-Solar, M.C.; Jungerman, J.A.; Paulson, D.W. Cyclotron production of Thallium-201 via the ^{205}Tl (p, 5n) $^{201}\text{Pb} \rightarrow ^{201}\text{Tl}$ reaction. In Proceedings of the International Symposium on Radiopharmaceuticals, Seattle, WA, USA, 18–23 March 1979; Society of Nuclear Medicine: New York, NY, USA; pp. 779–789.
165. Misiak, R.; Walczak, R.; Waś, B.; Bartyzel, M.; Mietelski, J.W.; Bilewicz, A. ^{47}Sc production development by cyclotron irradiation of ^{48}Ca . *J. Radioanal. Nucl. Chem.* **2017**, *313*, 429–434. [[CrossRef](#)]
166. Abel, E.P.; Domnanich, K.; Clause, H.K.; Kalman, C.; Walker, W.; Shusterman, J.A.; Greene, J.; Gott, M.; Severin, G.W. Production, collection, and purification of ^{47}Ca for the generation of ^{47}Sc through isotope harvesting at the national superconducting cyclotron laboratory. *ACS Omega* **2020**, *5*, 27864–27872. [[CrossRef](#)]
167. Lawrence, J.H. Nuclear physics and therapy: Preliminary report on a new method for the treatment of leukemia and polycythemia. *Radiology* **1940**, *35*, 51–60. [[CrossRef](#)]
168. Hupf, H.B.; Beaver, J.E. Cyclotron production of carrier-free gallium-67. *Int. J. Appl. Radiat. Isot.* **1970**, *21*, 75–76. [[CrossRef](#)]
169. do Carmo, S.J.C.; Scott, P.J.H.; Alves, F. Production of radiometals in liquid targets. *EJNMMI Radiopharm. Chem.* **2020**, *5*, 2. [[CrossRef](#)]
170. Skliarova, H.; Cisternino, S.; Cicoria, G.; Marengo, M.; Cazzola, E.; Gorgoni, G.; Palmieri, V. Medical Cyclotron Solid Target Preparation by Ultrathick Film Magnetron Sputtering Deposition. *Instruments* **2019**, *3*, 21. [[CrossRef](#)]
171. McNeil, B.L.; Robertson, A.K.; Fu, W.; Yang, H.; Hoehr, C.; Ramogida, C.F.; Schaffer, P. Production, purification, and radiolabeling of the $^{203}\text{Pb}/^{212}\text{Pb}$ theranostic pair. *EJNMMI Radiopharm. Chem.* **2021**, *6*, 6. [[CrossRef](#)]
172. Gracheva, N.; Carzaniga, T.S.; Schibli, R.; Braccini, S.; van der Meulen, N.P. ^{165}Er : A new candidate for Auger electron therapy and its possible cyclotron production from natural holmium targets. *Appl. Radiat. Isot.* **2020**, *159*, 109079. [[CrossRef](#)]
173. Nelson, B.J.B.; Wilson, J.; Andersson, J.D.; Wuest, F. High yield cyclotron production of a novel $^{133/135}\text{La}$ theranostic pair for nuclear medicine. *Sci. Rep.* **2020**, *10*, 22203. [[CrossRef](#)] [[PubMed](#)]
174. Dey, M.K.; Gupta, A.D.; Chakrabarti, A. Design of ultra-light superconducting proton cyclotron for production of isotopes for medical applications. *Proc. Cyclotr.* **2013**, *2013*, 447–450.
175. Waites, L.H.; Alonso, J.R.; Conrad, J. IsoDAR: A cyclotron-based neutrino source with applications to medical isotope production. *AIP Conf. Proc.* **2019**, *2160*, 040001.
176. Waites, L.H.; Alonso, J.; Conrad, J.M.; Koser, D.; Winklehner, D. Tools for the Development and Applications of the IsoDAR Cyclotron. *Energy (MeV/Nucl.)* **2021**, *60*, 30.
177. Pramudita, A. Linacs for medical isotope production. In *Proceeding on the scientific meeting and presentation on accelerator technology and its applications: Physics, nuclear reactor, Yogyakarta, Indonesia, 13 December 2011*; National Nuclear Energy Agency: Tangerang, Jawa Barat, Indonesia, 2012; Volume 47, pp. 11–16.
178. Griswold, J.R.; Medvedev, D.G.; Engle, J.W.; Copping, R.; Fitzsimmons, J.; Radchenko, V.; Cooley, J.; Fassbender, M.; Denton, D.; Murphy, K.; et al. Large scale accelerator production of ^{225}Ac : Effective cross sections for 78–192 MeV protons incident on ^{232}Th targets. *Appl. Radiat. Isot.* **2016**, *118*, 366–374. [[CrossRef](#)]
179. Zhuikov, B.L.; Ermolaev, S.V. Radioisotope research and development at the Linear Accelerator of the Institute for Nuclear Research of RAS. *Phys.-Uspekhi* **2021**, *64*, 1311. [[CrossRef](#)]
180. Antipov, K.; Ayzatsky, M.; Akchurin, Y.I.; Boriskin, V.; Beloglasov, V.; Biller, E.; Demidov, N.; Dikiy, N.; Dovbnya, A.; Dovbush, L.; et al. Electron linacs in NSC KIPT: R&D and application. *Вопросы Атомной Науки И Техники* **2001**, *37*, 40–47.
181. Danagulyan, A.S.; Hovhannisyan, G.H.; Bakhshiyani, T.M.; Avagyan, R.H.; Avetisyan, A.E.; Kerobyan, I.A.; Dallakyan, R.K. Formation of medical radioisotopes ^{111}In , $^{117\text{m}}\text{Sn}$, ^{124}Sb , and ^{177}Lu in photonuclear reactions. *Phys. At. Nucl.* **2015**, *78*, 447–452. [[CrossRef](#)]
182. Courtney, W.; Sowder, K.; McGyver, M.; Stevenson, N.; Brown, D. The challenges of commercial isotope production on a linear accelerator. *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2007**, *261*, 739–741. [[CrossRef](#)]

183. Matthews, M.; Saey, P.; Bowyer, T.; Vandergrift, G.; Cutler, N.R.C.; Ponsard, B.; Mikolajczak, R.; Tsipenyuk, Y.; Solin, L.; Fisher, D.; et al. *Workshop on Signatures of Medical and Industrial Isotope Production: A Review*; Pacific Northwest National Laboratory: Richland, Washington, USA, 2010.
184. Dikiy, N.P.; Dovbnya, A.N.; Medvedyeva, E.P.; Tur, Y.D. *Experience of Technetium-99m Generation for Nuclear Medicine on Electron Linac*; VANT: Kharkov, Ukraine, 1997; pp. 165–167.
185. Uvarov, V.L.; Dikiy, N.P.; Dovbnya, A.N.; Medvedyeva, Y.P.; Pugachov, G.D.; Tur, Y.D. Electron Accelerator's Based Production of Technetium-99m for Nuclear Medicine. *Bull. Amer. Phys. Soc.* **1997**, *42*, 1338.
186. De Jong, M. Producing medical isotopes using X-rays. *Sin Proc. IPAC 2012*, *12*, 3177.
187. Chemerisov, S.; Bailey, J.; Heltemes, T.; Jonah, C.; Makarashvili, V.; Tkac, P.; Rotsch, D.; Virgo, M.; Vandegrift, G. *Results of the Six-and-a-Half Day Electron-Accelerator Irradiation of Enriched Mo-100 Targets for the Production of Mo-99*; Argonne National Lab.(ANL): Argonne, IL, USA, 2016.
188. Takeda, T.; Fujiwara, M.; Kurosawa, M.; Takahashi, N.; Tamura, M.; Kawabata, T.; Fujikawa, Y.; Suzuki, K.N.; Abe, N.; Kubota, T.; et al. ^{99m}Tc production via the (γ, n) reaction on natural Mo. *J. Radioanal. Nucl. Chem.* **2018**, *318*, 811–821. [[CrossRef](#)]
189. Szpunar, B.; Rangacharyulu, C.; Date, S.; Ejiri, H. Estimate of production of medical isotopes by photo-neutron reaction at the Canadian light source. *Nucl. Instrum. Methods Phys. Res. Sect. A Accel. Spectrometers Detect. Assoc. Equip.* **2013**, *729*, 41–50. [[CrossRef](#)]
190. Babcock, C.; Goodacre, T.D.; Amani, P.; Au, M.; Bricault, P.; Brownell, M.; Cade, B.; Chen, K.; Egoriti, L.; Johnson, J.; et al. Offline target and ion source studies for TRIUMF-ARIEL. *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2020**, *463*, 464–467. [[CrossRef](#)]
191. Danon, Y.; Block, R.C.; Testa, R.; Testa, R.; Moore, H. Medical isotope production using a 60 MeV linear electron accelerator. *Trans.-Am. Nucl. Soc.* **2008**, *98*, 894.
192. Yagi, M.; Kondo, K. Preparation of carrier-free ^{67}Cu by the $^{68}\text{Zn}(\gamma, p)$ reaction. *Int. J. Appl. Radiat. Isot.* **1978**, *29*, 757–759. [[CrossRef](#)]
193. Hovhannisyanyan, G.H.; Bakhshiyanyan, T.M.; Dallakyan, R.K. Photonuclear production of the medical isotope ^{67}Cu . *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2021**, *498*, 48–51. [[CrossRef](#)]
194. Gopalakrishna, A.; Suryanarayana, S.; Naik, H.; Nayak, B.; Patil, B.; Devraju, S.; Upreti, R.; Kinshikar, R.; Deshpande, D.; Maletha, P.; et al. Production of ^{99}Mo and ^{64}Cu in a mixed field of photons and neutrons in a clinical electron linear accelerator. *J. Radioanal. Nucl. Chem.* **2018**, *317*, 1409–1417. [[CrossRef](#)]
195. Maslov, O.D.; Sabel'nikov, A.V.; Dmitriev, S.N. Preparation of ^{225}Ac by ^{226}Ra (γ, n) photonuclear reaction on an electron accelerator, MT-25 microtron. *Radiochemistry* **2006**, *48*, 195–197. [[CrossRef](#)]
196. Robertson, A.K.H.; Ramogida, C.F.; Schaffer, P.; Radchenko, V. Development of ^{225}Ac radiopharmaceuticals: TRIUMF perspectives and experiences. *Curr. Radiopharm.* **2018**, *11*, 156–172. [[CrossRef](#)]
197. Inagaki, M.; Sekimoto, S.; Tanaka, W.; Tadokoro, T.; Ueno, Y.; Kani, Y.; Ohtsuki, T. Production of ^{47}Sc , ^{67}Cu , ^{68}Ga , ^{105}Rh , ^{177}Lu , and ^{188}Re using electron linear accelerator. *J. Radioanal. Nucl. Chem.* **2019**, *322*, 1703–1709. [[CrossRef](#)]
198. Radel, R.; Sengbusch, E.; Piefer, G. Recent Progress on the PNL Accelerator-Based Intense Fusion Neutron Source. *Trans. Am. Nucl. Soc.* **2016**, *114*, 11–12.
199. Rotsch, D.A.; Brown, M.A.; Nolen, J.A.; Brossard, T.; Henning, W.F.; Chemerisov, S.D.; Gromov, R.G.; Greene, J. Electron linear accelerator production and purification of scandium-47 from titanium dioxide targets. *Appl. Radiat. Isot.* **2018**, *131*, 77–82. [[CrossRef](#)] [[PubMed](#)]
200. Melville, G.; Allen, B.J. Cyclotron and linac production of Ac-225. *Appl. Radiat. Isot.* **2009**, *67*, 549–555. [[CrossRef](#)]
201. Deshpande, A.; Dixit, T.; Bhat, S.; Jadhav, P.; Kottawar, A.; Krishnan, R.; Thakur, K.; Vidwans, M.; Waingankar, A. Design of High Energy Linac for Generation of Isotopes for Medical Applications. In Proceedings of the IPAC 2021-12th International Particle Accelerator Conference, Campinas, SP, Brazil, 24–28 May 2021; JACoW Publishing: Geneva, Switzerland, 2021; pp. 2472–2474.
202. Leung, K.N. New compact neutron generator system for multiple applications. *Nucl. Technol.* **2020**, *206*, 1607–1614. [[CrossRef](#)]
203. Kononov, V.N.; Bokhovko, M.V.; Kononov, O.E.; Soloviev, N.A.; Chu, W.T.; Nigg, D. Accelerator-based fast neutron sources for neutron therapy. *Nucl. Instrum. Methods Phys. Res. Sect. A Accel. Spectrometers Detect. Assoc. Equip.* **2006**, *564*, 525–531. [[CrossRef](#)]
204. Cloth, P.; Conrads, H. Neutronics of a dense-plasma focus—An investigation of a fusion plasma. *Nucl. Sci. Eng.* **1977**, *62*, 591–600. [[CrossRef](#)]
205. Csikai, J.; Dóczy, R. Applications of neutron generators. *Handb. Nucl. Chem.* **2011**, *3*, 363.
206. Reijonen, J. Compact neutron generators for medical, home land security, and planetary exploration. In Proceedings of the 2005 Particle Accelerator Conference, Knoxville, TN, USA, 16–20 May 2005; pp. 49–53.
207. Dovbnya, A.N.; Kuplennikov, E.L.; Tsymba, V.A.; Krasil'nikov, V.V. Possibility of ^{99m}Tc production at neutron generator. *Вопросы Атомной Науки И Техники* **2009**, *5*, 64–66.
208. Pagdon, K.; Gentile, C.; Cohen, A.; Ascione, G.; Baker, G. Production of Tc-99m from naturally occurring molybdenum absent uranium. In Proceedings of the 2011 IEEE/NPSS 24th Symposium on Fusion Engineering, Chicago, IL, USA, 26–30 June 2011; pp. 1–4.
209. Mausolf, E.J.; Johnstone, E.V.; Mayordomo, N.; Williams, D.L.; Guan, E.Y.Z.; Gary, C.K. Fusion-Based Neutron Generator Production of Tc-99m and Tc-101: A Prospective Avenue to Technetium Theranostics. *Pharmaceuticals* **2021**, *14*, 875. [[CrossRef](#)]

210. National Academies of Sciences, Engineering, and Medicine. *Molybdenum-99 for Medical Imaging*; National Academies Press: Washington, DC, USA, 2016.
211. Youker, A.J.; Chemerisov, S.D.; Tkac, P.; Kalensky, M.; Heltemes, T.A.; Rotsch, D.A.; Vandegrift, G.F.; Krebs, J.F.; Makarashvili, V.; Stepinski, D.C. Fission-produced Mo-99 without a nuclear reactor. *J. Nucl. Med.* **2017**, *58*, 514–517. [[CrossRef](#)]
212. Leung, K.N.; Leung, J.K.; Melville, G. Feasibility study on medical isotope production using a compact neutron generator. *Appl. Radiat. Isot.* **2018**, *137*, 23–27. [[CrossRef](#)] [[PubMed](#)]
213. Badwar, S.; Ghosh, R.; Lawriniang, B.M.; Vansola, V.; Sheela, Y.; Naik, H.; Naik, Y.; Suryanarayana, S.V.; Jyrwa, B.; Ganesan, S. Measurement of formation cross-section of ^{99}Mo from the ^{98}Mo (n, γ) and ^{100}Mo (n, 2n) reactions. *Appl. Radiat. Isot.* **2017**, *129*, 117–123. [[CrossRef](#)] [[PubMed](#)]
214. Auditore, L.; Amato, E.; Baldari, S. Theoretical estimation of ^{64}Cu production with neutrons emitted during ^{18}F production with a 30 MeV medical cyclotron. *Appl. Radiat. Isot.* **2017**, *122*, 229–234. [[CrossRef](#)] [[PubMed](#)]
215. Pandit-Taskar, N.; Batraki, M.; Divgi, C.R. Radiopharmaceutical therapy for palliation of bone pain from osseous metastases. *J. Nucl. Med.* **2004**, *45*, 1358–1365.
216. Kim, S.K.; Choi, H.D. New technique for Producing Therapeutic Radioisotope ^{89}Sr . In Proceedings of the Korean Nuclear Society Conference, jeju, Korea, 26–26 May 2005; Korean Nuclear Society: Seoul, Korea; pp. 751–752.
217. Molla, N.I.; Basunia, S.; Miah, M.R.; Hossain, S.M.; Rahman, M.M.; Spellerberg, S.; Qaim, S.M. Radiochemical Study of ^{45}Sc (n, p) ^{45}Ca and ^{89}Y (n, p) ^{89}Sr Reactions in the Neutron Energy Range of 13.9 to 14.7 MeV. *Radichim. Acta* **1998**, *80*, 189–192. [[CrossRef](#)]
218. Capogni, M.; Capone, M.; Pietropaolo, A.; Fazio, A.; Dellepiane, G.; Falconi, R.; Colangeli, A.; Palomba, S.; Valentini, G.; Fantuzzi, M.; et al. ^{64}Cu production by 14 MeV neutron beam. *J. Neutron Res.* **2020**, *22*, 257–264. [[CrossRef](#)]
219. Voyles, A.; Basunia, M.; Batchelder, J.; Bauer, J.; Becker, T.; Bernstein, L.; Matthews, E.; Renne, P.; Rutte, D.; Unzueta, M.; et al. Measurement of the ^{64}Zn , ^{47}Ti (n, p) cross sections using a DD neutron generator for medical isotope studies. *Nucl. Instrum. Methods Phys. Res. Sect. B Beam Interact. Mater. At.* **2017**, *410*, 230–239. [[CrossRef](#)]
220. Mellard, S.C.; Biegalski, S.R. MCNP based simulations for the optimization of radioxenon via DD and DT neutron generators from ^{132}Xe . *J. Radioanal. Nucl. Chem.* **2018**, *318*, 313–322. [[CrossRef](#)]
221. Weicheng, Z. Short-lived medical isotopes produced by 14MeV neutron generator. *At. Energy Sci. Technol* **1981**, *03*, 366–368. (In Chinese)
222. Yuntao, L.; Shizhong, A.; Jixin, L. Current Situation and Development Trend of Nuclear Technology Application. *Sci. Technol. Rev.* **2022**, *40*, 88–97. (In Chinese)
223. Huang, Z.; Wang, J.; Ma, Z.; Lu, X.; Wei, Z.; Zhang, S.; Liu, Y.; Zhang, Z.; Zhang, Y.; Yao, Z. Design of a compact D–D neutron generator. *Nucl. Instrum. Methods Phys. Res. Sect. A Accel. Spectrometers Detect. Assoc. Equip.* **2018**, *904*, 107–112. [[CrossRef](#)]
224. Jixin, L.; Yuqing, C.; Guang, L.; Xuesong, D.; Yijia, S.; Laicheng, Q.; Yuping, L.; Hua, J.; Guiqun, L. Production process of ^{64}Cu by C-30 cyclotron. In *Annual Report for China Institute of Atomic Energy*; China Institute of Atomic Energy: Beijing, China, 2013.