

## Supporting Information

### Synthesis, Quantification, and Characterization of Fatty Acid Amides from *In Vitro* and *In Vivo* Sources

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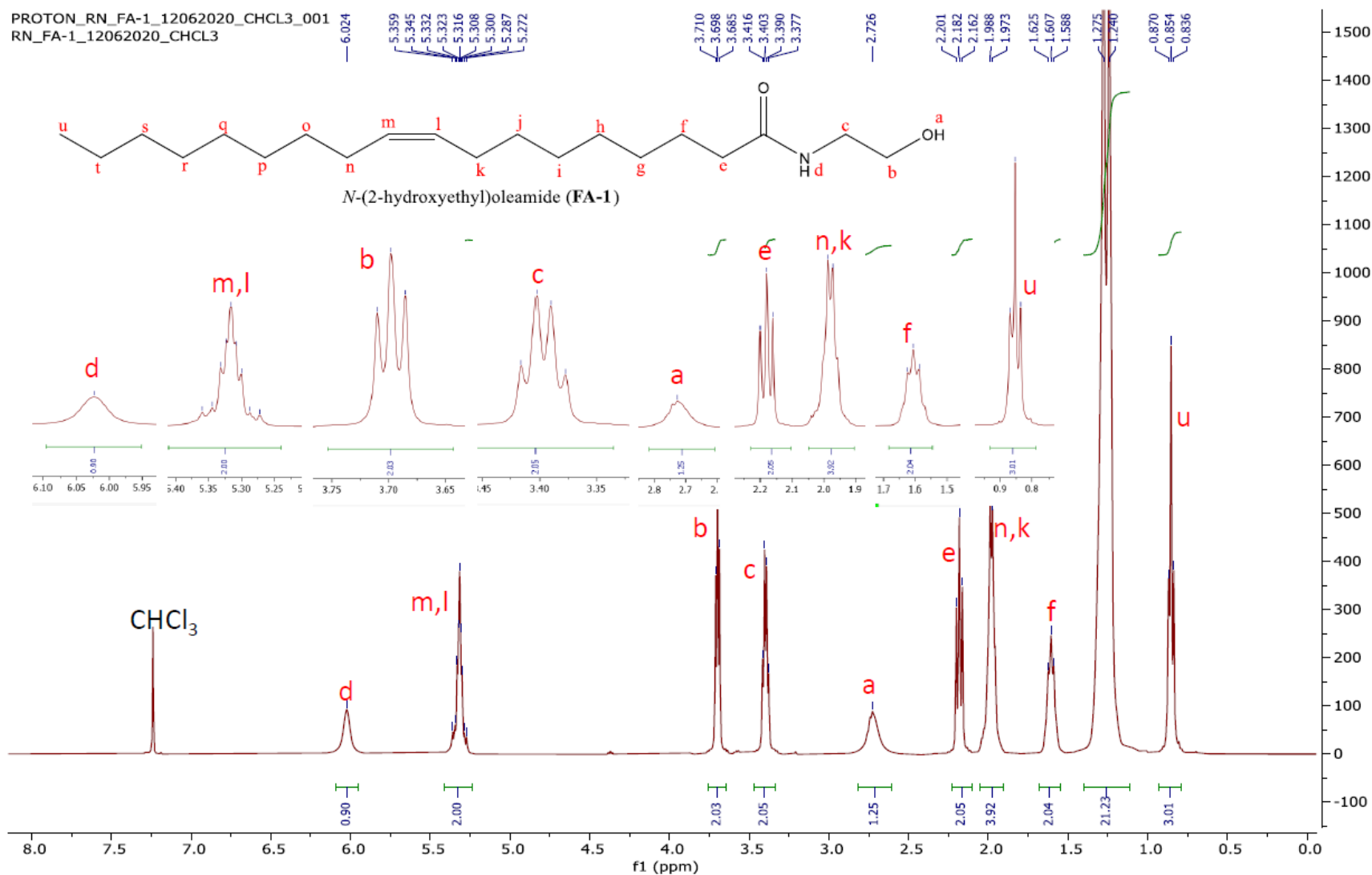
**Table S1.**  
**Identification and Quantification of Fatty Acid Amides**  
**Produced by *Drosophila melanogaster* Larvae by LC/QTOF-MS<sup>1</sup>**

Fatty Acid Amide	Standard		Endogenousfrom Larvae		EndogenousAmount (pmoles/gram of tissue)
	[M+H] <sup>+</sup> <i>m/z</i>	RT <sup>2</sup> (min)	[M+H] <sup>+</sup> <i>m/z</i>	RT <sup>2</sup> (min)	
<b><i>N</i>-Acylglycines</b>					
<i>N</i> -Palmitoylglycine	314.2635	5.968	314.2524	5.940	19.5 ± 7.0
<i>N</i> -Oleoylglycine	340.2785	6.067	340.2666	5.874	24.8 ± 12.0
<i>N</i> -Linoleoylglycine	338.2634	5.950	338.2399	5.794	12.1 ± 3.7
<i>N</i> -Archidonoylglycine	362.2630	5.800	362.2531	5.934	27.1 ± 14.7
<b><i>N</i>-Acylethanolamines</b>					
<i>N</i> -Oleylethanolamine	326.2998	6.051	326.2781	6.007	1.3 ± 1.5
Anandamide	348.2840	5.768	348.2593	5.641	1.5 ± 0.8
<b><i>N</i>-Acyldopamines<sup>3</sup></b>					
<i>N</i> -Palmitoyldopamine	392.3089	6.117	392.3356	6.237	3.8 ± 2.5
<i>N</i> -Oleoyldopamine	418.3243	6.196	418.3263	6.134	1.9 ± 0.5
<i>N</i> -Archidonoylglycine	440.3271	5.934	440.3624	5.990	2.8 ± 0.6
<b><i>N</i>-Acylserotonins</b>					
<i>N</i> -Palmitoylserotonin	415.3245	6.159	415.3047	6.123	2.2 ± 0.9
<i>N</i> -Oleoylserotonin	441.3397	6.234	441.3072	6.217	1.3 ± 0.3
<i>N</i> -Archidonoylserotonin	463.3319	5.976	463.3492	6.083	0.8 ± 0.4
<b>Primary Fatty Acid Amides</b>					
Palmitamide	256.2587	6.176	256.2498	6.148	20.4 ± 14.9
Palmitoleamide	254.2434	5.830	254.2322	5.805	19.2 ± 5.3
Oleamide	282.2741	6.250	282.2706	6.195	50.0 ± 31.3
Linoleamide	280.2585	5.950	280.2523	5.965	5.1 ± 4.2

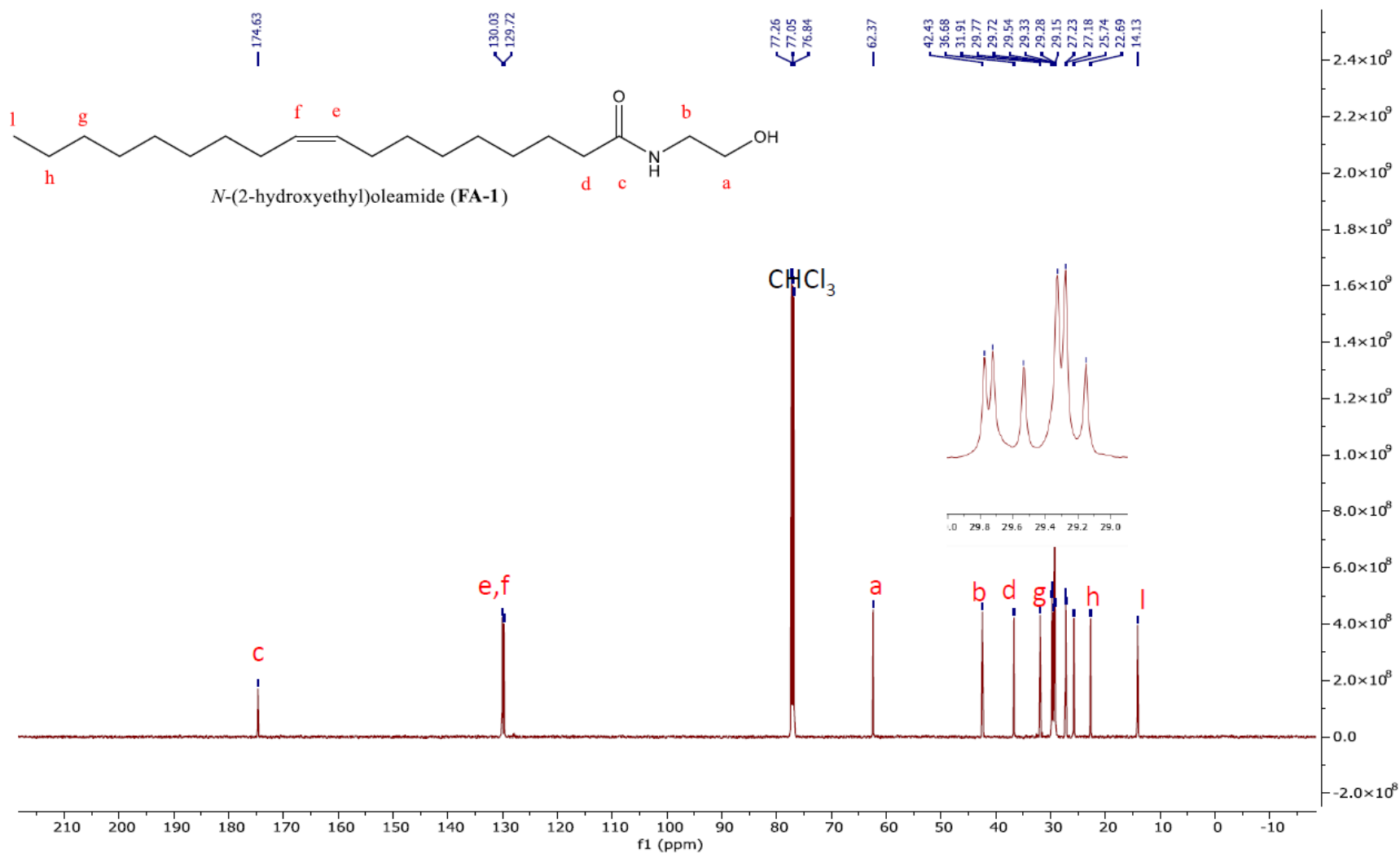
<sup>1</sup>The fatty acid amides were extracted, analyzed, and quantified as described by Jeffries *et al.* [20].

<sup>2</sup>RT is retention time

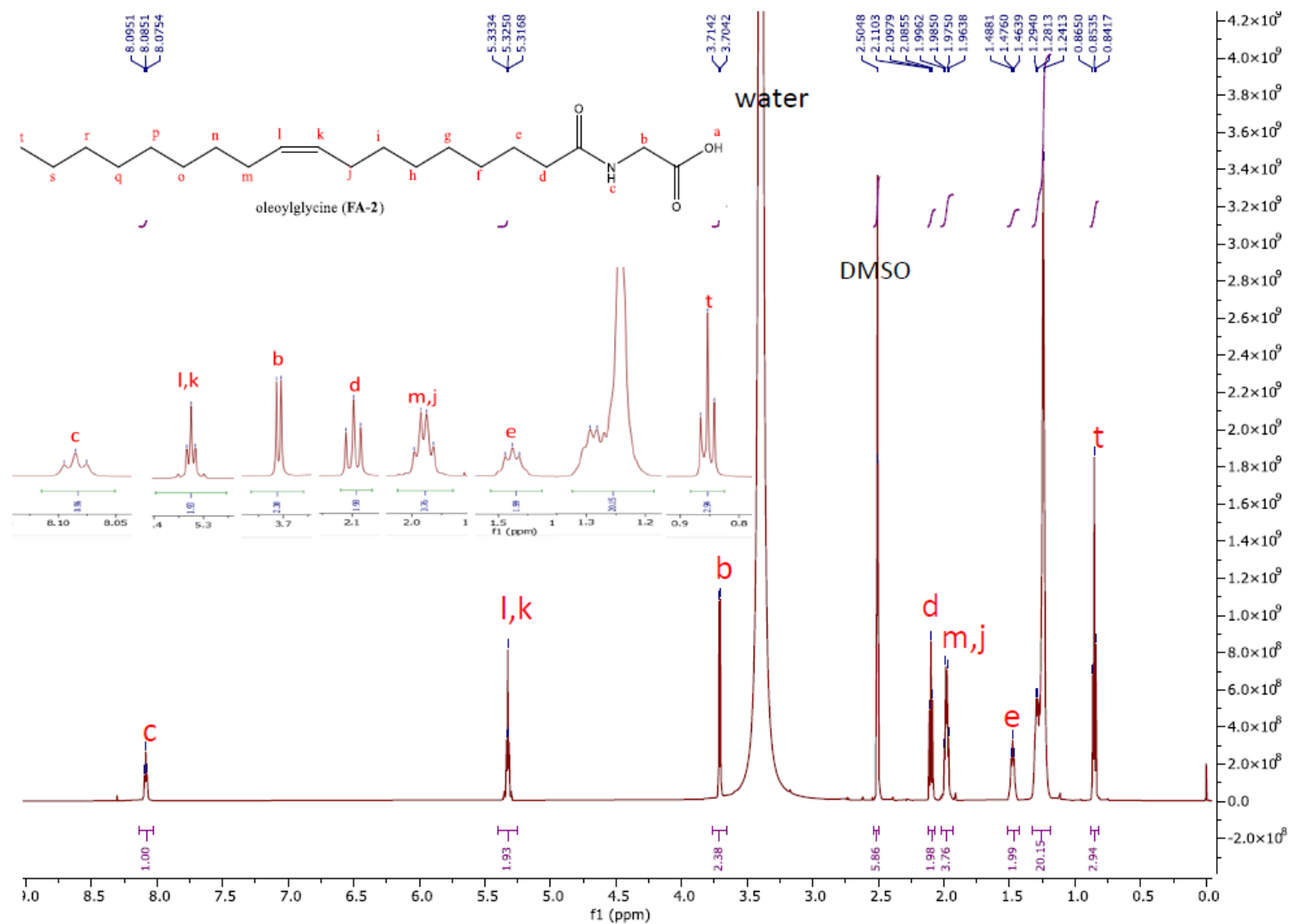
<sup>3</sup>Identified and quantified as the *N*-acyldopamine quinone.



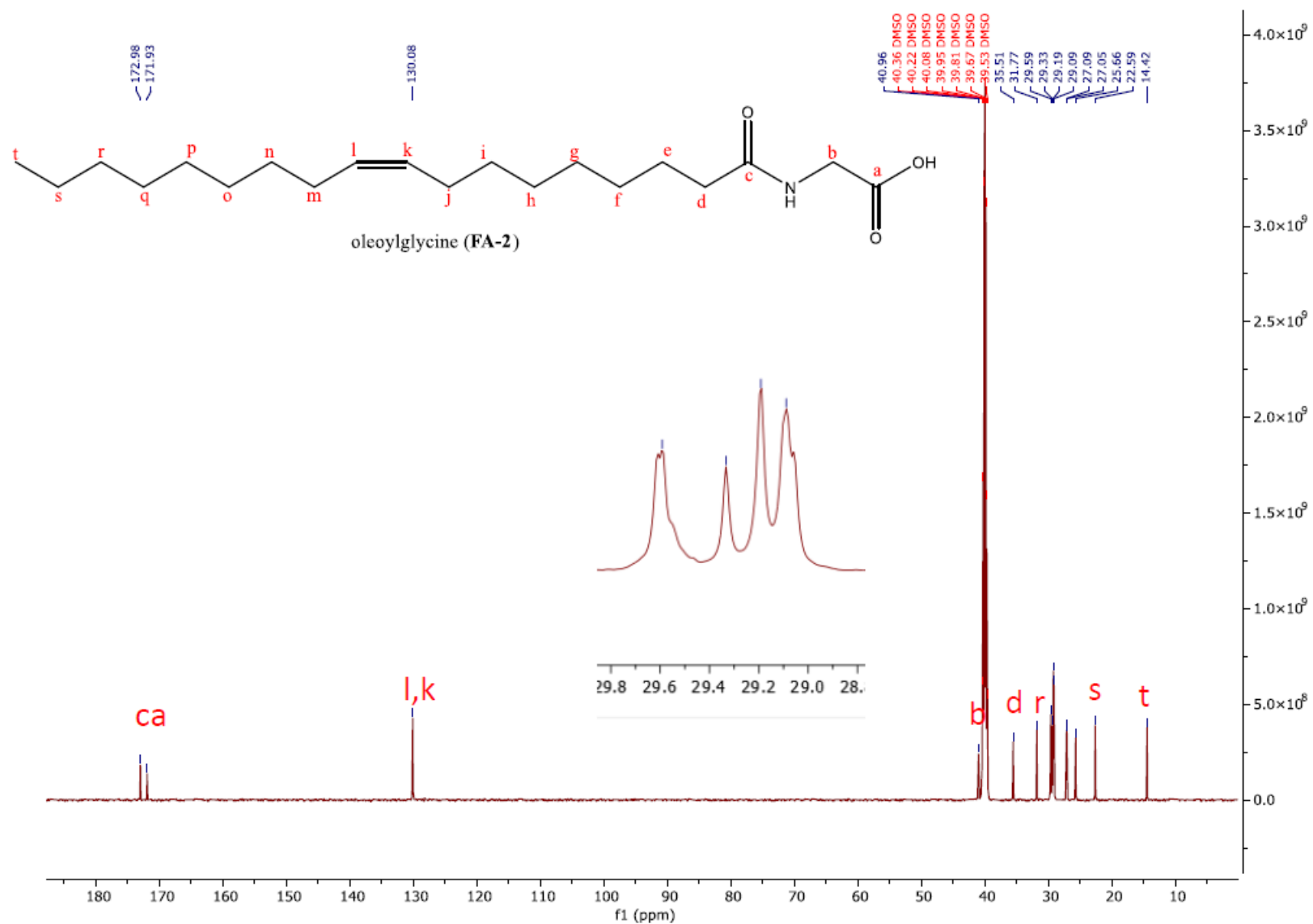
**Figure S1:** <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of FA-1, the inset shows multiplicity of the signals.



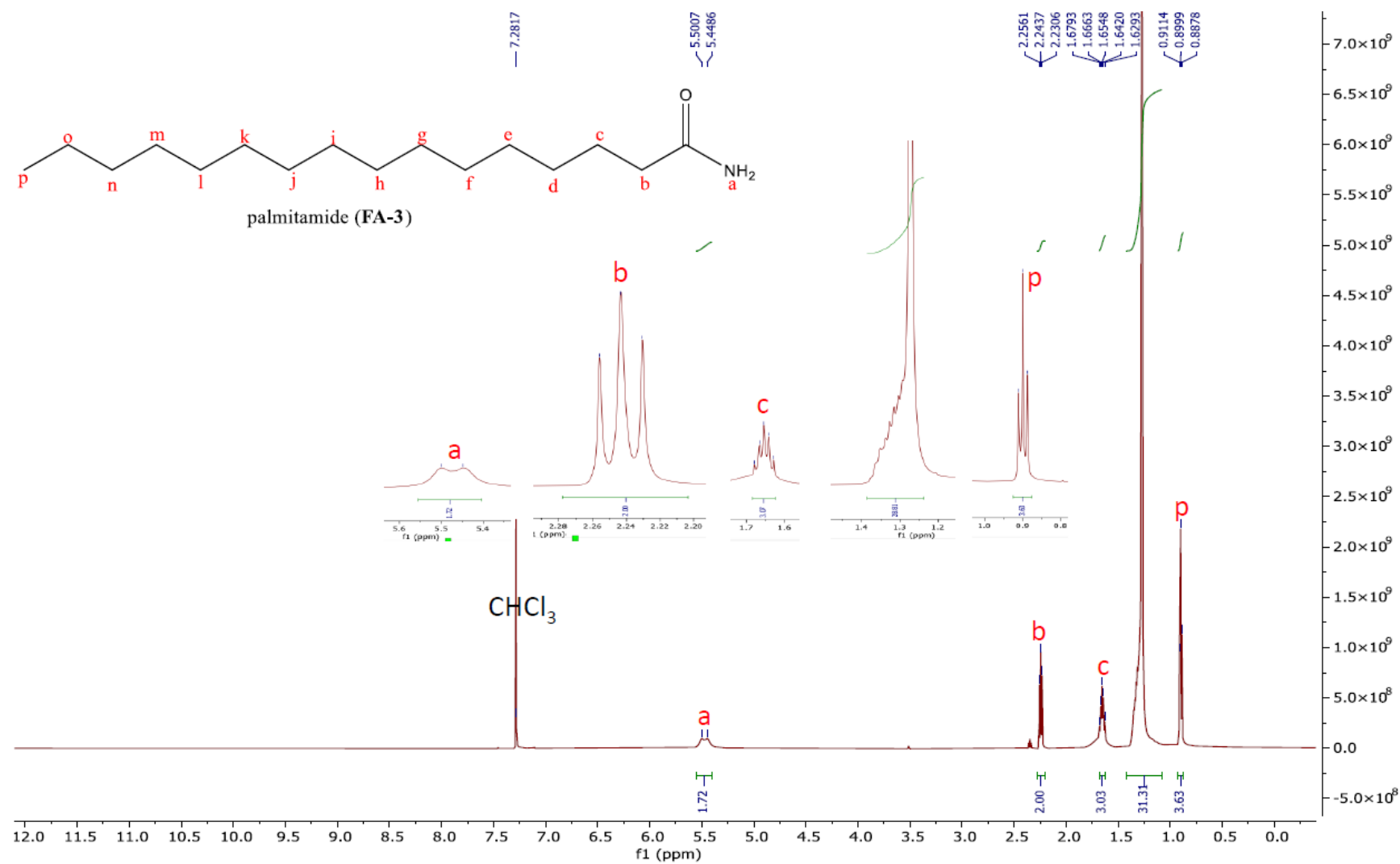
**Figure S2:** <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) of **FA-1**, the inset shows clustered signals.



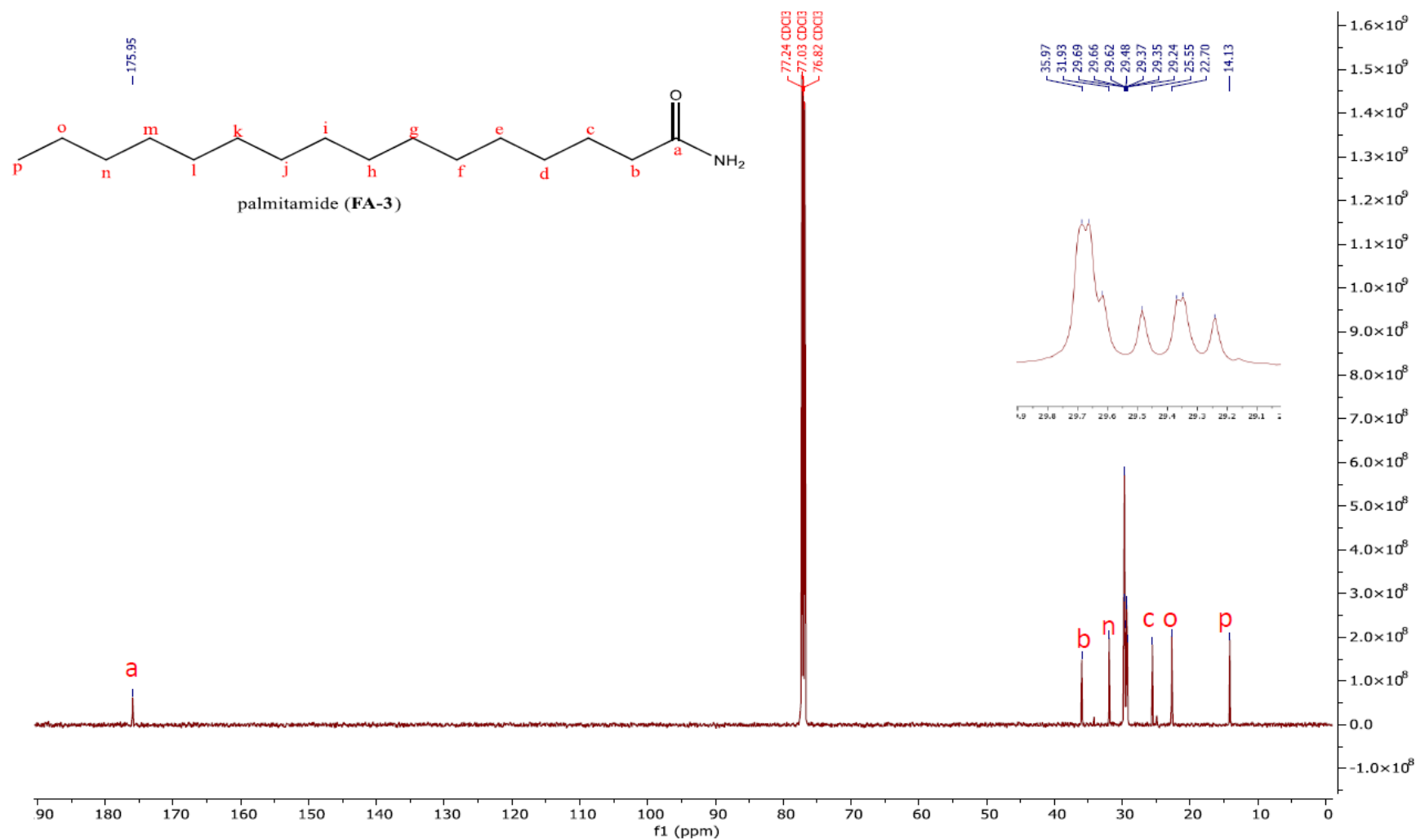
**Figure S3:**  $^1\text{H}$  NMR (600 MHz, DMSO) of **FA-2**, the inset shows multiplicity of the signals.



**Figure S4:**  $^{13}\text{C}$  NMR (151 MHz, DMSO) of **FA-2**, the inset shows clustered signals.

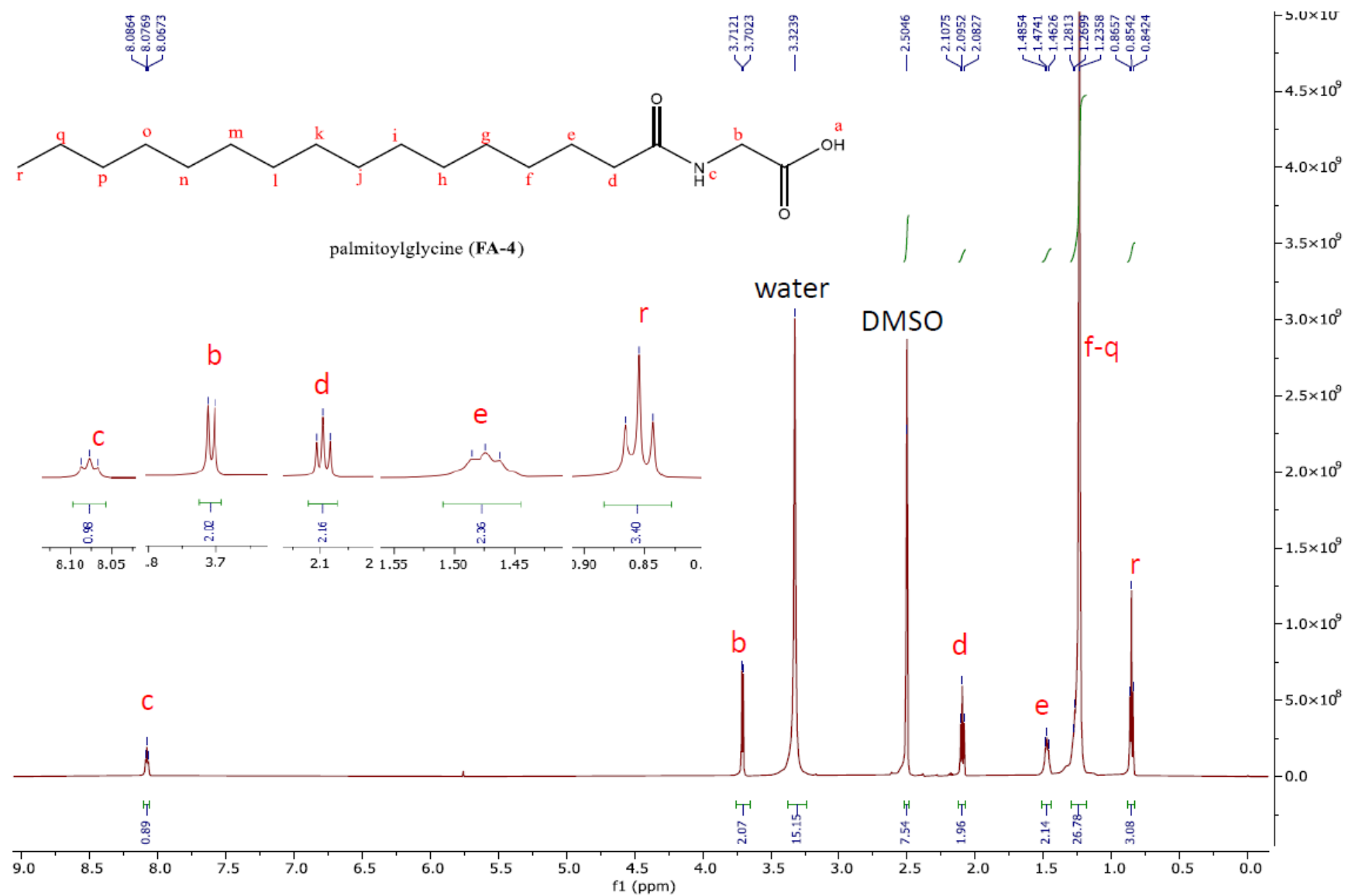


**Figure S5:** <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>) of **FA-3**, the inset shows multiplicity of the signals.

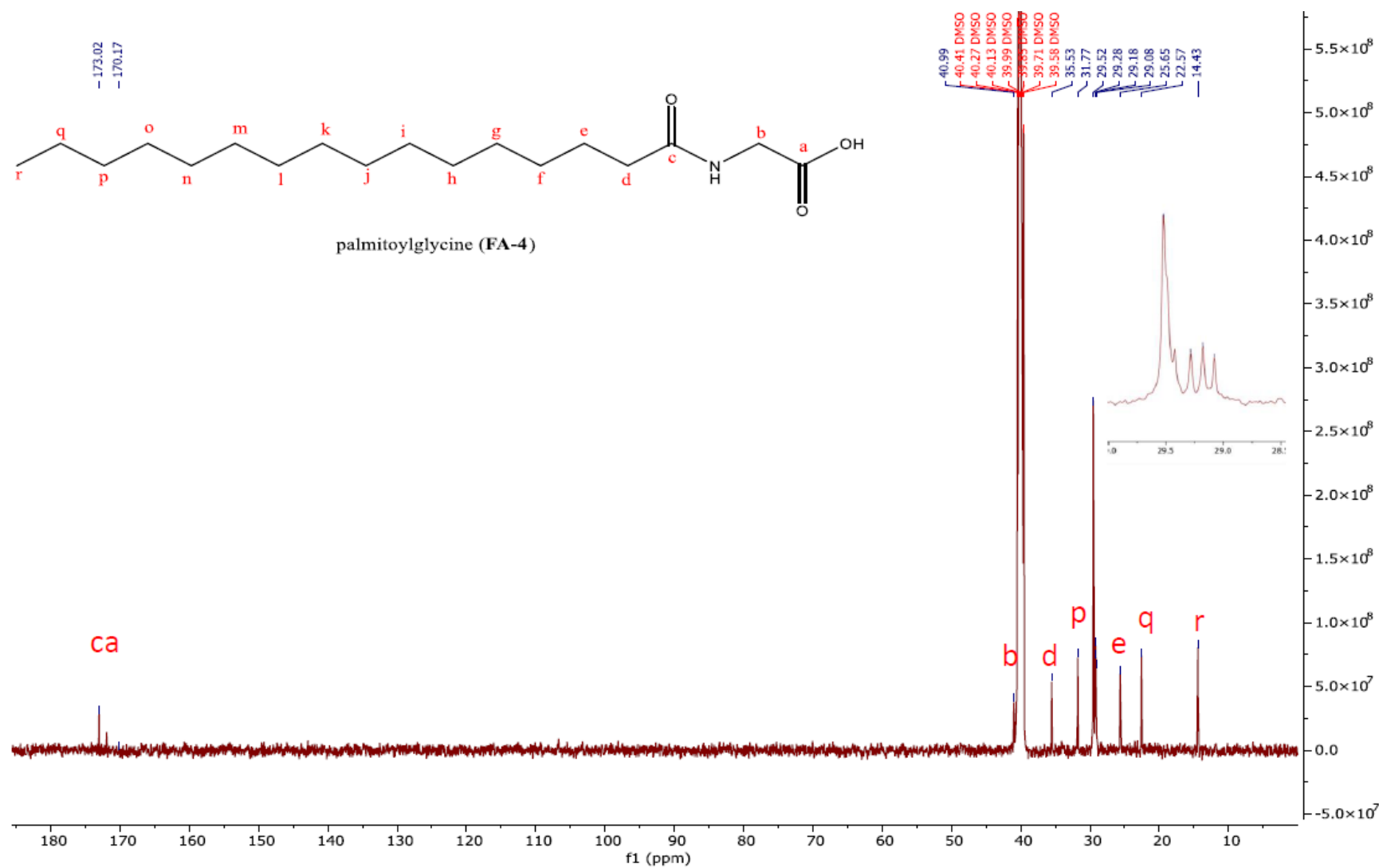


**Figure S6:**  $^{13}\text{C}$  NMR (151 MHz,  $\text{CDCl}_3$ ) of **FA-3**, the inset shows clustered signals.

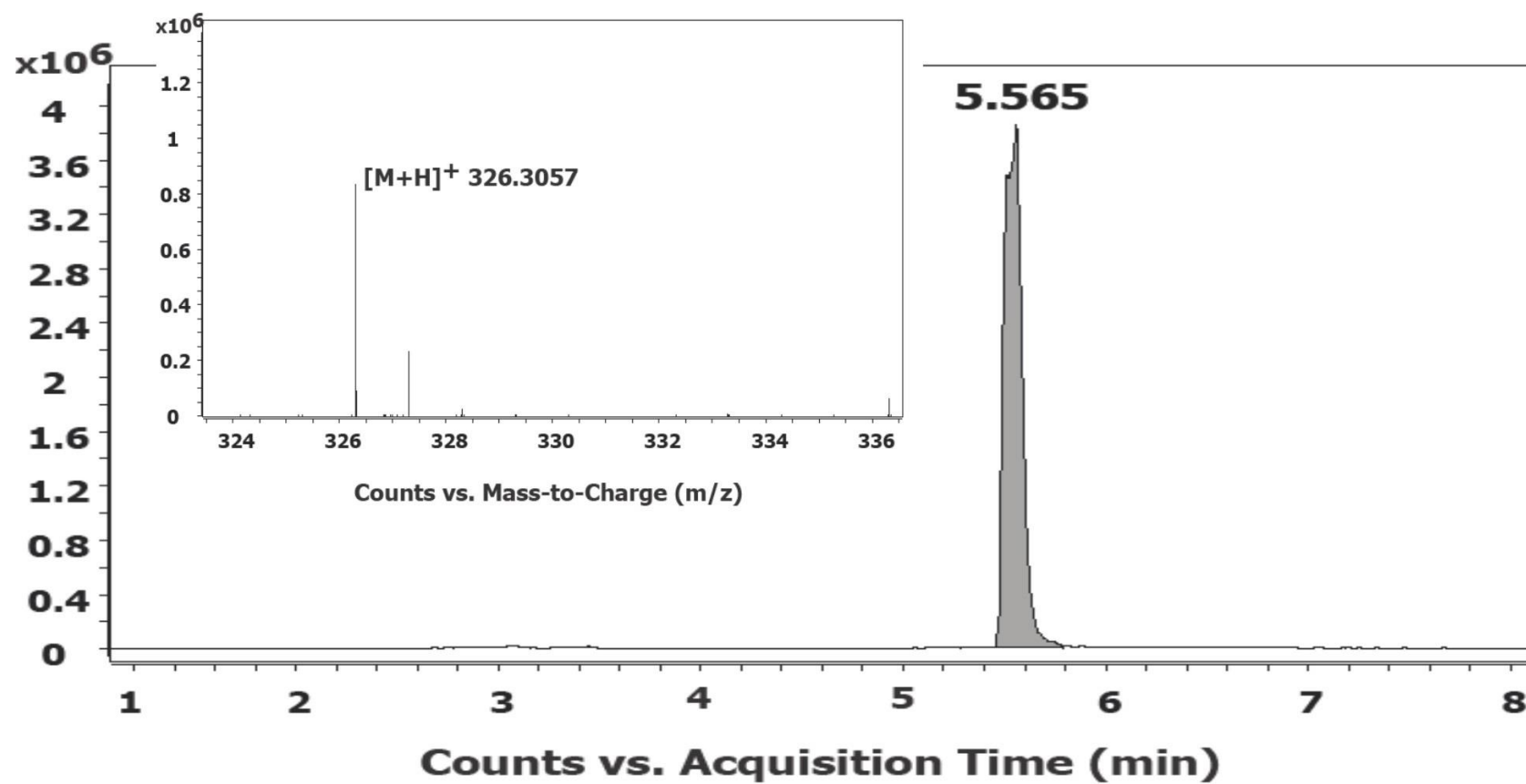




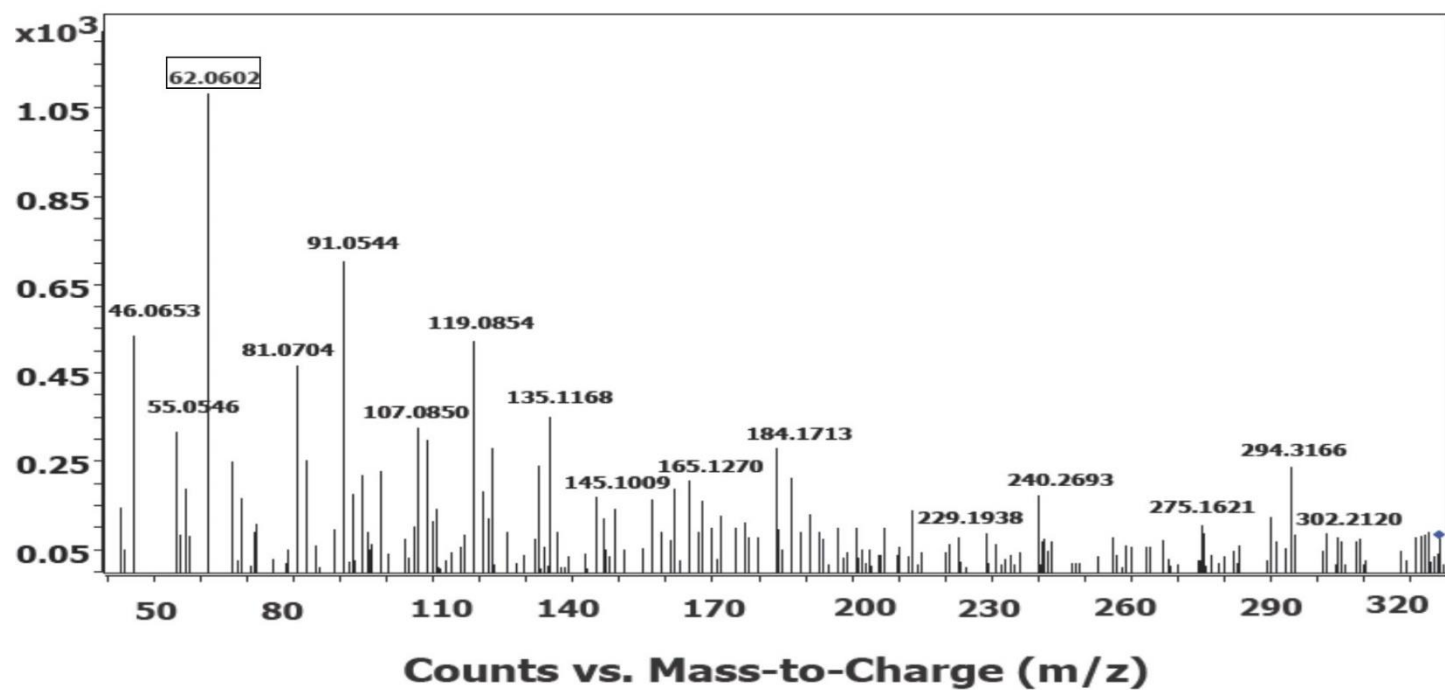
**Figure S7:**  $^1\text{H}$  NMR (600 MHz, DMSO) of **FA-4**, the inset shows multiplicity of the signals.



**Figure S8:**  $^{13}\text{C}$  NMR (151 MHz, DMSO) of **FA-4**, the inset shows clustered signals.



**Figure S9:** The EIC peak and mass spectrum of the *N*-oleoylethanolamine standard



**Figure S10:** Targeted MS/MS spectra of endogenous *N*-oleoylethanolamine in the *T. castaneum* extract at 20 eV collision energy displaying expected primary fragment ion, m/z 62.0602.