

Review

Chemical Diversity and Biological Activity of Secondary Metabolites Isolated from Indonesian Marine Invertebrates

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Abstract: Marine invertebrates have been reported to be an excellent resource of many novel bioactive compounds. Studies reported that Indonesia has remarkable yet underexplored marine natural products, with a high chemical diversity and a broad spectrum of biological activities. This review discusses recent updates on the exploration of marine natural products from Indonesian marine invertebrates (i.e., sponges, tunicates, and soft corals) throughout 2007–2020. This paper summarizes the structural diversity and biological function of the bioactive compounds isolated from Indonesian marine invertebrates as antimicrobial, antifungal, anticancer, and antiviral, while also presenting the opportunity for further investigation of novel compounds derived from Indonesian marine invertebrates.

Keywords: Indonesia; marine natural product; marine invertebrates; soft corals; sponges; tunicates; biodiversity; biological activity



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1. Introduction

A wide range of natural products (NPs) has been isolated from various marine organisms, especially marine invertebrates such as sponges, tunicates, soft corals, bryozoans, and nudibranchs. These marine invertebrates are excellent sources of NPs with vast chemical structures and potential biological activities [1–3]. During 2012–2017, no less than 550–700 new compounds have been reported from marine invertebrates [4], in which half of these compounds were isolated from marine sponges [5]. Among those, 4% and 22% of the compounds were identified in 2017 and 2016, respectively [4,5]. Between 1998 to 2018, one hundred and fourteen secondary metabolites were isolated from the marine sponges of the genus *Suberea* [6]. Meanwhile, a hundred and seventy compounds were isolated from soft corals of the genus *Dendronephthya* alone throughout 1999–2019 [7]. Soft corals belonging to the genus *Xenia* are rich in terpenoids, with 199 compounds isolated from 1977–2019 [8]. To date, approximately 30,000–40,000 marine natural products (MNPs) have been identified, with the majority of the compounds exhibiting cytotoxic and anticancer properties [4,5,9].

The biological potential of MNPs from marine invertebrates has been proven to be a valuable source for drug discovery and development. Most of the approved commercial marine-based drugs are of marine invertebrate origin. Eight marine drugs have been approved by the US Food and Drug Administration (FDA) and European Medicines Agency (EMA). The first approved marine drug is Ziconotide (Prialt®) discovered in the marine snail *Conus magus*. This peptide-derived marine drug is commonly used as an analgesic drug for the management of severe chronic pain through the intrathecal route. The second one is Omega-3-acid-ethyl esters (Lovaza®) derived from fish oil and used as an anti-hypertriglyceridemia drug. The third is Vidarabine (Vira-A®) derived from the

sponge *Cryptotethya crypta*, while the fourth is Iota-carrageenan (Carragelose[®]), derived from red macroalgae. These two were registered as antivirals. The other four drugs were approved for cancer treatment, i.e., (1) *viz.* Cytarabine (Cytosar-U[®] and Depocyt[®]) discovered in sponge *Cryptotethya crypta*, (2) Trabectedin (Yondelis[®]) isolated from the tunicate *Ecteinascidia turbinata*, (3) Eribulin mesylate (Halaven[®]) discovered from sponge *Halichondria okadai*, and (4) Brentuximab vedotin (Adcetris[®]) derived from the sea hare *Dolabella auricularia*. Additionally, approximately 30 MNPs were reported in different clinical trial stages, mainly derived from marine invertebrates [9–11].

Indonesia is the world's largest archipelagic country with 17,500 islands and a long coastline of 81,000 km. This extraordinary geographic attribute offers a highly diverse variety of marine organisms, resulting in Indonesia being known as a mega-biodiversity of marine organisms. Like other living organisms, marine species synthesize metabolites, either primary or secondary, to support their lives. Living in an extreme environment often induces these organisms to synthesize multiple secondary metabolites with unique chemical properties. A part of their defense mechanism, these metabolites have also been reported to have diverse biological activities that are important for drug discovery and development [12].

The study of Indonesian MNPs was started in 1972 by Engelbrecht et al., who discovered a compound called 25-hydroxy-24 ξ -methylcholesterol derived from a soft coral collected from Nias Island, Indonesia [13]. Following that, Cornery et al. reported two novel cytotoxic compounds from the Indonesian sponge *Hyatella* sp. called *viz.* laulimalide and isolaulimalide, which were active against the KB cell line with an IC₅₀ value of 15 ng/mL [14]. Since then, research on Indonesian MNPs has expanded significantly. From the 1970s to the year 2017, about 732 MNPs were isolated from Indonesian sea waters. They were mainly produced by sponges (Porifera), tunicates (Chordata), and soft corals (Cnidaria) [15]. In the past decade, hundreds of novel compounds have been discovered from Indonesian marine organisms, many of which showing potent biological activity [16,17].

This review presents recent updates on Indonesian MNPs isolated from three marine invertebrates (sponges, tunicates, and soft corals), reported from 2007 to 2020, covering the chemical diversity and biological activity.

2. Marine Invertebrates

2.1. Sponges

Among sponges, alkaloids were reported as the most isolated bioactive compounds, followed by terpenoids, peptides, and polyketides (Table 1). Among the isolated alkaloids from sponges, manzamines are mostly reported to exhibit a broad spectrum of biological activities such as cytotoxic, antimicrobial, antimalarial, antiviral, anti-inflammatory, anti-atherosclerotic, insecticidal, and proteasome inhibitor [18]. Their structure has a fused tetra- or pentacyclic ring attached to a carboline moiety.

Table 1. Summary of the marine natural products isolated from the marine sponges from Indonesian oceans.

Compound	Compound Class	Species	Biological Activity	Ref
Acanthomanzamine A (1)	Alkaloid	<i>Acanthostrongylophora ingens</i>	Cytotoxic against human cervical HeLa cells; inhibition of proteasome; cholesterol ester accumulation inhibitor	[18]
Acanthomanzamine B (2)	Alkaloid	<i>A. ingens</i>	Cytotoxic against human cervical HeLa cells; inhibition of proteasome; cholesterol ester accumulation inhibitor	[18]

Table 1. Cont.

Compound	Compound Class	Species	Biological Activity	Ref
Acanthomanzamine C (3)	Alkaloid	<i>A. ingens</i>	n.a.	[18]
Acanthomanzamine D (4)	Alkaloid	<i>A. ingens</i>	Cytotoxic against human cervical HeLa cells; inhibition of proteasome; cholesterol ester accumulation inhibitor	[18]
Acanthomanzamine E (5)	Alkaloid	<i>A. ingens</i>	Cytotoxic against human cervical HeLa cells; inhibition of proteasome; cholesterol ester accumulation inhibitor	[18]
Acantholactam (6)	Alkaloid	<i>A. ingens</i>	Inhibition of proteasome	[19]
Pre- <i>neo</i> -kauluamine (7)	Alkaloid	<i>A. ingens</i>	Inhibition of proteasome	[19]
Acanthocyclamine A (8)	Alkaloid	<i>A. ingens</i>	Antimicrobial against <i>E. coli</i> ; inhibitor of amyloid β -42 production	[20,21]
<i>Epi</i> -tetrahydrohalicyclamine B (9)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Tetrahydrohalicyclamine B (10)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Halicyclamine B (11)	Alkaloid	<i>A. ingens</i>	Antimicrobial against <i>S. aureus</i>	[21]
Chloromethylhalicyclamine B (12)	Alkaloid	<i>A. ingens</i>	Protein kinase CK1 δ/ϵ inhibitor	[21]
Cyclo (D-Pro-L-Phe) (13)	Alkaloid	<i>A. ingens</i>	Protein kinase CDK2/cyclin A inhibitor	[21]
Cyclo (L-Pro-Gly) (14)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Cyclo (L-Pro-L-Ala) (15)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Cyclo (D-Pro-L-Val) (16)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Cyclo (L-Pro-Ser) (17)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Cyclo (D-Pro-L-Ile) (18)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Cyclo (L-Pro-L-Tyr) (19)	Alkaloid	<i>A. ingens</i>	n.a.	[21]
Ingenine C (20)	Alkaloid	<i>A. ingens</i>	Cytotoxic against human breast MCF-7 and colorectal HCT116 cancer cells	[22]
Ingenine D (21)	Alkaloid	<i>A. ingens</i>	Cytotoxic against human breast MCF-7 and colorectal HCT116 cancer cells	[22]
Dispacamide E (22)	Alkaloid	<i>Stylissa massa</i>	Protein kinase inhibitor (GSK-3, DYRK1A, and CK-1)	[23]
Ethyl 3,4-dibromo-1 <i>H</i> -pyrrole-2-carboxylate (23)	Alkaloid	<i>S. massa</i>	n.a.	[23]
12- <i>N</i> -methyl stevensine (24)	Alkaloid	<i>Stylissa</i> sp.	Cytotoxic against mouse lymphoma L5187Y cancer cell line	[24]
12- <i>N</i> -methyl-2-debromostevensine (25)	Alkaloid	<i>Stylissa</i> sp.	n.a.	[24]
3-debromolatondaine B methyl ester (26)	Alkaloid	<i>Stylissa</i> sp.	n.a.	[24]
3-debromolatondaine A (27)	Alkaloid	<i>Stylissa</i> sp.	n.a.	[24]
Crambescidin 345 (28)	Alkaloid	<i>Clathria bulbotoxa</i>	Cytotoxic against the human epidermal A431 carcinoma cell line	[25]
Crambescidin 361 (29)	Alkaloid	<i>C. bulbotoxa</i>	Cytotoxic against the human epidermal A431 carcinoma cell line	[25]

Table 1. Cont.

Compound	Compound Class	Species	Biological Activity	Ref
Crambescidin 373 (30)	Alkaloid	<i>C. bulbotoxa</i>	Cytotoxic against the human epidermal A431 carcinoma cell line	[25]
Methyldorimidazole (31)	Alkaloid	<i>Leucetta chagosensis</i>	n.a.	[26]
Preclathridine B (32)	Alkaloid	<i>L. chagosensis</i>	n.a.	[26]
Naamidine H (33)	Alkaloid	<i>L. chagosensis</i>	Cytotoxic against human cervical HeLa cells	[27,28]
Naamidine I (34)	Alkaloid	<i>L. chagosensis</i>	Cytotoxic against human cervical HeLa cells	[27]
Spironaamidine (35)	Alkaloid	<i>Leucetta microraphis</i>	Antibacterial activity against <i>B. cereus</i>	[28]
Variabine A (36)	Alkaloid	<i>Luffariella variabilis</i>	n.a.	[29]
Variabine B (37)	Alkaloid	<i>L. variabilis</i>	Inhibition of proteasome and Ubc13 (E2)–Uev1A interaction	[29]
Sagitol C (38)	Alkaloid	<i>Oceanapia</i> sp.	Cytotoxic against mouse lymphoma L5187Y, human cervical HeLa, and rat pheochromocytoma PC12 cells	[30]
Cortistatin E (39)	Alkaloid	<i>Corticium complex</i>	n.a.	[31]
Cortistatin F (40)	Alkaloid	<i>C. complex</i>	n.a.	[31]
Cortistatin G (41)	Alkaloid	<i>C. complex</i>	n.a.	[31]
Cortistatin H (42)	Alkaloid	<i>C. complex</i>	n.a.	[31]
Cortistatin J (43)	Alkaloid	<i>C. complex</i>	Cytostatic antiproliferative activity against human umbilical vein endothelial cells (HUVECs)	[32]
Cortistatin K (44)	Alkaloid	<i>C. complex</i>	n.a.	[32]
Cortistatin L (45)	Alkaloid	<i>C. complex</i>	n.a.	[32]
11-Methoxy-3H-[1,6]naphthyridino[6,5,4-def]quinoxalin-3-one (46)	Alkaloid	<i>Aptos suberitoides</i>	n.a.	[33]
2,11-Dimethoxy-3H-[1,6]naphthyridino[6,5,4-def]quinoxalin-3-one (47)	Alkaloid	<i>A. suberitoides</i>	n.a.	[33]
5-Benzoydemethylaaptamine (48)	Alkaloid	<i>A. suberitoides</i>	Cytotoxic against mouse lymphoma L5187Y cancer cell line	[33]
3-Aminodemethyl(oxy)aaptamine (49)	Alkaloid	<i>A. suberitoides</i>	n.a.	[33]
2-methoxy-3-oxoaaptamine (50)	Alkaloid	<i>Aptos</i> sp.	Antibacterial against <i>Mycobacterium smegmatis</i>	[34]
19-Hydroxypsammalyisin E (51)	Alkaloid	<i>Aplysinella strongylata</i>	Antimalarial against <i>P. falciparum</i>	[35]
Psammalyisin K (52)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin K dimethoxy acetal (53)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin L (54)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin M (55)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin N (56)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin O (57)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin P (58)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
19-Hydroxypsammalyisin P (59)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin Q (60)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
19-Hydroxypsammalyisin Q (61)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin R (62)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin S (63)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]

Table 1. Cont.

Compound	Compound Class	Species	Biological Activity	Ref
19-Hydroxypsammalyisin S (64)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin T (65)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
19-Hydroxypsammalyisin T (66)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin U (67)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
19-Hydroxypsammalyisin U (68)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin V (69)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Psammalyisin W (70)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
19-Hydroxypsammalyisin W (71)	Alkaloid	<i>A. strongylata</i>	n.a.	[35]
Lamellodysidine A (72)	Terpenoid	<i>Lamellodysidea herbacea</i>	n.a.	[36]
Lamellodysidine B (73)	Terpenoid	<i>L. herbacea</i>	n.a.	[36]
<i>O,O</i> -dimethylingshuiolide A (74)	Terpenoid	<i>L. herbacea</i>	n.a.	[36]
11- <i>epi-O,O</i> -dimethylingshuiolide A (75)	Terpenoid	<i>L. herbacea</i>	n.a.	[36]
18-nor-3,17-dihydroxy-spongia-3,13(16),14-trien-2-one (76)	Terpenoid	<i>Spongia</i> sp.	Inhibition of aromatase	[37]
18-nor-3,5,17-trihydroxy-spongia-3,13(16),14-trien-2-one (77)	Terpenoid	<i>Spongia</i> sp.	n.a.	[37]
Songiapyridine (78)	Terpenoid	<i>Spongia</i> sp.	n.a.	[37]
20,24-bishomo-25-norscalarane 1 (79)	Terpenoid	<i>Carteriospongia foliascens</i>	Antiproliferative activity against human prostate PC3, colorectal LoVo, colorectal CACO-2, and breast MDA-468 cancer cells; inhibition of RCE-protease	[38]
20,24-bishomo-25-norscalarane 2 (80)	Terpenoid	<i>C. foliascens</i>	n.a.	[38]
20,24-bishomoscalarane ketals 3 (81)	Terpenoid	<i>C. foliascens</i>	Antiproliferative activity against human prostate PC3, colorectal LoVo, colorectal CACO-2, and breast MDA-468 cancer cells; inhibition of RCE-protease	[38]
20,24-bishomoscalarane ketals 4 (82)	Terpenoid	<i>C. foliascens</i>	Antiproliferative activity against human prostate PC3, colorectal LoVo, colorectal CACO-2, and breast MDA-468 cancer cells; inhibition of RCE-protease	[38]
nakijiquinone V (83)	Terpenoid	<i>Dactylospongia elegans</i>	n.a.	[39]
Halioxepine (84)	Terpenoid	<i>Haliclona</i> sp.	Cytotoxic against rat bladder tumour NBT-T2; antioxidant activity	[40]
Melophluoside A (85)	Terpenoid	<i>Melophlus sarasinorum</i>	Cytotoxic against human cervical HeLa cells	[41]
Melophluoside B (86)	Terpenoid	<i>M. sarasinorum</i>	Cytotoxic against human cervical HeLa cells	[41]
Jaspamide Q (87)	Peptide	<i>Jaspis splendens</i>	Cytotoxic against mouse lymphoma L5187Y cancer cell line	[42]
Jaspamide R (88)	Peptide	<i>J. splendens</i>	Cytotoxic against mouse lymphoma L5187Y cancer cell line	[42]

Table 1. Cont.

Compound	Compound Class	Species	Biological Activity	Ref
Sulfinyltheonellapeptolide (89)	Peptide	<i>Theonella swinhoei</i>	Antiproliferative activity against human liver HepG2 cancer cell line	[43]
Theonellapeptolide If (90)	Peptide	<i>T. swinhoei</i>	Antiproliferative activity against human liver HepG2 cancer cell line	[43]
Celebeside A (91)	Peptide	<i>Siliquariaspongia mirabilis</i>	Cytotoxic against HCT116; anti-HIV	[44]
Celebeside B (92)	Peptide	<i>S. mirabilis</i>	n.a.	[44]
Celebeside C (93)	Peptide	<i>S. mirabilis</i>	n.a.	[44]
Theopapuamide B (94)	Peptide	<i>S. mirabilis</i>	Cytotoxic against human colorectal HCT116 cancer cell line; anti-HIV	[44]
Theopapuamide C (95)	Peptide	<i>S. mirabilis</i>	Cytotoxic against human colorectal HCT116 cancer cell line	[44]
Theopapuamide D (96)	Peptide	<i>S. mirabilis</i>	n.a.	[44]
Haloirciniamide A (97)	Peptide	<i>Ircinia</i> sp.	n.a.	[45]
Seribunamide A (98)	Peptide	<i>Ircinia</i> sp.	n.a.	[45]
Manadoperoxide A (99)	Polyketide	<i>Plakortis</i> cfr. <i>simplex</i>	Antimalarial against <i>P. falciparum</i>	[46]
Manadoperoxide B (100)	Polyketide	<i>Plakortis</i> cfr. <i>simplex</i>	Antimalarial against <i>P. falciparum</i>	[46]
Manadoperoxide C (101)	Polyketide	<i>Plakortis</i> cfr. <i>simplex</i>	Antimalarial against <i>P. falciparum</i>	[46]
Manadoperoxide D (102)	Polyketide	<i>Plakortis</i> cfr. <i>simplex</i>	Antimalarial against <i>P. falciparum</i>	[46]
Callyspongiolide (103)	Macrolide	<i>Callyspongia</i> sp.	Cytotoxic against Jurkat J16 T and Ramos B lymphocytes	[47]
Clathruhoate (104)	Steroid	<i>Clathria</i> sp.	n.a.	[48]
Saranoside S (105)	Saponin	<i>Petrosia</i> sp.	n.a.	[49]

To date, more than 80 menzamine-derived alkaloids have been isolated from sponges. This report showed 21 alkaloids were isolated from marine sponge *A. ingens*, five of which were identified as novel manzamine alkaloids, acanthomanzamines A-E (1–5). These five acanthomanzamines were isolated from the marine sponge *A. ingens* collected in Mantehage, North Sulawesi [18]. The acanthomanzamines A (1) and B (2) were the first compounds to contain the 1,2,3,4-tetrahydroisoquinoline-6,7-diol moiety instead of the β -carboline moiety, while the acanthomanzamine C (3) contains the hexahydrocyclopenta[b]-pyrrol-4(2H)-one ring system, and the acanthomanzamines D (4) and E (5) have the oxazolidine and two methyloxazolidine rings, respectively (Figure 1). In terms of biological activity, the acanthomanzamines 1 and 2 demonstrated more potent cytotoxic activity against cervical cancer HeLa cells (IC_{50} values 4.2 and 5.7 μ M, respectively) than the acanthomanzamines 4 and 5 (IC_{50} values 15 and >20 μ M, respectively). However, the acanthomanzamines 4 and 5 showed better proteasome inhibition against the proteasomal chymotrypsin-like activity (IC_{50} values of 0.63 and 1.5 μ M, respectively). These findings suggested that the presence of 1,2,3,4-tetrahydroisoquinoline-6,7-diol probably enhanced the cytotoxicity. Meanwhile, compounds containing β -carboline perform better on chymotrypsin-like activity. Additionally, the acanthomanzamines 1, 2, 4, and 5 inhibited the accumulation of the cholesterol ester at 20 μ M in macrophages with 48%, 73%, 73%, and 61%, respectively [18].

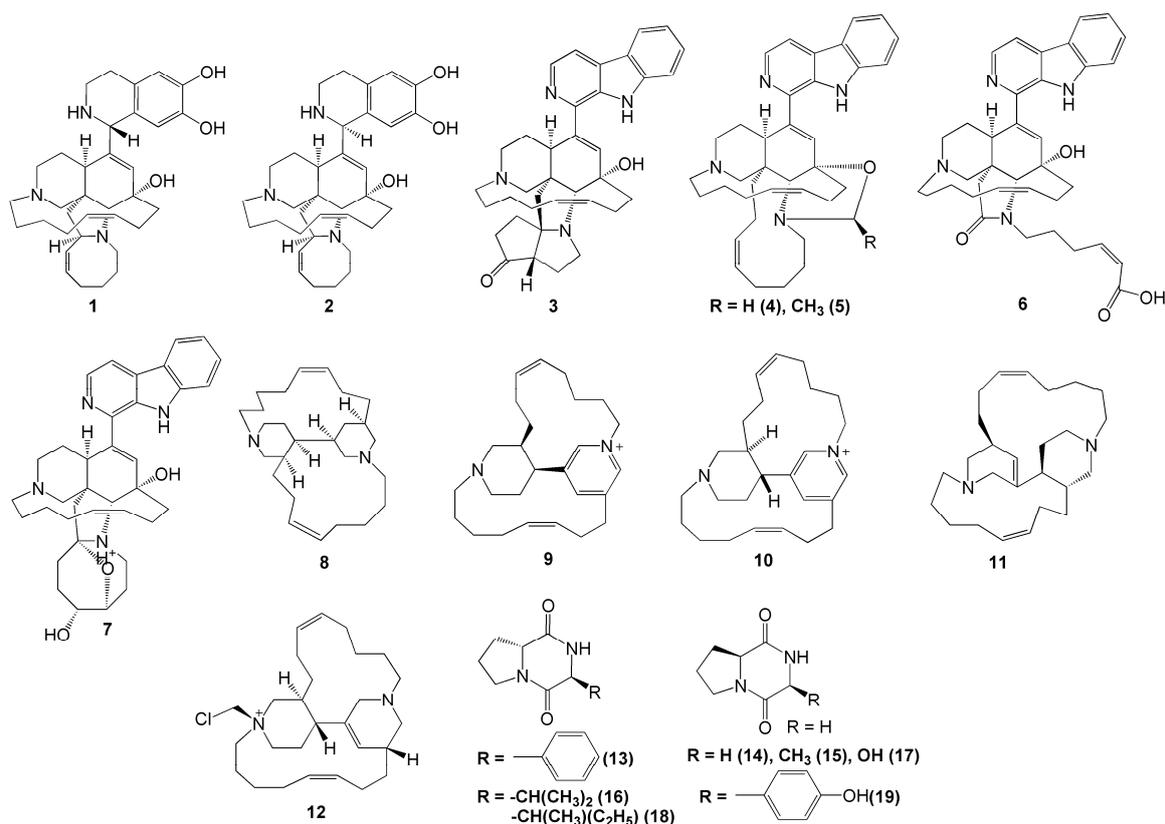


Figure 1. Chemical structures of 1–19.

The sponge *A. ingens* also produces other types of nitrogen-containing molecules, i.e., acantholactam (6), pre-*neo*-kauluamine (7), acathocyclamine A (8), *epi*-tetradehydrohalicyclamine B (9), tetradehydrohalicyclamine B (10), halicyclamine B (11), chloromethylhalicyclamine B (12), cyclo (D-Pro-L-Phe) (13), cyclo (L-Pro-Gly) (14), cyclo (L-Pro-L-Ala) (15), cyclo (D-Pro-L-Val) (16), cyclo (L-Pro-Ser) (17), cyclo (D-Pro-L-Ile) (18), and cyclo (L-Pro-L-Tyr) (19). Alkaloid compounds 6 and 7 were isolated from *A. ingens* collected in Bajotalawaan, North Sulawesi [19]. Compound 7 exhibited proteasome inhibitory activity with an IC_{50} value of 0.34 μ M, whereas compound 6 showed little to no activity [19]. These findings indicated that the eight-membered ring in the manzamines plays a key role in their bioactivity.

A novel 3-alkylpiperidine alkaloid (8) was also isolated from *A. ingens* collected from Wakatobi Marine National Park in Southeast Sulawesi [20]. Recently, the alkaloid compounds 9–19 were also isolated from *A. ingens* collected from South Sulawesi along with compound 8 [21]. Moreover, compound 8 was reported to have specific antimicrobial activity against *E. coli* and showed an inhibitory effect on amyloid β -42 production induced by A β 5 without cytotoxicity at 26 μ M. Compounds 8 and 11 exhibited antibacterial activity at 100 μ g/disc against *E. coli* and *S. aureus*. In addition to that, compound 12 was reported to have selective inhibitory activity against the protein kinase CK1 δ/ϵ with an IC_{50} value of 6 μ M, while the diketopiperazine compound 13 showed a selective kinase inhibitory activity against CDK2/cyclin A with an IC_{50} value of 1 μ M. However, no bioactivity was reported from compounds 14–19 [21].

A study reported two new pyrimidine- β -carboline alkaloids—namely, ingenines C (20) and D (21), successfully isolated from *A. ingens* obtained in Sulawesi [22]. The ingenines 20 and 21 exhibited cytotoxic activities towards the human breast MCF-7 and colorectal HCT116 cancer cells (IC_{50} values of 4.33 and 6.05 μ M, and 2.90 and 3.35 μ M, respectively) [22].

Two bromopyrrole alkaloids, dispacamide E (22) and ethyl 3,4-dibromo-1*H*-pyrrole-2-carboxylate (23), were isolated from methanolic extract of the marine sponge *S. massa*

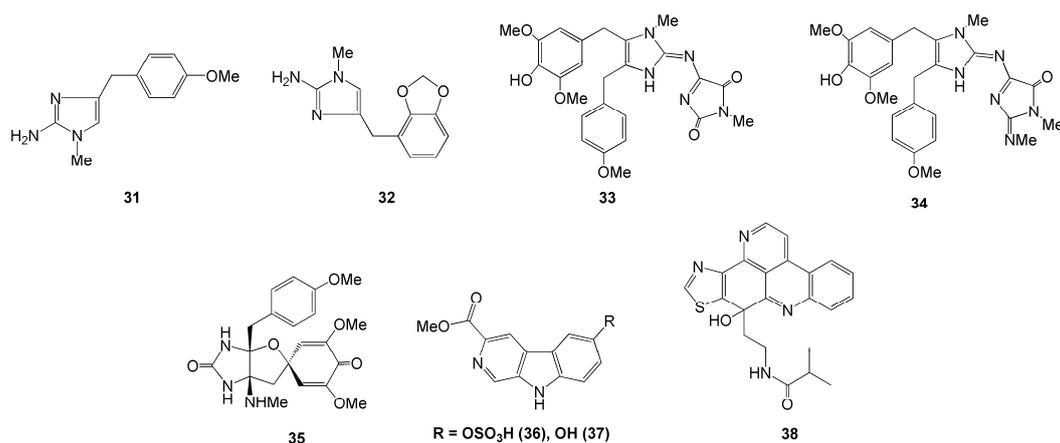


Figure 3. Chemical Structures of 31–38.

Two new β -carboline alkaloids, variabines A (36) and B (37), were isolated from Indonesian marine sponge *Luffariaella variabilis* collected in North Sulawesi [29]. Compound 36 was the first sulfated β -carboline alkaloid and the sulfonated derivative of compound 37. A hydroxy group in compound 37 replaces the sulfate group at C-6 in compound 36. Compound 37 was found to inhibit the chymotrypsin-like activity of the proteasome and Ubc13 (E2)-Uev1A interaction with IC_{50} values of 16.5 and 20.6 μ M, respectively. In contrast, compound 36 showed a weak effect on proteasome. This result suggested that the sulfate group in variabines decreases chymotrypsin-like activity [29].

A novel pyridoacridine alkaloid, sagitol C (38), was isolated from the ethyl acetate fraction of *Oceaniapia* sp. collected in Ambon, Maluku Islands [30]. Alkaloid 38 was found to exhibit cytotoxic activity against the mouse lymphoma L5187Y cancer cell line, HeLa cell, and the rat pheochromocytoma PC12 cell lines with ED_{50} values of 0.7, 0.9, and 2.3 μ M, respectively [30].

Seven novel anti-angiogenic steroidal alkaloids (Figure 4) were isolated from the sponge *Corticium simplex* collected in Flores Island, East Nusa Tenggara [31,32], namely cortistatins E-H (39–42) and J-L (43–45). These compounds have unique abeo-9(10-19)-stigmastane-type steroidal alkaloids. Compounds 38–42 have oxabicyclo[3.2.1]octene and *N*-methyl piperidine or 3-methylpyridine units in the side chain, while compounds 43–45 have an isoquinoline unit. Compound 43 showed cytostatic antiproliferative activity against human umbilical vein endothelial cells (HUVECs) at 8 nM [31,32].

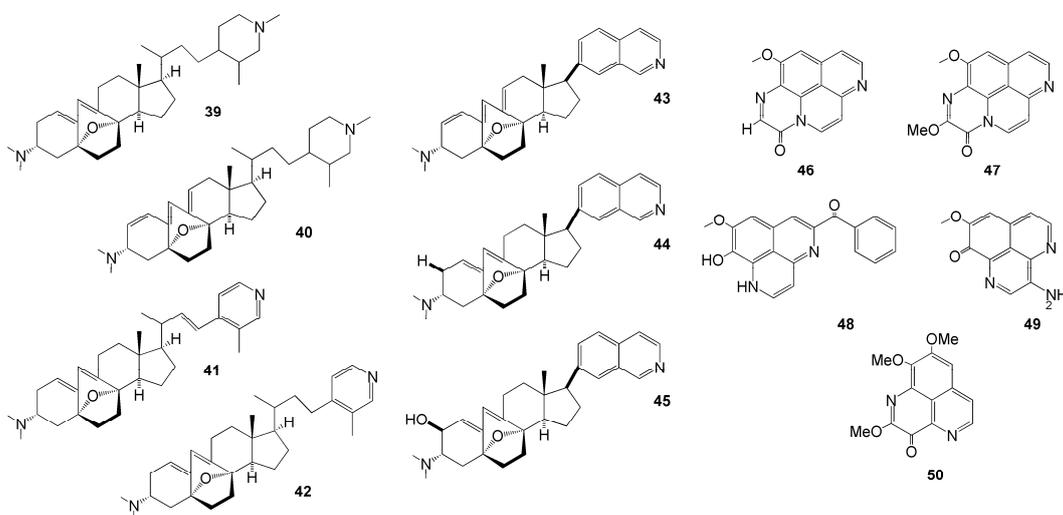


Figure 4. Chemical structures of 39–50.

The genus *Aaptos* is also an abundant source of novel aaptamine alkaloids (Table 1). Four new aaptamine derivatives were isolated from *A. suberitoides* collected in Ambon, Maluku Islands [33]—namely, 11-methoxy-3*H*-[1,6]naphthyridino [6,5,4-*def*]quinoxalin-3-one (46), 2,11-dimethoxy-3*H*-[1,6]naphthyridino [6,5,4-*def*]quinoxalin-3-one (47), 5-benzoyl-demethylaaptamine (48), and 3-aminodemethyl(oxy)aaptamine (49). Compound 47 is the 11-methoxy derivative of compound 46 and represents a benzoyl-aaptamine skeleton that has not been previously identified [33]. Concerning the biological activity, compound 48 exhibited cytotoxic activity against the mouse lymphoma L5187Y cancer cell line with an IC₅₀ value of 5.5 μM [33]. Another aaptamine derivative isolated from *Aaptos* sp. collected from Kupang, East Nusa Tenggara [34],—namely, 2-methoxy-3-oxoaaptamine (50), demonstrated an anti-mycobacterial activity under both aerobic and hypoxic conditions, both with a MIC value of 23 μM [34].

Twenty-one novel psammalyisin derivatives were successfully isolated from *A. strongylata* collected in Tulamben Bay, Bali [35]. Compounds 19-Hydroxypsammalyisin E (51), psammalyisin K (52), psammalyisin K dimethoxy acetal (53), psammalyisin L (54), and M (55), possess modified aromatic ring substituents. Meanwhile, psammalyisin N-P (56–58), 19-hydroxypsammalyisin P (59), psammalyisin Q (60), 19-hydroxypsammalyisin Q (61), psammalyisin R-S (62–63), 19-hydroxypsammalyisin S (64), psammalyisin T (65), and 19-hydroxypsammalyisin T (66) exhibit various side chains that are saturated fatty acid side chains (Figure 5). On the other hand, psammalyisin U (67), 19-hydroxypsammalyisin U (68), psammalyisin V-W (69–70), and 19-hydroxypsammalyisin W (71) have monoenoic fatty acid side chains. Among these 21 compounds, 19-hydroxypsammalyisin E (51), having the *N*-substitution of the ethylamino moiety, showed the best antimalarial activity with modest inhibition against *P. falciparum* (IC₅₀ = 6.4 μM) [35].

Several new terpenes were isolated from Indonesian marine sponges from the Dictyoceratida order (Figure 6), such as *L. herbacea* [36], *Spongia* sp. [37], and *Carterospongia foliascens* [38]. Four new sesquiterpenes were isolated from *L. herbacea* collected in Manado, North Sulawesi—namely, lamellodysidines A (72) and B (73), *O,O*-dimethylingshuiolide A (74), and 11-*epi-O,O*-dimethylingshuiolide A (75) [36]. Compound 72 was the first compound identified with a unique bridged polycyclic framework, and compound 73 is a novel nitrogenous sesquiterpene, while compounds 74 and 75 were obtained as an inseparable mixture due to their 11-epimeric nature [36].

Sponges from the genus *Spongia* are known as a rich source of unique terpenoids (Table 1). This is evident in the successful isolation of three novel terpenoids from *Spongia* sp. collected in Bunaken Marine Park, North Sulawesi [37]. These three compounds were identified as diterpene—namely, 18-nor-3,17-dihydroxyspongia-3,13(16),14-trien-2-one (76), 18-nor-3,5,17-trihydroxyspongia-3,13(16),14-trien-2-one (77), and spongiapyridine (78). Compound 77 possesses an unusual D-ring substitute, a pyridyl ring system, in the place of δ-lactone (Figure 6). In terms of the biological activity of these four compounds, only compound 76 possessed moderate activity on aromatase inhibition with an IC₅₀ of 34 μM and quinone reductase 1 induction with the concentration needed to double the enzymatic response of 11.2 μM [37].

Four new scalarane-based sesterterpenoids were isolated from *C. foliascens* associated with the coral reefs of Palau Barang Lompo, near Makasar, South Sulawesi [38]. The closely related compounds were identified as 20,24-bishomo-25-norscalaranes 1 (79) and 2 (80), and 20,24-bishomoscalaranes ketals 3 (81) and 4 (82) (Figure 6). Compounds 81 and 82 were isolated as an inseparable mixture. Compound 80 showed little to no activity toward RCE-protease inhibition compared to 79. Meanwhile, a combination of 80 and 82 (plus 81, as its inseparable mixture) had inhibition activity with IC₅₀ values of 38 and 4.2 μg/mL, respectively [38]. Compound 79 and the mixture of 81 and 82 also showed inhibition against tumour cell lines (human prostate PC3, colorectal LoVo, colorectal adenocarcinoma CACO-2, and breast MDA-468 cancer cell lines) with IC₅₀ values ranging from 2.9–9.5 μg/mL [38].

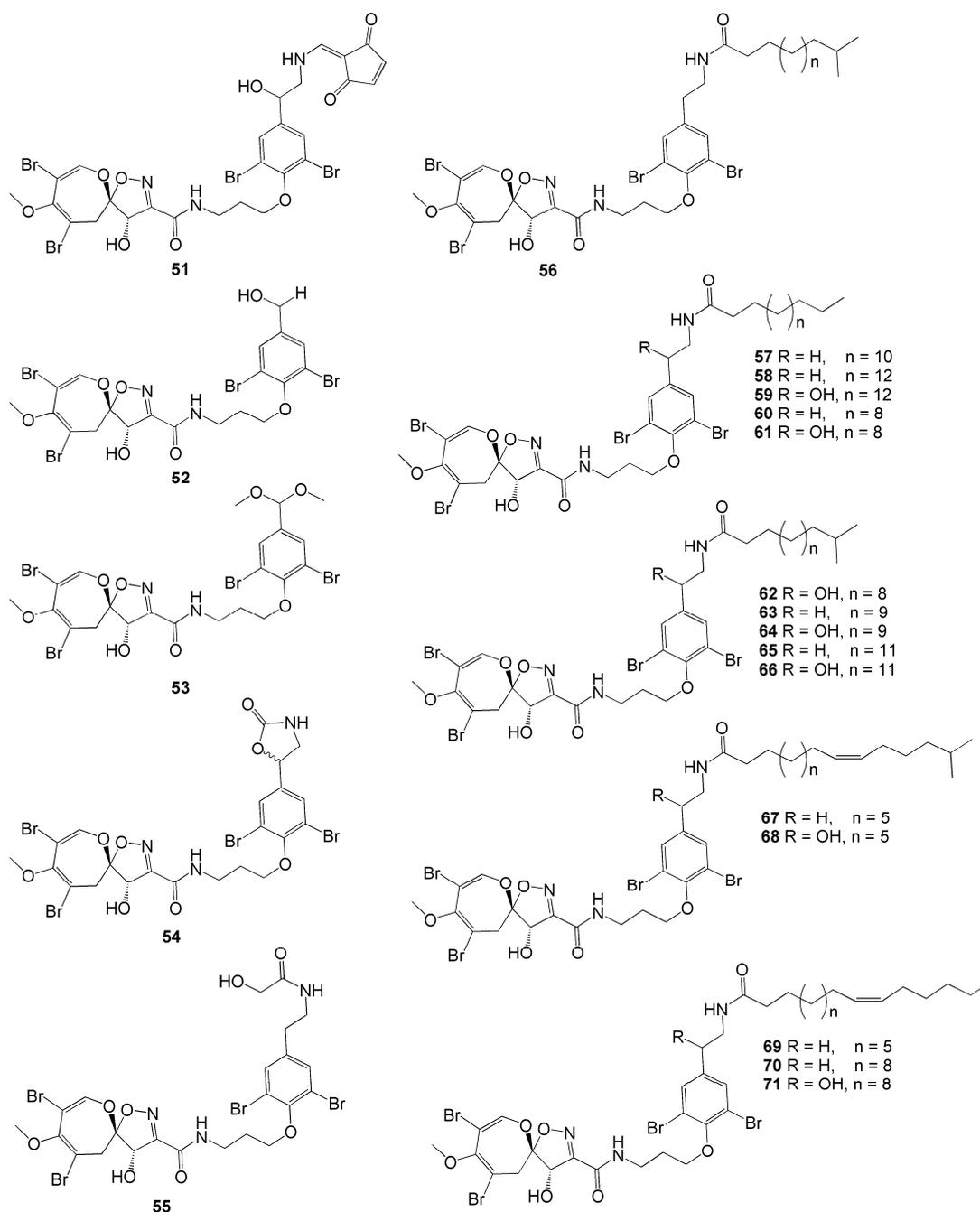


Figure 5. Chemical structures of 51–71.

Other compounds isolated from Sulawesi included nakijiquinone V (**83**), isolated from *Dactylosporgia elegans* from Tahuna, Sangihe Islands, North Sulawesi [39]. It has three methyl groups attached to a decalin system with an exocyclic double bond [39] (Figure 6). Another was a new meroditerpene—namely, Halioxepine (**84**), which was isolated from an Indonesian marine sponge from the genus *Haliclona* collected in Baubau, Southeast Sulawesi [40]. This compound showed moderate cytotoxicity against the rat bladder tumour NBT-T2 cells with IC_{50} of 11.6 μ M and antioxidant activity against 1,1-diphenyl-2-picrylhydrazyl (DPPH) with IC_{50} of 7.7 μ M [40]. Recently, two new terpenoids, melophluosides A (**85**) and B (**86**), were successfully isolated from *Melophlus sarassinorum* collected in Siladen, North Sulawesi. These new compounds belong to the triterpene galactosides of the pouoside class, with compound **85** as the first without an oxygenated

group at C-11. Both compounds exhibited moderate cytotoxic activity against HeLa cells with IC₅₀ values of 11.6 and 9.7 mM, respectively [41].

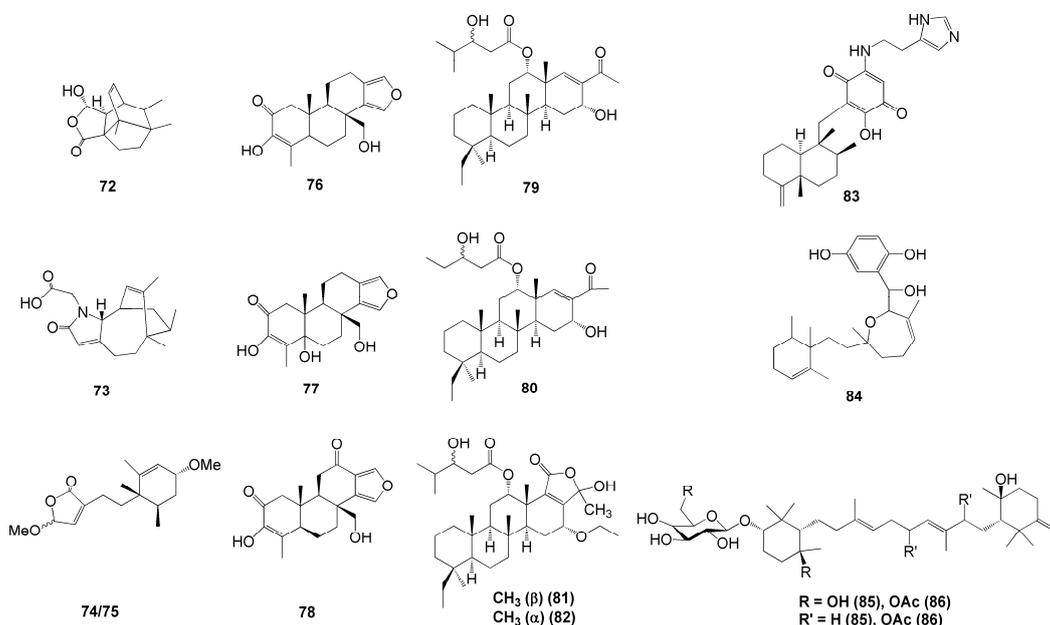


Figure 6. Chemical structures of 72–86.

Two new cyclopeptide jaspamide derivatives, jaspamide Q (87) and jaspamide R (88), were identified from the marine sponge *J. splendens* collected in neighbouring islands near East Kalimantan [42]. They exhibited potent cytotoxic activity against the mouse lymphoma L5187Y cancer cell line with IC₅₀ values of <126.8 μM and <203 μM, respectively [42].

Novel tridecapeptides of the theonellaepetolide family were isolated from *T. swinhoei* collected from Bunaken Marine Park in Manado, North Sulawesi—namely, sulfanyltheonellaepetolide (89) and theonellaepetolide If (90) [43]. These compounds differ in the abrine moiety. Compound 89 is a theonellaepetolide with a methylsulfanylacetyl group at the *N*-terminus, while compound 90 was the first theonellaepetolide with four valine residues (Figure 7). Both compounds showed significant antiproliferative activity against human liver HepG2 cancer cells with the same IC₅₀ value of 3 μM [43].

S. mirabilis is also one of the rich sources of diverse secondary metabolites, as exemplified by the isolation of six new depsipeptides with two different structural classes named celesbesides A–C (91–93) from the species collected in Sulawesi [44]. Compounds 91–93 are cyclic depsipeptides with a polyketide moiety and five amino acid residues (Figure 7). The celesbesides A and B (91, 92) possess a 3-carbamoyl threonine and a phosphoserine residue, which is quite uncommon. *S. mirabilis* also produced the peptides theopapuamides B–D (94–96), which are undeca-peptides with an *N*-terminal fatty acid moiety, with theopapuamide D (96) containing a rare homoisoleucine residue (Figure 7). These compounds exhibited several bioactivities. Theopapuamides B and C (94, 95) were found to have relatively strong antifungal activity against amphotericin B-resistant *C. albicans* [44]. Additionally, celesbeside A (91) and theopapuamide B (94) neutralized HIV-1 in a single-round infectivity assay with an IC₅₀ value of 2.1 ± 0.4 μM and 499.7 ± 0.3 μg/mL [44]. Celebeside 91 and theopapuamides 94 and 95 also demonstrated cytotoxicity against human colon carcinoma cells with IC₅₀ values 9.9 μM, 1.3 μM, and 2.5 μM, respectively [44]. Although they were potent against human colon HCT116 carcinoma cells, celesbesides A, B, and C (91–93) were also found to be cytotoxic for healthy cell lines [44].

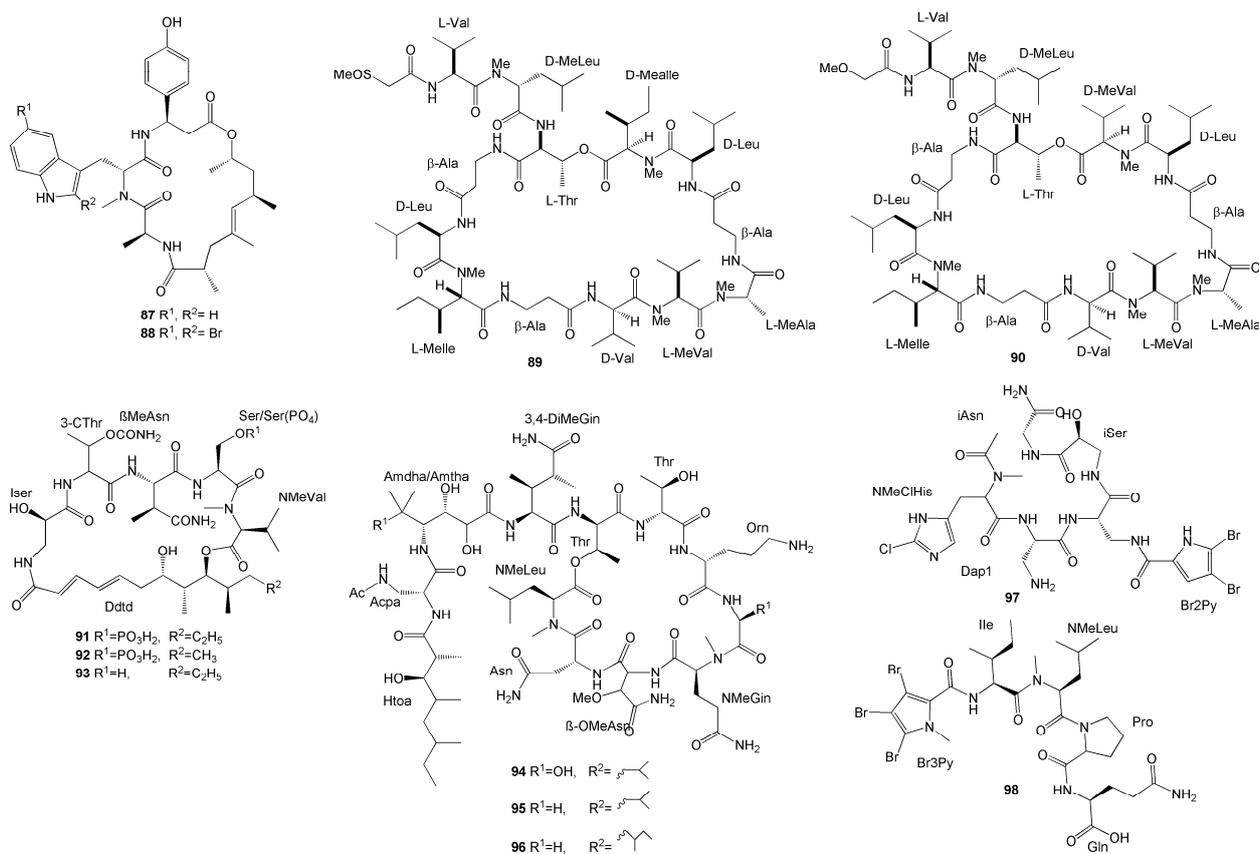


Figure 7. Chemical structures of 87–98.

Haloirciniamide A (97) and seribunamide A (98), new polyhalogenated peptides, have been isolated from *Ircinia* sp. collected from the coast of Thousand Islands. Compound 97 was the first dibromopyrrole cyclopeptide with a chlorohistidine ring, while compound 98 possess a rare tribromopyrrole ring. Unfortunately, both compounds did not show significant cytotoxicity against four human tumour cell lines [45].

Four new endoperoxycetal polyketides, manadoperoxides A–D (99–102), were isolated from the sponge *Plakortis* cfr. *simplex* obtained from the Bunaken Marine Park of Manado, North Sulawesi [46]. In these compounds, the methoxy group at C-6 is replaced by either a methyl or ethyl group instead of a peroxyketal-type (Figure 8), making them slightly different from those previously isolated from the same species in the Caribbean. All compounds isolated from that sponge *Plakortis* cfr. *simplex* showed moderate antimalarial activity against D10 and W2 strains of *Plasmodium falciparum* [46].

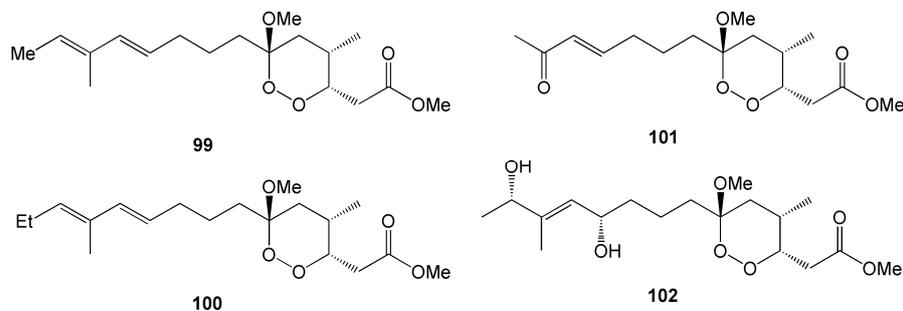


Figure 8. Chemical structures of 99–102.

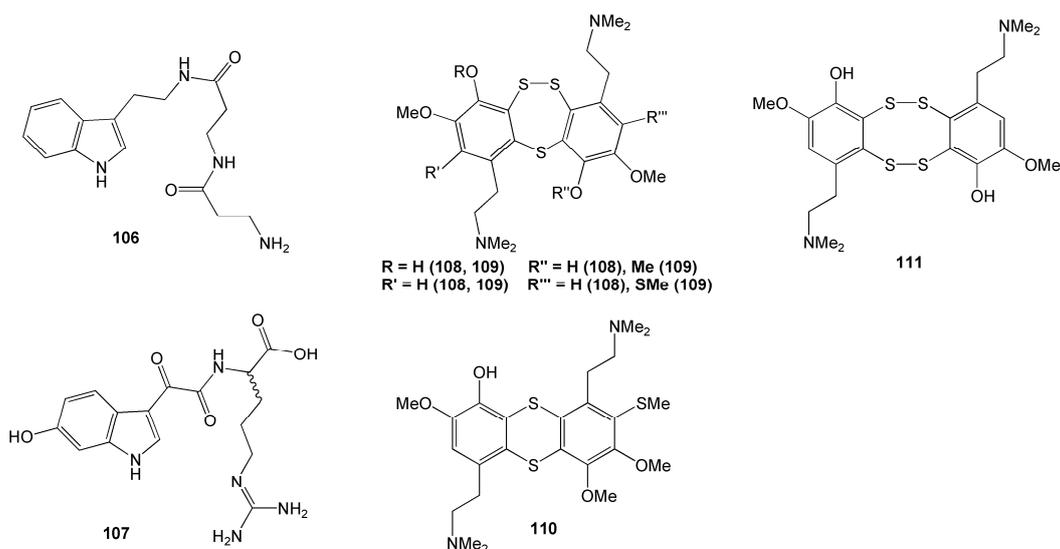


Figure 10. Chemical structures of **106–111**.

Table 2. Summary of the marine natural products isolated from the marine tunicates from Indonesian oceans.

Compound	Compound Class	Species	Biological Activity	Ref
Leptoclinidamide (106)	Alkaloid	<i>Leptoclinides dubius</i>	n.a.	[50]
(-)-leptoclinidamine B (107)	Alkaloid	<i>L. dubius</i>	n.a.	[50]
Lissoclibadin 4 (108)	Alkaloid	<i>Lissoclinum</i> cf. <i>badium</i>	Antibacterial against <i>S. aureus</i> and <i>E. coli</i>	[51]
Lissoclibadin 5 (109)	Alkaloid	<i>Lissoclinum</i> cf. <i>badium</i>	Antibacterial against <i>S. aureus</i> and <i>E. coli</i> ; anti-yeast activity against <i>S. cerevisiae</i>	[51]
Lissoclibadin 6 (110)	Alkaloid	<i>Lissoclinum</i> cf. <i>badium</i>	Antibacterial against <i>S. aureus</i> and <i>E. coli</i> ; anti-yeast activity against <i>S. cerevisiae</i>	[51]
Lissoclibadin 7 (111)	Alkaloid	<i>Lissoclinum</i> cf. <i>badium</i>	Antibacterial against <i>S. aureus</i> and <i>E. coli</i> ; anti-yeast activity against <i>S. cerevisiae</i>	[51]
Polycarpathiamine A (112)	Alkaloid	<i>Polycarpa aurata</i>	Cytotoxic against mouse lymphoma L5187Y cancer cell line	[52,53]
Polycarpathiamine B (113)	Alkaloid	<i>P. aurata</i>	n.a.	[52]
Polyaurine A (114)	Alkaloid	<i>P. aurata</i>	Egg deformation of <i>Schistosoma mansoni</i>	[53]
Polyaurine B (115)	Alkaloid	<i>P. aurata</i>	n.a.	[53]
Ethyl 2-(4-methoxyphenyl)-2-oxoacetate (116)	benzoyl derivatives	<i>P. aurata</i>	n.a.	[53]
Methyl 2-(4-hydroxyphenyl)-2-oxoacetate (117)	benzoyl derivatives	<i>P. aurata</i>	n.a.	[53]
Mollamide B (118)	Peptide	<i>Didemnum molle</i>	Cytotoxic against human non-small cell lung H460, breast MCF-7, and glioblastoma SF268 cells; antimalarial against <i>P. falciparum</i> ; antiparasitic against <i>Leishmania donovani</i> ; antiviral against HIV-1	[54]

Table 2. Cont.

Compound	Compound Class	Species	Biological Activity	Ref
Mollamide C (119)	Peptide	<i>D. molle</i>	Cytotoxic against mouse lymphocytic leukaemia L1210, human colorectal HCT116, lung H125, and murine colon MC 38 cancer cells	[54]

Another two novel alkaloids have recently been isolated from another *P. aurata* from the coast of Siladen, North Sulawesi [53]. Featuring a *p*-methoxyphenyl group, these compounds were named polyaurines A (114) and B (115). Compound 114 deformed eggs of parasite *Schistosoma mansoni*, yet it was not toxic to mammalian cells [53]. Two new benzoyl derivatives from this species, ethyl 2-(4-methoxyphenyl)-2-oxoacetate (116) and methyl 2-(4-hydroxyphenyl)-2-oxoacetate (117), were also isolated (Figure 11).

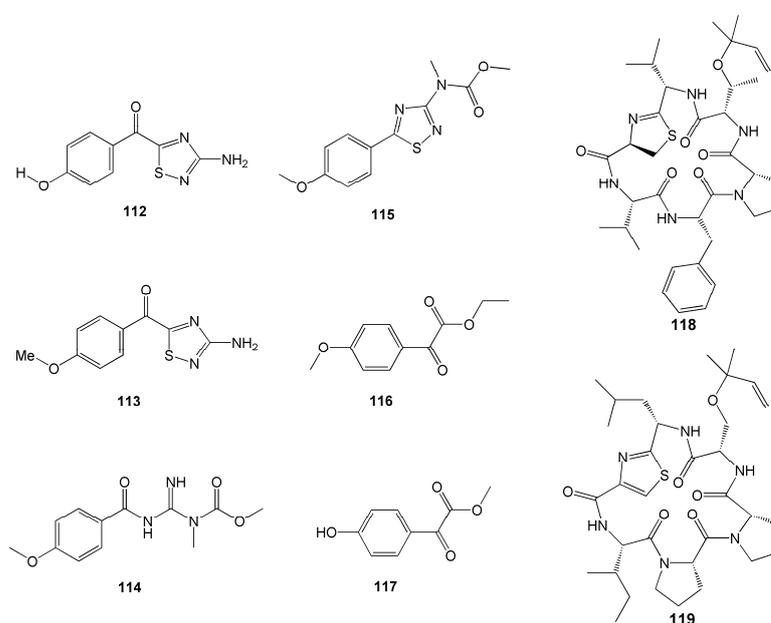


Figure 11. Chemical structures of 112–119.

Two new cyclic hexapeptides, mollamides B (118) and C (119) (Figure 11), were isolated from *D. mole* found in Manado Bay, Indonesia. Compound 118 was found to have an antiparasitic activity against *Plasmodium falciparum* D6 clone ($IC_{50} = 2.9 \mu M$), *P. falciparum* W2 clone ($IC_{50} = 3 \mu M$), and *Leishmania donovani* ($IC_{50} = 25.9 \mu M$ and $IC_{90} = 50.3 \mu M$) [54]. Compound 118 also displayed antiviral activity against HIV-1 in human PBM cells with an EC_{50} value of $48.7 \mu M$ and anticancer activity against the non-small lung H460, the breast MCF-7, and the human glioblastoma CNS SF268 cancer cell lines [54]. Furthermore, compound 119 showed anticancer activity with a unit zone differential value of 100 against mouse lymphocytic leukaemia L1210, human colon HCT116, and human lung H125 cells. This compound also showed a differential value of 250 against murine colon 38 [54]. However, compounds 118 and 119 did not show anti-inflammatory activity either in vitro in a cell-based assay through a cyclooxygenase enzyme (COX-2) activity assay or in vivo in rat neonatal microglia [54]. Furthermore, compounds 118 and 119 did not show antimicrobial activity against MRSA, *Mycobacterium intracellulare*, *Candida albicans*, *C. glabrata*, *C. krusei*, nor *Cryptococcus neoformans* [54].

2.3. Soft Corals

Terpenoids were often isolated from soft corals, with varying degrees of bioactivities (Table 3). Sarcofuranocembrenolides A (**120**) and B (**121**) were isolated from the soft coral *Sarcophyton* sp. collected in North Sulawesi [55]. Cembranoid **120** is a bisnorcembrenolide featuring a unique carbon skeleton of 8,19-bisnorfuranocembrenolide (Figure 12). On the other hand, cembranoid **121** is a furanocembrenolide with a C₁ unit (C-20) attached to C-10. In the ordinary cembranoids, the C₁ unit is attached to C-12 [55].

Table 3. Summary of the marine natural products isolated from the marine soft corals from Indonesian oceans.

Compound	Compound Class	Species	Biological Activity	Ref
Sarcofuranocembrenolide A (120)	Terpenoid	<i>Sarcophyton</i> sp.	n.a.	[55]
Sarcofuranocembrenolide B (121)	Terpenoid	<i>Sarcophyton</i> sp.	n.a.	[55]
Chloroscabrolide A (122)	Terpenoid	<i>Simularia</i> sp.	n.a.	[56]
Chloroscabrolide B (123)	Terpenoid	<i>Simularia</i> sp.	n.a.	[56]
Prescabrolide (124)	Terpenoid	<i>Simularia</i> sp.	n.a.	[56]
Cladielloide A (125)	Terpenoid	<i>Cladiella</i> sp.	n.a.	[57]
Cladielloide B (126)	Terpenoid	<i>Cladiella</i> sp.	Cytotoxic against lymphocytic leukaemia CCRF-CEM cells; inhibition of superoxide anion generation; inhibition of elastase release	[57]
3,4-epoxy-nephtenol acetate (127)	Terpenoid	<i>Nephtea</i> sp.	Antiproliferative activity against human glioblastoma SF268, breast MCF-7, and non-small cell lung H460 cancer cells	[58]
Sangiangol A (128)	Terpenoid	<i>Anthelia</i> sp.	Cytotoxic rat bladder tumour NBT-T2 cell line	[59]
Sangiangol B (129)	Terpenoid	<i>Anthelia</i> sp.	Cytotoxic rat bladder tumour NBT-T2 cell line	[59]
Loboanthamine (130)	Alkaloid	<i>Lobophytum</i> sp.	n.a.	[60]

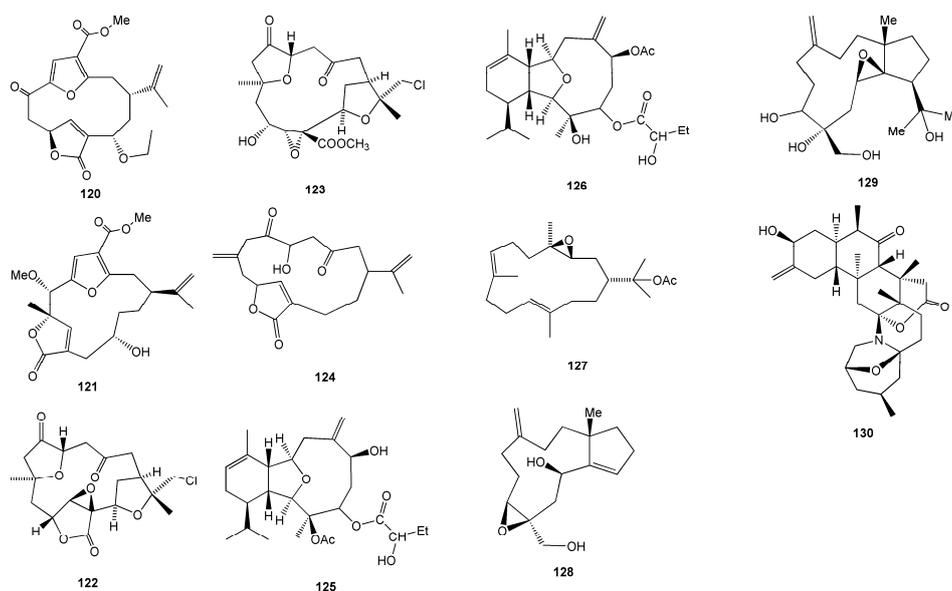


Figure 12. Chemical structures of 120–130.

Three C-4 norcembranoids-type macrocyclic diterpenoids, namely chloroscabrolide A (**122**), chloroscabrolide B (**123**), and prescabrolide (**124**), were isolated from *Simularia* sp. collected from Bunaken Marine Park, Manado, North Sulawesi [56]. Compounds **122** and **123** are two of very few chlorinated compounds from soft coral metabolites and the second

example within the class of cembranoids (Figure 12). These two compounds also feature an oxygen bridge connecting C-13 and C-15, which is quite unusual. Meanwhile, the terpenoid prescarbolid (124) is believed to be the precursor of the scabrolide/leptocladolide family of cembranoids [56].

Two new isomeric eunicellin-type diterpenoids were isolated from an Indonesian octocoral *Cladiella* sp. in the West Pacific Ocean [57]. Cladielloide A (125) and cladielloide B (126) both possess a 2-hydroxybutyryloxy group in their structures (Figure 13). Compound 126 exhibited a potent inhibitory effect on superoxide anion generation and elastase release by human neutrophils at 22 μ M. It also showed moderate cytotoxicity against the acute lymphocytic leukaemia CCRF-CEM tumour cells with an IC₅₀ value of 10.1 μ M [57].

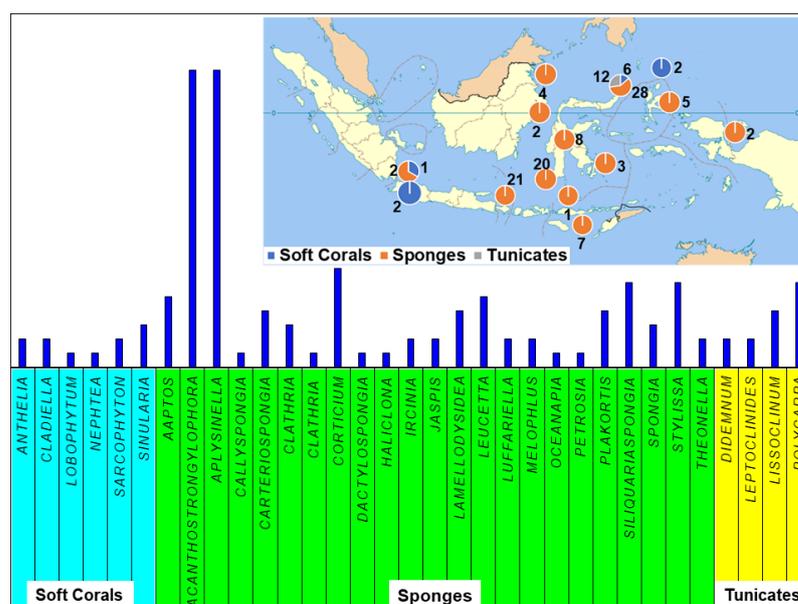


Figure 13. The distribution of sample origin and the division of compound class by genus.

A terpenoid compound, 3,4-epoxy-nephtenol (127), was isolated from the soft coral *Nephthea* sp. found in Seribu Islands, DKI Jakarta [58]. Compound 127 showed weak inhibitory growth against three human tumour cell lines, i.e., human glioblastoma SF268, human breast MCF-7, and human non-small cell lung H460 cancer cells [58].

The latest finding on terpenoids from soft coral was discovered from *Anthelia* sp. collected at Banten, West Java, which successfully isolated two new dolabellane diterpenoids, sangiangol A (128) and sangiangol B (129). These two compounds were found to show weak toxicity against NBT-T2 rat bladder epithelial cells (BRC-1370) at 18 and 28.2 μ M, respectively [59]. The first reported zoanthamine-type alkaloid from a marine invertebrate other than zoanthids was named loboanthamine (130), isolated from the Indonesian soft coral *Lobophytum* sp. collected from the Bunaken Marine Park, Manado, North Sulawesi [60]. However, there is no report on the bioactivity of the compounds 120–125 and 130 (Table 3).

3. Conclusions

Indonesian marine biodiversity holds immense potential for drug discovery and bioprospecting, supporting the continuous exploration and investigation of Indonesian MNPs from marine invertebrates such as sponges, tunicates, and soft corals. This is further justified by no less than a hundred and thirty novel compounds reported in this review from 43 publications throughout 2007–2020, the majority of which were isolated from the Indonesian marine invertebrates collected from the North region of Sulawesi, Indonesia (Figure 13).

Indonesian sponges have been known as a major source of many novel MNPs. In this review, sponges were found to contribute to 105 novel compounds reported between 2007–2020, the majority being alkaloids (Figure 14). Most of the sponge alkaloids were isolated from the genera *Aplysinella* and *Acanthostrongulophora*. The next large groups of metabolites isolated from Indonesian sponges were terpenes and peptides. Nearly all the sponge terpenoids were isolated from the order Dictyoceratida, in which the dominant genera were *Lamellodysidea* and *Carteriospongia*.

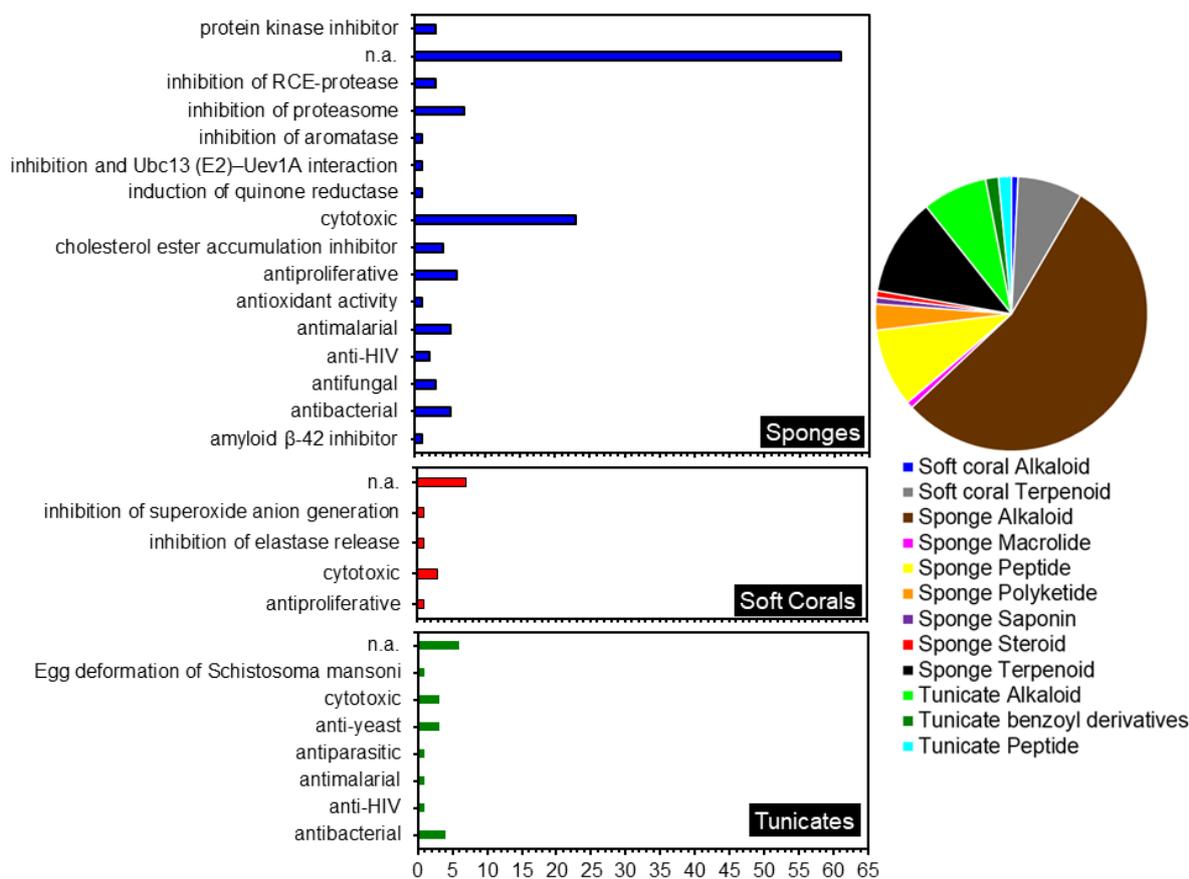


Figure 14. Biological activities of compounds isolated from Indonesian marine invertebrates (left) and the division of genus by compound class (right).

Meanwhile, all the peptides, with the exception of two peptides from *Ircinia* sp., were isolated from the order Tetractinellida. On the other hand, macrolides, steroids, and saponins were the least frequently isolated metabolites from Indonesian sponges. The sponge *Plakortis simplex* was the sole contributor of the polyketides isolated from Indonesia (Table 1).

During 2007–2020, ten alkaloids were reported from Indonesian tunicates, mainly from the family *Didemnidae* and *Styelidae*. Soft corals are also a good source of novel metabolites and are well-known as a producer of terpenoids, particularly the group of terpenes and cembranoid diterpenes. Soft coral-derived terpenoids have received significant attention in Indonesian MNPs research (Table 3). Compared to other invertebrates with alkaloids as the most abundant secondary metabolites, only one alkaloid was found in Indonesian soft coral species in this review.

Indonesian soft corals and tunicates yielded far lower novel secondary metabolites than sponges reported throughout 2007–2020. Therefore, this review highlighted the opportunity to explore further the chemical diversity and the biological activity of tunicates from Indonesian waters.

Diverse biological activities were shown by fifty per cent of the compounds isolated from the three Indonesian marine invertebrates discussed (Figure 14). The majority of compounds in this review were reported to possess cytotoxic or antiproliferative activity against various cancer cell lines. Four of these compounds showed a remarkable cytotoxic activity with IC₅₀ values of less than 1 µM, namely compounds **30**, **38**, **103** and **112**. Furthermore, four compounds showed IC₅₀ values between 1–4 µM, and another eight showed moderate cytotoxic with an IC₅₀ value of less than 10 µM. Most of these compounds belong to the alkaloid group, followed by peptides and terpenoids. It is, therefore, evident that the exploration of potential anticancer drugs from Indonesian marine resources warrants further investigation.

The bioactivity of compounds isolated from the Indonesian marine invertebrates (sponges, tunicates, and soft corals) as antimicrobial, antifungal, or antiviral was also described in this review. Unfortunately, little or no such activities were yet to be found from many compounds derived from the three Indonesian marine invertebrates, demonstrating the gap in the knowledge of their beneficial biological activity other than their cytotoxic activity. This knowledge gap opens up the opportunity for further research focusing on exploring and harnessing the potential of Indonesian marine invertebrates as sources of compounds with antimicrobial, antifungal, or antiviral activities, as well as further exploration of Indonesian marine biodiversity for the discovery of novel bioactive compounds.

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