Supplementary

Key factor study for generic long-acting PLGA microspheres based on a reverse

engineering of Vivitrol[®]

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Verification of HPLC methodology

Table S1 Verification of HPLC method to determine naltrexone*

		Results
System applicability		After 6 injections of the reference solution, the RSD of retention time and peak area is less than 1%, and the trailing factor is between 0.9 and 1.1. In addition, the separation degree is more than 4.
Linearity and range		When naltrexone is in the range of 1-100 μ g/mL, the linear equation is y = 35.772x + 43.091, and the linear regression coefficient r=0.9998, indicating a good linear relationship between concentration and peak area. The limit of detection is 100 ng/mL and the limit of quantitation is 1 μ g/mL.
	Repeatability	The RSD of 6 samples of solution was 0.27% (< 1.0%).
Precision	Intermediate precision	Another experimenter measured the samples with different instruments on other dates, and the RSD of six samples was 0.43% (< 2.0%).
Recovery rate		Both of the intra-group and inter-group recovery rate are between 98% and 102%, and the RSD of them is 0.49% (< 2.0%).

	The blank solvent and excipient solution did not interfere with the
Specificity	determination of naltrexone, and the resolution between the main peak
	and the adjacent peak was $5 (> 1.5)$.

*The verification results of HPLC methodology meet the requirements of ICH Q2.

Verification of Microsphere determination methodology Table S2 Particle size and distribution of microspheres determined by different amounts of microspheres

microspheres						
	50 mg	100 mg	150 mg	200 mg	300 mg	
D ₁₀ (µm)	-	-	19.01	19.43	19.48	
D ₅₀ (µm)	-	-	35.03	35.02	35.74	
D ₉₀ (µm)	-	-	62.46	60.06	62.75	
Span	-	-	1.24	1.16	1.21	

When the weight of microsphere is less than 100 mg, the determination condition cannot be reached. When the weight of microsphere is 200 mg, the span value measured is relatively small, and the particle size result is stable. While using a smaller amount of microspheres, it can achieve a certain shading, and will not precipitate and agglomerate. Therefore, the amount of microsphere is determined to be 200 mg.

Table S3 Particle size and distribution of microspheres determined by different suspending

	0.1 % PVA	Injection diluent	0.1 % Tween-20	0.5 % Tween -20	1.0 % Tween - 20
D ₁₀ (µm)	20.05	18.21	20.73	19.01	19.08
D50(µm)	36.43	35.25	37.19	35.03	34.90
D ₉₀ (µm)	63.06	68.58	62.71	62.46	60.09
Span	1.28	1.43	1.23	1.24	1.18

When 1.0% Tween-20 is used as suspension agent, the span value is the lowest, indicating that the particle size distribution is good. The selected suspension agent is 1% Tween-20. The suspending agent alone does not affect the determination of this method.

Table S4 Particle size and distribution of microspheres with different stirring speed

	1500 rpm	1800 rpm	2100 rpm
D ₁₀ (µm)	19.22	20.81	19.26
D ₅₀ (µm)	35.71	36.83	35.36
D ₉₀ (µm)	65.46	71.55	62.21
Span	1.19	1.18	1.12

When the rotational speed is 2100 rpm, the span value is the smallest, and the agglomeration phenomenon at low rotational speed is avoided.

		1		
Times	D ₁₀ (µm)	D50(µm)	D ₉₀ (µm)	Span
1	47.14	79.74	128.33	1.02
2	46.85	79.31	127.28	1.01
3	46.91	79.37	127.41	1.01
4	46.93	79.48	127.67	1.02
5	46.97	79.60	128.17	1.01
6	46.96	79.75	128.73	1.01
RSD/%	0.09	0.17	0.52	0.51

Table S5 Precision of particle size determination method

One sample of Vivitrol[®] was prepared, and the RSD of six measurements were all less than 1.0%, indicating that the precision of this method was good.

	$D_{10}(\mu m)$	$D_{50}(\mu m)$	D ₉₀ (µm)	Span	
1	47.04	77.32	124.54	1.00	1
2	47.59	77.60	126.58	1.02	
3	46.78	77.38	125.42	1.02	
4	46.98	78.18	126.83	1.02	
5	47.79	77.61	125.80	1.01	
6	46.55	78.20	126.96	1.03	
RSD/%	0.93	0.45	0.68	1.02	

Table S6 Repeatability of particle size determination method

Six samples of Vivitrol[®] were prepared in parallel, and each sample was repeatedly determined for 3 times. The RSD values were all less than 2.0%, which indicated that the method had good repeatability.

Molecular weight of GNM with d	lifferent PLGA
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Table S7 Molecular	weight by G	SNM prepared	from PLGA with	different molecul	lar weight
	0 1	1 1			0

	Mw	PD
GNM-PLGA (8,000 Da)	47695±273	1.52 ± 0.02
PLGA (8,000 Da)	83416±1557	1.73 ± 0.05
GNM-PLGA (140,000 Da)	81381±2475	1.93 ± 0.16
PLGA (140,000 Da)	140928 ± 774	1.83 ± 0.03

We prepared the GNM using PLGA which Mw is 63416±1557 Da, but the detected Mw in the resulted microsphere was only 27695±273 Da (Table S9). When the Mw of PLGA used in preparation increased to 140928±774 Da, the detected Mw of PLGA in GNM became similar with that of Vivitrol[®] as 81381±2475 Da, which meant that 42.25% molecular weight loss happened in the preparation process.

Morphology of microspheres with different solvent



Figure. S1 Morphology of microspheres when the solvent is DCM or EA. DCM: Dichloromethane, EA: ethyl acetate.

It showed significant difference in morphology with DCM or EA. Microspheres prepared using DCM seemed relatively round, no concavity but porous on the surface, and had apparent difference with that using EA.

Morphology of pressed microsphere before the water contact angle determined



Figure. S2 Morphology of pressed microsphere before the water contact angle determined

Table S8 Contact angle of microspheres before and after drug delivery

	Contact angle (°)
Blank Microspheres	62.19±0.40
GNM	55.93±0.38