

## Supplement Data

### N-Ethyl-2-pyrrolidinone-substituted Flavanols from White Tea Using Centrifugal Countercurrent Chromatography Off-Line ESI-MS Profiling and Semi-preparative Liquid Chromatography

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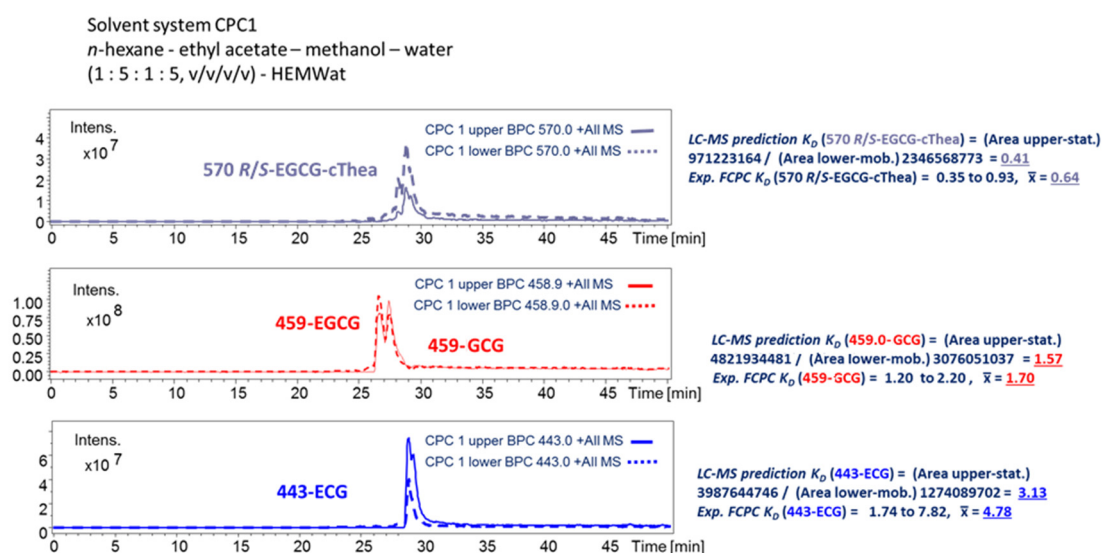
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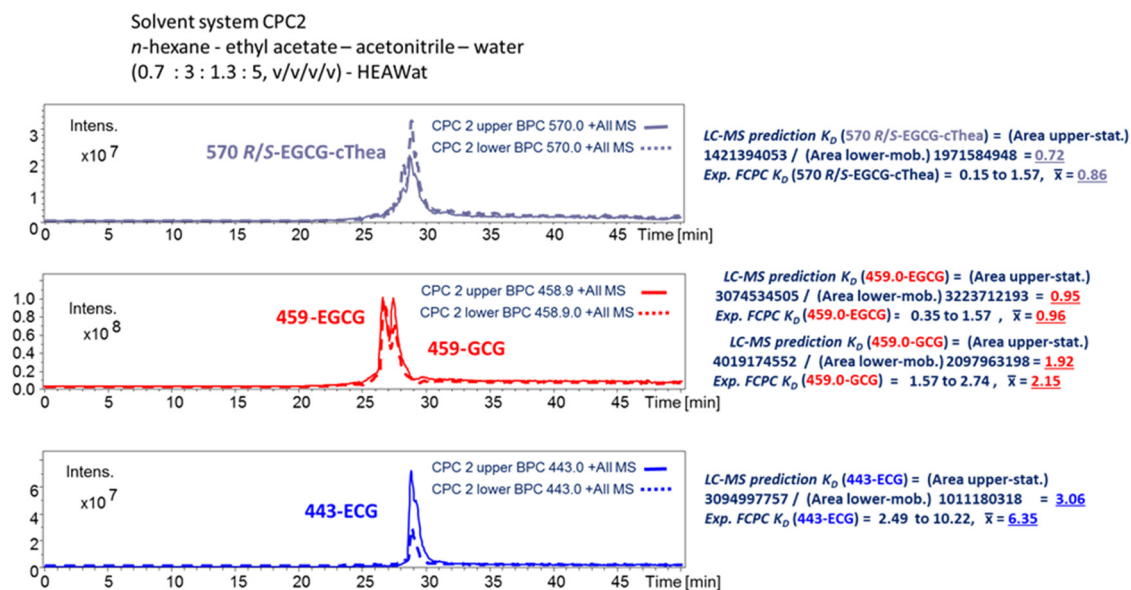
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**Supplement Figure S1a/1b.** Prediction of suitable CPC solvent systems for the white tea sample by LC-ESI-MS analysis of target metabolite distributions in the phase layers of shake-flask experiments by calculation of specific metabolite partition ratio values  $K_D$ .

Formula LC-ESI-MS prediction of partition ratio values : cf. results Table 1

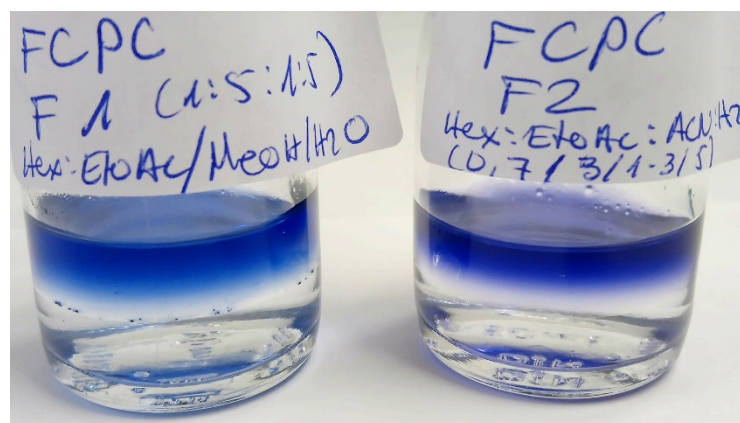
$K_D$ -value = peak area A upper phase / peak area A lower phase (Eq. 1) [Ito, 1996]





**Suppl. Figure S1a/1b:** Selected single ion traces (ESI pos. mode) for target metabolites in the phase layers and calculated  $K_D$ -values.

**Supplement Figure S2. Evaluation of the chosen biphasic solvent systems (CPC-1 and CPC-2) by use of Reichardt's-dye**



Reichardt's dye was suggested by Abbott & Kleiman (1991) for evaluation and classification of polarities of upper and lower phase layers in solvent systems being used for countercurrent chromatography (CCC) and therefore suitable in general for all-liquid separation techniques. Reichardt's dye absorbs light in the Vis-range ( $\lambda$  400-900 nm) depending on solute polarities.

The two biphasic solvent systems were prepared in small volumes being used for CPC-1 (HEMWat 1 : 5 : 1 : 5, v/v/v/v) and CPC-2 (HEAWat 0.7 : 3 : 1.3 : 5, v/v/v/v). 1 mg of Reichardt's dye was added for each shake flask experiment. The wavelength absorbance in the blue range of the upper layers indicated a similar non-polar character for the two CPC solvent systems. The principal difference in the solvent system composition is the exchange of methanol against acetonitrile. This change of a solvent

component was compensated by the ratios adjusted to the metabolite distributions in the respective phase layers (as seen by similar  $K_D$ -value predictions in Suppl. Fig. S1).

### Equations. Calculation of CPC-chromatographic separation parameters

In the performed CPC experiments, the chromatographic elution time was converted over *elution/retention volumes*  $V_R$  into their respective *partition ratio* values  $K_D$  (cf. **equations 2 - 7**). The  $K_D$ -based projection enables a better comparison between different liquid/liquid chromatography based machine designs such as *countercurrent chromatography* (CCC) [Grecco, 2019] and centrifugal partition chromatography (CPC). The experimental  $V_R$  values from the CPC runs for the tea metabolites were determined with high accuracy by the used *off-line* ESI-MS injection profiling of consecutive fractions as described below (also cf. 3.5).

$$\text{Retention volume } V_R = \text{elution time [min]} \times \text{flow rate [mL/ min]} \quad (\text{Eq. 2})$$

The  $S_F$  values of the used solvent systems were determined by equation (Eq. 3a-b) using  $V_C$  (200 mL), and  $V_M$  resulting in the  $S_F$  value measured at the hydrodynamic equilibrium.

$$V_S = (V_C - V_M) \quad (\text{Eq. 3a}) \quad S_F = V_S / V_C \times 100\% \quad (\text{Eq. 3b})$$

CPC-1:  $S_F = 84.5\%$

CPC-2:  $S_F = 67.0\%$

$V_S$  : retained experimental stationary phase volume

$V_C$  : CPC column volume/capacity (200 mL)

$V_M$  : volume of mobile phase take up to the coil at equilibrium of FCPC

$V_{CM}$ : volume with change of elution to extrusion-mode (*switch volume*)

$S_F$  : stationary phase retention

$K_D$ : partition ratio

The determined  $S_F$ -value in the experiment is corrected by the *extra column volume*  $V_{Ext}$  (10 mL) of the connecting periphery tubing in the CPC set-up, using equations (Eq. 4 - 6)

$$\text{Corrected } V_M = V_M - V_{ext} \quad (\text{Eq. 4})$$

CPC-1: *corr.*  $V_M = 31 \text{ mL} - 10 \text{ mL} = 21 \text{ mL}$

CPC-2: *corr.*  $V_M = 66 \text{ mL} - 10 \text{ mL} = 56 \text{ mL}$

$$\text{Corrected } V_S = V_C - \text{corrected } V_M \quad \text{with CPC column volume } V_C = 200 \text{ mL} \quad (\text{Eq. 5})$$

CPC-1: *corr.*  $V_S = 200 \text{ mL} - 21 \text{ mL} = 179 \text{ mL}$

CPC-2: *corr.*  $V_S = 200 \text{ mL} - 56 \text{ mL} = 144 \text{ mL}$

It is important to note that a high  $S_F$  value directly correlate to a higher resolution and efficiency of the CPC-separation.

$$\text{corrected } S_F = \text{corrected } V_S / V_C \quad (\text{Eq. 6})$$

CPC-1: *corr.*  $S_F = 179 \text{ mL} / 200 \text{ mL} \times 100\% = 89.5\%$

CPC-2: *corr.*  $S_F = 144 \text{ mL} / 200 \text{ mL} \times 100\% = 76.0\% \text{ mL}$

The metabolite and solvent system specific partition ratio  $K_D$  values in the CPC run were calculated by the equations (Eq. 7a, 7b) (results cf. Suppl. Table 1). [Berthod et al.]

During *elution-mode* :  $K_D = (V_R - \text{corrected } V_M) / \text{corrected } V_s$  (Eq. 7a) (cf. Fig. 3a, 3b)

In the *extrusion-mode* from begin of single phase recovery:

$$K_D = V_{CM} / (V_{CM} + V_C - V_R) \text{ [Berthod et al.]} \quad (\text{Eq. 7b}) \text{ (cf. Fig. 3a, 3b)}$$

The *separation factor*  $\alpha$  and *resolution factors*  $R_s$  depend on the distances between peaks and the peak widths that will be compared. The calculation of both factors depend on the determined  $K_D$  values.

$$\alpha = K_{D2} / K_{D1} \text{ (with } K_{D2} > K_{D1}) \quad (\text{Eq. 8}) \text{ (cf. Table 2)}$$

To calculate the *resolution factor* ( $R_s$ ) of the two peaks together with their respective  $K_D$  values, the measured peak widths at the baseline were used, as seen in the next formula:

$$R_s = 2 (K_{D2} - K_{D1}) / (W_2 + W_1) \quad (\text{Eq. 9}) \text{ (cf. Table 2)}$$

$W_n$ : peak width at baseline

## **References**

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