

Article

Bile-Acid-Appended Triazolyl Aryl Ketones: Design, Synthesis, In Vitro Anticancer Activity and Pharmacokinetics in Rats

Devesh S. Agarwal ¹, Samrat Mazumdar ², Kishan S. Italiya ², Deepak Chitkara ² and Rajeev Sakhuja ^{1,*}

¹ Department of Chemistry, Birla Institute of Technology and Science, Pilani 333 031, India; deveshagarwal88@gmail.com

² Department of Pharmacy, Birla Institute of Technology and Science, Pilani 333 031, India; samratm64@gmail.com (S.M.); italiyakishan2010@gmail.com (K.S.I.); deepak.chitkara@pilani.bits-pilani.ac.in (D.C.)

* Correspondence: rajeev.sakhuja@pilani.bits-pilani.ac.in

Abstract: A library of bile-acid-appended triazolyl aryl ketones was synthesized and characterized by detailed spectroscopic techniques such as ¹H and ¹³C NMR, HRMS and HPLC. All the synthesized conjugates were evaluated for their cytotoxicity at 10 μM against MCF-7 (human breast adenocarcinoma) and 4T1 (mouse mammary carcinoma) cells. In vitro cytotoxicity studies on the synthesized conjugates against MCF-7 and 4T1 cells indicated one of the conjugate **6cf** to be most active against both cancer cell lines, with IC₅₀ values of 5.71 μM and 8.71 μM, respectively, as compared to the reference drug docetaxel, possessing IC₅₀ values of 9.46 μM and 13.85 μM, respectively. Interestingly, another compound **6af** (IC₅₀ = 2.61 μM) was found to possess pronounced anticancer activity as compared to the reference drug docetaxel (IC₅₀ = 9.46 μM) against MCF-7. In addition, the potent compounds (**6cf** and **6af**) were found to be non-toxic to normal human embryonic kidney cell line (HEK 293), as evident from their cell viability of greater than 86%. Compound **6cf** induces higher apoptosis in comparison to **6af** (46.09% vs. 33.89%) in MCF-7 cells, while similar apoptotic potential was observed for **6cf** and **6af** in 4T1 cells. The pharmacokinetics of **6cf** in Wistar rats showed an MRT of 8.47 h with a half-life of 5.63 h. Clearly, these results suggest **6cf** to be a potential candidate for the development of anticancer agents.

Keywords: bile acid; anticancer; cytotoxicity; apoptosis; pharmacokinetic study



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1. Introduction

Cancer is presently a major health concern around the globe, leading to an alarming increase in the number of deaths, after cardiovascular disease, which is further expected to elevate to 12 million by 2030, as per WHO report [1–4]. Among all, breast cancer is the second most treacherous and common form of malignant tumor found in 23% of all forms of female cancers [5,6]. Cancer treatment procedures, such as hormone and radiation therapy, immunotherapy, combination chemotherapy and surgery, have been implemented to achieve reasonable success in this battle of mankind against cancer [7]. Among these, chemotherapy has proved to be one of the most promising pathways to overcome cancer; however, concerns such as selectivity, resistance and bioavailability of existing chemotherapeutic agents and associated side effects limit its exemplification as an ideal cancer-treating procedure [8]. Thus, the search for selective anticancer agents with lower side effects and better efficacy remains a prime target of medicinal chemists around the globe.

In this realm, some of the endogenous steroids and secondary bile acids have proven their repute as valuable cytotoxic agents [9]. For example, tauroursodeoxycholic acid (TUDCA) and ursodeoxycholic acid (UDCA) have shown significant apoptotic effects on various cancer cell lines [10–13]. Ursodeoxycholic acid (UDCA) has exhibited remarkable cytotoxicity against human oral squamous carcinoma (HSC-3), cultured animal/human

tumor cells and HepG2 human hepatoma cells in combination with doxorubicin and also prevented colorectal adenoma recurrence [14,15]. Bile acids have also served as handy tools for the prodrug approach. For example, dihydroartemisinin–bile-acid hybridization has resulted in enhancement of dihydroartemisinin anticancer activity [16]. Strikingly, considerable research has been fueled toward developing steroidal heterocycles, in view of their broad spectrum of biological activities and added advantage of hydrophobic steroidal behavior capable of interacting with cell membranes [17].

In particular, 1,2,3-triazole provides favorable properties to binding molecular targets in a biological environment due to its metabolic stability and hydrogen-bonding capability [18]. Thus, the synthesis of diverse triazolyl steroids has received special interest due to a wide range of pharmacological properties such as anticancer [17,19] and antimicrobial [20,21]. Triazole-derived terpenoids/steroids such as oleanolic acid [22,23], betulinic acid [24–28], eteinic acid [29], nor-testosterone [30], androst-5-ene [31], estrone [29,32], 5 α -androstane [33,34], cholesterol [20,21,35], 2-methoxyestradiol [36] and pregnenanes [37] have showcased interesting cytotoxic behavior against a variety of cancer cell lines (Figure 1, I–IV). However, bile-acid-appended triazoles have rarely been explored for their cytotoxic efficacy [38]. Drasar and coworkers documented one such report that discloses the synthesis and cytotoxicity of two ribbon-type pyridyl-based triazolyl cholic acid dimers, of which the ester analog demonstrated significant cytotoxic activity in low micromolar concentrations against lymphoblastic (CCRF-CEM) and myeloid leukemia (K562) cell lines (Figure 1, V) [29]. Perrone’s group reported cytotoxic studies on C-24-triazolyl-linked bile-acid–nucleoside conjugates, where a chenodeoxycholic-acid-linked deoxyadenosine derivative exhibited an IC₅₀ value of 16.2 \pm 2.2 μ M against leukemia cell line (K562) (Figure 1, VI) [39]. The same group reported the synthesis of triazolyl-linked bile-acid–deoxyadenosine conjugates and evaluated their cytotoxic activity against two leukemia cell lines (Jurkat and K562), colon cancer cell line (HCT116), ovarian cancer cell line (A2780) and human fibroblast cell line. The best compound in this series exhibited an IC₅₀ value of 8.51 \pm 4.05 μ M and 10.47 \pm 2.64 μ M against two leukemia cancer cell lines K562 and Jurkat, respectively (Figure 1, VII) [40].

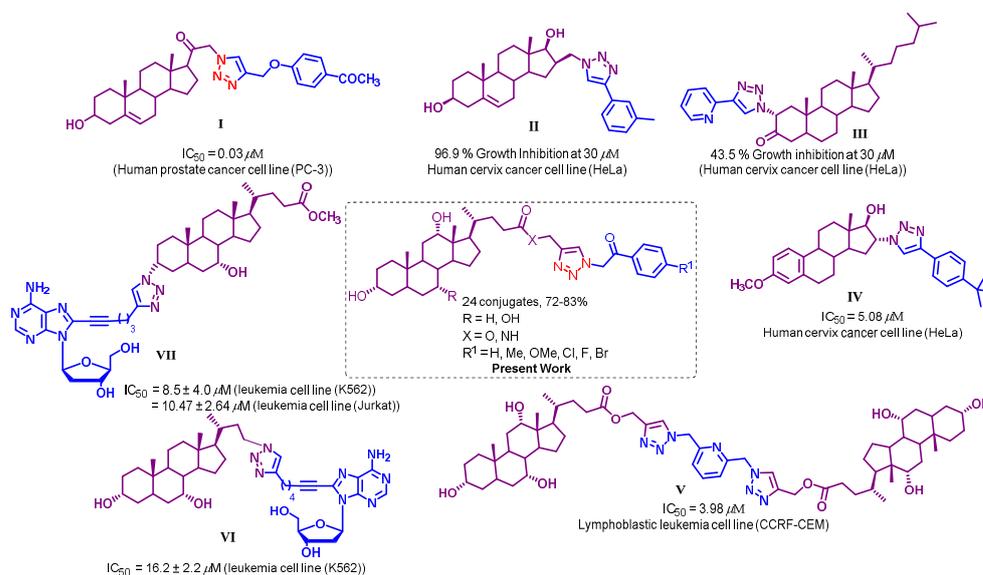


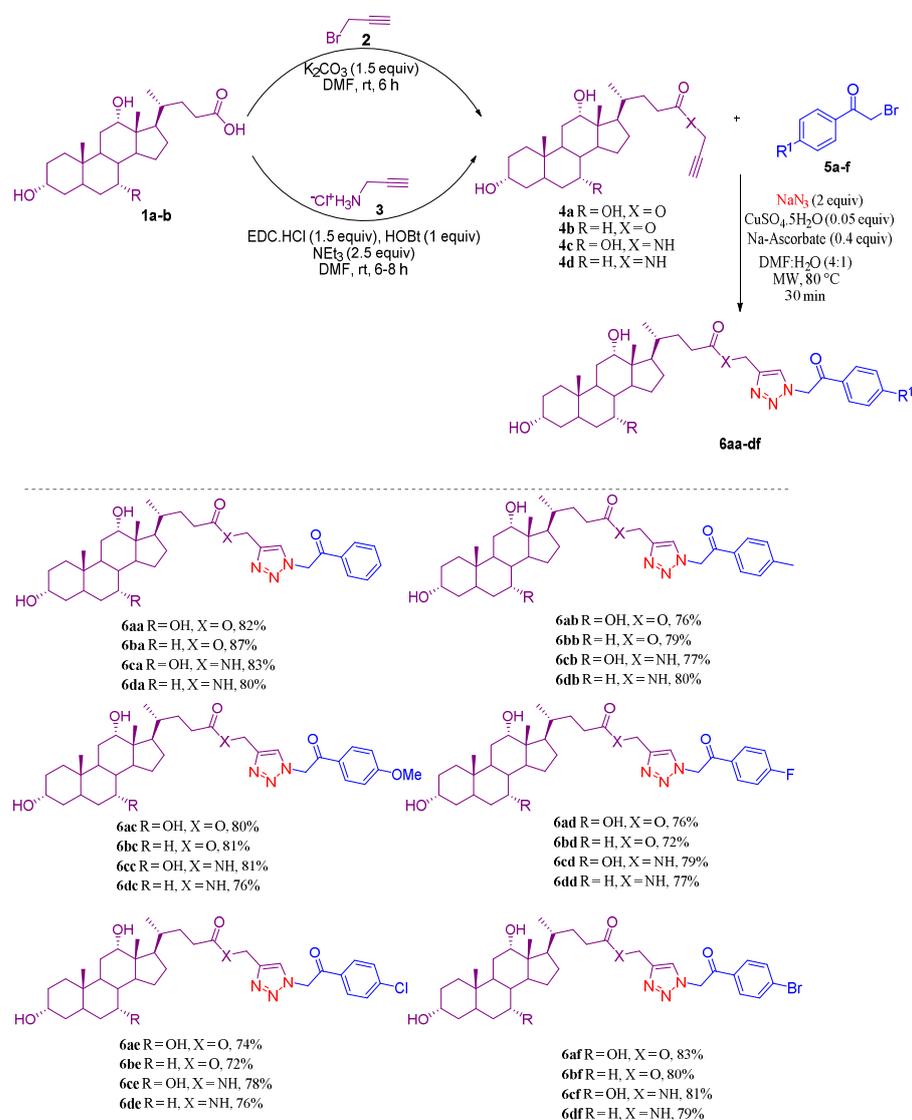
Figure 1. Representative examples of triazole-derived steroids and bile acids as anticancer agents.

In continuation to our program for the synthesis of C-24-functionalized bile acids as anticancer agents [41–43] and the aforementioned properties of triazolyl steroids, we present a synthetic approach for the synthesis of bile-acid-appended triazolyl aryl ketones. Their cytotoxic potency was examined against two breast cancer cell lines (MCF-7 and 4T1). In addition, in vivo pharmacokinetic study was also performed.

2. Results and Discussion

2.1. Chemistry

From the outset of the proposed work, the synthesis of targeted bile-acid-appended triazolyl aryl ketones commenced with the preparation of cholic acid and deoxycholic acid propargyl esters (**4a,b**) and amides (**4c,d**) by coupling cholic acid (**1a**)/deoxycholic acid (**1b**) with propargyl bromide (**2**)/propargyl amine hydrochloride (**3**), respectively, using reported single-step protocols (Scheme 1) [41,44]. Thereafter, a Cu-catalyzed multicomponent reaction between CA and DCA propargyl esters/amides (**4a–d**) with α -bromoacetophenones (**5a–f**) and sodium azide in aqueous DMF under microwave irradiation at 80 °C comfortably afforded the desired bile-acid-appended triazolyl aryl ketones (**6aa–6df**) in excellent yields (Scheme 1). All the synthesized compounds were completely characterized on the basis of ^1H NMR, ^{13}C NMR and HRMS. As a representative example, the assignment of hydrogen and carbons in **6aa** was also performed using COSY, HSQC and HMBC (SI). The ^1H and ^{13}C NMR assignments for the representative proton/carbon signals of **6aa** are given in Table 1, and selective correlations are showcased in Figure 2 on the basis of the ^1H , ^{13}C HMBC spectrum, please see Supplementary Materials.



Scheme 1. Synthesis of bile-acid-appended triazolyl aryl ketones.

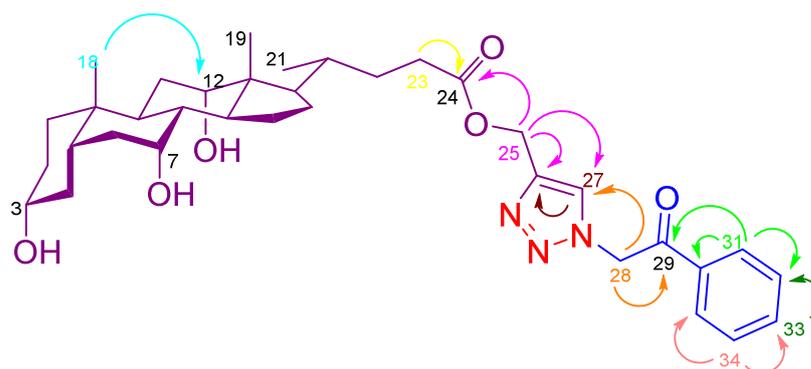


Figure 2. Selective correlations based on ^1H , ^{13}C -HMBC spectrum of **6aa**.

Table 1. ^1H and ^{13}C NMR assignments for the representative proton/carbon signals of **6aa**.

S. No.	Labeling	^1H NMR	^{13}C NMR
1.	18	0.57 (s, 3H)	12.8
2.	19	0.80 (s, 3H)	23.1
3.	21	0.92 (d, $J = 6.2$ Hz, 3H)	17.4
4.	3	3.17 (d, $J = 5.1$ Hz, 1H)	70.9
5.	7	3.61 (brs, 1H)	66.7
6.	12	3.78 (d, $J = 3.8$ Hz, 1H)	71.5
7.	OH	4.33 (d, $J = 4.3$ Hz, 1H)	-
8.	OH	4.12 (d, $J = 3.8$ Hz, 1H)	-
9.	OH	4.01 (d, $J = 3.3$ Hz, 1H)	-
10.	25 (COO-CH ₂)	5.18 (s, 2H)	57.5
11.	28 (N-CH ₂)	6.20 (s, 2H)	56.3
12.	27 (Triazole-H)	8.11 (s, 1H)	126.9
13.	24 (COO)	-	173.6
14.	29 (CO)	-	192.5

2.2. Biological Evaluation

2.2.1. Cytotoxic Activity

All the synthesized compounds (**6aa–df**) were studied for their anticancer activity in two cancer lines viz. human breast adenocarcinoma (MCF-7) and mouse mammary carcinoma (4T1) cells at 10 μM concentration (Table 2). Most of the compounds showcased moderate-to-good activity against both cell lines as compared to the standard drug (docetaxel). Among all, compound **6af** was found to be the most active (26.52% cell viability at 10 μM) against MCF-7 cells. In addition, compounds **6bf** and **6cf** were also found to be active against human breast cancer cell line (MCF-7), exhibiting cell viabilities of 44.43% and 37.53%, respectively, at 10 μM . In 4T1 cells, **6cf** exhibited 49.27% cell viability at 10 μM . Triazolyl aryl ketones appended with cholic acid at the expense of an ester bond (**6aa**, **6ab**, **6ac**, **6ad**, **6af**) were found to be more active than their corresponding amide surrogates and deoxycholic acid ester/amide conjugates, except **6be** and **6ce**. In general, *para*-substitution (Me, OMe, F, Cl, Br) on the aryl ketone showcased lower cell viability as compared to the unsubstituted analogs. The analogs containing electron-withdrawing groups (F, Cl, Br) on aryl ketone were found to be more active as compared to the ones containing electron-donating groups (Me, OMe). Among halo-substituted analogs, triazolyl bromo-substituted aryl ketones appended to cholic acid and deoxycholic acid via an ester bond (**6af**, **6bf**) and amide bond (**6cf**) were found to be more active in inhibiting the growth of MCF-7 cells. While in 4T1 cells, triazolyl bromo-substituted aryl ketones appended to cholic acid and deoxycholic acid via an amide bond (**6cf**, **6df**) were found to be more active. In addition, adsorption, distribution, metabolism and excretion (ADME) properties and physiochemical properties of the synthesized analogs were calculated using molinspiration cheminformatics [45,46]. Additionally, percentage absorption and drug-likeness model

score were also calculated using the reported formula [47] and Molsoft [48], respectively. As indicated by the TPSA values (between 60 and 160 Å²), all the analogs (**6aa–df**) possessed better intestinal absorption ability over the blood–brain barrier (BBB) penetration power. Similarly, all the analogs (**6aa–df**) possessed a positive drug-likeness score between 0.60 and 1.14, indicating them to be ideal drug candidates. For instance, the most active analogs **6af**, **6bf** and **6cf** were found to possess relatively good drug-likeness scores of 0.94, 0.85 and 0.88, respectively.

Table 2. In vitro cytotoxicity of compounds (**6aa–6df**) against two different cancer cell lines and normal human kidney cell line.

Com. No.	% Cell Viability at 10 µM			milog P ^d	TPSA ^e	% ABS ^f	Drug-Likeness Score
	MCF-7 ^a	4T1 ^b	HEK 293 ^c				
6aa	77.32	74.32	86.08	4.54	134.78	62.50	0.75
6ba	91.90	97.81	88.92	5.45	114.55	69.48	0.65
6ca	92.06	85.16	90.13	3.89	137.57	61.53	0.69
6da	91.20	99.56	97.86	4.81	117.34	68.51	0.60
6ab	57.35	72.93	87.63	4.99	134.78	62.50	0.82
6bb	67.60	85.69	89.17	5.90	114.55	69.48	0.72
6cb	61.86	96.74	86.00	4.34	137.57	61.53	0.75
6db	78.69	53.67	89.43	5.26	117.34	68.51	0.66
6ac	55.59	80.02	87.88	4.59	144.01	59.31	0.98
6bc	69.62	68.40	90.77	5.51	123.78	66.29	0.89
6cc	95.36	56.79	96.75	3.95	146.81	58.35	0.96
6dc	68.52	64.92	86.53	4.87	126.58	65.32	0.88
6ad	66.48	47.70	86.79	4.70	134.78	62.50	1.07
6bd	79.62	52.35	89.37	5.62	114.55	69.48	0.97
6cd	88.72	70.65	97.51	4.06	137.57	61.53	1.01
6dd	73.76	89.90	81.61	4.97	117.34	68.51	0.92
6ae	79.62	58.92	96.21	5.21	134.78	62.50	1.14
6be	68.52	76.01	88.37	6.13	114.55	69.48	1.05
6ce	69.16	50.59	92.69	4.57	137.57	61.53	1.09
6de	86.85	62.89	90.45	5.49	117.34	68.50	1.01
6af	26.52	55.91	88.20	5.34	134.78	62.50	0.94
6bf	44.43	60.86	86.82	6.26	114.55	69.48	0.85
6cf	37.53	49.27	87.82	4.70	137.57	61.53	0.88
6df	86.45	48.38	93.46	5.62	117.34	68.51	0.79
DTX^g	46.47	56.88	-	-	-	-	-

^a Human breast adenocarcinoma, ^b Mouse mammary carcinoma, ^c Human embryonic kidney 293 cells, ^d Logarithm of compound partition coefficient between n-octanol and water, ^e Topological polar surface area, ^f Percentage absorption calculated using the formula %ABS = 109 – (0.345 × TPSA), ^g Docetaxel.

Further, all the compounds (**6aa–df**) tested on a normal human embryonic kidney cell line (HEK 293) indicated cell viability to be greater than 85% and thus were found to be non-toxic against normal cells (Table 2). In particular, the most active derivatives **6af**, **6bf** and **6cf** possessing cell viability of 88.20, 86.82 and 87.82 appeared to be quite safer on normal human embryonic kidney cells.

IC₅₀ values of the most potent compounds **6af**, **6bf** and **6cf** were further evaluated against the two cancer cell lines by employing MTT assay (Table 3). Interestingly, **6af**, **6bf** and **6cf** showed IC₅₀ values in the range of 2.61–18.26 μM against the MCF-7 cancer cell line and 8.76–12.84 μM against the 4T1 cancer cell line. Compounds **6af** (IC₅₀ = 2.6 μM) and **6cf** (IC₅₀ = 5.71 μM) were found to possess pronounced anticancer activity as compared to the reference drug docetaxel (IC₅₀ = 9.46 μM) against human breast adenocarcinoma (MCF-7), while all the three compounds (**6af**, **6bf**, **6cf**) were found to be more active with respect to docetaxel (IC₅₀ = 13.85 μM) against rat mammary carcinoma (4T1). Further, these compounds did not induce cell death in HEK 293 cells.

Table 3. IC₅₀ (μM) values of the active compounds in two different cancer cell lines.

Compound	IC ₅₀ (μM)	
	MCF-7 ^a	4T1 ^b
6af	2.61 ± 0.70	12.84 ± 1.80
6bf	18.26 ± 1.48	9.68 ± 1.59
6cf	5.71 ± 1.00	8.76 ± 1.29
DTX	9.46 ± 0.98	13.85 ± 1.07

^a Human breast adenocarcinoma, ^b Mouse mammary carcinoma.

2.2.2. Apoptotic Study

The apoptotic effect of **6af** and **6cf** was evaluated by the Annexin V/PI staining method. Following treatment of MCF-7 cells with **6af** and **6cf** at 2.61 μM and 5.71 μM, respectively, it was observed that compound **6cf** was capable of inducing higher apoptosis in comparison to **6af** (46.09% vs. 33.89%) (Figure 3a,b,e). Meanwhile, in 4T1 cells, both **6af** (at 12.84 μM) and **6cf** (at 8.76 μM) produced total apoptosis of 19.02% and 19.56%, indicating similar apoptotic potential in 4T1 cells (Figure 3c,d,e).

Of the two compounds, the most active compound **6cf** was chosen for the in vivo pharmacokinetic study.

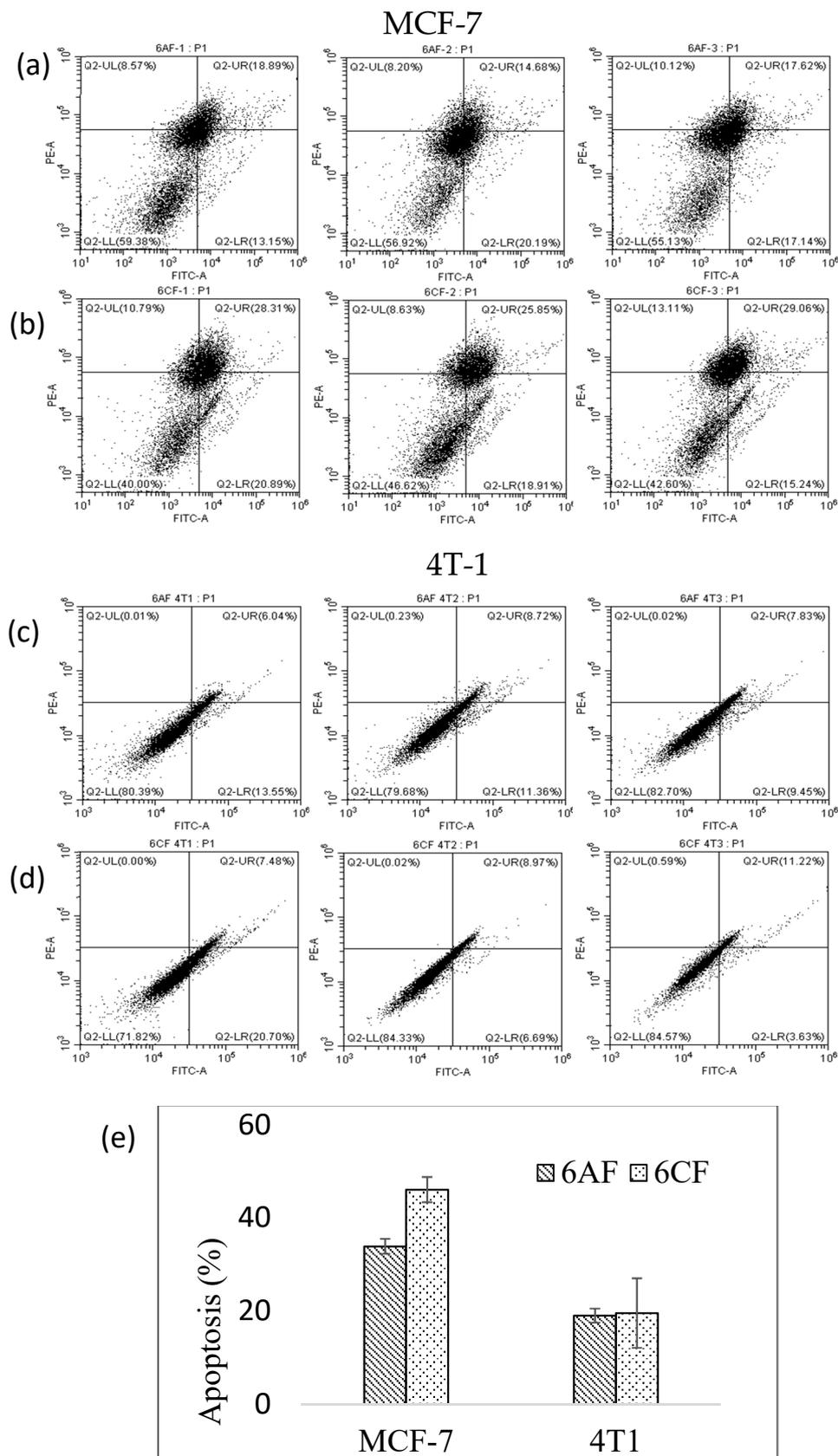


Figure 3. Apoptosis assay of **6af** and **6cf** in MCF-7 and 4T1 cells. (a,b) Flow cytometry plots for apoptosis in MCF-7 cells treated with **6af** and **6cf**, respectively. (c,d) Flow cytometry plots for apoptosis in 4T1 cells treated with **6af** and **6cf**, respectively. Upper left (necrotic cells), lower left (live cells), lower right (early apoptotic cells) and upper right (late apoptotic cells). (e) Graph showing the apoptosis (%) induced by **6af** and **6cf** in MCF-7 and 4T1 cells.

2.2.3. Pharmacokinetic Study of 6cf

The relationship between the pharmacokinetic parameters and in vitro cytotoxicity study could be useful in determining the starting dose for the initial clinical trials for anticancer drugs. The compound **6cf** was found to have an IC_{50} (μM) of 5.71 and 8.76 μM in MCF-7 and 4T1 cells, respectively. The pharmacokinetic study was performed at a dose of 10 mg/kg i.v. bolus in rats that showed the initial concentration of 1752.69 ng/mL ($\sim 2.56 \mu M$) with a half-life of 5.63 h. The mean plasma concentration–time profile of **6cf** after a single dose of 10 mg/kg (intravenously) in rats is presented in Figure 4. Different pharmacokinetic parameters were evaluated by a non-compartmental model approach using Phoenix WinNonlin software as shown in Table 4. The initial concentration (C_0) was found to be 1752.69 ± 66.52 ng/mL. The AUC_{0-last} calculated based on the trapezoidal rule was found to be 1995.306 ± 87.43 ng.h/mL. The mean residence time (MRT) was found to be 8.47 ± 0.96 h. The **6cf** half-life was found to be 5.63 ± 0.54 h [49]. Although it may not be feasible to predict the in vivo concentrations at the tumor site from the plasma concentrations, the pharmacokinetic data provide initial insights into the mean residence time of the drug candidate and may be useful in predicting the dose relationship with the pharmacological/toxic effect after in vivo assessment in the tumor models. Thus, further assessment in tumor models to advance this molecule is warranted.

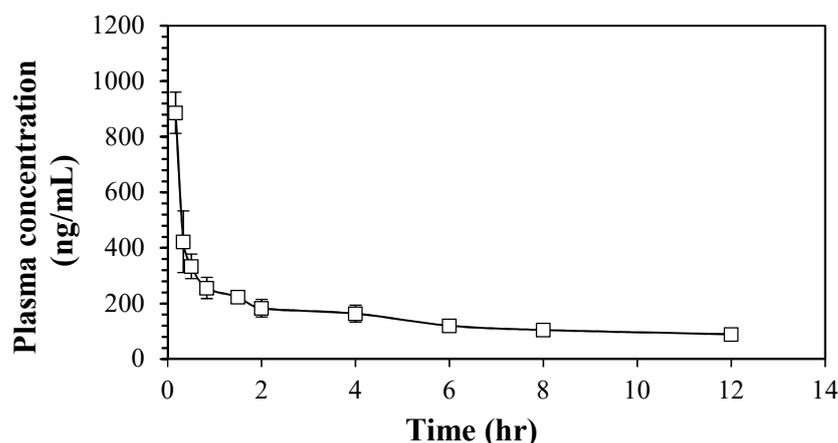


Figure 4. The pharmacokinetic profile for **6cf** in rat plasma after i.v. bolus at dose of 10 mg/kg administration to rat.

Table 4. The non-compartmental pharmacokinetic parameters for **6cf** in rat plasma after i.v. bolus at dose of 10 mg/kg administration to rat.

Parameters	6cf (Mean \pm SEM)
Initial Concentration, C_0 (ng/mL)	1752.69 ± 66.52
Half-Life, $t_{1/2}$ (h)	5.63 ± 0.539
Elimination Rate Constant, K_e (1/h)	0.13 ± 0.01
Area Under the Curve (0 to 12 h), AUC_{0-last} (ng.h/mL)	1995.306 ± 87.43
Area Under the Curve (0 to infinity), $AUC_{0-\infty}$ (ng.h/mL)	2690.50 ± 113.20
Area Under the First Moment Curve (0 to 12 h), $AUMC_{0-last}$ (ng.h/mL)	8155.94 ± 311.78
Area Under the First Moment Curve (0 to infinity), $AUMC_{0-\infty}$ (ng.h/mL)	22276.39 ± 2023.334
Mean Residence Time, MRT (h)	8.47 ± 0.96

3. Materials and Methods

All the chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA), Alfa Aesar (Haverhill, MA, USA) and Spectrochem India Pvt. Ltd. (Mumbai, India) and used without further purification. The solvents were purchased from Merck (Burlington, MA,

USA) and were distilled and dried before use. Nuclear magnetic resonance spectra were recorded on Bruker (Zurich, Switzerland) 400 spectrometer. The ^1H NMR experiments were reported in δ units, parts per million (ppm), and were measured relative to residual chloroform (7.26 ppm) or $\text{DMSO-}d_6$ (2.5 ppm) in the deuterated solvent. The ^{13}C NMR spectra were reported in ppm relative to deuteriochloroform (77.0 ppm) or $\text{DMSO-}d_6$ (39.5 ppm). All coupling constants J were reported in Hz. The following abbreviations were used to describe peak splitting patterns when appropriate: s = singlet, d = doublet, t = triplet, dd = doublet of doublet, m = multiplet and brs = broad singlet. Melting points were determined on a capillary point apparatus equipped with a digital thermometer and were uncorrected. Reactions were monitored by using thin-layer chromatography (TLC) on 0.2 mm silica gel F254 plates (Merck). The chemical structures of final products were confirmed by a high-resolution ESI/APCI hybrid quadrupole time-of-flight mass spectrometer. High-resolution mass spectrometry (HRMS) was performed with a Waters SYNAPT G2 HDMS instrument using time-of-flight (TOF-MS) with ESI/APCI-hybrid quadrupole. The purity of final products was confirmed by high-performance liquid chromatography (HPLC), using the following chromatographic conditions: liquid chromatographic conditions, a Thermo Fisher Rapid Separation (RS) UHPLC System (Ultimate 3000, Waltham, MA, USA) equipped with a pump (LPG-3400SD), Diode Array Detector (DAD) (DAD-3000, Thermo Fisher, Waltham, MA, USA) and autosampler (ACC-3000T, Thermo Fisher, Waltham, MA, USA) were used for purity analysis. The UHPLC system was equilibrated for approximately 40 min before beginning the sample analysis. Control of hardware and data handling was performed using Chromeleon software version 7.2 SR4 (Thermo Fisher, Waltham, MA, USA). Column: Inertsil[®] (GL Sciences, Tokyo, Japan) ODS C18 column (250 \times 4.6 mm, 5 μm). Mobile phase: ACN: water (95:05 % *v/v*); flow rate: 1 mL/min; detection wavelength: 259 nm; retention time: 3–5 min.

3.1. General Procedure for the Synthesis of CA and DCA Propargyl Amides (4c,d)

To a stirred solution of bile acid (CA (**1a**) or DCA (**1b**), 2.0 g, 1 equiv) in DMF (10 mL), triethyl amine (2.5 equiv) was added at 0 $^\circ\text{C}$, and subsequently EDC.HCl (1.5 equiv) and HOBT (1 equiv) were added. The reaction mixture was stirred for 15 min at 0 $^\circ\text{C}$, after which propargyl amine hydrochloride (1.5 equiv) was added. The reaction was stirred at room temperature for 6–8 h and was monitored by TLC. After the completion of the reaction, the reaction mixture was poured over crushed ice, and the resulted precipitate was filtered, washed with cold water, recrystallized from ethyl acetate/hexanes and triturated with diethyl ether to afford bile acid propargyl amide (**4c,d**), please see Supplementary Materials.

(4*R*)-*N*-(Prop-2-yn-1-yl)-4-((3*R*,7*R*,10*S*,12*S*,13*R*)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1*H*-cyclopenta[*a*]phenanthren-17-yl)pentanamide (**4c**): White solid; Yield: 90% (1.96 g); mp: 277–279 $^\circ\text{C}$ (Lit. [41] 276–278 $^\circ\text{C}$); ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 8.23 (t, J = 5.5 Hz, 1H, NH_{Amide}), 4.36 (d, J = 4.3 Hz, 1H, OH_{CA}), 4.13 (d, J = 3.5 Hz, 1H, OH_{CA}), 4.04 (d, J = 3.4 Hz, 1H, OH_{CA}), 3.82 (dd, J = 5.6, 2.5 Hz, 2H), 3.78 (d, J = 3.4 Hz, 1H, H-12_{CA}), 3.64–3.57 (m, 1H, H-7_{CA}), 3.24–3.14 (m, 1H, H-3_{CA}), 3.08 (t, J = 2.5 Hz, 1H, $\text{CH}_{\text{Alkyne}}$), 2.25–2.12 (m, 2H), 2.04–1.92 (m, 2H), 1.84–1.59 (m, 6H), 1.48–1.06 (m, 14H), 0.92 (d, J = 6.4 Hz, 3H, Me-21_{CA}), 0.81 (s, 3H, Me-19_{CA}), 0.58 (s, 3H, Me-18_{CA}); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 172.9 (C=O), 81.9, 73.2, 71.5 (C-12_{CA}), 70.9 (C-3_{CA}), 66.7 (C-7_{CA}), 46.6, 46.2, 42.0, 41.8, 35.8, 35.6, 35.3, 34.9, 32.7, 32.0, 30.9, 29.0, 28.2, 27.8, 26.7, 23.3, 23.1 (C-19_{CA}), 17.6 (C-21_{CA}), 12.8 (C-18_{CA}).

(4*R*)-4-((3*R*,10*S*,12*S*,13*R*)-3,12-Dihydroxy-10,13-dimethylhexadecahydro-1*H*-cyclopenta[*a*]phenanthren-17-yl)-*N*-(prop-2-yn-1-yl)pentanamide (**4d**): White solid; Yield: 89% (1.94 g); mp: 182–184 $^\circ\text{C}$ (Lit. [41] 184–186 $^\circ\text{C}$); ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 8.23 (t, J = 5.5 Hz, 1H, NH_{Amide}), 4.50 (brs, 1H, OH_{DCA}), 4.22 (brs, 1H, OH_{DCA}), 3.82 (dd, J = 5.6, 2.5 Hz, 2H), 3.79 (d, J = 2.7 Hz, 1H, H-12_{DCA}), 3.47 (brs, 1H, H-3_{DCA}), 3.09 (t, J = 2.5 Hz, 1H, $\text{CH}_{\text{Alkyne}}$), 2.13–1.95 (m, 2H), 1.83–1.74 (m, 2H), 1.72–1.42 (m, 8H), 1.39–1.06 (m, 14H), 0.91 (d, J = 6.4 Hz, 3H, Me-21_{DCA}), 0.85 (s, 3H, Me-19_{DCA}), 0.59 (s, 3H, Me-18_{DCA}); ^{13}C NMR (100 MHz, $\text{DMSO-}d_6$) δ 172.8 (C=O), 81.9, 73.2, 71.5 (C-12_{DCA}), 70.4 (C-3_{DCA}), 47.9, 46.7,

46.4, 42.1, 36.7, 36.1, 35.6, 35.5, 34.3, 33.4, 32.6, 32.0, 30.7, 29.1, 28.2, 27.7, 27.4, 26.6, 24.0, 23.6 (C-19_{DCA}), 17.5 (C-21_{DCA}), 12.9 (C-18_{DCA}).

3.2. General Procedure for the Synthesis of Bile-Acid-Appended Triazolyl Aryl Ketones (6aa–df)

Bile acid propargyl ester/propargyl amide (**4a–d**) (100 mg, 1 equiv), NaN₃ (2 equiv), CuSO₄·5H₂O (0.05 equiv), sodium ascorbate (0.4 equiv) and substituted α -bromo acetone/phenacyl bromide (**5a–f**) (2 equiv) were added in a microwave vial containing DMF:H₂O (4 mL:1 mL) mixture. The reaction mixture was stirred under microwave irradiation for 30 min at 80 °C, and the progress of the reaction was monitored by TLC (MeOH:DCM, 1% *v/v*). After the completion of the reaction, the mixture was quenched by adding crushed ice. The aqueous layer was extracted using ethyl acetate (2 × 20 mL). The combined organic layer was dried over anhydrous sodium sulfate, concentrated under reduced pressure and subjected to flash column chromatography (SiO₂ (100–200 mesh), DCM:MeOH, 99:1 *v/v*) to yield pure bile-acid-appended triazolyl aryl ketone (**6aa–6df**), please see Supplementary Materials.

(1-(2-Oxo-2-phenylethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6aa**): Off-white solid; Yield: 82% (0.111 g); mp: 108–109 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.11 (s, 1H, H_{Triazole}), 8.10–8.03 (m, 2H, H_{Ar}), 7.80–7.70 (m, 1H, H_{Ar}), 7.66–7.58 (m, 2H, H_{Ar}), 6.20 (s, 2H), 5.18 (s, 2H), 4.33 (d, *J* = 4.3 Hz, 1H, OH_{CA}), 4.12 (d, *J* = 3.8 Hz, 1H, OH_{CA}), 4.01 (d, *J* = 3.3 Hz, 1H, OH_{CA}), 3.78 (d, *J* = 3.8 Hz, 1H, H-12_{CA}), 3.61 (brs, 1H, H-7_{CA}), 3.17 (d, *J* = 5.1 Hz, 1H, H-3_{CA}), 2.43–2.27 (m, 2H), 2.23–2.13 (m, 2H), 1.97–1.63 (m, 7H), 1.47–1.12 (m, 13H), 0.92 (d, *J* = 6.2 Hz, 3H, Me-21_{CA}), 0.80 (s, 3H, Me-19_{CA}), 0.57 (s, 3H, Me-18_{CA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 192.5 (C=O), 173.6 (C=O), 142.4, 134.7, 134.6, 129.5, 128.6, 126.9, 71.5 (C-12_{CA}), 70.9 (C-3_{CA}), 66.7 (C-7_{CA}), 57.5, 56.3, 46.6, 46.2, 42.0, 41.8, 35.8, 35.5, 35.3, 34.9, 31.1, 30.9, 29.0, 27.7, 26.7, 23.1 (C-19_{CA}), 17.4 (C-21_{CA}), 12.8 (C-18_{CA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical formula C₃₅H₅₀N₃O₆⁺ 608.3694: found: 608.3685; Anal. RP-HPLC *t*_R = 3.773 min, purity 95.46%; [α]_D²⁰ = +20 (c 1.0, MeOH).

(1-(2-Oxo-2-phenylethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6ba**): Off-white solid; Yield: 87% (0.119 g); mp: 143–144 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.11 (s, 1H, H_{Triazole}), 8.07 (d, *J* = 7.9 Hz, 2H, H_{Ar}), 7.75 (t, *J* = 7.4 Hz, 1H, H_{Ar}), 7.65–7.57 (d, *J* = 7.5 Hz, 2H, H_{Ar}), 6.20 (s, 2H), 5.17 (s, 2H), 4.53 (d, *J* = 4.2 Hz, 1H, OH_{DCA}), 4.24 (d, *J* = 3.9 Hz, 1H, OH_{DCA}), 3.78 (brs, 1H, H-12_{DCA}), 3.49 (brs, 1H, H-3_{DCA}), 2.46–2.30 (m, 2H), 2.25–2.13 (m, 2H), 1.74–1.44 (m, 9H), 1.37–1.14 (m, 13H), 0.90 (d, *J* = 6.3 Hz, 3H, Me-21_{DCA}), 0.84 (s, 3H, Me-19_{DCA}), 0.57 (s, 3H, Me-18_{DCA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 192.6 (C=O), 173.6 (C=O), 142.3, 134.7, 134.5, 129.5, 128.6, 126.9, 71.5 (C-12_{DCA}), 70.4 (C-3_{DCA}), 57.5, 56.3, 47.9, 46.6, 46.5, 42.1, 36.7, 36.1, 35.6, 35.4, 34.3, 33.4, 31.1, 30.7, 29.0, 27.6, 27.4, 26.6, 23.6 (C-19_{DCA}), 17.3 (C-21_{DCA}), 12.9 (C-18_{DCA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical formula C₃₅H₅₀N₃O₅⁺ 592.3745 found: 592.3733; Anal. RP-HPLC *t*_R = 4.553 min, purity 94.55%; [α]_D²⁰ = +16 (c 1.0, MeOH).

(4R)-N-((1-(2-Oxo-2-phenylethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (**6ca**): Off-white solid; Yield: 83% (0.112 g); mp: 148–149 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.39 (t, *J* = 5.8 Hz, 1H, NH_{Amide}), 8.09–8.05 (m, 2H, H_{Ar}), 7.84 (s, 1H, H_{Triazole}), 7.77–7.72 (m, 1H, H_{Ar}), 7.61 (t, *J* = 7.7 Hz, 2H, H_{Ar}), 6.15 (s, 2H), 4.39 (brs, 1H, OH_{CA}), 4.33 (d, *J* = 5.7 Hz, 2H), 4.12 (d, *J* = 3.7 Hz, 1H, OH_{CA}), 4.03 (d, *J* = 3.5 Hz, 1H, OH_{CA}), 3.78 (d, *J* = 3.8 Hz, 1H, H-12_{CA}), 3.60 (brs, 1H, H-7_{CA}), 3.18 (s, 1H, H-3_{CA}), 2.19–2.10 (m, 2H), 2.05–1.94 (m, 2H), 1.80–1.60 (m, 6H), 1.49–1.15 (m, 14H), 0.93 (d, *J* = 6.1 Hz, 3H, Me-21_{CA}), 0.79 (s, 3H, Me-19_{CA}), 0.56 (s, 3H, Me-18_{CA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 192.7 (C=O), 173.2 (C=O), 145.6, 134.7, 134.6, 129.5, 128.6, 124.9, 71.5 (C-12_{CA}), 70.9 (C-3_{CA}), 66.7 (C-7_{CA}), 56.2, 54.7, 46.6, 46.2, 41.9, 41.8, 35.6, 35.3, 34.8, 34.6, 32.1, 30.8, 29.0, 27.8, 26.7, 23.1 (C-19_{CA}), 17.6 (C-21_{CA}), 12.8 (C-18_{CA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical

formula: $C_{35}H_{51}N_4O_5^+$ 607.3854 found: 607.3840; Anal. RP-HPLC $t_R = 3.780$ min, purity 95.86%; $[\alpha]_D^{20} = +58$ (c 1.0, MeOH).

(4R)-4-((3R,10S,12S,13R)-3,12-Dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)-N-((1-(2-oxo-2-phenylethyl)-1H-1,2,3-triazol-4-yl)methyl)pentanamide (**6da**): Off-white solid; Yield: 80% (0.110 g); mp: 135–136 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.39 (t, $J = 5.7$ Hz, 1H, NH_{Amide}), 8.06 (d, $J = 7.1$ Hz, 2H, H_{Ar}), 7.84 (s, 1H, $H_{Triazole}$), 7.74 (t, $J = 7.4$ Hz, 1H, H_{Ar}), 7.61 (t, $J = 7.7$ Hz, 2H, H_{Ar}), 6.15 (s, 2H), 4.53 (d, $J = 4.2$ Hz, 1H, OH_{DCA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.22 (d, $J = 4.0$ Hz, 1H, OH_{DCA}), 3.78 (brs, 1H, H-12 $_{DCA}$), 3.48 (brs, 1H, H-3 $_{DCA}$), 2.17–1.98 (m, 2H), 1.76–1.44 (m, 10H), 1.37–1.14 (m, 14H), 0.91 (d, $J = 6.3$ Hz, 3H, Me-21 $_{DCA}$), 0.82 (s, 3H, Me-19 $_{DCA}$), 0.56 (s, 3H, Me-18 $_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 192.7 (C=O), 173.1 (C=O), 145.6, 134.7, 134.6, 129.4, 128.6, 124.8, 71.5 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 56.2, 47.9, 46.7, 46.4, 42.1, 36.7, 36.1, 35.6, 35.5, 34.6, 34.3, 33.4, 32.8, 32.1, 30.7, 29.1, 27.7, 27.4, 26.6, 24.0, 23.6 (C-19 $_{DCA}$), 17.5 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{51}N_4O_4^+$ 591.3905 found: 591.3883; Anal. RP-HPLC $t_R = 3.610$ min, purity 96.50%; $[\alpha]_D^{20} = +9$ (c 1.0, MeOH).

(1-(2-Oxo-2-(*p*-tolyl)ethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6ab**): White solid; Yield: 76% (0.103 g); mp: 121–124 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.10 (s, 1H, $H_{Triazole}$), 7.97 (d, $J = 8.3$ Hz, 2H, H_{Ar}), 7.42 (d, $J = 8.0$ Hz, 2H, H_{Ar}), 6.15 (s, 2H), 5.16 (s, 2H), 4.38 (d, $J = 4.4$ Hz, 1H, OH_{CA}), 4.15 (d, $J = 3.5$ Hz, 1H, OH_{CA}), 4.05 (d, $J = 3.3$ Hz, 1H, OH_{CA}), 3.77 (d, $J = 3.4$ Hz, 1H, H-12 $_{CA}$), 3.60 (brs, 1H, H-7 $_{CA}$), 3.21–3.14 (m, 1H, H-3 $_{CA}$), 2.42 (s, 3H, Me $_{Ar}$), 2.30–2.18 (m, 2H), 2.14–1.92 (m, 2H), 1.84–1.55 (m, 8H), 1.45–1.66 (m, 12H), 0.91 (d, $J = 6.1$ Hz, 3H, Me-21 $_{CA}$), 0.79 (s, 3H, Me-19 $_{CA}$), 0.56 (s, 3H, Me-18 $_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 192.0 (C=O), 173.6 (C=O), 145.3, 142.3, 132.0, 130.0, 128.7, 126.9, 71.4 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 57.5, 56.2, 46.5, 46.2, 42.0, 41.8, 35.4, 35.3, 34.8, 31.1, 31.0, 30.8, 29.0, 27.7, 26.6, 23.1 (C-19 $_{CA}$), 21.8 (Me $_{Ar}$), 17.3 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); Anal. RP-HPLC $t_R = 4.010$ min, purity 99.37%; HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{36}H_{52}N_3O_6^+$ 622.3851 found: 622.3837; $[\alpha]_D^{20} = +29$ (c 1.0, MeOH).

(1-(2-Oxo-2-(*p*-tolyl)ethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6bb**): White solid; Yield: 79% (0.110 g); mp: 123–125 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.10 (s, 1H, $H_{Triazole}$), 7.97 (d, $J = 8.2$ Hz, 2H, H_{Ar}), 7.42 (d, $J = 8.0$ Hz, 2H, H_{Ar}), 6.15 (s, 2H), 5.16 (s, 2H), 4.53 (d, $J = 4.3$ Hz, 1H, OH_{DCA}), 4.24 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.78 (d, $J = 3.8$ Hz, 1H, H-12 $_{DCA}$), 3.47 (brs, 1H, H-3 $_{DCA}$), 2.42 (s, 3H, Me $_{Ar}$), 2.39–2.24 (m, 2H), 1.85–1.50 (m, 10H), 1.47–1.15 (m, 14H), 0.90 (d, $J = 6.2$ Hz, 3H, Me-21 $_{DCA}$), 0.83 (s, 3H, Me-19 $_{DCA}$), 0.57 (s, 3H, Me-18 $_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 192.0 (C=O), 173.6 (C=O), 145.3, 142.3, 132.0, 130.0, 128.7, 126.9, 71.5 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 57.5, 56.2, 47.9, 46.6, 46.5, 42.0, 36.7, 36.1, 35.6, 35.4, 34.3, 33.4, 31.1, 30.7, 29.0, 27.6, 27.4, 26.6, 24.0, 23.5 (C-19 $_{DCA}$), 21.8, 17.3 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M]^+$ calcd for chemical formula: $C_{36}H_{52}N_3O_5^+$ 606.3901 found: 606.3874; Anal. RP-HPLC $t_R = 4.813$ min, purity 95.99% $[\alpha]_D^{20} = +47$ (c 1.0, MeOH).

(4R)-N-((1-(2-Oxo-2-(*p*-tolyl)ethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (**6cb**): White solid; Yield: 77% (0.107 g); mp: 129–131 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.39 (t, $J = 5.7$ Hz, 1H, NH_{Amide}), 7.96 (d, $J = 8.2$ Hz, 2H, H_{Ar}), 7.82 (s, 1H, $H_{Triazole}$), 7.41 (d, $J = 8.1$ Hz, 2H, H_{Ar}), 6.11 (s, 2H), 4.38 (d, $J = 4.3$ Hz, 1H, OH_{CA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.13 (d, $J = 3.5$ Hz, 1H, OH_{CA}), 4.04 (d, $J = 3.3$ Hz, 1H, OH_{CA}), 3.77 (d, $J = 3.6$ Hz, 1H, H-12 $_{CA}$), 3.59 (brs, 1H, H-7 $_{CA}$), 3.23–3.16 (m, 1H, H-3 $_{CA}$), 2.41 (s, 3H, Me $_{Ar}$), 2.22–2.11 (m, 2H), 2.06–1.94 (m, 2H), 1.82–1.58 (m, 7H), 1.50–1.11 (m, 13H), 0.92 (d, $J = 6.2$ Hz, 3H, Me-21 $_{CA}$), 0.78 (s, 3H, Me-19 $_{CA}$), 0.55 (s, 3H, Me-18 $_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 192.1 (C=O), 173.2 (C=O), 145.6, 145.3, 132.1, 130.0, 128.7, 124.8, 71.5 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 56.1, 46.6, 46.2, 42.0, 41.8, 35.8, 35.6, 35.3, 34.8, 34.6, 32.8, 32.1, 30.8, 29.0, 27.8, 26.7, 23.1 (C-19 $_{CA}$), 21.7 (Me $_{Ar}$), 17.6 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for

chemical formula: $C_{36}H_{53}N_4O_5^+$ 621.4010 found: 621.4006; Anal. RP-HPLC $t_R = 3.770$ min, purity 95.06%; $[\alpha]_D^{20} = +19$ (c 1.0, MeOH).

(4R)-4-((3R,10S,12S,13R)-3,12-Dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)-N-((1-(2-oxo-2-(p-tolylethyl)-1H-1,2,3-triazol-4-yl)methyl)pentanamide) (6db): White solid Yield: 80% (0.112 g); mp: 112–115 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.37 (t, $J = 5.8$ Hz, 1H, NH_{Amide}), 7.97 (d, $J = 8.2$ Hz, 2H, H_{Ar}), 7.82 (s, 1H, $H_{Triazole}$), 7.41 (d, $J = 8.1$ Hz, 2H, H_{Ar}), 6.11 (s, 2H), 4.48 (d, $J = 4.2$ Hz, 1H, OH_{DCA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.20 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.77 (d, $J = 3.8$ Hz, 1H, H-12 $_{DCA}$), 3.37 (brs, 1H, H-3 $_{DCA}$), 2.42 (s, 3H, Me_{Ar}), 2.15–1.95 (m, 2H), 1.82–1.42 (m, 11H), 1.37–1.13 (m, 13H), 0.92 (d, $J = 6.3$ Hz, 3H, Me-21 $_{DCA}$), 0.82 (s, 3H, Me-19 $_{DCA}$), 0.56 (s, 3H, Me-18 $_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 192.1 (C=O), 173.0 (C=O), 145.6, 145.2, 132.1, 130.0, 128.7, 124.8, 71.5 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 56.1, 47.9, 46.7, 46.4, 42.1, 36.1, 35.5, 34.6, 34.3, 33.4, 32.8, 32.1, 29.1, 27.7, 27.4, 26.6, 24.0, 23.5 (C-19 $_{DCA}$), 21.8 (Me_{Ar}), 17.5 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{36}H_{53}N_4O_4^+$ 605.4035 found: 605.4061; Anal. RP-HPLC $t_R = 4.100$ min, purity 93.82%; $[\alpha]_D^{20} = +$ (c 1.0, MeOH).

(1-(2-(4-Methoxyphenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (6ac): Off-white solid; Yield: 80% (0.114 g); mp: 176–177 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.10 (s, 1H, $H_{Triazole}$), 8.05 (d, $J = 8.9$ Hz, 2H, H_{Ar}), 7.13 (d, $J = 9.0$ Hz, 2H, H_{Ar}), 6.13 (s, 2H), 5.17 (s, 2H), 4.36 (brs, 1H, OH_{CA}), 4.14 (d, $J = 3.5$ Hz, 1H, OH_{CA}), 4.04 (d, $J = 3.3$ Hz, 1H, OH_{CA}), 3.88 (s, 3H, OMe_{Ar}), 3.77 (d, $J = 3.1$ Hz, 1H, H-12 $_{CA}$), 3.60 (brs, 1H, H-7 $_{CA}$), 3.22–3.15 (m, 1H, H-3 $_{CA}$), 2.40–2.24 (m, 2H), 2.23–2.10 (m, 2H), 1.80–1.61 (m, 6H), 1.49–1.09 (m, 14H), 0.91 (d, $J = 6.1$ Hz, 3H, Me-21 $_{CA}$), 0.80 (s, 3H, Me-19 $_{CA}$), 0.57 (s, 3H, Me-18 $_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 190.8 (C=O), 173.6 (C=O), 164.4, 142.3, 131.1, 127.4, 126.9, 114.7, 71.4 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 57.5, 56.2, 55.9, 46.5, 46.2, 42.0, 41.8, 35.8, 35.5, 35.3, 34.8, 31.1, 31.0, 30.9, 29.0, 27.7, 26.7, 23.3, 23.1 (C-19 $_{CA}$), 17.4 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{36}H_{52}N_3O_7^+$ 638.3800 found: 638.3798; Anal. RP-HPLC $t_R = 4.003$ min, purity 99.43%; $[\alpha]_D^{20} = +31$ (c 1.0, MeOH).

(1-(2-(4-Methoxyphenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (6bc): Off-white solid; Yield: 81% (0.116 g); mp: 144–45 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.09 (s, 1H, $H_{Triazole}$), 8.05 (d, $J = 8.8$ Hz, 2H, H_{Ar}), 7.13 (d, $J = 8.9$ Hz, 2H, H_{Ar}), 6.12 (s, 2H), 5.17 (s, 2H), 4.52 (d, $J = 4.3$ Hz, 1H, OH_{DCA}), 4.22 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.88 (s, 3H, OMe_{Ar}), 3.78 (d, $J = 4.2$ Hz, 1H, H-12 $_{DCA}$), 3.46 (brs, 1H, H-3 $_{DCA}$), 2.43–2.31 (m, 2H), 2.29–2.15 (m, 2H), 1.78–1.55 (m, 10H), 1.35–1.20 (m, 12H), 0.91 (d, $J = 6.2$ Hz, 3H, Me-21 $_{DCA}$), 0.84 (s, 3H, Me-19 $_{DCA}$), 0.57 (s, 3H, Me-18 $_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 190.8 (C=O), 173.6 (C=O), 164.4, 142.3, 131.1, 127.4, 126.9, 114.7, 71.5 (C-12 $_{DCA}$), 70.5 (C-3 $_{DCA}$), 57.5, 56.2, 55.9, 47.9, 46.6, 46.5, 42.0, 36.7, 36.1, 35.6, 35.4, 34.3, 33.4, 31.0, 30.7, 29.0, 27.6, 27.4, 26.5, 23.5 (C-19 $_{DCA}$), 17.3 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{36}H_{52}N_3O_6^+$ 622.3851 found: 622.3853; Anal. RP-HPLC $t_R = 4.797$ min, purity 97.61%; $[\alpha]_D^{20} = +11$ (c 1.0, MeOH).

(4R)-N-((1-(2-(4-Methoxyphenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (6cc): Off-white solid; Yield: 81% (0.115 g); mp: 158–159 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.38 (t, $J = 5.8$ Hz, 1H, NH_{Amide}), 8.04 (d, $J = 8.9$ Hz, 2H, H_{Ar}), 7.82 (s, 1H, $H_{Triazole}$), 7.12 (d, $J = 9.0$ Hz, 2H, H_{Ar}), 6.08 (s, 2H), 4.36 (brs, 1H, OH_{CA}), 4.34–4.29 (m, 2H), 4.12 (d, $J = 3.5$ Hz, 1H, OH_{CA}), 4.03 (d, $J = 3.5$ Hz, 1H, OH_{CA}), 3.87 (s, 3H, OMe_{Ar}), 3.77 (d, $J = 3.5$ Hz, 1H, H-12 $_{CA}$), 3.59 (brs, 1H, H-7 $_{CA}$), 3.22–3.16 (m, 1H, H-3 $_{CA}$), 2.20–2.13 (m, 2H), 2.04–1.95 (m, 2H), 1.81–1.63 (m, 6H), 1.48–1.29 (m, 8H), 1.27–1.17 (m, 6H), 0.92 (d, $J = 6.2$ Hz, 3H, Me-21 $_{CA}$), 0.78 (s, 3H, Me-19 $_{CA}$), 0.55 (s, 3H, Me-18 $_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 190.9 (C=O), 173.1 (C=O), 164.3, 145.6, 131.0, 127.5, 124.8, 114.7, 71.5 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 56.2, 55.8, 46.6, 46.2, 42.0, 41.8, 35.8, 35.6, 35.3, 34.8, 34.6, 32.1, 30.9, 29.0, 27.8, 26.7, 23.3, 23.1 (C-19 $_{CA}$), 17.5 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS

(ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{36}H_{53}N_4O_6^+$ 637.3960 found: 637.3961; Anal. RP-HPLC $t_R = 4.097$ min, purity 93.92%; $[\alpha]_D^{20} = +47$ (c 1.0, MeOH).

(4R)-4-((3R,10S,12S,13R)-3,12-Dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)-N-((1-(2-(4-methoxyphenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)pentanamide (**6dc**): Off-white solid; Yield: 76% (0.109 g); mp: 114–115 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.38 (t, $J = 5.7$ Hz, 1H, NH_{Amide}), 8.05 (d, $J = 8.9$ Hz, 2H, H_{Ar}), 7.82 (s, 1H, $H_{Triazole}$), 7.12 (d, $J = 9.0$ Hz, 2H, H_{Ar}), 6.08 (s, 2H), 4.51–4.49 (m, 1H, OH_{DCA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.21 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.88 (s, 3H, OMe_{Ar}), 3.78 (d, $J = 4.0$ Hz, 1H, $H-12_{DCA}$), 3.40 (brs, 1H, $H-3_{DCA}$), 2.01–2.03 (m, 2H), 2.00–1.80 (d, $J = 8.3$ Hz, 2H), 1.77–1.47 (m, 10H), 1.–1.18 (m, 12H), 0.92 (d, $J = 6.3$ Hz, 3H, $Me-21_{DCA}$), 0.82 (s, 3H, $Me-19_{DCA}$), 0.56 (s, 3H, $Me-18_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 190.9 (C=O), 173.1 (C=O), 164.3, 145.6, 131.0, 127.5, 124.8, 114.7, 71.5 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 56.2, 55.8, 47.9, 46.7, 46.4, 42.1, 36.7, 36.1, 35.6, 35.5, 34.6, 34.3, 33.4, 32.8, 32.1, 30.7, 29.1, 27.7, 27.4, 26.6, 23.5 (C-19 $_{DCA}$), 17.5 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{36}H_{53}N_4O_5^+$ 621.4010 found: 621.4009; Anal. RP-HPLC $t_R = 4.097$ min, purity 94.90%; $[\alpha]_D^{20} = +21$ (c 1.0, MeOH).

(1-(2-(4-Fluorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6ad**): Off-white solid; Yield: 76% (0.106 g); mp: 152–153 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.19–8.09 (m, 3H, $H_{Triazole}$ & H_{Ar}), 7.46 (t, $J = 8.7$ Hz, 2H, H_{Ar}), 6.19 (s, 2H), 5.17 (s, 2H), 4.35 (d, $J = 4.3$ Hz, 1H, OH_{CA}), 4.13 (d, $J = 3.4$ Hz, 1H, OH_{CA}), 4.03 (d, $J = 3.7$ Hz, 1H, OH_{CA}), 3.77 (brs, 1H, $H-12_{CA}$), 3.61 (brs, 1H, $H-7_{CA}$), 3.22–3.16 (m, 1H, $H-3_{CA}$), 2.41–2.24 (m, 2H), 2.21–2.14 (m, 2H), 1.83–1.67 (m, 5H), 1.64–1.40 (m, 5H), 1.35–1.17 (m, 10H), 0.91 (d, $J = 5.9$ Hz, 3H, $Me-21_{CA}$), 0.80 (s, 3H, $Me-19_{CA}$), 0.57 (s, 3H, $Me-18_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.3 (C=O), 173.6 (C=O), 142.4, 131.8, 132.7, 126.9, 116.7, 116.5, 71.4 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 57.5, 56.2, 46.5, 46.2, 42.0, 35.8, 35.5, 35.3, 34.8, 31.1, 31.1, 30.9, 29.0, 27.7, 26.7, 23.3, 23.1 (C-19 $_{CA}$), 17.4 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{49}N_3O_6F^+$ 626.3600 found: 626.3600; Anal. RP-HPLC $t_R = 3.783$ min, purity 95.12%; $[\alpha]_D^{20} = +51$ (c 1.0, MeOH).

(1-(2-(4-Fluorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6bd**): Off-white solid; Yield: 72% (0.101 g); mp: 127–128 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.19–8.14 (m, 2H, H_{Ar}), 8.10 (s, 1H, $H_{Triazole}$), 7.46 (t, $J = 8.9$ Hz, 2H, H_{Ar}), 6.19 (s, 2H), 5.17 (s, 2H), 4.49 (d, $J = 4.3$ Hz, 1H, OH_{DCA}), 4.22 (d, $J = 4.0$ Hz, 1H, OH_{DCA}), 3.78 (d, $J = 4.1$ Hz, 1H, $H-12_{DCA}$), 3.43–3.40 (m, 1H, $H-3_{DCA}$), 2.40–2.22 (m, 2H), 1.81–1.70 (m, 5H), 1.62–1.44 (m, 5H), 1.36–1.11 (m, 14H), 0.91 (d, $J = 6.1$ Hz, 3H, $Me-21_{DCA}$), 0.84 (s, 3H, $Me-19_{DCA}$), 0.57 (s, 3H, $Me-18_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.3 (C=O), 173.6 (C=O), 142.4, 131.8, 131.7, 126.9, 116.7, 116.5, 71.5 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 57.5, 56.2, 47.9, 46.6, 46.5, 42.1, 36.7, 36.1, 35.6, 35.4, 34.3, 33.4, 31.1, 30.7, 29.0, 27.6, 27.4, 26.6, 24.0, 23.6 (C-19 $_{DCA}$), 17.3 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{49}N_3O_5F^+$ 610.3651 found: 610.3650; Anal. RP-HPLC $t_R = 5.333$ min, purity 96.72%; $[\alpha]_D^{20} = +8$ (c 1.0, MeOH).

(4R)-N-((1-(2-(4-Fluorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (**6cd**): Off-white solid; Yield: 79% (0.110 g); mp: 137–138 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.40 (t, $J = 5.7$ Hz, 1H, NH_{Amide}), 8.15 (dd, $J = 8.8, 5.5$ Hz, 2H, H_{Ar}), 7.83 (s, 1H, $H_{Triazole}$), 7.45 (t, $J = 8.9$ Hz, 2H, H_{Ar}), 6.15 (s, 2H), 4.44 (brs, 1H, OH_{CA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.14 (brs, 1H, OH_{CA}), 4.06–4.01 (m, 1H, OH_{CA}), 3.77 (d, $J = 3.0$ Hz, 1H, $H-12_{CA}$), 3.61–3.58 (m, 1H, $H-7_{CA}$), 3.21–3.16 (m, 1H, $H-3_{CA}$), 2.16–2.12 (m, 2H), 2.03–1.99 (m, 2H), 1.80–1.60 (m, 7H), 1.44–1.12 (m, 13H), 0.92 (d, $J = 6.2$ Hz, 3H, $Me-21_{CA}$), 0.78 (s, 3H, $Me-19_{CA}$), 0.55 (s, 3H, $Me-18_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.4 (C=O), 173.2 (C=O), 145.7, 131.8, 131.7, 124.8, 116.7, 116.4, 71.5 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 56.1, 46.6, 46.2, 42.0, 41.8, 35.8, 35.6, 35.3, 34.8, 34.6, 32.8, 32.1, 30.8, 29.0, 27.8, 26.7, 23.1 (C-19 $_{CA}$), 17.6 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical

formula: $C_{35}H_{50}N_4O_5F^+$ 625.3760 found: 625.3754; Anal. RP-HPLC $t_R = 4.000$ min, purity 94.54%; $[\alpha]_D^{20} = +22$ (c 1.0, MeOH).

(4R)-4-((3R,10S,12S,13R)-3,12-Dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)-N-((1-(2-(4-fluorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)pentanamide (**6dd**): Off-white solid; Yield: 77% (0.117 g); mp: 126–127 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.40–8.36 (m, 1H, NH_{Amide}), 8.18–8.14 (m, 2H, H_{Ar}), 7.83 (s, 1H, $H_{Triazole}$), 7.46 (d, $J = 8.8$ Hz, 2H, H_{Ar}), 6.15 (s, 2H), 4.49 (d, $J = 4.3$ Hz, 1H, OH_{DCA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.20 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.77 (d, $J = 3.8$ Hz, 1H, H-12 $_{DCA}$), 3.43–3.40 (m, 1H, H-3 $_{DCA}$), 2.18–1.98 (m, 2H), 1.80–1.66 (m, 6H), 1.64–1.44 (m, 5H), 1.35–1.15 (m, 13H), 0.92 (d, $J = 6.4$ Hz, 3H, Me-21 $_{DCA}$), 0.82 (s, 3H, Me-19 $_{DCA}$), 0.56 (s, 3H, Me-18 $_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.4 (C=O), 173.1 (C=O), 145.7, 131.8, 131.7, 124.8, 116.7, 116.4, 71.5 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 56.1, 47.9, 46.7, 46.4, 42.1, 36.7, 36.1, 35.6, 35.5, 34.6, 34.3, 33.4, 32.8, 32.1, 30.7, 29.1, 27.7, 27.4, 26.6, 24.0, 23.5 (C-19 $_{DCA}$), 17.5 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{50}N_4O_4F^+$ 609.3811 found: 609.3807; Anal. RP-HPLC $t_R = 4.030$ min, purity 95.47%; $[\alpha]_D^{20} = +60$ (c 1.0, MeOH).

(1-(2-(4-Chlorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6ae**): Off-white solid; Yield: 74% (0.106 g); mp: 153–154 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.10 (s, 1H, $H_{Triazole}$), 8.08 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 7.70 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 6.19 (s, 2H), 5.17 (s, 2H), 4.39 (brs, 1H, OH_{CA}), 4.15 (d, $J = 3.5$ Hz, 1H, OH_{CA}), 4.05 (d, $J = 3.3$ Hz, 1H, OH_{CA}), 3.77 (d, $J = 3.4$ Hz, 1H, H-12 $_{CA}$), 3.60 (brs, 1H, H-7 $_{CA}$), 3.18 (brs, 1H, H-3 $_{CA}$), 2.42–2.24 (m, 2H), 2.22–2.09 (m, 2H), 1.82–1.59 (m, 7H), 1.49–1.14 (m, 13H), 0.91 (d, $J = 6.1$ Hz, 3H, Me-21 $_{CA}$), 0.80 (s, 3H, Me-19 $_{CA}$), 0.56 (s, 3H, Me-18 $_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.7 (C=O), 173.6 (C=O), 142.4, 139.6, 133.3, 130.6, 129.6, 126.8, 71.5 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 57.5, 56.3, 46.5, 46.2, 42.0, 41.8, 35.8, 35.4, 35.3, 34.8, 31.2, 31.1, 30.8, 29.0, 27.7, 26.7, 23.1 (C-19 $_{CA}$), 17.4 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{49}N_3O_6Cl^+$ 642.3304 found: 642.3291; Anal. RP-HPLC $t_R = 4.107$ min, purity 97.01%; $[\alpha]_D^{20} = +27$ (c 1.0, MeOH).

(1-(2-(4-Chlorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6be**): Off-white solid; Yield: 72% (0.104 g); mp: 92–93 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.10 (d, $J = 2.8$ Hz, 2H, H_{Ar}), 8.07 (s, 1H, $H_{Triazole}$), 7.69 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 6.19 (s, 2H), 5.17 (s, 2H), 4.52 (d, $J = 4.2$ Hz, 1H, OH_{DCA}), 4.22 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.78 (d, $J = 3.7$ Hz, 1H, H-12 $_{DCA}$), 3.48 (brs, 1H, H-3 $_{DCA}$), 2.43–2.15 (m, 2H), 1.82–1.43 (m, 11H), 1.37–1.08 (m, 13H), 0.90 (d, $J = 6.2$ Hz, 3H, Me-21 $_{DCA}$), 0.84 (s, 3H, Me-19 $_{DCA}$), 0.57 (s, 3H, Me-18 $_{DCA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.7 (C=O), 173.6 (C=O), 142.4, 139.6, 133.3, 130.6, 129.6, 126.9, 71.4 (C-12 $_{DCA}$), 70.4 (C-3 $_{DCA}$), 57.5, 56.3, 47.9, 46.6, 46.5, 42.0, 36.8, 36.1, 35.6, 35.4, 34.3, 33.4, 31.1, 31.1, 29.0, 27.6, 27.4, 26.6, 23.6 (C-19 $_{DCA}$), 17.3 (C-21 $_{DCA}$), 12.9 (C-18 $_{DCA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{49}N_3O_5Cl^+$ 626.3355 found: 626.3336; Anal. RP-HPLC $t_R = 5.343$ min, purity 96.73%; $[\alpha]_D^{20} = +18$ (c 1.0, MeOH).

(4R)-N-((1-(2-(4-Chlorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (**6ce**): Off-white solid; Yield: 78% (0.112 g); mp: 154–155 °C; 1H NMR (400 MHz, DMSO- d_6) δ 8.40 (t, $J = 5.7$ Hz, 1H, NH_{Amide}), 8.07 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 7.83 (s, 1H, $H_{Triazole}$), 7.69 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 6.15 (s, 2H), 4.39 (brs, 1H, OH_{CA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.14 (brs, 1H, OH_{CA}), 4.05 (brs, 1H, OH_{CA}), 3.77 (brs, 1H, H-12 $_{CA}$), 3.60 (brs, 1H, H-7 $_{CA}$), 3.21–3.17 (m, 1H, H-3 $_{CA}$), 2.24–2.16 (m, 2H), 2.14–2.07 (m, 2H), 1.81–1.61 (m, 7H), 1.44–1.20 (m, 13H), 0.92 (d, $J = 6.2$ Hz, 3H, Me-21 $_{CA}$), 0.78 (s, 3H, Me-19 $_{CA}$), 0.54 (s, 3H, Me-18 $_{CA}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 191.8 (C=O), 173.2 (C=O), 145.7, 139.6, 133.3, 130.5, 129.6, 124.8, 71.5 (C-12 $_{CA}$), 70.9 (C-3 $_{CA}$), 66.7 (C-7 $_{CA}$), 56.2, 46.6, 46.2, 42.0, 41.8, 35.8, 35.6, 35.3, 34.9, 34.8, 34.6, 32.8, 32.7, 32.1, 32.0, 30.8, 29.0, 28.2, 27.8, 23.3 (C-19 $_{CA}$), 17.5 (C-21 $_{CA}$), 12.8 (C-18 $_{CA}$); HRMS (ESI): m/z $[M + H]^+$ calcd for chemical formula: $C_{35}H_{50}N_4O_5Cl^+$ 641.3464 found: 641.3457; Anal. RP-HPLC $t_R = 3.607$ min, purity 97.57%; $[\alpha]_D^{20} = +16$ (c 1.0, MeOH).

(4R)-N-((1-(2-(4-Chlorophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (**6de**): Off-white solid; Yield: 76% (0.110 g); mp: 100–101 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.39 (t, *J* = 5.8 Hz, 1H, NH_{Amide}), 8.07 (d, *J* = 8.6 Hz, 2H, H_{Ar}), 7.82 (s, 1H, H_{Triazole}), 7.69 (d, *J* = 8.6 Hz, 2H, H_{Ar}), 6.15 (s, 2H), 4.52 (d, *J* = 4.1 Hz, 1H, OH_{DCA}), 4.32 (d, *J* = 5.7 Hz, 2H), 4.22 (d, *J* = 4.2 Hz, 1H, OH_{DCA}), 3.77 (brs, 1H, H-12_{DCA}), 3.47 (m, 1H, H-3_{DCA}), 2.20–2.09 (m, 2H), 2.05–1.98 (m, 2H), 1.78–1.51 (m, 11H), 1.35–1.18 (m, 11H), 0.91 (d, *J* = 6.3 Hz, 3H, Me-21_{DCA}), 0.82 (s, 3H, Me-19_{DCA}), 0.55 (s, 3H, Me-18_{DCA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 192.0 (C=O), 172.9 (C=O), 145.7, 139.6, 133.3, 130.5, 129.6, 124.8, 71.5 (C-12_{DCA}), 70.4 (C-3_{DCA}), 56.2, 47.9, 46.7, 46.4, 42.1, 36.8, 36.1, 35.6, 35.5, 34.6, 34.3, 33.4, 32.8, 32.1, 30.7, 29.1, 27.7, 27.4, 26.6, 24.0, 23.5 (C-19_{DCA}), 17.5 (C-21_{DCA}), 12.9 (C-18_{DCA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical formula: C₃₅H₅₀N₄O₄Cl⁺ 625.3515 found: 625.3492; Anal. RP-HPLC t_R = 4.040 min, purity 96.32%; [α]_D²⁰ = +47 (c 1.0, MeOH).

(1-(2-(4-Bromophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6af**): Off-white solid; Yield: 83% (0.127 g); mp: 159–160 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.10 (s, 1H, H_{Triazole}), 8.00 (d, *J* = 8.6 Hz, 2H, H_{Ar}), 7.84 (d, *J* = 8.6 Hz, 2H, H_{Ar}), 6.19 (s, 2H), 5.17 (s, 2H), 4.39 (d, *J* = 3.6 Hz, 1H, OH_{CA}), 4.15 (d, *J* = 3.3 Hz, 1H, OH_{CA}), 4.05 (d, *J* = 3.1 Hz, 1H, OH_{CA}), 3.77 (brs, 1H, H-12_{CA}), 3.60 (brs, 1H, H-7_{CA}), 3.23–3.16 (m, 1H, H-3_{CA}), 2.42–2.28 (m, 2H), 2.25–2.09 (m, 2H), 1.81–1.60 (m, 7H), 1.47–1.15 (m, 13H), 0.91 (d, *J* = 6.1 Hz, 3H, Me-21_{CA}), 0.80 (s, 3H, Me-19_{CA}), 0.56 (s, 3H, Me-18_{CA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 191.9 (C=O), 173.6 (C=O), 142.4, 133.6, 132.5, 130.6, 128.9, 126.9, 71.4 (C-12_{CA}), 70.9 (C-3_{CA}), 66.7 (C-7_{CA}), 57.5, 56.3, 46.5, 46.2, 42.0, 41.8, 35.8, 35.5, 35.3, 34.8, 31.1, 30.9, 29.0, 27.7, 26.7, 23.3, 23.1 (C-19_{CA}), 17.3 (C-21_{CA}), 12.8 (C-18_{CA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical formula: C₃₅H₄₉N₃O₆Br⁺ 686.2799 found: 686.2780; Anal. RP-HPLC t_R = 4.197 min, purity 96.62%; [α]_D²⁰ = +11 (c 1.0, MeOH).

(1-(2-(4-Bromophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl (4R)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanoate (**6bf**): Off-white solid; Yield: 80% (0.124 g); mp: 176–177 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.10 (s, 1H, H_{Triazole}), 8.00 (d, *J* = 8.7 Hz, 2H, H_{Ar}), 7.84 (d, *J* = 8.6 Hz, 2H, H_{Ar}), 6.19 (s, 2H), 5.17 (s, 2H), 4.50 (d, *J* = 4.3 Hz, 1H, OH_{DCA}), 4.23 (d, *J* = 4.1 Hz, 1H, OH_{DCA}), 3.78 (d, *J* = 3.8 Hz, 1H, H-12_{DCA}), 3.43 (brs, 1H, H-3_{DCA}), 2.40–2.20 (m, 2H), 1.80–1.44 (m, 10H), 1.36–1.16 (m, 14H), 0.90 (d, *J* = 6.2 Hz, 3H, Me-21_{DCA}), 0.84 (s, 3H, Me-19_{DCA}), 0.57 (s, 3H, Me-18_{DCA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 191.9 (C=O), 173.6 (C=O), 142.4, 133.6, 132.5, 130.6, 128.9, 126.9, 71.5 (C-12_{DCA}), 70.4 (C-3_{DCA}), 57.5, 56.3, 47.9, 46.6, 46.5, 42.1, 36.7, 36.1, 35.6, 35.4, 34.3, 33.4, 33.1, 31.1, 30.7, 29.0, 27.6, 27.4, 26.6, 24.0, 23.6 (C-19_{DCA}), 17.3 (C-21_{DCA}), 12.9 (C-18_{DCA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical formula: C₃₅H₄₉N₃O₅Br⁺ 670.2850 found: 670.2855; Anal. RP-HPLC t_R = 5.507 min, purity 98.37%; [α]_D²⁰ = +81 (c 1.0, MeOH).

(4R)-N-((1-(2-(4-Bromophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,7R,10S,12S,13R)-3,7,12-trihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide (**6cf**): Off-white solid; Yield: 81% (0.124 g); mp: 144–145 °C; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.39 (t, *J* = 5.8 Hz, 1H, NH_{Amide}), 7.99 (d, *J* = 8.3 Hz, 2H, H_{Ar}), 7.86–7.81 (m, 3H, H_{Triazole}&H_{Ar}), 6.15 (s, 2H), 4.36 (d, *J* = 4.3 Hz, 1H, OH_{CA}), 4.32 (d, *J* = 5.7 Hz, 2H), 4.12 (d, *J* = 3.5 Hz, 1H, OH_{CA}), 4.03 (d, *J* = 3.4 Hz, 1H, OH_{CA}), 3.79–3.76 (m, 1H, H-12_{CA}), 3.60 (brs, 1H, H-7_{CA}), 3.21–3.15 (m, 1H, H-3_{CA}), 2.18–2.12 (m, 2H), 2.04–1.94 (m, 2H), 1.83–1.58 (m, 8H), 1.50–1.15 (m, 12H), 0.92 (d, *J* = 6.2 Hz, 3H, Me-21_{CA}), 0.78 (s, 3H, Me-19_{CA}), 0.55 (s, 3H, Me-18_{CA}); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 192.1 (C=O), 173.1 (C=O), 145.7, 133.6, 132.5, 130.6, 128.8, 124.8, 71.5 (C-12_{CA}), 70.9 (C-3_{CA}), 66.7 (C-7_{CA}), 56.2, 46.6, 46.2, 42.0, 41.8, 35.6, 35.3, 34.8, 34.6, 32.8, 32.1, 30.9, 29.0, 27.8, 26.7, 23.3, 23.0 (C-19_{CA}), 17.5 (C-21_{CA}), 12.8 (C-18_{CA}); HRMS (ESI): *m/z* [M + H]⁺ calcd for chemical formula: C₃₅H₅₀N₄O₅Br⁺ 685.2959 found: 685.2956; Anal. RP-HPLC t_R = 3.720 min, purity 96.70%; [α]_D²⁰ = +13 (c 1.0, MeOH).

(4R)-N-((1-(2-(4-Bromophenyl)-2-oxoethyl)-1H-1,2,3-triazol-4-yl)methyl)-4-((3R,10S,12S,13R)-3,12-dihydroxy-10,13-dimethylhexadecahydro-1H-cyclopenta[a]phenanthren-17-yl)pentanamide

(**6df**): Off-white solid; Yield: 79% (0.122 g); mp: 135–136 °C; ^1H NMR (400 MHz, DMSO- d_6) δ 8.36 (t, $J = 5.7$ Hz, 1H, NH_{Amide}), 7.99 (d, $J = 8.6$ Hz, 2H, H_{Ar}), 7.88–7.80 (m, 3H, $\text{H}_{\text{Triazole}} \& \text{H}_{\text{Ar}}$), 6.14 (s, 2H), 4.48 (d, $J = 4.2$ Hz, 1H, OH_{DCA}), 4.32 (d, $J = 5.7$ Hz, 2H), 4.19 (d, $J = 4.1$ Hz, 1H, OH_{DCA}), 3.78 (brs, 1H, H-12 $_{\text{DCA}}$), 3.39 (brs, 1H, H-3 $_{\text{DCA}}$), 2.15–2.00 (m, 2H), 1.77–1.47 (m, 11H), 1.36–1.16 (m, 13H), 0.91 (d, $J = 6.3$ Hz, 3H, Me-21 $_{\text{DCA}}$), 0.82 (s, 3H, Me-19 $_{\text{DCA}}$), 0.55 (s, 3H, Me-18 $_{\text{DCA}}$); ^{13}C NMR (100 MHz, DMSO- d_6) δ 192.1 (C=O), 173.2 (C=O), 145.7, 133.6, 132.5, 130.6, 128.8, 124.8, 71.5 (C-12 $_{\text{DCA}}$), 70.9 (C-3 $_{\text{DCA}}$), 56.2, 46.6, 46.2, 42.0, 41.8, 35.7, 35.6, 35.3, 34.8, 32.8, 30.8, 27.8, 26.7, 23.1 (C-19 $_{\text{DCA}}$), 17.5 (C-21 $_{\text{DCA}}$), 12.8 (C-18 $_{\text{DCA}}$); HRMS (ESI): m/z $[\text{M} + \text{H}]^+$ calcd for chemical formula: $\text{C}_{35}\text{H}_{50}\text{N}_4\text{O}_4\text{Br}^+$ 669.3010 found: 669.3003; Anal. RP-HPLC $t_{\text{R}} = 3.937$ min, purity 98.65%; $[\alpha]_{\text{D}}^{20} = +17$ (c 1.0, MeOH).

3.3. Biological Assay

3.3.1. Cell Culture and MTT Assay

The cytotoxicity activity of all the conjugates (**6aa–6df**) was evaluated in vitro by MTT assay against two different breast cancer cell lines 4T1 (murine) and MCF-7 (human) and HEK 293 (human) as normal cell line. DTX was used as positive control. Cells were grown in DMEM supplemented with 10% FBS and 1% antibiotic solution and incubated at 5% CO_2 and 37 °C for 24 h. The stock solutions of all the conjugates were prepared in DMSO and diluted for further use. Briefly, 5×10^3 cells/well were seeded in 96-well cell culture plates and allowed to adhere for 24 h. Cell inhibition (%) was determined after 48 h exposure to the compounds at 1–25 μM concentration. After 48 h, MTT assay was performed and the yellow tetrazolium salt (MTT) was reduced in metabolically active cells to form insoluble purple formazan crystals, which were solubilized by the addition of DMSO. The optical density (OD) was recorded at 560 nm and 630 nm as reference wavelength. Percentage cell inhibition was determined by comparison with untreated cells [50,51].

3.3.2. Apoptotic Study

The extent of apoptosis induced by compounds **6af** and **6cf** in MCF-7 and 4T1 cells was quantified by flow cytometry according to the manufacturer's protocol. Briefly, cells were seeded in a 6-well plate at a cell density of 1×10^6 cells/well. After 24 h, the media was discarded and cells were treated with fresh media containing compounds **6af** and **6cf** at their respective IC_{50} concentrations for 48 h. After treatment, cells were trypsinized, harvested in PBS and collected by centrifugation for 5 min at 2000 rpm. Cells were then resuspended in 1X binding buffer and stained with FITC-labeled Annexin V Alexa Fluor 488 (5 μL) and propidium iodide (10 μL). Cells were analyzed using flow cytometer (Beckman Coulter), and data were analyzed with CytExpert software.

3.3.3. Pharmacokinetic Study of **6cf**

Wistar rats (male; 8–10 weeks, 200–240 g) were procured from Central Animal Facility, BITS Pilani (Pilani, India). Animal experiment protocol was approved by Institutional Animal Ethics Committee (IAEC/RES/24/03), BITS Pilani, Pilani, and all experiments were conducted as per CPCSEA guidelines. Rats were housed in well-ventilated cages at standard laboratory conditions with regular light/dark cycles for 12 h and fed with standard normal diet ad libitum.

The pharmacokinetic study of **6cf** was performed on Wistar rats. **6cf** solution (prepared in normal saline with 5% w/v tween 80) was administered intravenously at the dose of 10 mg/kg with maximum dosing volume of 300 μL to each rat without fasting ($n = 4$). After i.v. dosing, blood samples were collected for each preset time point at 10, 20, 30, 50 min, 1.5, 2, 4, 6, 8, 12 and 24 h. **6cf** plasma concentration–time profile was plotted and analyzed by non-compartmental model approach using Phoenix 2.1 WinNonlin (Pharsight Corporation, USA) to determine $t_{1/2}$, elimination half-life; C_0 , drug concentration in plasma at $t = 0$; AUC_{0-t} , area under curve from zero to the last time point; $\text{AUC}_{0-\infty}$, area under curve from zero to infinity; and MRT, mean residence time.

3.3.4. Determination of **6cf** in Rat Plasma

A simple liquid–liquid extraction (LLE) method was used for extraction of **6cf** from the rat plasma. A 200 μL aliquot of plasma sample containing **6cf** was taken in 5 mL glass tube, followed by the addition of 100 μL of internal standard (I.S.) (clobetasol, 2 $\mu\text{g}/\text{mL}$) solution. Samples were vortexed for 1 min, and then 2 mL of ethyl acetate was added as extracting solvent. The samples were vortexed for 5 min and centrifuged at 5000 rpm for 15 min at 4 $^{\circ}\text{C}$. The organic layer was collected and evaporated to dryness at 40 ± 0.5 $^{\circ}\text{C}$. The residue was reconstituted with 250 μL of mobile phase and vortexed for 1 min. Finally, 150 μL of sample was injected into HPLC for quantification.

3.3.5. Liquid Chromatographic Conditions

A Thermo Fisher Rapid Separation (RS) UHPLC System (Ultimate 3000) equipped with a pump (LPG-3400SD), Diode Array Detector (DAD) (DAD-3000) and autosampler (ACC-3000T) with 250 μL injection loop was used for purity analysis. The UHPLC system was equilibrated for approximately 40 min before beginning the sample analysis. Column temperature was 35 $^{\circ}$ throughout the analysis. **6cf** and I.S. were separated on Intersil[®] ODS (C18) column (250 \times 4.6 mm, 5 μm) with a mobile phase consisting of acetonitrile:water (60:40 % *v/v*) run in isocratic mode at a flow rate of 1 mL/min, detection wavelength 259 nm and injection volume of 150 μL . Retention time was found to be 6.2 and 12.2 min for **6cf** and clobetasol (I.S.), respectively. Control of hardware and data handling was performed using Chromeleon software version 7.2 SR4.

4. Conclusions

In summary, we synthesized a series of cholic-acid- and deoxycholic-acid-appended triazolyl aryl ketones in excellent yields via a Cu-catalyzed multi-component approach. All the synthesized conjugates were evaluated for their cytotoxicity against human breast adenocarcinoma (MCF-7) and mouse mammary carcinoma (4T1) cells at 10 μM , which highlighted three conjugates (**6af**, **6bf**, **6cf**) displaying interesting anticancer activity with IC_{50} values less than 19 μM on both tested cancer cell lines. Among these, the cholic-acid-appended triazolyl 4-bromophenyl ketone (**6cf**) connected via an amide bond was found to be active against both cancer cell lines with IC_{50} values of 5.71 μM and 8.71 μM , respectively, as compared to the reference drug possessing an IC_{50} value of 9.46 μM and 13.85 μM , respectively. Meanwhile, cholic-acid-appended triazolyl 4-bromophenyl ketone connected via an ester bond (**6af**) was found to be active against both cancer cell lines with IC_{50} values of 2.61 μM and 12.84 μM , respectively. Most of the conjugates showed low cytotoxicity toward the normal human embryonic kidney cell line (HEK 293) as evident from their cell viability data. Apoptosis studies of **6af** and **6cf** on MCF-7 cells at their respective IC_{50} values indicated induction of higher apoptosis by **6cf** in comparison to **6af** (46.09% vs. 33.89%). Meanwhile, in 4T1 cells, a similar apoptotic potential of the two compounds contributing to a total apoptosis of 19.02% and 19.56% in 4T1 cells was observed. Additionally, an MRT of 8.47 h with a half-life of 5.63 h was observed by *in vivo* pharmacokinetics studies of **6cf** in rats. In light of the present work, it appears that cytotoxicity is not only driven by the nature of the bile acid, but also by the electronic effect of the substituent present on the aryl moiety of aryl ketones. Clearly, the results suggest the potential of the studied conjugates in the development of anticancer drug candidates.

Supplementary Materials: Original ^1H and ^{13}C NMR spectra of **4c,d** and **6aa–6df**, COSY, HSQC and HMBC spectra of **6aa**, HRMS spectra of **6aa–6df** and HPLC chromatogram of **6aa–6df**.

Author Contributions: Conceptualization, R.S.; methodology, D.S.A.; resources, R.S. and D.C.; data curation, D.S.A., S.M. and K.S.I.; writing—original draft preparation, D.S.A.; writing—review and editing, R.S. and D.C.; supervision, R.S. and D.C.; project administration, R.S. and D.C.; funding acquisition, R.S. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest: The authors declare no conflict of interest.

Sample Availability: Samples of the compounds **6aa–df** are available from the authors.

References

1. Bach, P.B.; Jett, J.R.; Pastorino, U.; Tockman, M.S.; Swensen, S.J.; Begg, C.B. Computed tomography screening and lung cancer outcomes. *Jama* **2007**, *297*, 953–961. [[CrossRef](#)]
2. Gibbs, J.B. Mechanism-based target identification and drug discovery in cancer research. *Science* **2000**, *287*, 1969–1973. [[CrossRef](#)] [[PubMed](#)]
3. Arve, L.; Voigt, T.; Waldmann, H. Charting biological and chemical space: PSSC and SCONP as guiding principles for the development of compound collections based on natural product scaffolds. *Qsar Comb. Sci.* **2006**, *25*, 449–456. [[CrossRef](#)]
4. Gali, R.; Banothu, J.; Porika, M.; Velpula, R.; Hnamte, S.; Bavantula, R.; Abbagani, S.; Busi, S. Indolylmethylene benzo [h] thiazolo [2,3-b] quinazolinones: Synthesis, characterization and evaluation of anticancer and antimicrobial activities. *Bioorg. Med. Chem. Lett.* **2014**, *24*, 4239–4242. [[CrossRef](#)] [[PubMed](#)]
5. Sørli, T. Molecular portraits of breast cancer: Tumour subtypes as distinct disease entities. *Eur. J. Cancer* **2004**, *40*, 2667–2675. [[CrossRef](#)] [[PubMed](#)]
6. Siegel, R.; Ward, E.; Brawley, O.; Jemal, A. Cancer statistics, 2011: The impact of eliminating socioeconomic and racial disparities on premature cancer deaths. *Ca-Cancer J. Clin.* **2011**, *61*, 212–236. [[CrossRef](#)] [[PubMed](#)]
7. Chabner, B.A.; Roberts Jr, T.G. Chemotherapy and the war on cancer. *Nat. Rev. Cancer* **2005**, *5*, 65. [[CrossRef](#)]
8. Rosen, H.; Abribat, T. The rise and rise of drug delivery. *Nat. Rev. Drug Discov.* **2005**, *4*, 381. [[CrossRef](#)]
9. Kim, N.-D.; Im, E.-O.; Choi, Y.-H.; Yoo, Y.-H. Synthetic bile acids: Novel mediators of apoptosis. *Bmb Rep.* **2002**, *35*, 134–141. [[CrossRef](#)]
10. Carey, E.J.; Lindor, K.D. Chemoprevention of colorectal cancer with ursodeoxycholic acid: Cons. *Clin. Res. Hepatol. Gastroenterol.* **2012**, *36*, S61–S64. [[CrossRef](#)]
11. Serfaty, L.; Bissonnette, M.; Poupon, R. Ursodeoxycholic acid and chemoprevention of colorectal cancer. *Gastroenterol. Clin. Biol.* **2010**, *34*, 516–522. [[CrossRef](#)]
12. Serfaty, L. Chemoprevention of colorectal cancer with ursodeoxycholic acid: Pro. *Clin. Res. Hepatol. Gastroenterol.* **2012**, *36*, S53–S60. [[CrossRef](#)]
13. Tatsumura, T.; Sato, H.; Yamamoto, K.; Ueyama, T. Ursodeoxycholic acid prevents gastrointestinal disorders caused by anticancer drugs. *Jpn. J. Surg.* **1981**, *11*, 84–89. [[CrossRef](#)] [[PubMed](#)]
14. Pang, L.; Zhao, X.; Liu, W.; Deng, J.; Tan, X.; Qiu, L. Anticancer effect of ursodeoxycholic acid in human oral squamous carcinoma HSC-3 cells through the caspases. *Nutrients* **2015**, *7*, 3200–3218. [[CrossRef](#)]
15. Dyakova, L.; Culita, D.-C.; Marinescu, G.; Alexandrov, M.; Kalfin, R.; Patron, L.; Alexandrova, R. Metal Zn (II), Cu (II), Ni (II) complexes of ursodeoxycholic acid as putative anticancer agents. *Biotechnol. Biotechnol. Equip.* **2014**, *28*, 543–551. [[CrossRef](#)]
16. Marchesi, E.; Chinaglia, N.; Capobianco, M.L.; Marchetti, P.; Huang, T.-E.; Weng, H.-C.; Guh, J.-H.; Hsu, L.-C.; Perrone, D.; NavacchiaKomori, M.L. Dihydroartemisinin–bile acid hybridization as an effective approach to enhance dihydroartemisinin anticancer activity. *ChemMedChem* **2019**, *14*, 779–787. [[CrossRef](#)]
17. Gupta, A.; Kumar, B.S.; Negi, A.S. Current status on development of steroids as anticancer agents. *J. Steroid Biochem. Mol. Biol.* **2013**, *137*, 242–270. [[CrossRef](#)] [[PubMed](#)]
18. Agalave, S.G.; Maujan, S.R.; Pore, V.S. Click chemistry: 1,2,3-triazoles as pharmacophores. *Chem. Asian J.* **2011**, *6*, 2696–2718. [[CrossRef](#)]
19. Frank, E.; Molnar, J.; Zupko, I.; Kadar, Z.; Wolfling, J. Synthesis of novel steroidal 17 α -triazolyl derivatives via Cu (I)-catalyzed azide-alkyne cycloaddition, and an evaluation of their cytotoxic activity in vitro. *Steroids* **2011**, *76*, 1141–1148. [[CrossRef](#)]
20. Aly, M.R.E.S.; Saad, H.A.; Mohamed, M.A.M. Click reaction based synthesis, antimicrobial, and cytotoxic activities of new 1,2,3-triazoles. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 2824–2830. [[CrossRef](#)]
21. Aly, M.R.E.S.; Saad, H.A.; Abdel-Hafez, S.H. Synthesis, antimicrobial and cytotoxicity evaluation of new cholesterol congeners. *Beilstein J. Org. Chem.* **2015**, *11*, 1922–1932. [[CrossRef](#)] [[PubMed](#)]
22. Pertino, M.; Lopez, C.; Theoduloz, C. Schmeda-Hirschmann, G. 1,2,3-Triazole-substituted oleanolic acid derivatives: Synthesis and antiproliferative activity. *Molecules* **2013**, *18*, 7661–7674. [[CrossRef](#)] [[PubMed](#)]

23. Wei, G.; Luan, W.; Wang, S.; Cui, S.; Li, F.; Liu, Y.; Liu, Y.; Cheng, M. A library of 1,2,3-triazole-substituted oleanolic acid derivatives as anticancer agents: Design, synthesis, and biological evaluation. *Org. Biomol. Chem.* **2015**, *13*, 1507–1514. [[CrossRef](#)]
24. Bębenek, E.; Kadela-Tomanek, M.; Chrobak, E.; Latocha, M.; Boryczka, S. Novel triazoles of 3-acetylbetulin and betulone as anticancer agents. *Med. Chem. Res.* **2018**, *27*, 2051–2061. [[CrossRef](#)] [[PubMed](#)]
25. Majeed, R.; Sangwan, P.L.; Chinthakindi, P.K.; Khan, I.; Dangroo, N.A.; Thota, N.; Hamid, A.; Sharma, P.R.; Saxena, A.K.; Koul, S. Synthesis of 3-O-propargylated betulinic acid and its 1, 2, 3-triazoles as potential apoptotic agents. *Eur. J. Med. Chem.* **2013**, *63*, 782–792. [[CrossRef](#)]
26. Sidova, V.; Zoufaly, P.; Pokorný, J.; Dzubak, P.; Hajdúch, M.; Popa, I.; Urban, M. Cytotoxic conjugates of betulinic acid and substituted triazoles prepared by Huisgen Cycloaddition from 30-azidoderivatives. *PLoS ONE* **2017**, *12*, e0171621. [[CrossRef](#)]
27. Csuk, R.; Barthel, A.; Kluge, R.; Strohl, D. Synthesis, cytotoxicity and liposome preparation of 28-acetylenic betulin derivatives. *Bioorg. Med. Chem.* **2010**, *18*, 7252–7259. [[CrossRef](#)]
28. Csuk, R.; Barthel, A.; Sczepek, R.; Siewert, B.; Schwarz, S. Synthesis, encapsulation and antitumor activity of new betulin derivatives. *Arch. Pharm.* **2011**, *344*, 37–49. [[CrossRef](#)]
29. Jurášek, M.; Džubák, P.; Sedlák, D.; Dvořáková, H.; Hajdúch, M.; Bartůnek, P.; Drašar, P. Preparation, preliminary screening of new types of steroid conjugates and their activities on steroid receptors. *Steroids* **2013**, *78*, 356–361. [[CrossRef](#)]
30. Mohamed, Z.; El-Koussi, N.A.; Mahfouz, N.M.; Youssef, A.F.; Jaleel, G.A.A.; Shouman, S.A. Cu (I) catalyzed alkyne-azide 1, 3-dipolar cycloaddition (CuAAC): Synthesis of 17 α -[1-(substituted phenyl)-1,2,3-triazol-4-yl]-19-nor-testosterone-17 β -yl acetates targeting progestational and antipro-liferative activities. *Eur. J. Med. Chem.* **2015**, *97*, 75–82. [[CrossRef](#)]
31. Kadar, Z.; Kovacs, D.; Frank, E.; Schneider, G.; Huber, J.; Zupko, I.; Bartok, T.; Wolfling, J. Synthesis and in vitro antiproliferative activity of novel androst-5-ene triazolyl and tetrazolyl derivatives. *Molecules* **2011**, *16*, 4786–4806. [[CrossRef](#)] [[PubMed](#)]
32. Molnar, J.; Frank, E.; Minorics, R.; Kadar, Z.; Ocsovszki, I.; Schonecker, B.; Wolfling, J.; Zupko, I. A click approach to novel D-ring-substituted 16 α -triazolylestrone derivatives and characterization of their antiproliferative properties. *PLoS ONE* **2015**, *10*, e0118104. [[CrossRef](#)]
33. Kadar, Z.; Molnar, J.; Schneider, G.; Zupko, I.; Frank, E. A facile ‘click’ approach to novel 15 β -triazolyl-5 α -androstane derivatives, and an evaluation of their antiproliferative activities in vitro. *Bioorg. Med. Chem.* **2012**, *20*, 1396–1402. [[CrossRef](#)] [[PubMed](#)]
34. Kadar, Z.; Baji, A.; Zupko, I.; Bartok, T.; Wolfling, J.; Frank, E. Efficient approach to novel 1 α -triazolyl-5 α -androstane derivatives as potent antiproliferative agents. *Org. Biomol. Chem.* **2011**, *9*, 8051–8057. [[CrossRef](#)] [[PubMed](#)]
35. Kadar, Z.; Frank, E.; Schneider, G.; Molnar, J.; Zupko, I.; Koti, J.; Schonecker, B.; Wolfling, J. Efficient synthesis of novel A-ring-substituted 1, 2, 3-triazolylcholestane derivatives via catalytic azide-alkyne cycloaddition. *Arkivoc* **2012**, *3*, 279–296. [[CrossRef](#)]
36. Solum, E.J.; Vik, A.; Hansen, T.V. Synthesis, cytotoxic effects and tubulin polymerization inhibition of 1,4-disubstituted 1,2,3-triazole analogs of 2-methoxyestradiol. *Steroids* **2014**, *87*, 46–53. [[CrossRef](#)]
37. Banday, A.H.; Shameem, S.A.; Gupta, B.; Kumar, H.S. D-ring substituted 1,2,3-triazolyl 20-keto pregnenanes as potential anticancer agents: Synthesis and biological evaluation. *Steroids* **2010**, *75*, 801–804. [[CrossRef](#)]
38. Navacchia, M.L.; Marchesi, E.; Perrone, D. Bile acid conjugates with anticancer activity: Most recent research. *Molecules* **2021**, *26*, 25. [[CrossRef](#)]
39. Navacchia, M.L.; Marchesi, E.; Mari, L.; Chinaglia, N.; Gallerani, E.; Gavioli, R.; Capobianco, M.L.; Perrone, D. Rational Design of Nucleoside–Bile Acid Conjugates Incorporating a Triazole Moiety for Anticancer Evaluation and SAR Exploration. *Molecules* **2017**, *22*, 1710. [[CrossRef](#)]
40. Perrone, D.; Bortolini, O.; Fogagnolo, M.; Marchesi, E.; Mari, L.; Massarenti, C.; Navacchia, M.L.; Sforza, F.; Varani, K.; Capobianco, M.L. Synthesis and in vitro cytotoxicity of deoxyadenosine–bile acid conjugates linked with 1,2,3-triazole. *New J. Chem.* **2013**, *37*, 3559–3567. [[CrossRef](#)]
41. Agarwal, D.S.; Krishna, V.S.; Sriram, D.; Yogeewari, P.; Sakhuja, R. Clickable conjugates of bile acids and nucleosides: Synthesis, characterization, in vitro anticancer and antituberculosis studies. *Steroids* **2018**, *139*, 35–44. [[CrossRef](#)] [[PubMed](#)]
42. Agarwal, D.S.; Singh, R.P.; Lohitesh, K.; Jha, P.N.; Chowdhury, R.; Sakhuja, R. Synthesis and evaluation of bile acid amides of α -cyanostilbenes as anticancer agents. *Mol. Divers.* **2018**, *22*, 305–321. [[CrossRef](#)] [[PubMed](#)]
43. Agarwal, D.S.; Anantaram, H.S.; Sriram, D.; Yogeewari, P.; Nanjegowda, S.H.; Mallu, P.; Sakhuja, R. Synthesis, characterization and biological evaluation of bile acid-aromatic/heteroaromatic amides linked via amino acids as anti-cancer agents. *Steroids* **2016**, *107*, 87–97. [[CrossRef](#)] [[PubMed](#)]
44. Vatmurge, N.S.; Hazra, B.G.; Pore, V.S.; Shirazi, F.; Deshpande, M.V.; Kadreppa, S.; Chattopadhyay, S.; Gonnade, R.G. Synthesis and biological evaluation of bile acid dimers linked with 1, 2, 3-triazole and bis- β -lactam. *Org. Biomol. Chem.* **2008**, *6*, 3823–3830. [[CrossRef](#)]
45. Jarrahpour, A.; Fathi, J.; Mimouni, M.; Hadda, T.; Sheikh, J.; Chohan, Z.; Parvez, A. Petra, osiris and molinspiration (POM) together as a successful support in drug design: Antibacterial activity and biopharmaceutical characterization of some azo Schiff bases. *Med. Chem. Res.* **2012**, *21*, 1984–1990. [[CrossRef](#)]

46. Ertl, P.; Rohde, B.; Selzer, P. Fast calculation of molecular polar surface area as a sum of fragment-based contributions and its application to the prediction of drug transport properties. *J. Med. Chem.* **2000**, *43*, 3714–3717. [[CrossRef](#)]
47. Gündüz, M.G.; Uğur, S.B.; Güney, F.; Özkul, C.; Krishna, V.S.; Kaya, S.; Sriram, D.; Doğan, S.D. 1,3-Disubstituted urea derivatives: Synthesis, antimicrobial activity evaluation and in silico studies. *Bioorg. Chem.* **2020**, *102*, 2020. [[CrossRef](#)]
48. Bartzatt, R.; Donigan, L. Applying pattern recognition methods to analyze the molecular properties of a homologous series of nitrogen mustard agents. *AAPS PharmSciTech* **2006**, *7*, 35. [[CrossRef](#)]
49. Hao, T.; Ling, Y.; Wu, M.; Shen, Y.; Gao, Y.; Liang, S.; Gao, Y.; Qian, S. Enhanced oral bioavailability of docetaxel in rats combined with myricetin: In situ and in vivo evidences. *Eur. J. Pharm. Sci.* **2017**, *101*, 71–79. [[CrossRef](#)]
50. Sharma, S.; Mazumdar, S.; Italiya, K.S.; Date, T.; Mahato, R.I.; Mittal, A.; Chitkara, D. Cholesterol and Morpholine Grafted Cationic Amphiphilic Copolymers for miRNA-34a Delivery. *Mol. Pharm.* **2018**, *15*, 2391–2402. [[CrossRef](#)]
51. Italiya, K.S.; Mazumdar, S.; Sharma, S.; Chitkara, D.; Mahato, R.I.; Mittal, A. Self-assembling lisofylline-fatty acid conjugate for effective treatment of diabetes mellitus. *Nanomedicine* **2019**, *15*, 175–187. [[CrossRef](#)] [[PubMed](#)]