Supplementary Materials: Nucleic Acid Delivery by Solid Lipid Nanoparticles Containing Switchable Lipids: Plasmid DNA vs. Messenger RNA

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pDNA in SLN4



Figure S1. Binding, protection and release capacity of pDNA-DX-SLN4. Vector was able to bind and release the pDNA. However, vector showed low capacity of protection of pDNA.



Figure S2. Flow cytometry analysis of cellular uptake of pDNA-vectors using Nile Red-labeled SLNs in ARPE-19. A: Intracellular uptake at 37 °C. B: Intracellular uptake at 4 °C. The higher the displacement to the right, the higher the fluorescence intensity in the cells. At 37° C both pDNA-DX-SLN1 and pDNA-HA-SLN1 were more displaced than those with SLN2. However, at 4° C, both pDNA-DX-SLN2 and pDNA-HA-SLN2 were more displaced to the right than those with SLN1.



Figure S3. Flow cytometry analysis of cellular uptake of mRNA-vectors using Nile Red-labeled SLNs in ARPE-19. A: Intracellular uptake at 37 °C. B: Intracellular uptake at 4 °C The higher the displacement to the right, the higher the fluorescence intensity in the cells. At both 37 °C and 4 °C, all SLN1-based vectors the histograms were more displaced to the right than those formulated with SLN2.



Figure S4. Flow cytometry analysis of cellular uptake of pDNA-vectors using Nile Red-labeled SLNs in HEK-293 cells. The higher the displacement to the right, the higher the fluorescence intensity in the cells. A: Intracellular uptake at 37 °C. B: Intracellular uptake at 4 °C. The displacement to the right was much higher at 37 °C than at 4 °C with all the vectors.



Figure S5. Flow cytometry analysis of cellular uptake of mRNA-vectors using Nile Red-labeled SLNs in HEK-293 cells. A: Intracellular uptake at 37 °C. B: Intracellular uptake at 4 °C. The higher the displacement to the right, the higher the fluorescence intensity in the cells. SLN1-based vectors were more displaced to the right than SLN2-based vectors at both temperatures, except mRNA-P0.5-SLN2 and mRNA-P1-SLN2 formulations at 4° C, which were more displaced than mRNA-P-SLN1.



Figure S6. Study of the binding, protection and release of pDNA-based vectors formulated with SLN1 and SLN2 after 2 months of storage. It can be seen that at 2 months vectors were able to bind the pDNA, but the protection ability decreased respect to time 0.



Figure S7. Study of the binding, protection and release of mRNA-based vectors formulated with SLN1 and SLN2 after 2 months of storage. It can be seen that at 2 months vectors were able to bind the mRNA, but the protection capacity decreased.



Figure S8. Microscopy images of the intracellular disposition in ARPE-19 cells of mRNA combined with P, SLN1 and the combination of both components. Images were captured with a Leica DM IL LED Fluo inverted microscope (Leica Microsystems CMS GmbH, Wetzlar, Germany). Red color: mRNA; blue color: cell nuclei. It can be seen that SLN1 alone did not condense the mRNA as much as P did.



Figure S9. Microscopy images of ARPE-19 cells 24 h, 48 h, 72 h, 96 h and 11 days after transfection with pDNA-DX-SLN1 and mRNA-P0.25-SLN1 vectors. Green color corresponds to the GFP. Images were captured with a Leica DM IL LED Fluo inverted microscope (Leica Microsystems CMS GmbH, Wetzlar, Germany). Scale bar: 20 μ m. mRNA formulations showed the highest transfection efficacy at 24-48 h, and the protein expression decreased notably at 96 h, whereas pDNA formulations showed the maximum transfection at 72-96 h. In both cases it lasted at least 11 days.



Figure S10. Optimization of the dose of mRNA-based vectors in ARPE-19 cells. A: percentage of transfected cells. B: Intensity of fluorescence of transfected cells. At the dose of 2.5 μ g, vectors prepared with SLN1 showed transfection percentages of almost 100%. Therefore, this dose was discarded because it was difficult to optimize the formulations from such a high percentage. The dose of 0.8 μ g in all cases gave very low percentages and intensities of transfection. Accordingly, the dose of 1.5 μ g of mRNA was selected for the different studies carried out with freshly prepared vectors in ARPE-19 cells.