# Supporting information:

# Fluorimetric and CD recognition between various ds-DNA/RNA depends on a cyanine connectivity in cyanine-guanidiniocarbonyl-pyrrole conjugate

Tamara Šmidlehner,<sup>1,4</sup> Marta Košćak,<sup>1</sup> Ksenija Božinović,<sup>2</sup> Dragomira Majhen,<sup>2</sup> Carsten Schmuck, <sup>+3</sup> and Ivo Piantanida<sup>1,\*</sup>

- <sup>1</sup> Division of Organic Chemistry and Biochemistry, Ruđer Bošković Institute, Bijenička Cesta 54, 10000 Zagreb, Croatia; <u>marta.koscak@irb.hr (M.K.), pianta@irb.hr</u> (I.P.).
- <sup>2</sup> Division of Molecular Biology, Ruđer Bošković Institute, Bijenička cesta 54, 10 000 Zagreb, Croatia. ksenija.bozinovic@irb.hr (K.B.), <u>dragomira.majhen@irb.hr</u> (D.M.)
- <sup>3</sup> University of Duisburg-Essen, Institute of Organic Chemistry, Essen, Germany.
- <sup>4</sup> Present address: National Institute of Chemistry, Hajdrihova 19, POBox 660, SI-1001 Ljubljana, Slovenia, tamara.smidlehner@ki.si\_(T.Š.).
- \* Correspondence: pianta@irb.hr (I.P.); Tel.: +385-1-4571-326



#### 1. Structural properties of studied DNA and RNA

- 2. Physico-chemical properties of aqueous solutions
- 3. Study of interactions of 1 and 2 with double-stranded DNA/RNA

# 1. Structural properties of studied DNA and RNA

Polynucleotides were purchased as noted: poly dGdC – poly dGdC, poly dAdT – poly dAdT, poly A – poly U, *calf thymus* (ct)-DNA (Aldrich) and dissolved in sodium cacodylate buffer, I = 0.05 M, pH=7.0. The ct-DNA was additionally sonicated and filtered through a 0.45 mm filter to obtain mostly short (ca. 100 base pairs) rod-like B-helical DNA fragments [1]. The polynucleotide concentration was determined spectroscopically [2] as the concentration of phosphates (corresponds to c(nucleobase)).

Structure type	Groove width [Å]		Groove depth [Å]	
	major	minor	major	minor
<sup>[a]</sup> poly rA – poly rU	3.8	10.9	13.5	2.8
<sup>[b]</sup> ct-DNA (48% of GC-pairs)	11.4	3.3	7.5	7.9
<sup>[b]</sup> poly dAdT – poly dAdT	11.2	6.3	8.5	7.5
<sup>[c]</sup> poly dGdC – poly dGdC	13.5	9.5	10.0	7.2

Table S1. Groove widths and depths for selected nucleic acid conformation [3,4].

[a] A - helical structure

[b] B - helical structure

[c] B- helical structure with sterically blocked minor groove by amino groups of guanines

## 2. Physico-chemical properties of aqueous solutions

#### 2.1. Solubility

All compounds were dissolved in water to give stock solutions of 10<sup>-3</sup> M. The stock solutions where stored at -20 °C, and working aliquots kept at +25 °C. No visible precipitation or degradation was noticed over several months.

#### 2.2. UV/Vis and fluorescence spectra, stability

The experiments where performed in buffer solution (sodium cacodylate buffer, I = 0.05 M, pH = 7.0). The absorbancies of **1** and **2** buffered solutions were proportional to their concentration within the used concentration range.



**Figure S1. a)** Dependence of UV/Vis spectra on concentration of **1**, **b)** Dependence of Abs(504 nm) on c (**1**) at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S2. a)** Dependence of UV/Vis spectra on concentration of **1**, **b)** Dependence of Abs(504 nm) on c (**1**) at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S3. a)** Dependence of UV/Vis spectra on concentration of **2**, **b)** Dependence of Abs(504 nm) on c (**2**) at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S4. a)** Dependence of UV/Vis spectra on concentration of **2**, **b)** Dependence of Abs(504 nm) on c (**2**) at pH 5.0, sodium cacodylate buffer, I = 0.05 M.

# *Temperature dependence:*



**Figure S5.** Temperature dependence of UV/Vis spectra ( $c = 1.0 \times 10^{-5}$  M) at pH 7.0, sodium cacodylate buffer, I = 0.05 M. LEFT: 1; RIGHT: 2.

**Table S2.** Electronic absorption data of **1** and **2** determined from data on Figures S1-S4.

зотроина, рп	NIIII	E/ mmoi · cm-
1, pH 7.0	504	18018
1, pH 5.0	504	18868
2, pH 7.0	508	18629
2, pH 5.0	505	20917

#### Compound, pH $\lambda/nm \epsilon/mmol^{-1} cm^{2}$

# 3. Study of interactions with double-stranded DNA/RNA in aqueous medium

3.1 Fluorescence spectrophotometric titrations

3.1.1. Fluorescence Spectrophotometric titrations with 1

General conditions: slits 5-10; emission at 530 nm; excitation: 505 nm



**Figure S6. a)** Changes in fluorescence spectrum of **1** ( $c = 5.0 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with ctDNA; **b)** Dependence of **1** intensity at  $\lambda_{max} = 532$  nm on c(ctDNA), at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S7. a)** Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with ctDNA; **b)** Dependence of **1** intensity at  $\lambda_{max} = 531$  nm on c(ctDNA), at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S8. a)** Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dAdT)<sub>2</sub>; **b)** Dependence of **1** intensity at  $\lambda_{max} = 533$  nm on  $c(p(dAdT)_2)$ , at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S9. a)** Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dAdT)<sub>2</sub>; **b)** Dependence of **1** intensity at  $\lambda_{max} = 533$  nm on  $c(p(dAdT)_2)$ , at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S10.** a) Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dGdC)<sub>2</sub>; b) Dependence of **1** intensity at  $\lambda_{max} = 530$  nm on  $c(p(dGdC)_2)$ , at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S11. a)** Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dGdC)<sub>2</sub>; **b)** Dependence of **1** intensity at  $\lambda_{max} = 530$  nm on  $c(p(dGdC)_2)$ , at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S12.** a) Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with pApU; b) Dependence of **1** intensity at  $\lambda_{max} = 534$  nm on c(pApU), at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S13**. **a)** Changes in fluorescence spectrum of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with pApU; **b)** Dependence of **1** intensity at  $\lambda_{max} = 534$  nm on c(pApU), at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S14**. Changes in fluorescence of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon addition of polynucleotides at pH 7.0.



**Figure S15**. Changes in fluorescence of **1** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon addition of polynucleotides at pH 5.0.

### 3.1.2. Fluorescence Spectrophotometric titrations with 2 General conditions: slits 5-10; emission at 530 nm; excitation: 505 nm



**Figure S16. a)** Changes in fluorescence spectrum of **2** ( $c = 5.0 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with ctDNA; **b)** Dependence of **2** intensity at  $\lambda_{max} = 528$  nm on c(ctDNA), at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S17. a)** Changes in fluorescence spectrum of **2** ( $c = 5.0 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with ctDNA; **b)** Dependence of **2** intensity at  $\lambda_{max} = 527$  nm on c(ctDNA), at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S18. a)** Changes in fluorescence spectrum of **2** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dAdT)<sub>2</sub>; **b)** Dependence of **2** intensity at  $\lambda_{max} = 526$  nm on  $c(p(dAdT)_2)$ , at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S19.** a) Changes in fluorescence spectrum of 2 ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dAdT)<sub>2</sub>; b) Dependence of 2 intensity at  $\lambda_{max} = 526$  nm on  $c(p(dAdT)_2)$ , at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure 7.** a) Changes in fluorescence spectrum of 2 ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dGdC)<sub>2</sub>; b) Dependence of 2 intensity at  $\lambda_{max} = 528$  nm on  $c(p(dGdC)_2)$ , at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S20.** a) Changes in fluorescence spectrum of 2 ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with p(dGdC)<sub>2</sub>; b) Dependence of 2 intensity at  $\lambda_{max} = 528$  nm on  $c(p(dGdC)_2)$ , at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S21.** a) Changes in fluorescence spectrum of 2 ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with pApU; b) Dependence of 2 intensity at  $\lambda_{max} = 533$  nm on c(pApU), at pH 7.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S22.** a) Changes in fluorescence spectrum of 2 ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon titration with pApU; b) Dependence of 2 intensity at  $\lambda_{max} = 532$  nm on c(pApU), at pH 5.0, sodium cacodylate buffer, I = 0.05 M.



**Figure S23**. Changes in fluorescence of **2** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon addition of polynucleotides at pH 7.0.



**Figure S24**. Changes in fluorescence of **2** ( $c = 5 \times 10^{-7}$  M,  $\lambda_{exc} = 505$  nm) upon addition of polynucleotides at pH 5.0.

## 3.2. Circular dichroism (CD) experiments



### 3.2.1. CD titrations with 1

**Figure S25.** CD titration of a) ctDNA, b)  $p(dAdT)_2$ , c)  $p(dGdC)_2$ , d) pApU (c = 2 × 10<sup>-5</sup> M) with **1** at molar ratios r = [compound] / [polynucleotide] (pH 7.0, buffer sodium cacodylate, I = 0.05 M).



**Figure S26.** CD titration of a) ctDNA, b)  $p(dAdT)_2$ , c)  $p(dGdC)_2$ , d) pApU (c = 2 × 10<sup>-5</sup> M) with **1** at molar ratios r = [compound] / [polynucleotide] (pH 5.0, buffer sodium cacodylate, I = 0.05 M).

## 3.2.2. CD titrations with 2



**Figure S27.** CD titration of a) ctDNA, b)  $p(dAdT)_2$ , c)  $p(dGdC)_2$ , d) pApU (c = 2 × 10<sup>-5</sup> M) with **2** at molar ratios r = [compound] / [polynucleotide] (pH 7.0, buffer sodium cacodylate, I = 0.05 M).



**Figure S28.** CD titration of a) ctDNA, b)  $p(dAdT)_2$ , c)  $p(dGdC)_2$ , d) pApU (c = 2 × 10<sup>-5</sup> M) with **2** at molar ratios r = [compound] / [polynucleotide] (pH 5.0, buffer sodium cacodylate, I = 0.05 M).

### 3.3.1. ΔTm with 1

a)

a)



**Figure S29. a)** Melting curve of ctDNA upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **1** at pH 7.0 (buffer sodium cacodylate, l = 0.05 M), **b)** first derivation of absorbance on temperature.



**Figure S30.** a) Melting curve of ctDNA upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **1** at pH 5.0 (buffer sodium cacodylate, l = 0.05 M), b) first derivation of absorbance on temperature.



**Figure S31. a)** Melting curve of  $p(dAdT)_2$  upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **1** at pH 7.0 (buffer sodium cacodylate, I = 0.05 M), **b)** first derivation of absorbance on temperature



**Figure S32. a)** Melting curve of  $p(dAdT)_2$  upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **1** at pH 5.0 (buffer sodium cacodylate, I = 0.05 M), **b)** first derivation of absorbance on temperature



**Figure S33.** a) Melting curve of pApU upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **1** at pH 7.0 (buffer sodium cacodylate, l = 0.05 M), b) first derivation of absorbance on temperature

a)



**Figure S34.** a) Melting curve of pApU upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **1** at pH 5.0 (buffer sodium cacodylate, l = 0.05 M), b) first derivation of absorbance on temperature.



3.3.2. ΔTm with 2

a) b) **Figure S35. a)** Melting curve of ctDNA upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **2** at pH 7.0 (buffer sodium cacodylate, l = 0.05 M), **b)** first derivation of absorbance on temperature



**Figure S36. a)** Melting curve of ctDNA upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **2** at pH 5.0 (buffer sodium cacodylate, l = 0.05 M), **b)** first derivation of absorbance on temperature



**Figure S37. a)** Melting curve of  $p(dAdT)_2$  upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **2** at pH 7.0 (buffer sodium cacodylate, l = 0.05 M), **b)** first derivation of absorbance on temperature



**Figure S38. a)** Melting curve of  $p(dAdT)_2$  upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **2** at pH 5.0 (buffer sodium cacodylate, I = 0.05 M), **b)** first derivation of absorbance on temperature



**Figure S39.** a) Melting curve of pApU upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **2** at pH 7.0 (buffer sodium cacodylate, l = 0.05 M), b) first derivation of absorbance on temperature



**Figure S40.** a) Melting curve of pApU upon addition r = 0.1 and r = 0.2 ([compound/ [polynucleotide]) of **2** at pH 5.0 (buffer sodium cacodylate, l = 0.05 M), b) first derivation of absorbance on temperature.

899, doi: 10.1002/poc.680.

Saenger, W. *Principles of Nucleic Acid Structure*; *Springer-Verlag* **1983**, New York.
Cantor, C. R.; Schimmel; P. R. Biophysical Chemistry. *WH Freeman and Co.* **1980**, *3*, 1109-1181, San Francisco.

<sup>1.</sup> Chaires, J.B.; Dattagupta, N.; Crothers, D.M. Studies on interaction of anthracycline antibiotics and deoxyribonucleic acid: Equilibrium binding studies on interaction of daunomycin with deoxyribonucleic. *Biochemistry* **1982**, *21*, 3933–3940. 2. Tumir, L.-M.; Piantanida, I.; Cindrić Juranović, I.; et al New permanently charged phenanthridinium-nucleobase conjugates. Interactions with nucleotides and polynucleotides and recognition of ds-polyAH+. *J Phys Org Chem* **2003**, *16*, 891–