In vitro Antimycobacterial Activity and Physicochemical Characterization of Diaryl Ether Triclosan Analogues as Potential InhA Reductase Inhibitors

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Docking methodology

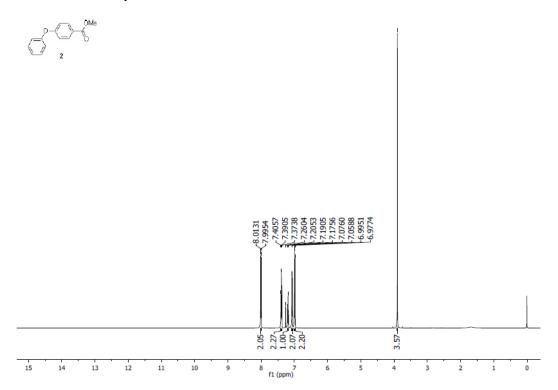
2

Selected ¹H and ¹³C NMR Spectra for some of our designed Compounds 3

Docking methodology

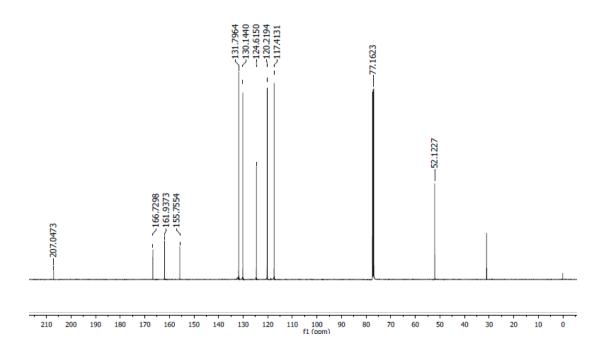
Discovery Studio 2.5 software (Accelrys Inc., San Diego, CA, USA) was used for docking analysis. Fully automated docking tool using "Dock ligands (CDOCKER)" protocol running on Intel (R) core (TM) i32370 CPU @ 2.4 GHz 2.4 GHz, RAM Memory 2 GB under the Windows 7.0 system. The crystal structure of InhA reductase enzyme (PDB code: 3FNH) was downloaded from protein data bank [32-34]. The docked compounds were built using Chem. 3D ultra 12.0 software [Chemical Structure Drawing Standard; Cambridge Soft corporation, USA (2010)], and copied to Discovery Studio 2.5 software. Automatic protein preparation module was used applying CHARMm force field. The binding site sphere has been defined automatically by the software. Herein, the prepared receptor is given as input for "input receptor molecule" parameter in the CDOCKER protocol parameter explorer. Force fields are applied on compounds Triclosan, **3**, **8b** and **8g** to get the minimum lowest energy structure. The obtained poses those have displayed the best ligand–InhA reductase enzyme interactions were selected and used for CDOCKER energy (protein-ligand interaction energies) calculations. Receptor–ligand interactions of the complexes were examined in 2D and 3D styles.

¹H NMR for compound 2



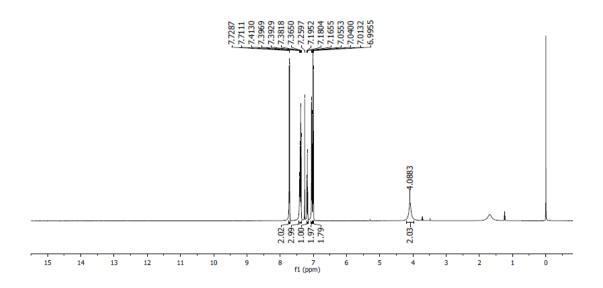
¹³C NMR for compound 2



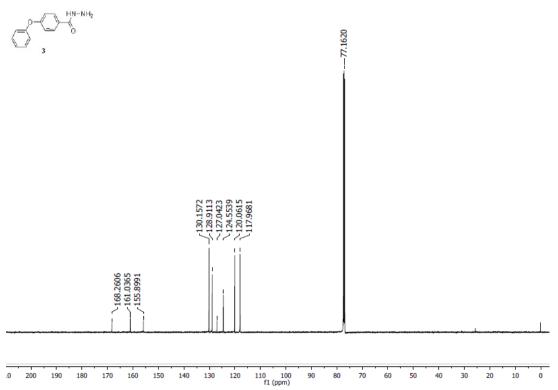


¹H NMR for compound 3

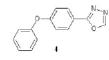


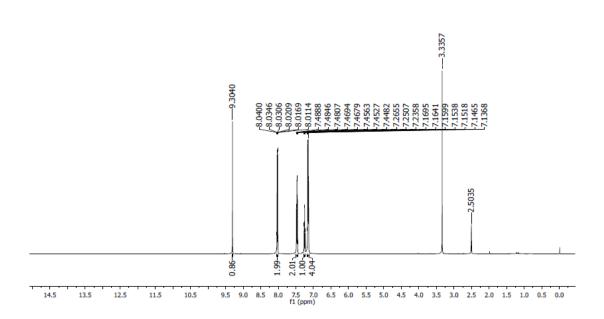


¹³C NMR for compound 3

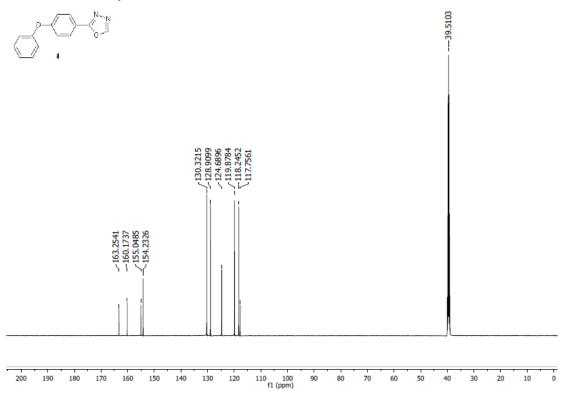


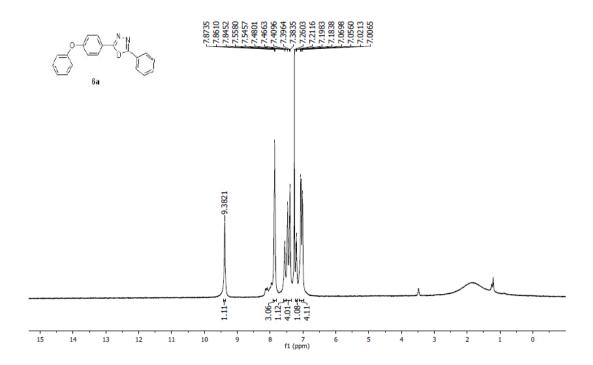
¹H NMR for compound 4



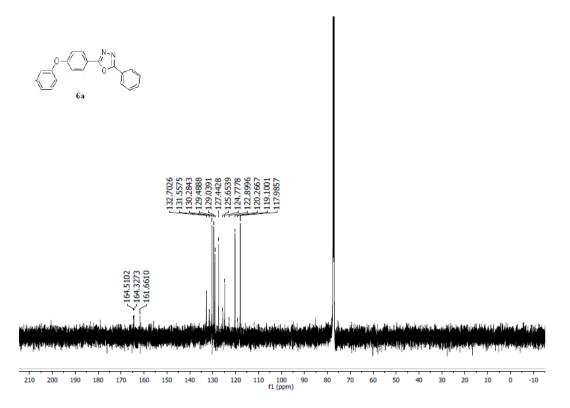


¹³C NMR for compound 4

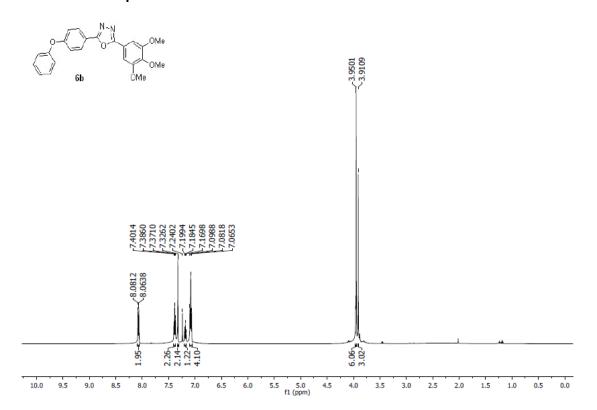




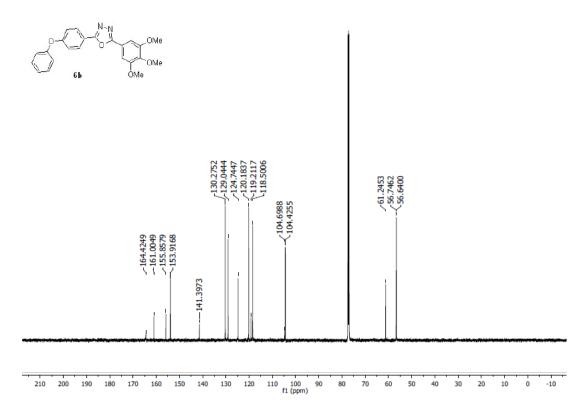




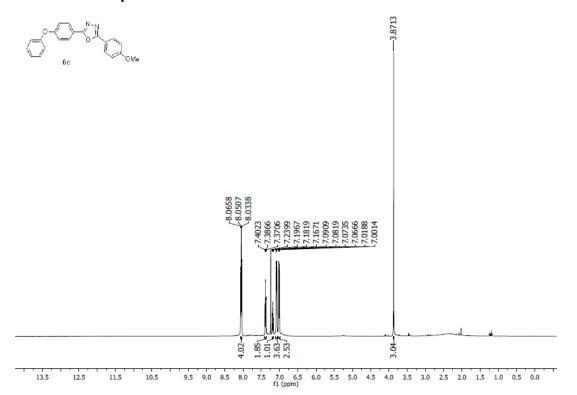
¹H NMR for compound 6b



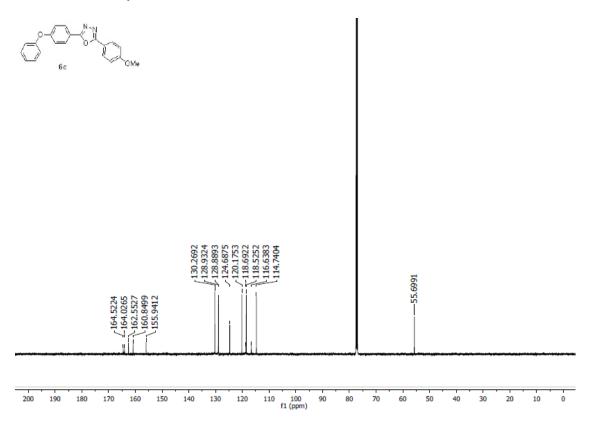
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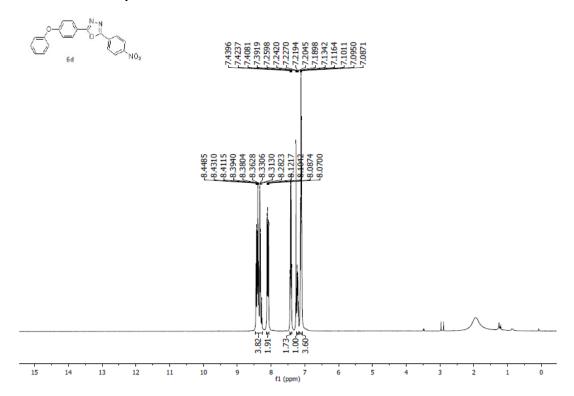
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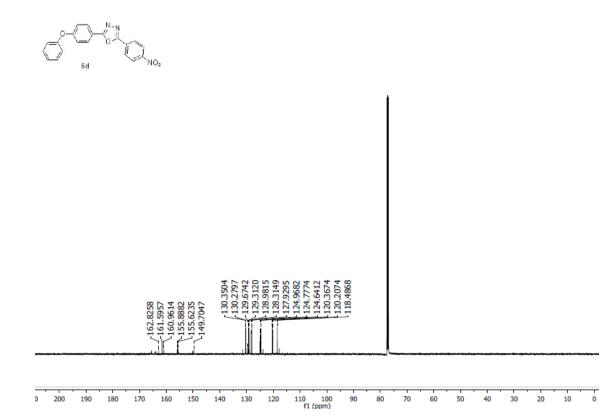
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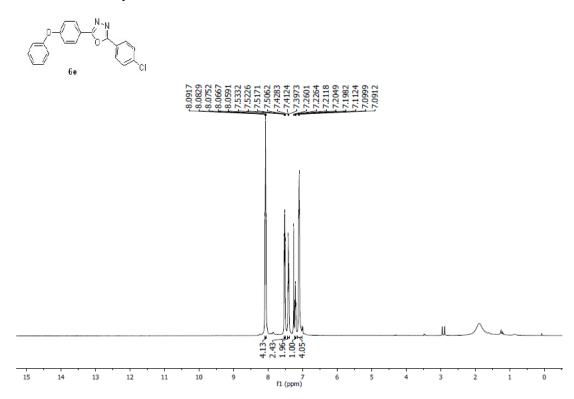
¹H NMR for compound 6d



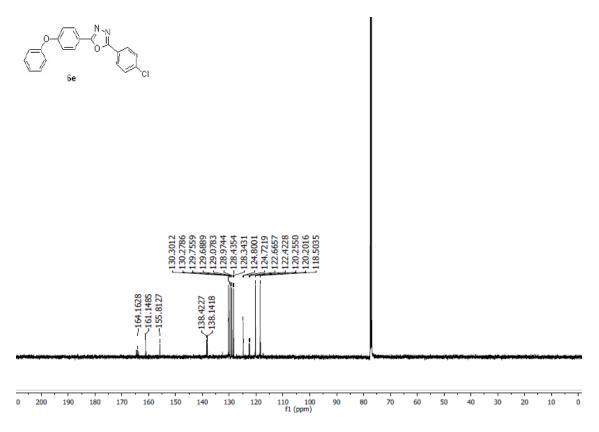
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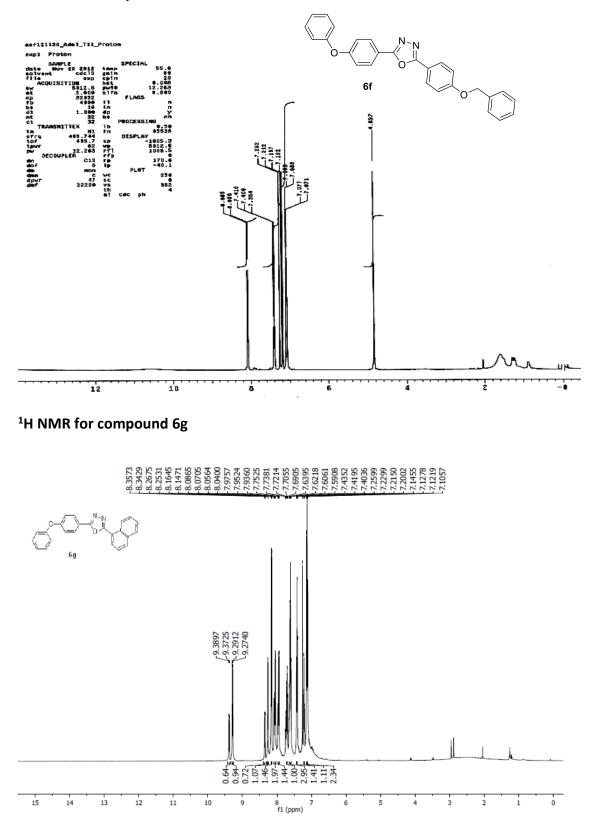
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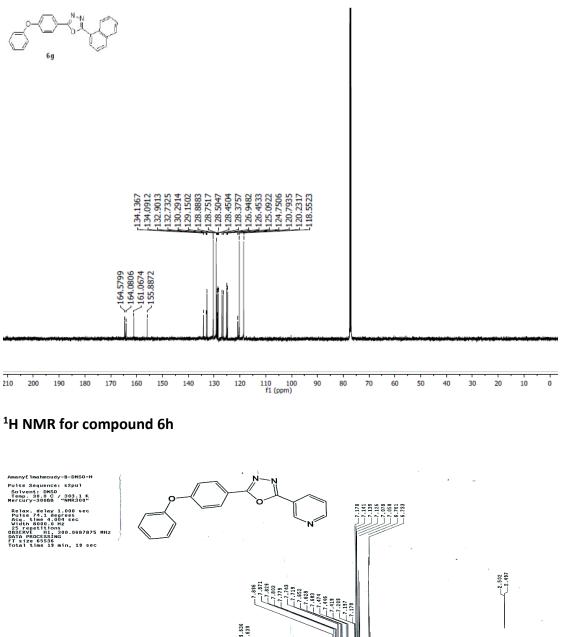
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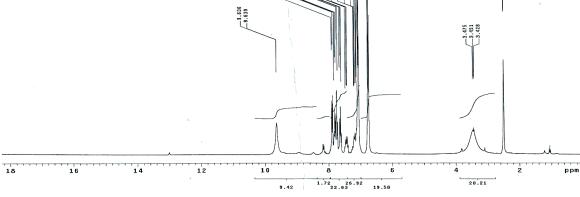


¹H NMR for compound 6f

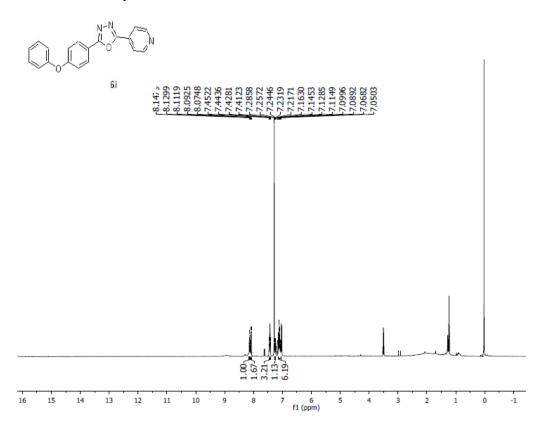


¹³C NMR for compound 6g

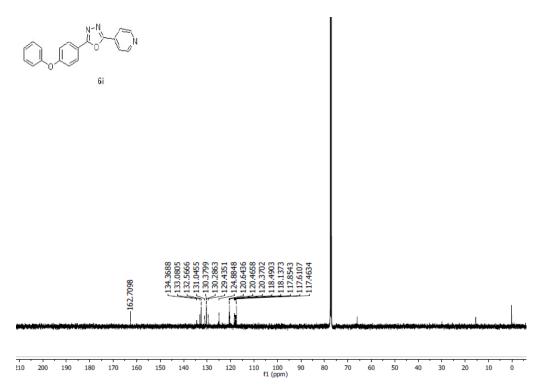




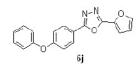
¹H NMR for compound 6i

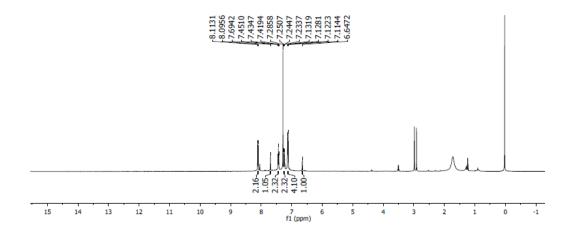


¹³C NMR for compound 6i

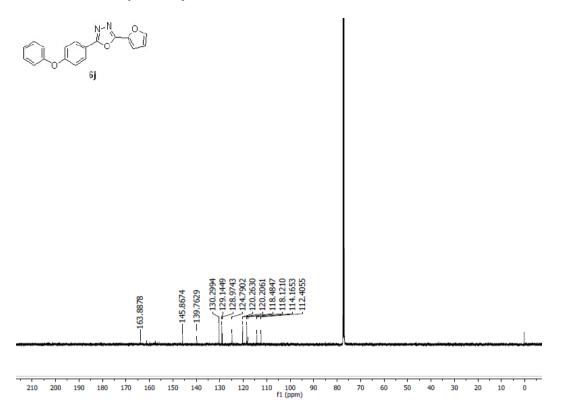


¹H NMR for compound 6j

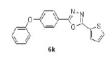


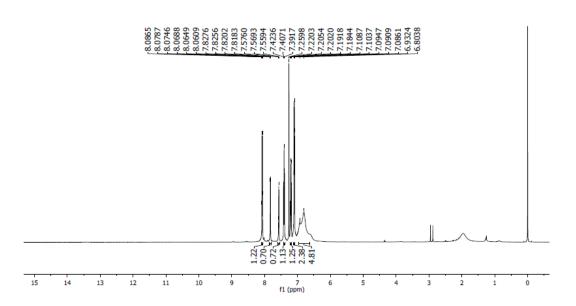


¹³C NMR for compound 6j

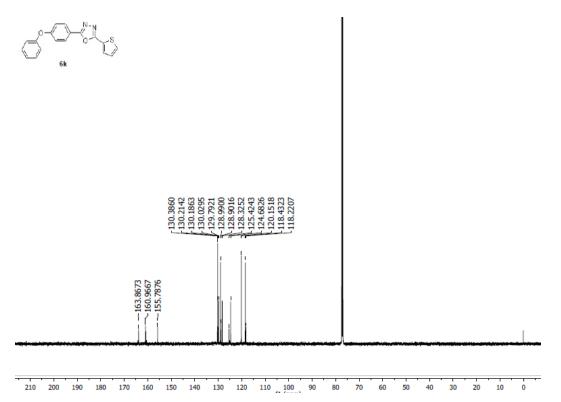


¹H NMR for compound 6k

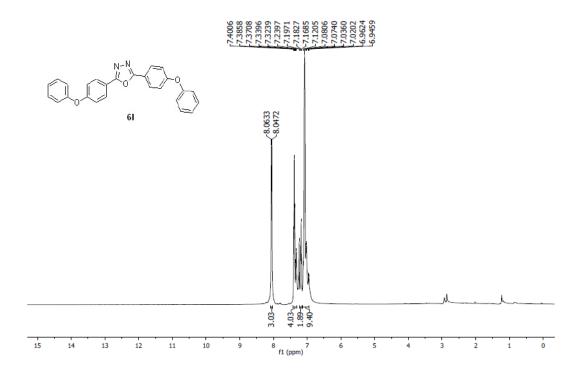




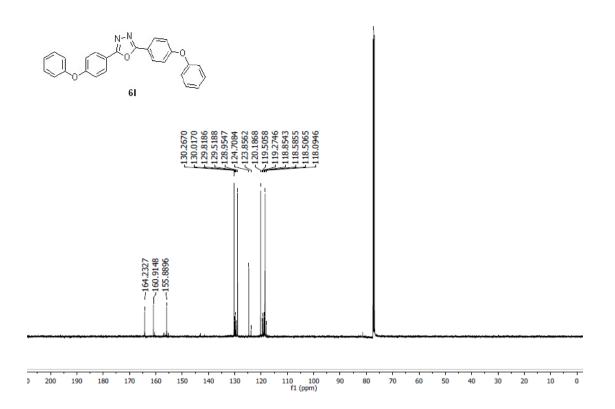
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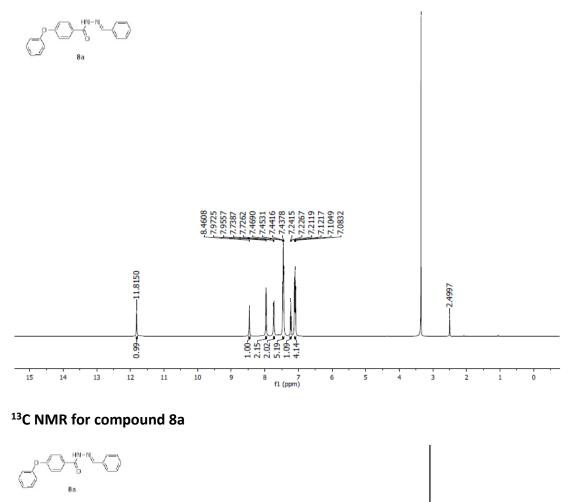
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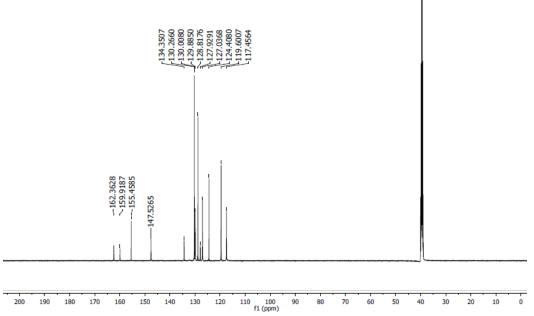


¹³C NMR for compound 6I

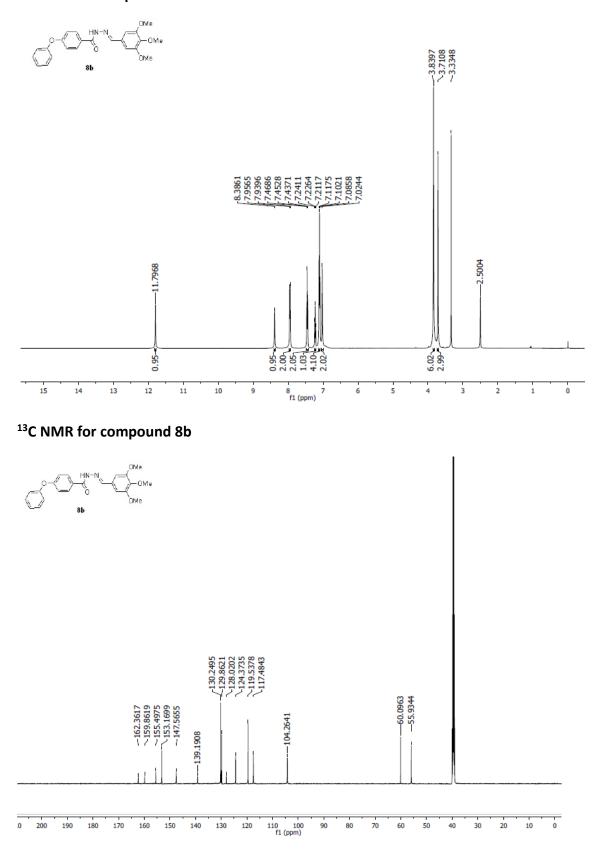


¹H NMR for compound 8a

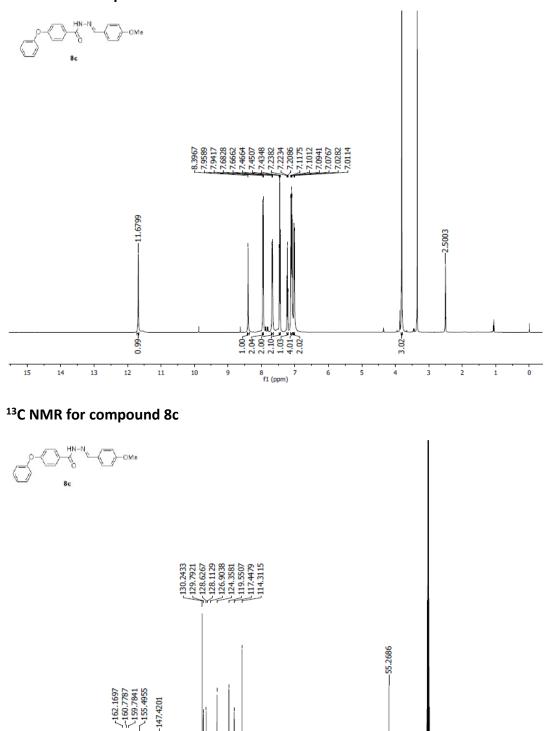




¹H NMR for compound 8b



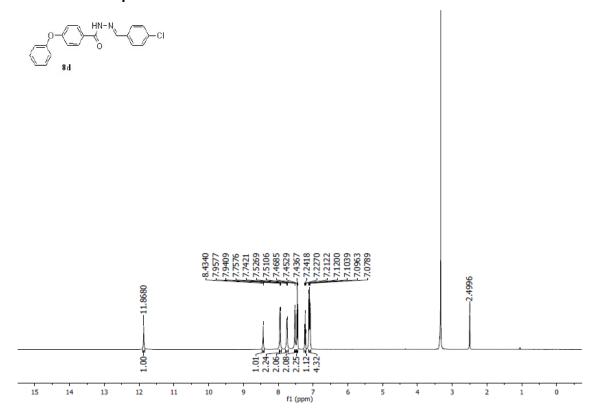
¹H NMR for compound 8c



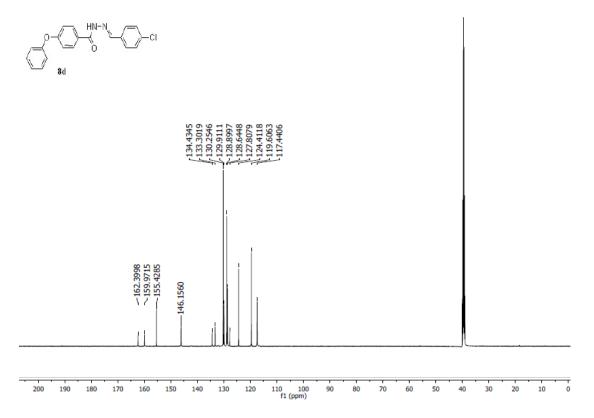
D 110 100 f1 (ppm)

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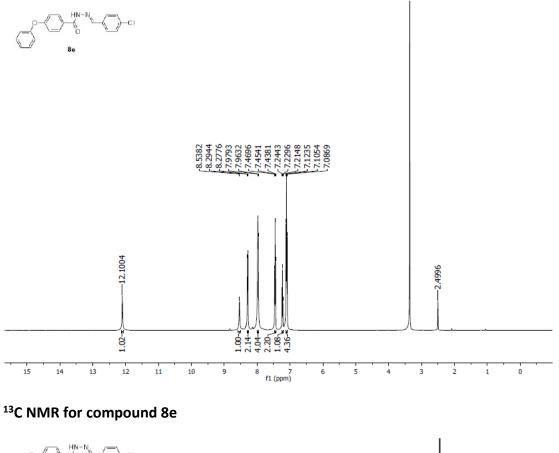
¹H NMR for compound 8d

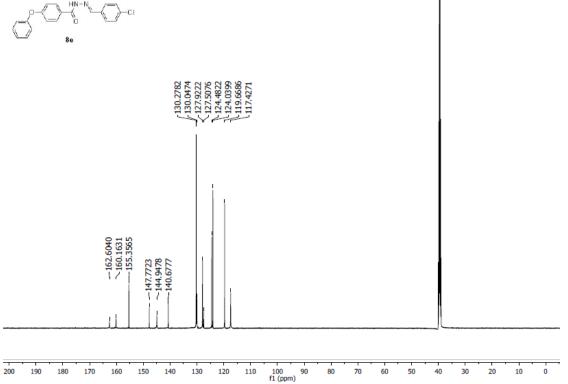


¹³C NMR for compound 8d

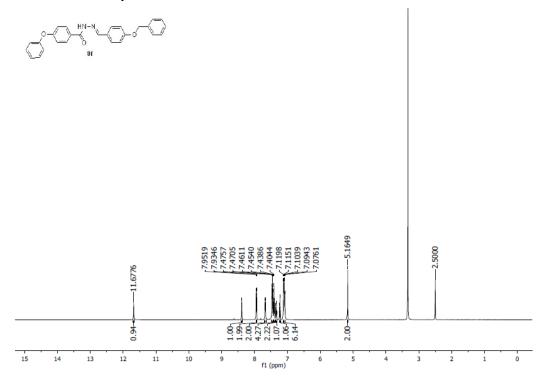


¹H NMR for compound 8e

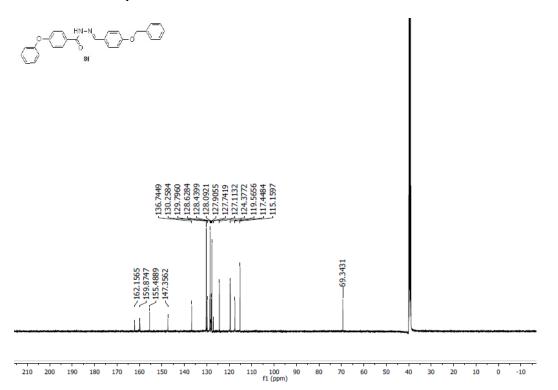


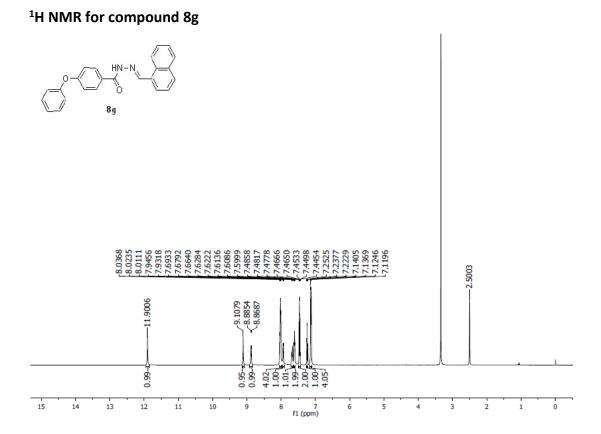




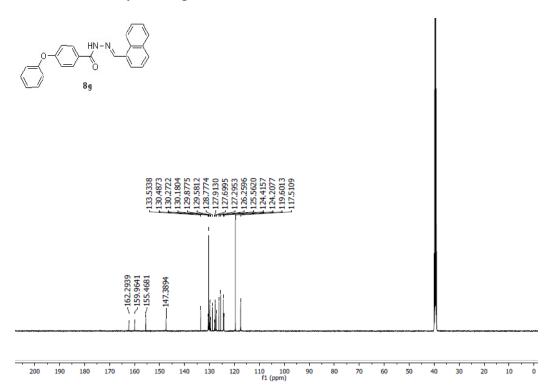


¹³C NMR for compound 8f

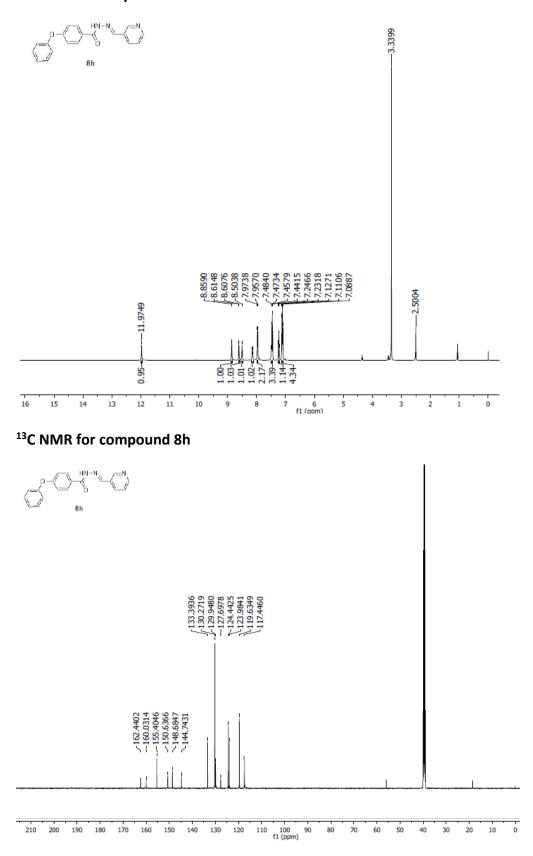




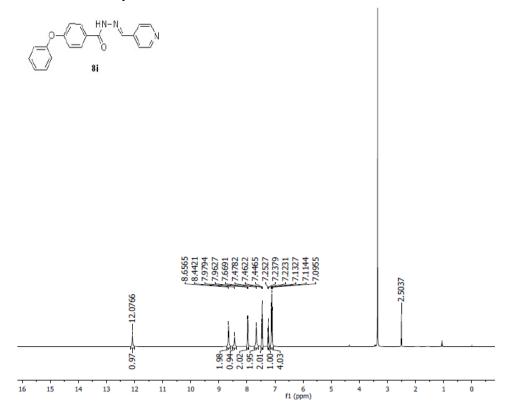
¹³C NMR for compound 8g



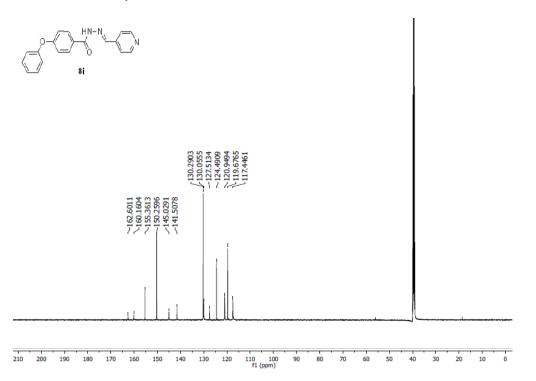
¹H NMR for compound 8h

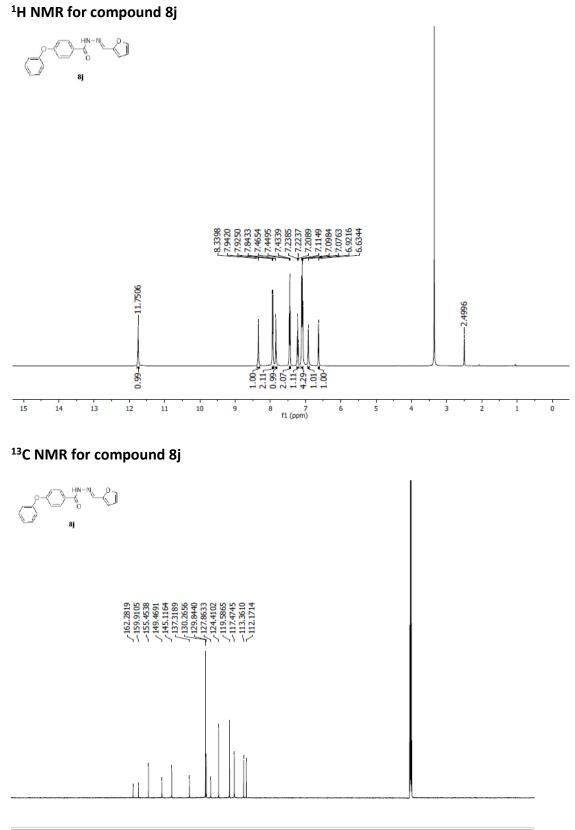






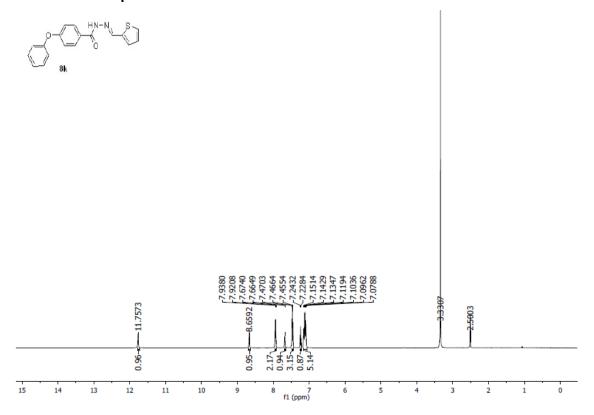
¹³C NMR for compound 8i





210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 f1 (ppm)

¹H NMR for compound 8k



¹³C NMR for compound 8k

