



Supplementary materials

2 **Conversion of D-fructose to**

3 5-acetoxymethyl-2-furfural using immobilized lipase

4 and cation exchange resin

5 Nhan Thanh Thien Huynh ^{1,2}, Kyung Won Lee ^{1,3}, Jin Ku Cho ^{1,2}, Yong Jin Kim ^{1,2}, Baekjin Kim ^{1,2}, 6 Jong Shik Shin ³, and Seunghan Shin ^{1,2,*}

- ¹ Green Chemistry & Engineering R&D Department, Korea Institute of Industrial Technology (KITECH), 89
 Yangdaegiro-gil, Ipjang-myeon, Seobuk-gu, Cheonan, Chungnam 31056, Korea
- 9 ² Department of Green Process and System Engineering, Korea University of Science and Technology (UST),
 217 Gajeong-ro, Yuseong-gu, Daejeon 34113, Korea
- 11 ³ Department of Biotechnology, Yonsei University, 50 Yonsei-ro, Seodaemun-gu, Seoul 03722, Korea
- 12 * Correspondence: shshin@kitech.re.kr. Tel: +82-41-5898-422. Fax: +82-41-5898-580
- 13 Received: date; Accepted: date; Published: date
- 14

15 **1.** Characterization methods:

16 1.1 Nuclear Magnetic Resonance (NMR):

NMR analysis was performed on a Bruker Spectrospin 300 (Bruker Corporation, Germany).
Figure S1 shows ¹H NMR of DAF (300 MHz, CDCl₃): 4.44-4.33 (m, 1H on C2), 4.30-4.23 (m, 2H of C5-CH₂), 4.21-4.13 (m, 2H on C3 and C4), 4.05-4.00 (m, 2H of C2-CH₂), 2.17-2.12 (m, 6H of 2
-COOCH₃; major anomer: 2.14 and 2.13; minor anomer: 2.17 and 2.12). ¹³C NMR of DAF (300 MHz, DMSO-d₆): 170.83-170.47 (d, 2C in -COO- groups), 100.86 (s, C5 in the ring), 78.85 (s, C2 in the ring), 76.49 (s, C4 in the ring), 75.57 (s, C3 in the ring), 66.16 (s, C6 in -CH₂ group), 64.62 (s, C1 in -CH₂ group), 21.04 (m, 2C in -CH₃ groups) (Figure S2).

AMF was also analyzed with NMR and Figure S4 shows ¹H NMR of AMF (300 MHz, THF-d₈): 9.62 (s, 1H on –CHO), 7.31-7.30 (d, 1H of C4 on furan ring), 6.68-6.67 (d, 1H of C3 on furan ring), 5.13 (s, 2H of –CH₂), 2.06 (s, 3H of –CH₃). ¹³C NMR of AMF (300 MHz, THF-d₈): 176.91 (s, C in –CHO group), 169.23 (s, C in –COO– group), 155.57 (s, C2 in the furan ring), 155.37 (s, C5 in the furan ring), 120.62 (s, C4 in the furan ring), 112.06 (s, C3 in the furan ring), 57.27 (s, C in –CH₂ group), 19.32 (s, C in –CH₃ group) (Figure S5).

30 1.2 High performance liquid chromatography (HPLC):

HPLC analysis of the dehydration products was performed on an Agilent 1200 series equipped
with a Bio-Rad Animex HPX-87P column and an RID detector (at 254 nm). H₂O was used as mobile
phase and the flow rate was adjusted at 0.6 mL/min. The retention times for AMF and HMF are 54.2

34 min. and 29.3 min. respectively (shown in figure S7).

35 1.3 Liquid chromatography combined with mass spectroscopy (LC-MS):

LC-MS analysis of the DAF was performed on an Agilent 1260 Infinity II series equipped with
 an InfinityLab LC/MSD mass spectrometer detector. HCOOH 0.001 M in H₂O was used as mobile
 phase and the flow rate was adjusted at 0.5 mL/min. DAF was ionized in API-ES positive mode and
 the mass spectrum was shown in Figure S3.

40 1.4 Gas chromatography combined with mass spectroscopy (GC-MS):

- 41 GC-MS analysis of the dehydration products was performed on a Shimadzu GC-2010 Plus
- 42 system equipped with QP2010 Ultra mass spectrometer detector. AMF and HMF formation (Figure
- 43 S8) is verified by comparing spectrum with GC-MS NIST 2011 database.

44 **2. Analysis results:**



45 46

Figure S1. ¹H NMR analysis of DAF





51 52

Figure S4. ¹H NMR analysis of AMF





53 54





61

62

Scheme S1. Plausible mechanism for the dehydration of DAF to AMF under acidic conditions



Scheme S2. Suggested pathway for the formation of AMF and HMF in the dehydration of DAF

63