

# Lipophilicity determination of antifungal isoxazolo[3,4-*b*]pyridin-3(1*H*)-ones and their N1-substituted derivatives with chromatographic and computational methods

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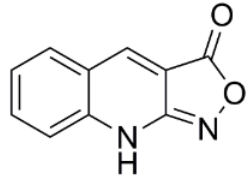
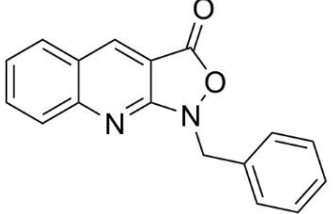
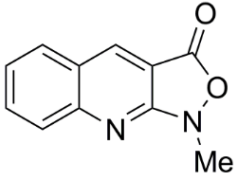
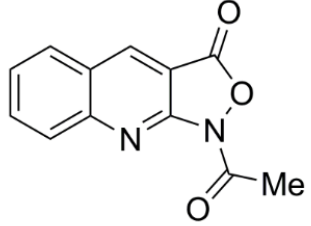
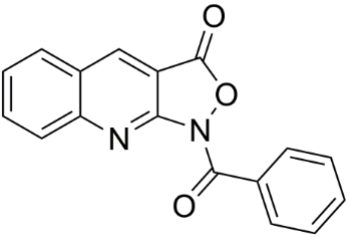
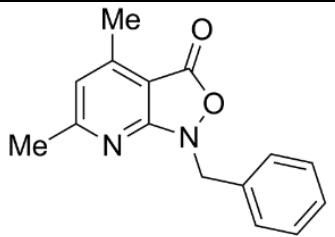
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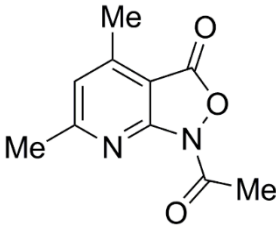
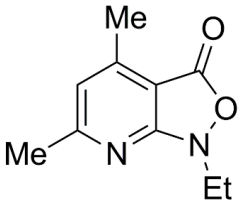
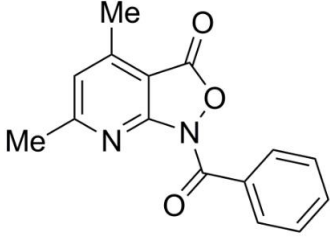
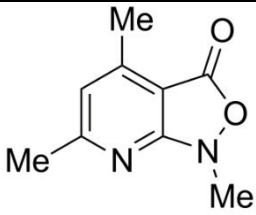
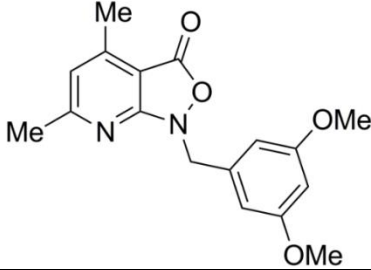
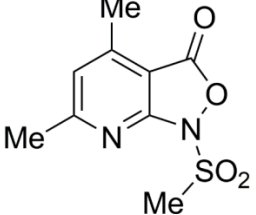
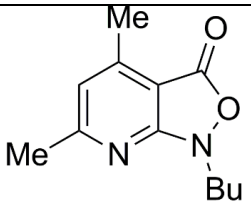
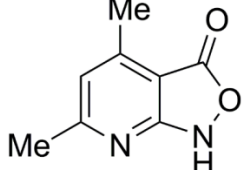
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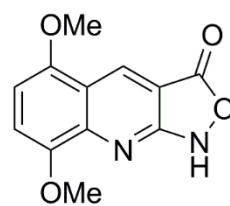
**Table 1S.** Chemical names and structural formulas of the studied of pyrido- and quinolino-isoxazolones

No.	Chemical name	Chemical structure
1	isoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
2	1-benzylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
3	1-methylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
4	1-acetylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
5	1-benzoylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
6	1-benzyl-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	

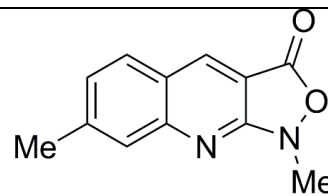
7	1-acetyl-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
8	1-ethyl-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
9	1-benzoyl-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
10	1,4,6-trimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
11	1-(3,5-dimethoxybenzyl)-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
12	4,6-dimethyl-1-(methylsulfonyl)isoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
13	1-butyl-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
14	4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	

15	4,6-dimethyl-1-propylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
16	4,6-dimethyl-1-(prop-2-yn-1-yl)isoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
17	1-(4-fluorobenzoyl)-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
18	1-(3,4,5-trimethoxybenzyl)-4,6-dimethylisoxazolo[3,4- <i>b</i> ]pyridin-3(1 <i>H</i> )-one	
19	6-methoxy-1-methylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
20	6-fluoro-1-methylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
21	6-chloro-1-methylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	
22	1,6,7-trimethylisoxazolo[3,4- <i>b</i> ]quinolin-3(1 <i>H</i> )-one	

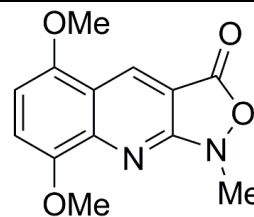
23 5,8-dimethoxyisoxazolo[3,4-*b*]quinolin-3(1*H*)-one



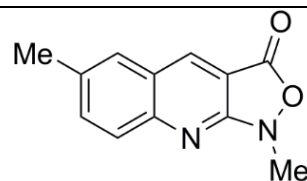
24 1,7-dimethylisoxazolo[3,4-*b*]quinolin-3(1*H*)-one



25 5,8-dimethoxy-1-methylisoxazolo[3,4-*b*]quinolin-3(1*H*)-one



26 1,6-dimethylisoxazolo[3,4-*b*]quinolin-3(1*H*)-one



**Table 2S.** Minimum, maximum and mean value of log*P*-s calculated by different software

	mean log P	minimal log P	maximal log P
log P Chem Draw	1.88	-0.42	3.10
Clog P Chem Draw	2.44	0.11	4.10
vlogp	2.45	0.95	3.43
LogPC Spartan	2.65	0.69	4.47
LogPV Spartan	0.76	-0.71	4.72
miLogP	1.95	0.54	3.16
log KOWWIN	2.29	-0.04	3.94
AlogPs	2.26	0.72	3.67
AClogP	2.46	1.24	3.52
AlogpP	2.56	0.96	3.95
MlogGP	2.51	1.02	3.70
XlogP2	2.11	0.72	3.44
XlogP3	2.58	0.92	3.96

**Table 3S.** Retention data for the investigated pyrido- and quinolino-isoxazolones obtained from the Soczewiński–Wachtmeister’s with statistical parameters

Stationary phase: C <sub>8</sub> bonded silica gel								
Mobile phase: MeOH-Water								
No	$R_M^0$	$\sigma R_M^0$	$m$	$\sigma m$	$R$	$R^2$	$F$	$s$
1	1.496	0.102	-2.198	0.151	0.991	0.981	210.480	0.063
2	3.120	0.146	-3.730	0.217	0.993	0.987	296.732	0.091
3	2.162	0.092	-2.805	0.137	0.995	0.991	418.648	0.057
4	1.893	0.067	-2.593	0.100	0.997	0.994	672.229	0.042
5	2.929	0.096	-3.592	0.144	0.997	0.994	626.162	0.060
6	3.293	0.214	-4.063	0.318	0.988	0.976	163.499	0.133
7	1.791	0.121	-2.477	0.180	0.990	0.979	189.270	0.075
8	2.175	0.089	-2.772	0.132	0.995	0.991	438.375	0.055
9	2.834	0.156	-3.539	0.231	0.992	0.983	233.815	0.097
10	1.919	0.094	-2.558	0.140	0.994	0.988	334.492	0.059
11	3.477	0.106	-4.192	0.158	0.997	0.994	706.077	0.066
12	2.238	0.164	-2.094	0.244	0.973	0.949	73.798	0.101
13	2.988	0.065	-3.555	0.096	0.999	0.997	1357.867	0.040
14	0.991	0.058	-1.831	0.086	0.996	0.991	455.085	0.036
15	2.722	0.103	-3.391	0.153	0.996	0.992	489.391	0.064
16	1.951	0.075	-2.734	0.111	0.997	0.993	602.508	0.047
17	2.864	0.133	-3.579	0.198	0.994	0.988	326.728	0.083
18	2.808	0.097	-3.476	0.144	0.997	0.993	583.604	0.060
19	2.421	0.049	-3.023	0.072	0.999	0.998	1746.354	0.030
20	2.123	0.040	-2.780	0.059	0.999	0.998	2223.849	0.025
21	2.568	0.079	-3.182	0.118	0.997	0.995	732.630	0.049
22	2.907	0.058	-3.498	0.087	0.999	0.998	1632.503	0.036
23	2.123	0.027	-2.664	0.041	1.000	0.999	4242.300	0.017
24	2.431	0.031	-3.033	0.046	1.000	0.999	4333.181	0.019
25	2.557	0.036	-3.026	0.054	0.999	0.999	3161.924	0.023
26	2.461	0.089	-2.990	0.133	0.996	0.992	508.969	0.055

Table 3S. (continued)

Stationary phase: C <sub>18</sub> bonded silica gel								
Mobile phase: MeOH-Water								
No	$R_M^0$	$\sigma R_M^0$	$m$	$\sigma m$	$R$	$R^2$	$F$	$s$
1	2.771	0.163	-3.153	0.200	0.999	0.997	1183.611	0.019
2	3.702	0.213	-4.031	0.262	0.994	0.987	236.093	0.083
3	2.045	0.060	-2.414	0.074	0.999	0.997	1056.838	0.023
4	1.935	0.035	-2.440	0.043	1.000	0.999	3170.983	0.014
5	3.143	0.117	-3.561	0.144	0.998	0.995	609.047	0.046
6	3.012	0.142	-3.363	0.175	0.996	0.992	367.840	0.055
7	1.616	0.053	-2.087	0.066	0.999	0.997	1013.956	0.021
8	2.098	0.041	-2.465	0.050	0.999	0.999	2383.061	0.016
9	2.901	0.056	-3.345	0.069	0.999	0.999	2359.717	0.022
10	1.775	0.071	-2.186	0.088	0.998	0.995	620.512	0.028
11	3.041	0.132	-3.324	0.162	0.996	0.993	419.007	0.051
12	3.017	0.669	-3.633	0.823	0.931	0.867	19.491	0.260
13	3.175	0.084	-3.457	0.104	0.999	0.997	1108.349	0.033
14	0.945	0.263	-1.841	0.323	0.957	0.915	32.394	0.102
15	2.770	0.058	-3.111	0.071	0.999	0.998	1927.377	0.022
16	1.948	0.056	-2.641	0.068	0.999	0.998	1494.420	0.022
17	3.158	0.038	-3.649	0.047	1.000	0.999	5987.251	0.015
18	3.151	0.052	-3.743	0.064	1.000	0.999	3382.991	0.020
19	2.509	0.094	-2.888	0.116	0.998	0.995	618.312	0.037
20	2.314	0.104	-2.778	0.128	0.997	0.994	473.594	0.040
21	2.412	0.171	-2.718	0.210	0.991	0.982	167.234	0.066
22	3.028	0.131	-3.319	0.161	0.996	0.993	425.171	0.051
23	2.066	0.101	-2.569	0.125	0.996	0.993	425.453	0.039
24	2.731	0.131	-3.107	0.162	0.996	0.992	369.059	0.051
25	2.664	0.137	-2.980	0.169	0.995	0.990	310.187	0.054
26	2.771	0.163	-3.153	0.200	0.994	0.988	247.242	0.063

$R$  the coefficient of correlation,  $R^2$  the coefficient of determination,  $F$  the value of the Snedecor's  $F$ -test,  $s$  the standard estimation error,  $p$  value < 0.05.



**Table 4S.** Antifungal activity toward references *Candida* species [1–3]

No.	<i>Candida albicans</i> ATCC 10231	<i>Candida glabrata</i> ATCC 66032	<i>Candida lusitanae</i> ATCC 34499	<i>Candida parapsilosis</i> ATCC 22019	<i>Candida tropicalis</i> ATCC 750
1	>128	>128	>128	>128	>128
2	>128	>128	>128	>128	>128
3	>128	>128	>128	>128	>128
4	>128	>128	>128	>128	>128
5	>128	>128	>128	>128	>128
6	100	>200	100	>200	>200
7	>200	>200	100	>200	>200
8	>200	>200	>200	>200	>200
9	50	50	50	50	50
10	>200	>200	>200	>200	>200
11	>200	>200	>200	100	>200
12	100	>200	100	>200	>200
13	nt	nt	nt	nt	nt
14	>200	>200	50	>200	100
15	100	>200	50	<6.2	25
16	100	>200	>200	<6.2	>200
17	50	100	25	<6.2	100
18	>200	>200	>200	100	>200
19	>128	>128	>128	>128	>128
20	>128	>128	>128	>128	>128
21	>128	>128	>128	>128	>128
22	>128	>128	>128	>128	>128
23	>128	>128	>128	>128	>128
24	>128	>128	>128	>128	>128
25	>128	>128	>128	>128	>128
26	>128	>128	>128	>128	>128

nt – not tested

1. Saczewski, J.; Fedorowicz, J.; Korcz, M.; Saczewski, F.; Wicher, B.; Gdaniec, M.; Konopacka, A. Experimental and theoretical studies on the tautomerism and reactivity of isoxazolo[3,4-b]quinolin-3(1H)-ones. *Tetrahedron* **2015**, *71*, 8975–8984.
2. Saczewski, J.; Fedorowicz, J.; Kedzia, A.; Ziolkowska-Klinkosz, M.; Jalińska, A. Synthesis and Antifungal Activity of Some 4,6-Dimethylisoxazolo[3,4-b]pyridin-3(1H)-one Derivatives. *Med. Chem. (Los. Angeles)*. **2016**, *12*, 640–646.
3. Saczewski, J.; Kedzia, A.; Jalińska, A. New derivatives of 4,6-dimethylisoxazolo[3,4-b] pyridin-3(1H)-one: Synthesis, tautomerism, electronic structure and antibacterial activity. *Heterocycl. Commun.* **2014**, *20*, 215–223.

**Table 5S.** Cross-validation – analysis of variance summary for the TLC C<sub>8</sub> QSRR model.

Source of ariance	<i>SS</i>	<i>df</i>	<i>MS</i>	<i>F</i>	Prob. > <i>F</i>
Total	16.512	17	0.971	49.44	4.66E-6
Fit	16.121	8	2.015		
Residual	0.391	9	0.0408		

*SS* – sum of squares, *df* – degrees of freedom, *MS* – median square, *F* – Fisher test statistic.

**Table 6S.** Cross-validation – analysis of variance summary for the TLC C<sub>18</sub> QSRR model.

Source of ariance	<i>SS</i>	<i>df</i>	<i>MS</i>	<i>F</i>	Prob. > <i>F</i>
Total	16.463	17	0.968	244.26	8.60E-9
Fit	16.354	8	2.044		
Residual	0.109	9	0.008		

*SS* – sum of squares, *df* – degrees of freedom, *MS* – median square, *F* – Fisher test statistic.

**Table 7S.** Cross-validation – analysis of variance summary for the HPLC log<sub>k</sub> QSRR model.

Source of ariance	<i>SS</i>	<i>df</i>	<i>MS</i>	<i>F</i>	Prob. > <i>F</i>
Total	16.429	17	0.966	31.56	2.62E-5
Fit	15.979	8	1.997		
Residual	0.451	9	0.063		

*SS* – sum of squares, *df* – degrees of freedom, *MS* – median square, *F* – Fisher test statistic.

**Table 8S.** Cross-validation – analysis of variance summary for the MEKC log<sub>k</sub> QSRR model.

Source of ariance	<i>SS</i>	<i>df</i>	<i>MS</i>	<i>F</i>	Prob. > <i>F</i>
Total	16.731	17	0.984	19.54	4.47E-5
Fit	16.185	10	1.619		
Residual	0.546	7	0.083		

*SS* – sum of squares, *df* – degrees of freedom, *MS* – median square, *F* – Fisher test statistic.

**Table 9S.** List of molecular descriptors with coefficients values in PLS models built for RP-LC and MEKC.

TLC C <sub>8</sub> QSRR model				
No.	Name	Coefficients	Descriptor full name	Block
1	PJI2	0.09	2D Petitjean shape index	Topological indices
2	X5Av	0.12	average valence connectivity index of order 5	Connectivity indices
3	MATS7i	-0.03	Moran autocorrelation of lag 7 weighted by ionization potential	2D autocorrelations
4	GATS3p	0.08	Geary autocorrelation of lag 3 weighted by polarizability	2D autocorrelations
5	GATS5p	-0.21	Geary autocorrelation of lag 5 weighted by polarizability	2D autocorrelations
6	JGI2	-0.12	mean topological charge index of order 2	2D autocorrelations
7	SpMax2_Bh(s)	-0.11	largest eigenvalue n. 2 of Burden matrix weighted by I-state	Burden eigenvalues
8	SpMax3_Bh(s)	0.11	largest eigenvalue n. 3 of Burden matrix weighted by I-state	Burden eigenvalues
9	Mor19s	-0.01	signal 19 / weighted by I-state	3D-MoRSE descriptors
10	E2u	-0.24	2nd component accessibility directional WHIM index / unweighted	WHIM descriptors
11	E1i	0.24	1st component accessibility directional WHIM index / weighted by ionization potential	WHIM descriptors
12	HATS4u	-0.30	leverage-weighted autocorrelation of lag 4 / unweighted	GETAWAY descriptors
13	HATS4v	-0.26	leverage-weighted autocorrelation of lag 4 / weighted by van der Waals volume	GETAWAY descriptors
14	R3p	0.20	R autocorrelation of lag 3 / weighted by polarizability	GETAWAY descriptors
15	R4p	0.27	R autocorrelation of lag 4 / weighted by polarizability	GETAWAY descriptors
16	R7p+	0.42	R maximal autocorrelation of lag 7 / weighted by polarizability	GETAWAY descriptors
17	CATS3D_08_AL	0.15	CATS3D Acceptor-Lipophilic at lag 08	CATS 3D
TLC C <sub>18</sub> QSRR model				
No	Name	Coefficients		
1	PJI2	0.04	2D Petitjean shape index	Topological indices
2	X5Av	0.27	average valence connectivity index of order 5	Connectivity indices
3	MATS2s	-0.06	Moran autocorrelation of lag 2 weighted by I-state	2D autocorrelations
4	SpMax3_Bh(s)	0.19	largest eigenvalue n. 3 of Burden matrix weighted by I-state	Burden eigenvalues
5	DISPp	-0.11	displacement value / weighted by polarizability	Geometrical descriptors
6	Mor10v	0.15	signal 10 / weighted by van der Waals volume	3D-MoRSE descriptors
7	Mor10p	0.11	signal 10 / weighted by polarizability	3D-MoRSE descriptors
8	E1e	0.08	1st component accessibility directional WHIM index / weighted by Sanderson electronegativity	WHIM descriptors
9	E2p	-0.20	2nd component accessibility directional WHIM index / weighted by polarizability	WHIM descriptors

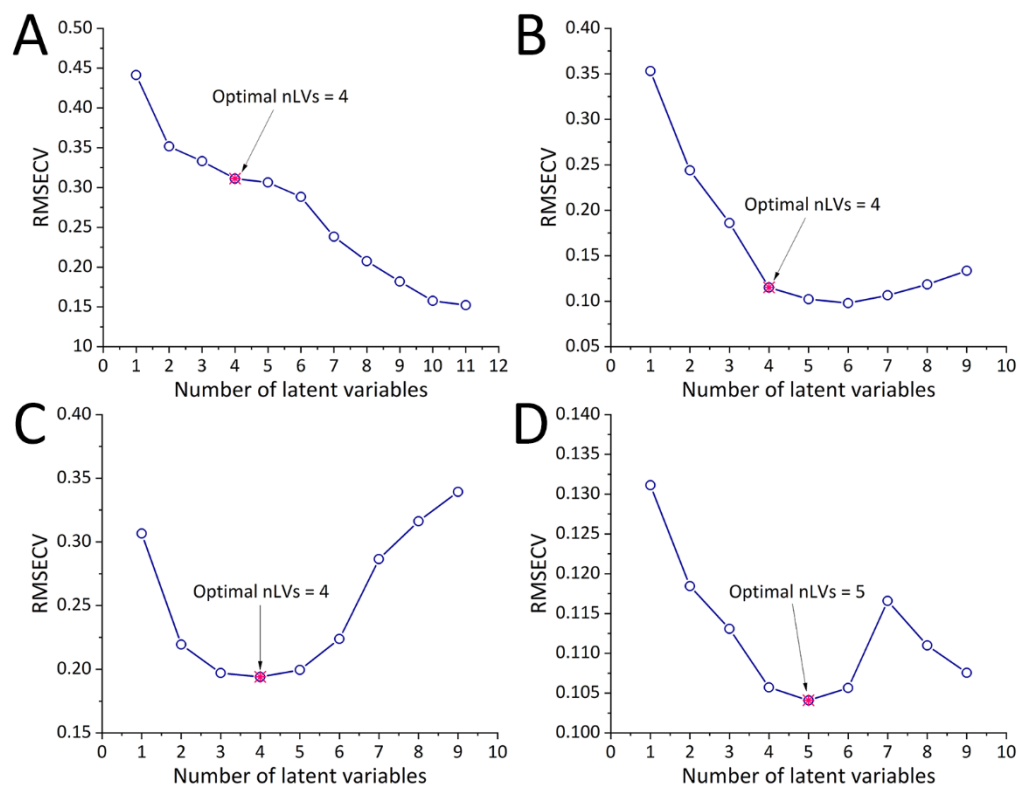
**Table 8S.** (continued)

10	Kv	0.18	K global shape index / weighted by van der Waals volume	WHIM descriptors
11	HATS1e	-0.18	leverage-weighted autocorrelation of lag 1 / weighted by Sanderson electronegativity	GETAWAY descriptors
12	HATS4i	-0.10	leverage-weighted autocorrelation of lag 4 / weighted by ionization potential	GETAWAY descriptors
13	R4v+	-0.08	R maximal autocorrelation of lag 4 / weighted by van der Waals volume	GETAWAY descriptors
14	R3p	0.24	R autocorrelation of lag 3 / weighted by polarizability	GETAWAY descriptors
15	R4p	0.15	R autocorrelation of lag 4 / weighted by polarizability	GETAWAY descriptors
16	CATS3D_03_DL	0.01	CATS3D Donor-Lipophilic at lag 03	CATS 3D
17	CATS3D_08_AL	0.08	CATS3D Acceptor-Lipophilic at lag 08	CATS 3D
HPLC				
No	Label	Coefficients		
1	ATSC3p	0.29	Centred Broto-Moreau autocorrelation of lag 3 weighted by polarizability	2D autocorrelations
2	MATS7i	-0.11	Moran autocorrelation of lag 7 weighted by ionization potential	2D autocorrelations
3	MATS2s	-0.03	Moran autocorrelation of lag 2 weighted by I-state	2D autocorrelations
4	SpMax2_Bh(s)	-0.19	largest eigenvalue n. 2 of Burden matrix weighted by I-state	Burden eigenvalues
5	SpMax3_Bh(s)	0.18	largest eigenvalue n. 3 of Burden matrix weighted by I-state	Burden eigenvalues
6	DISPi	-0.15	displacement value / weighted by ionization potential	Geometrical descriptors
7	E2u	-0.14	2nd component accessibility directional WHIM index / unweighted	WHIM descriptors
8	Ki	0.04	K global shape index / weighted by ionization potential	WHIM descriptors
9	HATS4u	-0.13	leverage-weighted autocorrelation of lag 4 / unweighted	GETAWAY descriptors
10	HATS4v	-0.05	leverage-weighted autocorrelation of lag 4 / weighted by van der Waals volume	GETAWAY descriptors
11	R4v+	-0.13	R maximal autocorrelation of lag 4 / weighted by van der Waals volume	GETAWAY descriptors
12	R3p	0.21	R autocorrelation of lag 3 / weighted by polarizability	GETAWAY descriptors
13	R4p	0.28	R autocorrelation of lag 4 / weighted by polarizability	GETAWAY descriptors
14	R7p+	0.18	R maximal autocorrelation of lag 7 / weighted by polarizability	GETAWAY descriptors
15	R4s	0.05	R autocorrelation of lag 4 / weighted by I-state	GETAWAY descriptors
16	R6s+	0.19	R maximal autocorrelation of lag 6 / weighted by I-state	GETAWAY descriptors
17	CATS3D_08_AL	0.20	CATS2D Acceptor-Lipophilic at lag 08	CATS 3D

**Table 9S.** (continued)

MECK				
No	Label	Coefficients		
1	VE1_Dz(p)	0.14	coefficient sum of the last eigenvector from Barysz matrix weighted by polarizability	2D matrix-based descriptors
2	Mor24s	-0.24	signal 24 / weighted by I-state	3D-MoRSE descriptors
3	E2u	0.52	2nd component accessibility directional WHIM index / unweighted	WHIM descriptors
4	E1e	0.24	1st component accessibility directional WHIM index / weighted by Sanderson electronegativity	WHIM descriptors
5	HATS1e	-0.56	leverage-weighted autocorrelation of lag 1 / weighted by Sanderson electronegativity	GETAWAY descriptors
6	HATS7p	0.06	leverage-weighted autocorrelation of lag 7 / weighted by polarizability	GETAWAY descriptors
7	R4v+	0.33	R maximal autocorrelation of lag 4 / weighted by van der Waals volume	GETAWAY descriptors
8	R4p	0.16	R autocorrelation of lag 4 / weighted by polarizability	GETAWAY descriptors
9	R4p+	0.17	R maximal autocorrelation of lag 4 / weighted by polarizability	GETAWAY descriptors
10	R6s+	0.31	R maximal autocorrelation of lag 6 / weighted by I-state	GETAWAY descriptors
11	CATS3D_08_AA	-0.49	CATS3D Acceptor-Acceptor at lag 08	CATS 3D
12	CATS3D_03_LL	-0.33	CATS3D Lipophilic-Lipophilic at lag 03	CATS 3D

**Figure 1S.** Optimization of the number of latent variables (LVs) for the consensus GA-PLS models for **A)** TLC with the  $C_8$  plate, **B)** TLC with the  $C_{18}$  plate, **C)** HPLC  $\log k_w$ , and **D)** MEKC  $\log k$  parameters. The optimal number of LVs is denoted in pink.



**Figure 2S.** Leave one out-cross validation (LOO-CV) predictive ability (on the training set) for **A)** TLC with the C<sub>8</sub> plate, **B)** TLC with the C<sub>18</sub> plate, **C)** HPLC log<sub>k<sub>w</sub></sub>, and **D)** MEKC log<sub>k</sub> parameters.

