

Review

Quinoa Secondary Metabolites and Their Biological Activities or Functions

Minyi Lin, Peipei Han, Yuying Li, Weixuan Wang, Daowan Lai  and Ligang Zhou * 

Department of Plant Pathology, College of Plant Protection, China Agricultural University, Beijing 100193, China

* Correspondence: lgzhou@cau.edu.cn; Tel.: +86-10-6273-1199

Received: 31 May 2019; Accepted: 3 July 2019; Published: 9 July 2019



Abstract: Quinoa (*Chenopodium quinoa* Willd.) was known as the “golden grain” by the native Andean people in South America, and has been a source of valuable food over thousands of years. It can produce a variety of secondary metabolites with broad spectra of bioactivities. At least 193 secondary metabolites from quinoa have been identified in the past 40 years. They mainly include phenolic acids, flavonoids, terpenoids, steroids, and nitrogen-containing compounds. These metabolites exhibit many physiological functions, such as insecticidal, molluscicidal and antimicrobial activities, as well as various kinds of biological activities such as antioxidant, cytotoxic, anti-diabetic and anti-inflammatory properties. This review focuses on our knowledge of the structures, biological activities and functions of quinoa secondary metabolites. Biosynthesis, development and utilization of the secondary metabolites especially from quinoa bran were prospected.

Keywords: quinoa (*Chenopodium quinoa*); secondary metabolites; biological activities; functions

1. Introduction

Quinoa (*Chenopodium quinoa* Willd.), a dicotyledonous plant belonging to Chenopodiaceae family, is one of the oldest native crops in the Andean region of South America, with approximately 7000 years of cultivation [1]. It has been considered as a pseudo-cereal because of the grain characteristics [2]. Consumption of seeds is the most common use of quinoa. Once the bran (also called hull or seed coats) containing saponins has been eliminated, the seeds can be consumed as entire grains or milled to flour for preparation of bread and pastry. The other parts such as leaves and stems were used as feed [2,3].

Quinoa has been recognized as a complete food due to a variety of vitamins, significant amounts of minerals, unsaturated fatty acids, dietary fiber, abounding proteins, and excellent balance of essential amino acids. The year 2013 was named “The International Year of Quinoa” by the UN. Quinoa has been introduced and cultivated all over the world in the past ten years [3–8].

Quinoa possesses a large number of secondary metabolites, such as phenolic acids, flavonoids, terpenoids, steroids, and nitrogen-containing compounds. These metabolites play various physiological and ecological roles against harmful microorganisms, birds and insects. They also exhibit features beneficial to humans, including anti-diabetic [9], anticancer [10], cytotoxic [11], antimicrobial [12], anti-inflammatory [13], immunoregulatory [14] and adjuvant activities [15].

To our knowledge, there are many reviews on quinoa, most of them are focused on the nutritional, functional and antinutritional aspects [16–18], abiotic stress responses [19], biodiversity and sustainability [20], or only a specific topic of quinoa secondary metabolites and their biological activities such as steroids [21,22] and triterpenoid saponins [23], but no review covers almost all secondary metabolites and their biological activities. In this review, we summarize and discuss quinoa secondary metabolites on their structural diversity, biological activities or functions during the past 40 years.

2. Phenolic Acids and Their Biological Activities or Functions

About 29 phenolic acid analogues have been identified in quinoa. According to their structural features, they can be classified as benzoic acid analogues (1–16) and cinnamic acid analogues (17–29). Benzoic acid (1) was derived from cinnamic acid (19) *in planta* in the biosynthetic pathway of phenolic acids [24]. Phenolic acid derivatives are present in either free or conjugated forms. The total of conjugated phenolic acids in quinoa were at comparable level as that of free ones, suggesting that conventional solvent extraction and chromatographic analysis of extractable phenolic acids might have significantly underestimated the total phenolic acid content in quinoa, as such methods only detect free phenolic acids [25].

Phenolic acids can be released by acid, alkaline, and enzymatic treatments from the conjugated forms. It was reported that at least 19 phenolic acids were released in the residue of quinoa which can enhance bioaccessibility [25]. Bound phenolic acid derivatives in conjugated forms were not affected by environmental stresses [26]. Higher content of phenolic acids showed stronger antioxidant and inhibitory activities of α -glucosidase and pancreatic lipase [25].

2.1. Benzoic Acid Analogues and Their Biological Activities or Functions

At least 16 benzoic acid analogues have been identified from quinoa. Their biological activities are listed in Table 1, and the structures are shown in Figure 1. Benzoic acid derivatives include benzoic acid (1), gallic acid (8), protocatechuic acid (10), syringic acid (12), vanillic acid (13), and their analogues. They are rich in the leaves and seeds of quinoa [25,27]. Though the benzoic acid analogues from quinoa have not been evaluated for their biological activities, these metabolites from other plant species have been reported to have antimicrobial [28,29], allelopathic [30], antioxidant [31], and antifeedant [32] activities (Table 1).

Table 1. Benzoic acid analogues and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|--------------------------------|--|---------|
| Benzoic acid (1) | Leaves and flour | - | [27,33] |
| 4-Hydroxybenzoic acid = <i>p</i> -Hydroxybenzoic acid (2) | Seeds | - | [25] |
| | Leaves and seeds | - | [27,34] |
| | | Antimicrobial activity | [28] |
| | | Allelopathic effect | [30] |
| 2,4-Dihydroxybenzoic acid (3) | Seeds | - | [25,35] |
| 2,5-Dihydroxybenzoic acid (4) | Seeds | - | [35] |
| 3,4-Dihydroxybenzoic acid (5) | Seeds | - | [35] |
| Canthoside A (6) | Flour | - | [33] |
| Ethyl- <i>m</i> -digallate (7) | Flour | - | [33] |
| | | Antifeedant activity | [32] |
| Gallic acid (8) | Leaves, sprouts and seeds | - | [27,34] |
| | | Antioxidant activity | [31] |
| | | Antibacterial activity | [36] |
| 1- <i>O</i> -Galloyl- β - <i>D</i> -glucoside (9) | Seeds and flour | - | [33] |
| Protocatechuic acid (10) | Sprouts and seeds | - | [25,37] |
| | | Antioxidant activity | [31] |
| | | Anticancer activity | [38] |
| | | Antibacterial activity | [39] |
| | | Antiulcer activity | [40] |
| | | Antiageing activity | [41] |
| | | Anti-inflammatory, antiibrotic, antiatherosclerotic, hyperlipidemic, analgesic, hepatoprotective and nephroprotective activities | [42] |
| | | Antiviral activity | [43] |

Table 1. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|---|------------|
| Protocatechuic acid 4- <i>O</i> -glucoside (11) | Flour | - | [33] |
| Syringic acid (12) | Leaves and seeds | Antioxidant activity | [44] |
| | | - | [25,26] |
| | | Allelopathic effect | [30] |
| | | Antioxidant activity | [31] |
| | | Antimicrobial activity | [45] |
| | | Hepatoprotective effect | [46] |
| Vanillic acid (13) | Leaves and seeds | Anti-inflammatory activity | [47] |
| | | - | [25,34] |
| | | Allelopathic effect | [30] |
| | | Hepatoprotective effect | [46] |
| | | Antioxidant and antimicrobial activities, and inhibitory activity on COX-I and COX-II | [48] |
| | | - | [49] |
| Vanillic acid glucosyl ester (14) | Seeds | - | [49] |
| Vanillic acid 4- <i>O</i> -glucoside (15) | Seeds | - | [35] |
| Vanillin (16) | Seeds and flour | - | [25,33,35] |
| | | Antioxidant activity | [50] |
| | | Antimicrobial activity | [51] |
| | | Antidepressant activity | [52] |
| | | Anti-angiogenic, anti-inflammatory and anti-nociceptive activities | [53] |
| | | - | [53] |

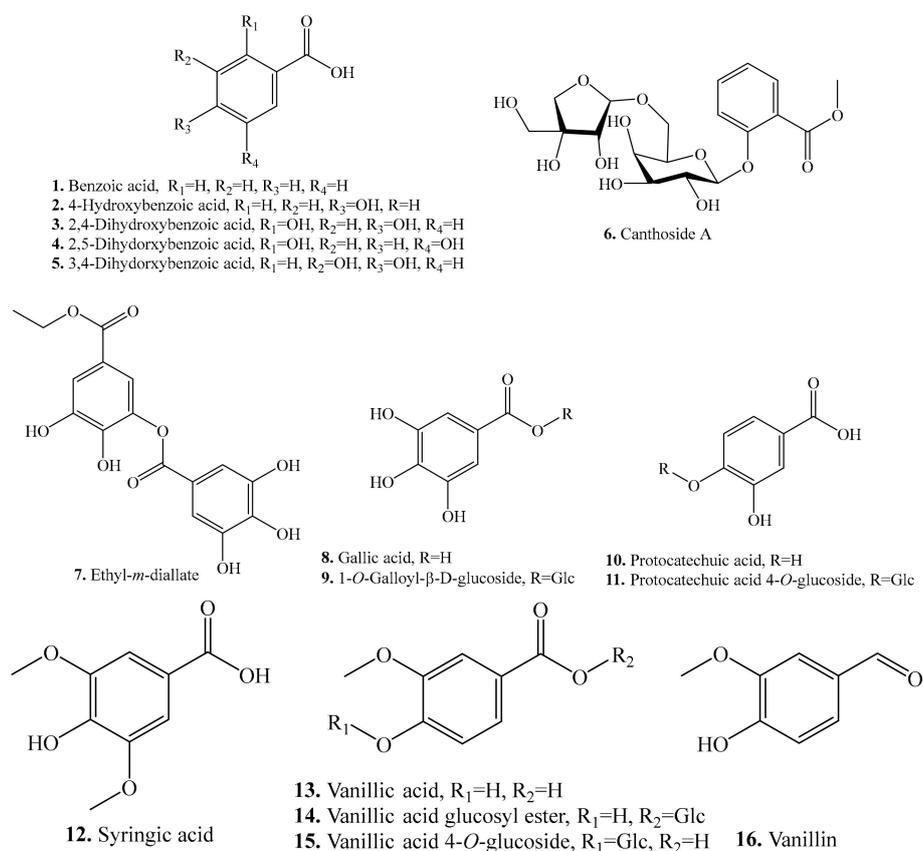


Figure 1. Structures of the benzoic acid analogues isolated from quinoa.

2.2. Cinnamic Acid Analogues and Their Biological Activities or Functions

Thirteen cinnamic acid analogues have been identified from quinoa. Their biological activities are listed in Table 2, and the structures are shown in Figure 2. These cinnamic acid derivatives include caffeic acid (17), chlorogenic acid (18), cinnamic acid (19), coumaric acid (20/21), ferulic acid (24), rosmarinic acid (28), sinapinic acid (29), and their analogues. Ferulic acid (24) and its derivatives were the predominant phenolics in bound form to be present in quinoa seeds [17].

Both ferulic acid (24) and sinapinic acid (29) had more phytotoxic effects on cucumber seedling as compared to the other tested phenolic acids [54]. The phenolic acids from quinoa were also isolated from other plant species which showed a variety of biological activities such as antimicrobial [28], allelopathic [30], antioxidant [31], anti-apoptotic [55], anti-diabetic [56] activities that are mentioned in Table 2.

Table 2. Cinnamic acid analogues and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|--------------------------------|--|------------|
| Caffeic acid (17) | Seeds | - | [25,34] |
| | | Antimicrobial activity | [29] |
| | | Allelopathic effect | [30] |
| | | Antioxidant activity | [31] |
| | | Anti-apoptotic activity | [55] |
| | | Inhibitory activity on xanthine oxidase | [57] |
| Chlorogenic acid (18) | Leaves and seeds | - | [25,26] |
| | | Antimicrobial activity | [29] |
| | | Antioxidant activity | [31] |
| | | Anti-diabetic activity | [56] |
| | | Hemolytic activity | [58] |
| | | Neuroprotective effects | [59] |
| | | Anti-obesity activity | [60] |
| | | Antihepatotoxic effect | [61] |
| Antibiofilm activity | [62] | | |
| Cinnamic acid (19) | Sprouts and seeds | - | [34] |
| <i>o</i> -Coumaric acid (20) | Leaves and seeds | - | [25,26] |
| | | Allelopathic effect | [30] |
| <i>p</i> -Coumaric acid (21) | Leaves and seeds | Antioxidant activity | [31] |
| | | - | [27,63] |
| <i>p</i> -Coumaric acid glucoside (22) | Seeds | Antilisterial activity | [64] |
| 8,5'-Diferulic acid (23) | Seeds | - | [35] |
| Ferulic acid (24) | Leaves, sprouts and seeds | - | [27,34,35] |
| | | Antimicrobial activity | [29] |
| | | Anti-apoptotic activity | [55] |
| | | Antioxidant activity | [65] |
| | | Cholesterol-lowering activity | [66] |
| | | Anti-thrombosis and anti-atherosclerosis effects | [67,68] |
| | | Anti-inflammatory activity | [69] |
| | | Anti-cancer activity | [70] |
| | | - | [33] |
| | | - | [35] |
| Ferulic acid 4- <i>O</i> -glucoside (25) | Flour | - | [33] |
| Isoferulic acid (26) | Seeds | - | [35] |
| | | Antioxidant activity | [71] |

Table 2. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|--------------------------------|---|------|
| 4'-Geranyloxyferulic acid (27) | Seeds | - | [72] |
| Rosmarinic acid (28) | Seeds | - | [25] |
| | | Antimicrobial activity | [73] |
| | | Anti-inflammatory activity | [74] |
| | | Antioxidant activity | [75] |
| | | Antimutagenicity activity | [76] |
| | | Antiviral and anti-inflammatory effects | [77] |
| Sinapinic acid = <i>trans</i> -Sinapic acid (29) | Leaves | - | [27] |
| | Seeds | - | [25] |
| | | Antioxidant activity | [44] |
| | | Anxiolytic-like effects | [78] |
| | | Cerebral protective and cognition-improving effects | [79] |

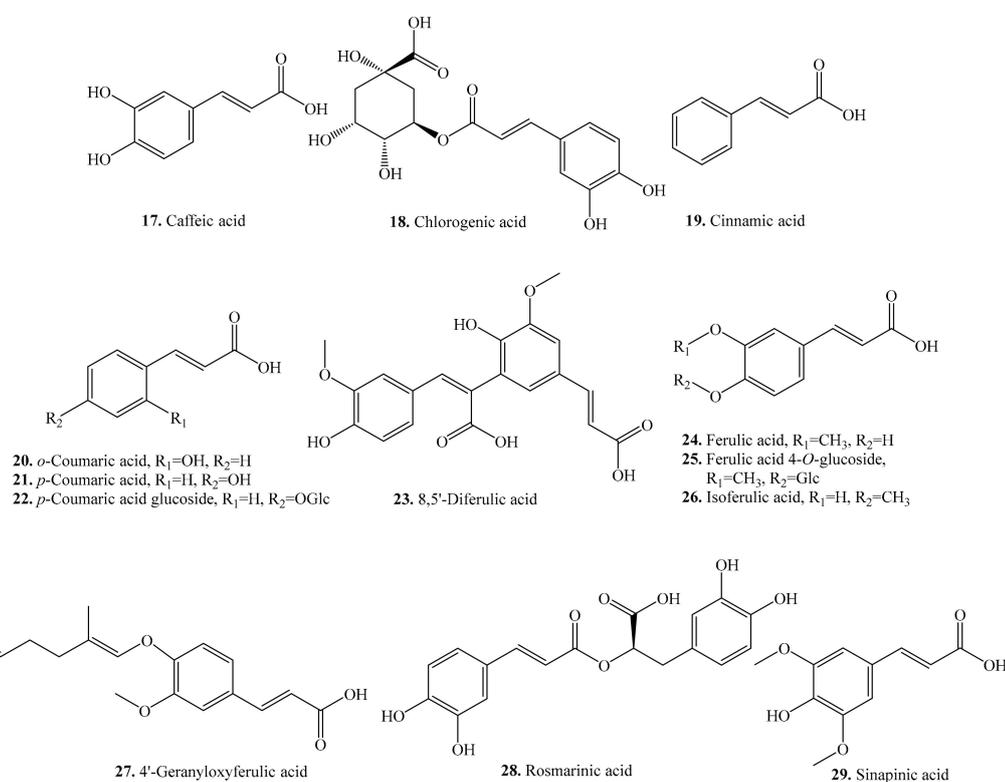


Figure 2. Structures of the cinnamic acid analogues isolated from quinoa.

3. Flavonoids and Their Biological Activities or Functions

Flavonoids are based upon a fifteen-carbon skeleton consisting of two benzene rings linked via a heterocyclic pyrene ring [80]. They contain aglycones and their glycosides. The main flavonoid aglycones are kaempferol (35) and quercetin (46). Other aglycones in quinoa include acacetin (30), myricetin (45), daidzein (62), and genistein (63). According to the structural features, quinoa flavonoids can be classified as flavones (30–33), flavonols (34–54), flavanones (or dihydroflavones, 55–57), flavanols (58–60), and isoflavones (61–65). Flavonoids play important roles in plants against the feeding insects and herbivores [81]. Flavonoids also have deterrent effects with respect to feeding and physiological behavior against some soil herbivorous nematodes [82].

3.1. Flavones and Their Biological Activities or Functions

Four flavones, namely acacetin (30), isovitexin (31), orientin (32) and vitexin (33), have been identified from quinoa. Their biological activities are listed in Table 3, and the structures are shown in Figure 3. Flavones were significantly richer in sprouts than in other parts of quinoa. Quinoa sprouts grown in the darkness contained vitexin (33) and substantial amounts of isovitexin (31), whereas those grown in daylight only contained isovitexin (31). It is remarkable that no isovitexin (31) was present in quinoa seeds [34]. Acacetin (30), isovitexin (31), orientin (32) and vitexin (33) were also isolated from other plant species which showed various biological activities such as antioxidant [83], anti-inflammatory [84] activities, which are listed in Table 3.

Table 3. Flavones and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. | | |
|--|--------------------------------|--|---------|--|------|
| Acacetin (30) | Flour | - | [33] | | |
| | | Antioxidant activity | [83] | | |
| | | Spasmolytic and antinociceptive activities | [85] | | |
| | | Antiproliferative activity | [86] | | |
| | | Antiherpetic activity | [87] | | |
| | | Anticancer activity | [88] | | |
| | | Anti-inflammatory and antinociceptive activities | [89] | | |
| | | Hypouricemic effect | [90] | | |
| | | Isovitexin (31) | Sprouts | - | [34] |
| | | | | Anti-inflammatory and antioxidant activities | [84] |
| Anti-neoplastic effect | [91] | | | | |
| Anti-tumour activity | [92] | | | | |
| Neuroprotective effect | [93] | | | | |
| Anxiolytic property | [94] | | | | |
| Anti-Alzheimer's disease | [95] | | | | |
| Reduced postprandial blood glucose | [96] | | | | |
| Inhibitory effect on α -glucosidase | [97] | | | | |
| Inhibitory activity on rat lens aldose reductase | [98] | | | | |
| Orientin (32) | Seeds | - | [34] | | |
| | | Anticancer activity | [7] | | |
| | | Anti-inflammatory activity | [99] | | |
| | | Antioxidant activity | [100] | | |
| | | Antiapoptosis activity | [101] | | |
| | | Antithrombotic and antiplatelet activities | [102] | | |
| | | Antiproliferative activity | [103] | | |
| Vitexin (33) | Sprouts and seeds | - | [34] | | |
| | | Anti-carcinogenic effect | [91] | | |
| | | Anxiolytic property | [94] | | |
| | | Anti-Alzheimer's disease property | [95] | | |
| | | Reduced postprandial blood glucose | [96] | | |
| | | Inhibitory effect on α -glucosidase | [97] | | |
| | | Induced apoptosis property | [104] | | |
| | | Agonist-induced regulation of vascular contractility | [105] | | |
| | | Antioxidant activity | [106] | | |
| | | Anti-inflammatory activity | [107] | | |
| | | Neuroprotective effect | [108] | | |
| | | Anti-depressant effect | [109] | | |
| | | Anti-convulsant effect | [110] | | |
| | | Antiepileptic effect | [111] | | |
| | | Anti-nociceptive effect | [112] | | |
| | | Anti-hypoxia/ischemia injury | [113] | | |
| | | Anti-ischemia/reperfusion injury | [114] | | |
| Anti-thyroid effect | [115] | | | | |
| Antimicrobial activity | [116] | | | | |
| Anti-viral effect | [117] | | | | |

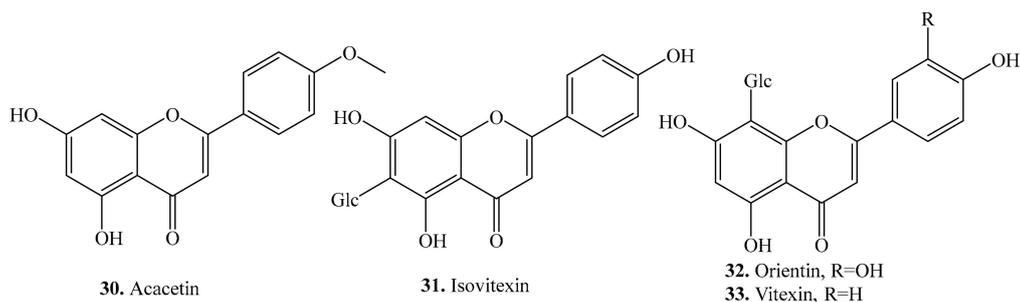


Figure 3. Structures of the flavones isolated from quinoa.

3.2. Flavonols and Their Biological Activities or Functions

About 21 flavonols have been identified in quinoa. Most of them are present in the seeds. Their biological activities are listed in Table 4, and their structures are shown in Figure 4.

Both kaempferol (35) and quercetin (46) are two main flavonols. They are in the form of glycosides present in quinoa. Structure-activity relationship of their antioxidant activity showed that the ability to quench free hydroxyl radicals increased with the amount of hydroxyl groups in the ring B. For example, myricetin (45) was a stronger antioxidant than kaempferol (35) [118]. In addition, the compounds with 3',4'-dihydroxy substituents in the ring B had much stronger antioxidative activities than those without *ortho*-dihydroxy substitution in the ring B [119]. Quercetin (46) was the strongest antioxidant among the flavonoids. Both isorhamnetin (34) and kaempferol (35) were the most abundant flavonoids in quinoa leaves, and it also contained large amounts of rutin (54) [27]. Four kaempferol 3-glycosides (38–41) exhibited moderate antioxidant activity while two quercetin 3-glycosides (50,51) showed strong antioxidant activity, suggesting that quinoa could represent an important source of free radical inhibitors [120].

Many flavonoids are characterized by antibacterial, antifungal and antiviral activities, not only against plant pathogens, but also against the pathogens for humans and animals (Table 4). Kaempferol (35) and its derivatives showed antibacterial activity against Gram-positive and Gram-negative bacteria, as well as against the fungus *Candida glabrata* [121,122].

About eight quercetin derivatives (46–53) have been identified in quinoa. Kaempferol (35), myricetin (45) and quercetin (46) acted as the deterrents against *Radopholus similis* and *Meloidogyne incognita* [82]. Quercetin-3-glucoside (47) and rutin (54) from *Pinus banksiana* inhibited the development of *Lymantria dispar* and increased its mortality [123]. Quercetin (46), quercetin 3-*O*-glucoside (47) and its six derivatives exhibited inhibitory activity on the shoot growth of *Arabidopsis thaliana* as well as on the spore germination of the fungus *Neurospora crassa* [124].

Table 4. Flavonols and their biological activities or functions.

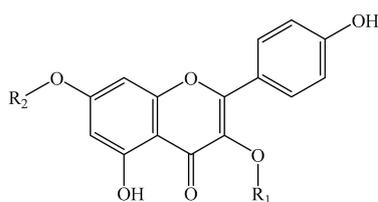
| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|------------------------|--------------------------------|---|-----------|
| Isorhamnetin (34) | Leaves | - | [27,125] |
| | | Chemopreventive activity | [126] |
| | | Antituberculosis activity | [127] |
| | | Antioxidant activity | [128] |
| | | Anti-tumor activity | [129,130] |
| | | Inhibitory activity on farnesyl protein transferase | [131] |
| | | Anti-inflammatory activity | [132] |
| Anticoagulant activity | [133] | | |

Table 4. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|--------------------------------|--|------------------|
| Kaempferol (35) | Leaves and seeds | - | [25,26,35] |
| | | Antibacterial activity | [36] |
| | | Antioxidant activity | [134] |
| | | Inhibit UVB-induced COX-2 expression | [135] |
| | | Anti-inflammatory activity | [136] |
| | | Stimulate osteoblastic activity | [137] |
| Kaempferol 3-glucoside (36) | Seeds | - | [35] |
| Kaempferol 3-galactoside (37) | Seeds | - | [35] |
| Kaempferol | Seeds | Antioxidant activity | [49,120,138,139] |
| 3-O-(2,6-di- α -L-rhamnopyranosyl)- β -D-galactopyranoside (38) | | | |
| Kaempferol | | | |
| 3-O- β -D-apiofuranosyl-(1 \rightarrow 2)-O- α -L-rhamnopyranosyl(1 \rightarrow 6)- β -D-galactopyranoside (39) | Seeds | Antioxidant activity | [49,120,138] |
| Kaempferol | Seeds | Antioxidant activity | [120,138] |
| 3-O- β -D-apiofuranosyl-(1 \rightarrow 2)- β -D-galactopyranoside (40) | | | |
| Kaempferol | Seeds | Antioxidant activity | [120] |
| 3-O- α -L-rhamnopyranosyl-(1 \rightarrow 2)- β -D-galactopyranoside (41) | | | |
| Kaempferol 3-O- β -D-glucuronic acid (42) | Seeds | Antioxidant activity | [49] |
| Kaempferol 3,7-dirhamnoside (43) | Seeds | - | [35] |
| Morin (44) | Sprouts and seeds | - | [34] |
| | | Anti-biofilm activity | [140] |
| | | Anti-inflammatory activity | [141] |
| | | Antitumor activity | [142] |
| | | Inhibitory effect on the expression of α 1 (I) collagen | [143] |
| | | Antioxidant activity | [144] |
| | | Anticancer activity | [145] |
| | | Inhibited the increase of ROS and reduced the apoptotic cell | [146] |
| | | Neuroprotective effect | [147] |
| | | Hepatoprotective activity | [148] |
| | | - | [63] |
| | | - | [36] |
| | | - | [149] |
| - | [150] | | |
| - | [151] | | |
| - | [152] | | |
| Quercetin (46) | Leaves and seeds | - | [27,35,125,139] |
| | | COX-I and COX-II inhibition activity | [48] |
| | | Stimulate osteoblastic activity | [137] |
| | | Antioxidant and prooxidant activities | [149] |
| | | Anti-inflammatory activity | [153] |
| | | Cytotoxic activity | [154] |
| Quercetin 3-O-glucoside (47) | Flour | - | [33] |
| Quercetin-3-rutinoside (48) | Seeds | - | [35] |
| Quercetin 3-arabinoside (49) | Seeds | - | [35] |

Table 4. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|---|-----------------------------------|
| Quercetin 3- <i>O</i> - β -D-apiofuranosyl-(1 \rightarrow 2)- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside (50) | Seeds | Antioxidant activity | [120,139] |
| Quercetin 3- <i>O</i> -(2,6-di- α -L-rhamnopyranosyl)- β -D-galactopyranoside (51) | Seeds | Antioxidant activity | [49,120] |
| Quercetin 3- <i>O</i> -(2,6-di- <i>O</i> - α -rhamnopyranosyl)- β -glucopyranoside (52) | Seeds | - | [139] |
| Quercetin 3- <i>O</i> - β -D-apiofuranosyl-(1 \rightarrow 2)- <i>O</i> - α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside-3,4-dimethyl ether (53) | Seeds | Antioxidant activity | [49] |
| Rutin (54) | Leaves, sprouts and seeds | - Anti-diabetic activity Antioxidant activity Antitumorogenic activity | [27,34] [56] [155] [156] |



35. Kaempferol, $R_1=H$, $R_2=H$

36. Kaempferol 3-*O*-glucoside, $R_1=Glc$, $R_2=H$

37. Kaempferol 3-*O*-galactoside, $R_1=Gal$, $R_2=H$

38. Kaempferol 3-*O*-(2,6-di- α -L-rhamnopyranosyl)- β -D-galactopyranoside,
 $R_1=Gal$ -(2,6-di- α -L-Rha), $R_2=H$

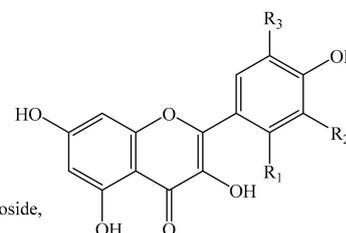
39. Kaempferol 3-*O*- β -D-apiofuranosyl(1 \rightarrow 2)-*O*- α -L-rhamnopyranosyl (1 \rightarrow 6)- β -D-galactopyranoside,
 $R_1=Gal$ (6 \rightarrow 1)Rha(2 \rightarrow 1)Apiose, $R_2=H$

40. Kaempferol 3-*O*- β -D-apiofuranosyl (1 \rightarrow 2)- β -D-galactopyranoside, $R_1=Gal$ (2 \rightarrow 1)Apiose, $R_2=H$

41. Kaempferol 3-*O*- α -L-rhamnopyranosyl (1 \rightarrow 2)- β -D-galactopyranoside, $R_1=Gal$ (2 \rightarrow 1)Rha, $R_2=H$

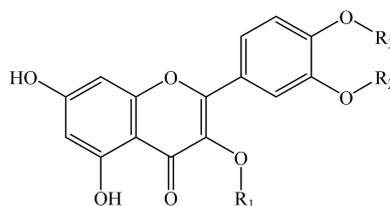
42. Kaempferol 3-*O*- β -D-glucuronic acid, $R_1=Glc$, $R_2=H$

43. Kaempferol 3,7-di- α -L-rhamnoside, $R_1=Rha$, $R_2=Rha$



44. Morin, $R_1=OH$, $R_2=H$, $R_3=H$

45. Myricetin, $R_1=H$, $R_2=OH$, $R_3=OH$



34. Isorhamnetin, $R_1=H$, $R_2=CH_3$, $R_3=H$

46. Quercetin, $R_1=H$, $R_2=H$, $R_3=H$

47. Quercetin 3-*O*-glucoside, $R_1=Glc$, $R_2=H$, $R_3=H$

48. Quercetin 3-*O*-rutinoside, $R_1=Rutinose$, $R_2=H$, $R_3=H$

49. Quercetin 3-*O*-arabinoside, $R_1=Ara$, $R_2=H$, $R_3=H$

50. Quercetin 3-*O*- β -D-apiofuranosyl-(1 \rightarrow 2)- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside,
 $R_1=Gal$ (6 \rightarrow 1)Rha(2 \rightarrow 1)Apiose, $R_2=H$, $R_3=H$

51. Quercetin 3-*O*-(2,6-di- α -L-rhamnopyranosyl)- β -D-galactopyranoside,
 $R_1=Gal$ -(2,6-di- α -L-Rha), $R_2=H$, $R_3=H$

52. Quercetin 3-*O*-(2,6-di-*O*- α -rhamnopyranosyl)- β -D-glucopyranoside,
 $R_1=Glc$ -(2,6-di-*O*- α -Rha), $R_2=H$, $R_3=H$

53. Quercetin 3-*O*- β -D-apiofuranosyl-(1 \rightarrow 2)-*O*- α -L-rhamnopyranosyl-(1 \rightarrow 6)- β -D-galactopyranoside-3,4-dimethyl ether, $R_1=Gal$ (6 \rightarrow 1)Rha(6 \rightarrow 1)Apiose, $R_2=CH_3$, $R_3=CH_3$

54. Rutin, $R_1=Rha$ (6 \rightarrow 1)Glc, $R_2=H$, $R_3=H$

Figure 4. Structures of the flavonols isolated from quinoa.

3.3. Flavanones and Their Biological Activities or Functions

Three flavanones hesperidin (55), neohesperidin (56), and naringin (57) were identified in quinoa seeds (Table 5 and Figure 5). Both hesperidin (55) and neohesperidin (56) were found in the sprouts [34]. These flavanones isolated from other plant species were screened to show a variety of biological activities such as neuroprotective [147], antioxidant [157], anti-inflammatory [158] and antifungal [159] activities.

Table 5. Flavanones and their biological activities.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|----------------------------|--------------------------------|--|-------|
| Hesperidin (55) | Seeds | - | [34] |
| | | Neuroprotective effect | [147] |
| | | Antioxidant and cytotoxic activities | [157] |
| | | Anti-inflammatory activity | [158] |
| | | Antifungal activity | [159] |
| | | Anti-proliferative and apoptotic activities | [160] |
| | | Protects the liver against drug-induced injury | [161] |
| Neohesperidin (56) | Seeds | Cardioprotective activity | [162] |
| | | - | [34] |
| | | Neuroprotective effect | [147] |
| | | Antifungal activity | [159] |
| | | Antioxidant activity | [163] |
| Naringin (57) | Seeds | Induces cell apoptosis | [164] |
| | | - | [35] |
| | | Antifungal activity | [159] |
| | | Antioxidative activity | [165] |
| | | Anti-osteoporosis activity | [166] |
| Anti-inflammatory activity | [167] | | |

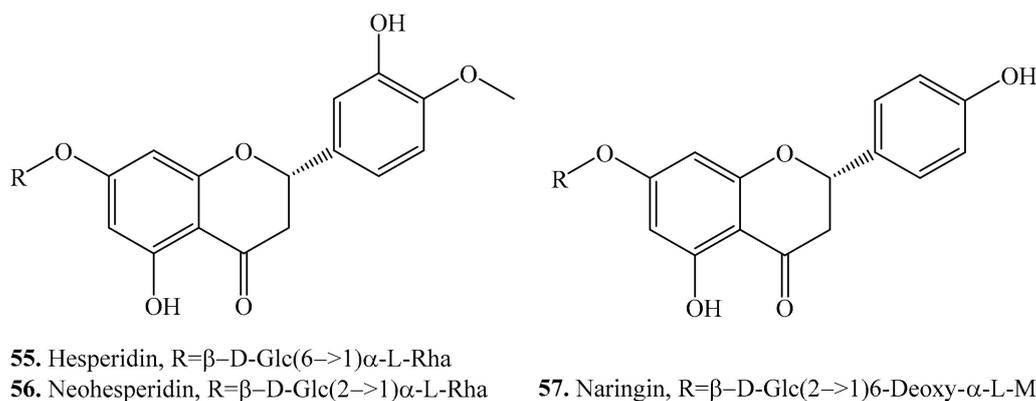


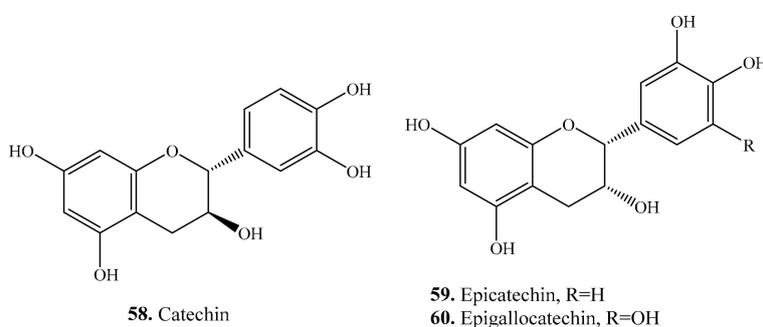
Figure 5. Structures of the flavanones isolated from quinoa.

3.4. Flavanols and Their Biological Activities or Functions

Three flavanols namely catechin (58), epicatechin (59), and epigallocatechin (60) were found in quinoa seeds. Their biological activities are listed in Table 6, and their structures are shown in Figure 6. They generally showed antioxidant [149,168] and antimutagenic [169] activities.

Table 6. Flavanols and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|-----------------------|--------------------------------|---------------------------------|---------|
| Catechin (58) | Seeds | - | [25] |
| | | Antioxidant activity | [149] |
| | | Antimutagenic activity | [169] |
| | | Anti-metastatic activity | [170] |
| | | Antifungal activity | [171] |
| | | Apoptosis-inducing activity | [172] |
| Epicatechin (59) | Seeds | - | [35] |
| | | Antimutagenic activity | [169] |
| | | Antioxidant activity | [173] |
| | | Antiproliferative activity | [174] |
| Epigallocatechin (60) | Seeds | - | [33,35] |
| | | Antioxidant activity | [168] |

**Figure 6.** Structures of the flavanols isolated from quinoa.

3.5. Isoflavones and Their Biological Activities or Functions

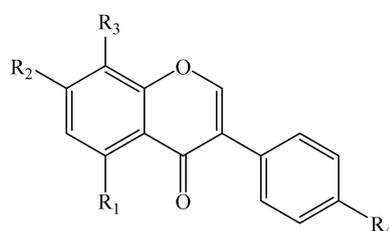
Five isoflavanones, i.e., biochanin (61), daidzein (62), genistein (63), prunetin (64), and puerarin (65) were found in quinoa (Table 7 and Figure 7). They showed antinematodal activities on *Radopholus similis* [82]. Isoflavones are recognized to be estrogenic compounds that are often associated with a reduced risk of cancers. The estrogenic activity can be enhanced after metabolization to more active compounds such as daidzein (62) and genistein (63) by gut microorganisms [175].

Table 7. Isoflavones and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|------------------|--------------------------------|--|-----------|
| Biochanin A (61) | Seeds | - | [35] |
| Daidzein (62) | Seeds | - | [176] |
| | | Antioxidant activity | [177] |
| | | Enhance adipocyte differentiation and PPAR γ transcriptional activities | [178] |
| | | Affected human nonhormone-dependent cervical cancer cells | [179] |
| | | Modulate in vitro rat uterine contractile activity | [180] |
| | | Anti-hypoxia activity | [181] |
| | | Antithrombotic and anti-allergic activities | [182] |
| | | Chemoprotective activity | [183] |
| | | Inhibits bone loss in ovariectomized mice | [184] |
| | | Antiproliferative activity | [185,186] |

Table 7. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|----------------|--------------------------------|--|-----------|
| Genistein (63) | Seeds | - | [176] |
| | | Antiproliferative activity on human breast cancer cells | [185,186] |
| | | Modulate in vitro rat uterine contractile activity | [180] |
| | | Antioxidant activity | [187] |
| | | Inhibitory activity on tyrosine-specific protein kinases | [188] |
| | | Antitumor activity | [189] |
| | | Cytotoxic activity and anticancer activities | [190] |
| | | Antitumor and antiangiogenic activities | [191,192] |
| | | Antibacterial activity | [193] |
| | | Inhibition of cyclooxygenase-2 activity | [194] |
| | | Antiproliferative activity | [195] |
| | | Antileukemic activity | [196] |
| Prunetin (64) | Seeds | Induction of quinone reductase activity | [197] |
| | | Induces growth arrest and suppresses telomerase activities | [198] |
| Puerarin (65) | Seeds | - | [25] |
| | | Anti-inflammatory activity | [199] |
| Puerarin (65) | Seeds | - | [35] |
| | | Antithrombotic and antiallergic activities | [182] |
| | | Anti-apoptosis activity | [200] |
| | | Antioxidant activity | [201] |
| | | Antihyperglycemic effect | [202] |



61. Biochanin A, R₁=OH, R₂=OH, R₃=H, R₄=OCH₃
 62. Daidzein, R₁=H, R₂=OH, R₃=H, R₄=OH
 63. Genistein, R₁=OH, R₂=OH, R₃=H, R₄=OH
 64. Prunetin, R₁=OH, R₂=OCH₃, R₃=H, R₄=OH
 65. Puerarin, R₁=OH, R₂=OH, R₃=Glc, R₄=OH

Figure 7. Structures of the the isoflavonoids isolated from quinoa.

4. Terpenoids and Their Biological Activities or Functions

The terpenoids in quinoa mainly include monoterpenoids and triterpenoids which are biosynthesized through the isoprenoid metabolic pathway. The monoterpenoids usually play functions as allelochemicals in quinoa. The triterpenoids are present in the seed coats (also called bran or hull), and have a characteristic bitter or astringent taste to protect it from birds and insects, and possess detergent properties [2]. The saponins are also of interest as valuable adjuvants and the first saponin-based vaccines have been introduced commercially [203].

4.1. Monoterpenoids and Their Biological Activities or Functions

Quinoa monoterpenoids and their biological activities are listed in Table 8. Their structures are shown in Figure 8. At least 15 monoterpenoids in the essential oils of quinoa from the East Mediterranean have been identified [204]. Penstebioside (74) was an iridoid glycoside isolated from the flour of

quinoa [33]. γ -Terpinene (78) was also isolated from rice to show antibacterial activity on *Xanthomonas oryzae* pv. *oryzae* (Xoo) [205].

Table 8. Monoterpenoids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--------------------------------------|--------------------------------|---------------------------------|-------|
| <i>cis</i> -Ascaridole (66) | Leaves | - | [204] |
| <i>cis</i> -Isoascaridole (67) | Leaves | - | [204] |
| Camphene (68) | Leaves | - | [204] |
| Camphor (69) | Leaves | - | [204] |
| <i>trans</i> -Carveol (70) | Leaves | - | [204] |
| <i>p</i> -Cymene (71) | Leaves | - | [204] |
| <i>p</i> -Mentha-1(7),8-diene (72) | Leaves | - | [204] |
| <i>trans-p</i> -Menth-2-en-1-ol (73) | Leaves | - | [204] |
| Penstebioside (74) | Flour | - | [33] |
| β -Pinene (75) | Leaves | - | [204] |
| Pinocarvone (76) | Leaves | - | [204] |
| α -Terpinene (77) | Leaves | - | [204] |
| γ -Terpinene (78) | Leaves | - | [204] |
| | | Antibacterial activity | [205] |
| Terpin-1-ol (79) | Leaves | - | [204] |
| α -Terpinyl acetate (80) | Leaves | - | [204] |

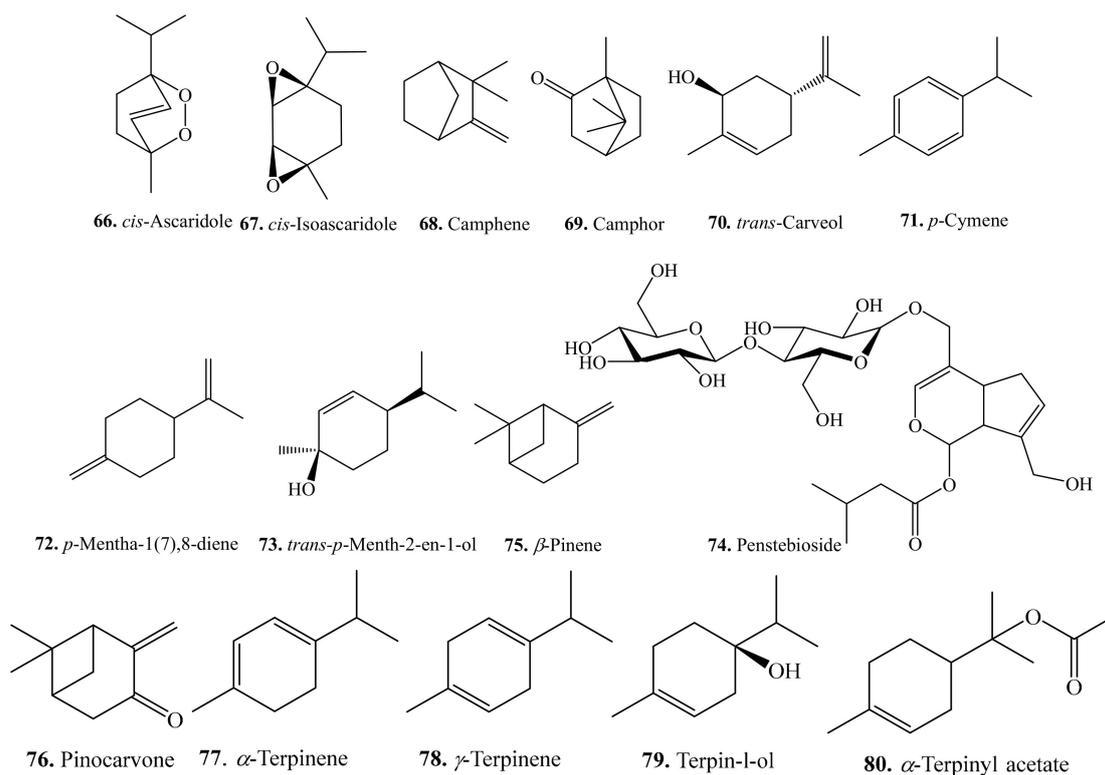


Figure 8. Structures of the monoterpenoids isolated from quinoa.

4.2. Sesquiterpenoids and Their Biological Activities or Functions

Only one sesquiterpene namely caryophyllene (81) was identified in quinoa [204]. Its structure is shown in Figure 9.

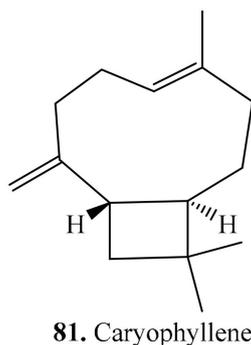


Figure 9. Structure of the sesquiterpenoid isolated from quinoa.

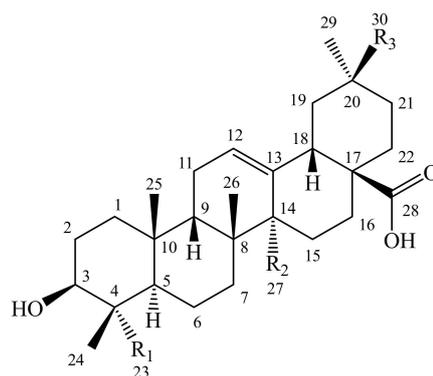
4.3. Triterpenoids and Their Biological Activities or Functions

Triterpenoids, including their aglycones (sapogenins) and glycosides (saponins), are mainly present in the bran to protect quinoa from pests and herbivores (i.e., birds and insects) and pathogenic microorganisms [206]. Quinoa saponins are characterized as the bitter metabolites. The quinoa could be classified into bitter and sweet varieties according to the triterpenoid saponin content, which is much lower in the sweet varieties and higher in the bitter ones [138,207].

The crude saponin fraction inhibited the growth of *Candida albicans* at 50 $\mu\text{g/mL}$ [208]. The alkali-transformed saponin from quinoa bran showed inhibition against halitosis-related bacterium *Fusobacterium nucleatum*, with a minimum inhibitory concentration (MIC) of 31.3 $\mu\text{g/mL}$. It could be used as an antibacterial agent to treat halitosis [209]. When the fungal pathogen *Botrytis cinerea* was treated with the saponin extracts, mycelial growth and conidial germination were significantly inhibited [210].

When golden apple snails (*Pomacea canaliculata*, GAS) were treated with the crude saponin under laboratory conditions in 24 h at approximately 33 $\mu\text{g/mL}$, they were completely killed [211]. Similarly, when giant apple snails (*Pomacea maculata*) were treated with saponins above 7 $\mu\text{g/mL}$ after 72 h, they were also 100% killed. Quinoa saponin could be a viable product to safely control *P. maculata* in rice fields [212]. Therefore, quinoa saponins could be developed into molluscicide. In addition, this molluscicide was found to be non-toxic to other non-target species such as goldfish (*Carassius auratus*) and tilapia (*Oreochromis mossambicus*), while providing adequate protection from *Pomacea* snails to newly sprouted rice seeds under laboratory conditions [211,213].

The quinoa triterpenoids contain either tetracycles or pentacycles in their core structures. Most of them are pentacyclic triterpenoids in the form of saponins. The saponins contain an aglycone (sapogenin) and one to three saccharide chains in their structures, and were classified according to the number of saccharide chains as mono-, di-, and tridesmosides. Liquid chromatography-tandem mass spectrometry (LC-MS/MS) allowed a complete preassignment and identification of the major saponins and aglycones [214]. The main aglycones (Figure 10), which are oleanolic acid (82), hederagenin (83), spergulagenic acid (84), serjanic acid (85), phytolaccagenic acid (86), gypsogenin (or named 3β -hydroxy-23-oxo-olean-12-en-28-oic acid) (87), 3β -hydroxy-27-oxo-olean-12-en-28-oic acid (88), and $3\beta,23,30$ -trihydroxy-olean-12-en-28-oic acid (89), and their glycosides are shown in Tables 2–9 [23,215–217]. They have a five-ring skeleton, and are biosynthesized from β -amyrin *in planta* (134) [23]. Among them, oleanolic acid is the major aglycone [218]. Sugars, which were glucose (Glc), glucuronic acid (GlcA), galactose (Gal), arabinose (Ara), and xylose (Xyl), can be linked to the aglycone at C-3, C-23 or C-28 [214].



82. Oleanolic acid, $R_1=CH_3$, $R_2=CH_3$, $R_3=CH_3$
 83. Hederagenin, $R_1=CH_2OH$, $R_2=CH_3$, $R_3=CH_3$
 84. Spergulagenic acid, $R_1=CH_3$, $R_2=CH_3$, $R_3=COOH$
 85. Serjanic acid, $R_1=CH_3$, $R_2=CH_3$, $R_3=COOCH_3$
 86. Phytolaccagenic acid, $R_1=CH_2OH$, $R_2=CH_3$, $R_3=COOCH_3$
 87. Gypsogenin, $R_1=CHO$, $R_2=CH_3$, $R_3=CH_3$
 88. 3 β -Hydroxy-27-oxo-olean-12-en-28-oic acid, $R_1=CH_3$, $R_2=CHO$, $R_3=CH_3$
 89. 3 β ,23,30-Trihydroxy-olean-12-en-28-oic acid, $R_1=CH_2OH$, $R_2=CH_3$, $R_3=CH_2OH$

Figure 10. Structures of the main triterpenoid aglycones in quinoa.

4.3.1. Oleanolic Acid Derivatives and Their Biological Activities or Functions

About 11 oleanolic acid analogues have been identified in quinoa. Their biological activities are listed in Table 9, and the structures are shown in Figure 11. The major sugars of the saccharide moieties are arabinose, glucose and galactose [219].

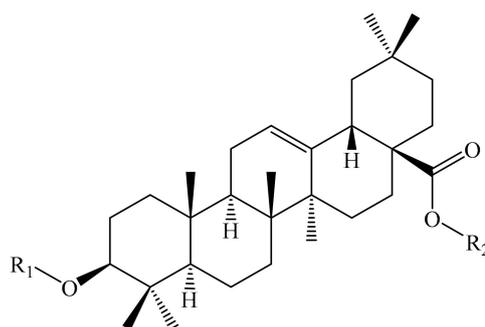
Oleanolic acid and its glycosides are mainly present in the bran (seeds) of quinoa. They showed a variety of biological activities such as antimicrobial [220,221], anti-HIV [222], anti-inflammatory [223,224], antioxidant [225], antifertility [226], antitumor or anticancer [227–229], antidiabetogenic [230], anticomplement [231] properties. They also exhibited inhibitory activities on serin protease and porcine pancreatic elastase [232].

Table 9. Oleanolic acid derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|---------------------------------|---|--|
| Oleanolic acid (82) | Seeds and bran | - Antimicrobial activity Anti-HIV activity Anti-inflammatory activity Antioxidant activity Antifertility activity Antitumor activity Inhibitory activities on serin protease and porcine pancreatic elastase | [217,233] [220,221] [222] [223] [225] [226] [227,228] [232] |
| Methyl oleanate (90) | Bran | Anti-inflammatory activity | [234] |
| 3-O- α -L-Arabinopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl oleanolic acid | Seeds | - | [235,236] |
| 28-O- β -D-glucopyranosyl ester (91) | | | |
| 3-O- β -D-Glucopyranosyl oleanolic acid (92) | Seeds | - Antidiabetogenic activity Anti-inflammatory activity Hemolytic activity | [237] [230] [224] [231] |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl oleanolic acid | Flowers, fruits, seeds and bran | - | [11,235,236] |
| 28-O- β -D-glucopyranosyl ester (93) | | | |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl oleanolic acid 28-O- β -D-glucopyranosyl ester (94) | Flowers, fruits, seeds and bran | - | [11,217,238] |

Table 9. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|------------------------------------|---------------------------------|------------------|
| 3- <i>O</i> -β- <i>D</i> -Glucopyranosyl oleanolic acid (95) | Seeds | - | [208,238] |
| 3- <i>O</i> -β- <i>D</i> -Glucuronopyranosyl oleanolic acid 28- <i>O</i> -β- <i>D</i> -glucopyranosyl ester (96) | Flowers, fruits, seeds and bran | Hemolytic activity | [11,208,217,236] |
| 3- <i>O</i> -β- <i>D</i> -Xylopyranosyl-(1→3)-β- <i>D</i> - glucuronopyranosyl oleanolic acid (97) | Seeds | - | [237] |
| 3- <i>O</i> -β- <i>D</i> -Xylopyranosyl(1→3)-6- methyl-β- <i>D</i> -glucuronopyranosyl oleanolic acid (98) | Seeds | - | [237] |
| 3- <i>O</i> -β- <i>D</i> -Xylopyranosyl-(1→3)-β- <i>D</i> - glucuronopyranosyl oleanolic acid 28- <i>O</i> -β- <i>D</i> -glucopyranosyl ester (99) | Flowers, fruits, seeds and bran | - | [11,217,237] |



82. Oleanolic acid, $R_1=H$, $R_2=H$
 90. Methyl oleanate, $R_1=H$, $R_2=CH_3$
 91. 3-*O*-α-*L*-Arabinopyranosyl-(1→3)-β-*D*-glucuronopyranosyl oleanolic acid 28-*O*-β-*D*-glucopyranosyl ester,
 $R_1=GlcA(3\rightarrow1)Ara$, $R_2=Glc$
 92. 3-*O*-β-*D*-Glucopyranosyl oleanolic acid, $R_1=Glc$, $R_2=H$
 93. 3-*O*-β-*D*-Glucopyranosyl-(1→3)-α-*L*-arabinopyranosyl oleanolic acid 28-*O*-β-*D*-glucopyranosyl ester,
 $R_1=Ara(3\rightarrow1)Glc$, $R_2=Glc$
 94. 3-*O*-β-*D*-Glucopyranosyl-(1→2)-β-*D*-glucopyranosyl-(1→3)-α-*L*-arabinopyranoside oleanolic acid 28-*O*-β-*D*-
 glucopyranosyl ester, $R_1=Ara(3\rightarrow1)Glc(2\rightarrow1)Glc$, $R_2=Glc$
 95. 3-*O*-β-*D*-Glucopyranosyl oleanolic acid, $R_1=GlcA$, $R_2=H$
 96. 3-*O*-β-*D*-Glucuronopyranosyl oleanolic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1=GlcA$, $R_2=Glc$
 97. 3-*O*-β-*D*-Xylopyranosyl-(1→3)-β-*D*-glucuronopyranosyl oleanolic acid, $R_1=GlcA(3\rightarrow1)Xyl$, $R_2=H$
 98. 3-*O*-β-*D*-Xylopyranosyl-(1→3)-6-methyl-β-*D*-glucuronopyranosyl oleanolic acid, $R_1=6-OMe\ GlcA(3\rightarrow1)Xyl$, $R_2=H$
 99. 3-*O*-β-*D*-Xylopyranosyl-(1→3)-β-*D*-glucuronopyranosyl oleanolic acid 28-*O*-β-*D*-glucopyranosyl ester,
 $R_1=GlcA(3\rightarrow1)Xyl$, $R_2=Glc$

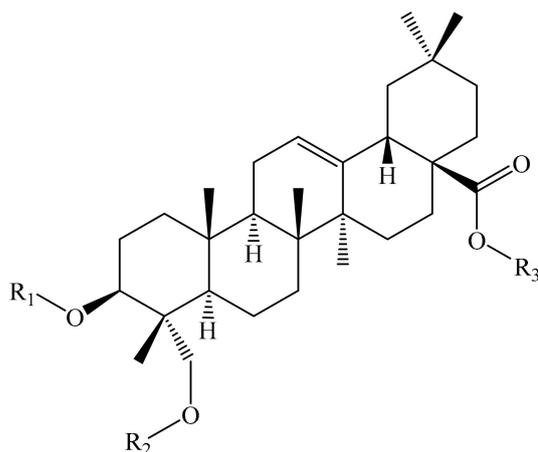
Figure 11. Structures of the oleanolic acid and its glycosides isolated from quinoa.

4.3.2. Hederagenin Derivatives and Their Biological Activities or functions

About 10 hederagenin analogues have been identified in quinoa. Their biological activities are listed in Table 10, and the structures are shown in Figure 12. Hederagenin (83) was the main aglycone of saponins from quinoa leaves [239]. Hederagenin glycosides existed in nature and possessed many biological activities such as molluscicidal [240], cytotoxic [241], antifungal [242], leishmanicidal [243], anti-inflammatory [244] activities, and they have been recently reported to show low cytotoxic properties for several human cancer cell lines with median effective concentration (EC_{50}) >30 μ M [245]. Hederagenin monodesmosides also showed strong haemolytic activity [208], hence the saponins have been considered as the serious antinutritional factors [246]. Hederagenin from the leaves of ivy (*Hedera helix*) induced apoptosis of LoVo cells through the mitochondrial apoptotic pathway, which indicated that hederagenin might be a promising therapeutic candidate for the prevention and treatment of human colon cancer [247].

Table 10. Hederagenin derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. | | |
|--|---------------------------------|--|------------------------------|------------------------|-----------|
| Hederagenin (83) | Seeds and bran | - | [215–217] | | |
| | | Inhibitory activity on serin protease, and porcine pancreatic elastase | [232] | | |
| | | Cytotoxic activity on P-388 mouse lymphoma, L-1210 mouse lymphomatic leukemia, HL-60 human promyelocytic leukemia and SNU-5 human stomach cancer cells | [248,249] | | |
| | | Haemolytic activity | [250] | | |
| | | Anti-inflammatory activity | [244] | | |
| | | Antidermatophytic activity | [251] | | |
| | | Antitrichomonas activity | [252] | | |
| | | Inducing apoptosis in human LoVo colon cells | [247] | | |
| | | 3-O- α -L-Arabinopyranosyl hederagenin (100) | Seeds | - | [208] |
| | | | | Molluscicidal activity | [140,253] |
| Cytotoxic activity on human carcinoma and melanoma cell lines DLD-1, PA1, A549, MCF7, PC3, and M4 | [241] | | | | |
| Antifungal activity | [242] | | | | |
| Leishmanicidal activity | [243] | | | | |
| 3-O- α -L-Arabinopyranosyl hederagenin 28-O- β -D-glucopyranosyl ester (101) | Flowers, fruits, seeds and bran | - | [11,208,216,217] | | |
| | | Antidermatophytic activity | [251] | | |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl hederagenin (102) | Seeds and bran | - | [208,216,238] | | |
| | | Cytotoxic activity on A549, SK-OV-3, SK-MEL-2, XF498 and HCT15 | [255] | | |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl hederagenin (103) | Bran | - | [216] | | |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranoside hederagenin 28-O- β -D-glucopyranosyl ester (104) | Flowers, fruits, seeds and bran | - | [11,208,216,217,236,238,256] | | |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl hederagenin 28-O- β -D-glucopyranosyl ester (105) | Flowers, fruits, seeds and bran | - | [11,216,238] | | |
| 3-O- β -D-Glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl hederagenin 28-O- β -D-glucopyranosyl ester (106) | Seeds | - | [256] | | |
| 3,23-Bis(O- β -D-glucopyranosyloxy)olean-12-en-28-oic acid 28-O- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl ester (107) | Seeds | - | [257] | | |
| 3-O- β -D-Glucuronopyranosyl hederagenin 28-O- β -glucopyranosyl ester (108) | Flowers, fruits, seeds and bran | - | [11,217,238] | | |
| 3-O- β -D-Xylopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl hederagenin 28-O- β -D-glucopyranosyl ester (109) | Bran | - | [217] | | |



- 83.** Hederagenin, $R_1=H$, $R_2=H$, $R_3=H$
100. 3-*O*- α -L-Arabinopyranosyl hederagenin, $R_1=Ara$, $R_2=H$, $R_3=H$
101. 3-*O*- α -L-Arabinopyranosyl hederagenin 28-*O*- β -D-glucopyranosyl ester, $R_1=Ara$, $R_2=H$, $R_3=Glc$
102. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl hederagenin, $R_1=Ara(3\rightarrow 1)Glc$, $R_2=H$, $R_3=H$
103. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl hederagenin, $R_1=Gal(3\rightarrow 1)Glc$, $R_2=H$, $R_3=H$
104. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranoside hederagenin 28-*O*- β -D-glucopyranosyl ester, $R_1=Ara(3\rightarrow 1)Glc$, $R_2=H$, $R_3=Glc$
105. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranosyl hederagenin 28-*O*- β -D-glucopyranosyl ester, $R_1=Gal(3\rightarrow 1)Glc$, $R_2=H$, $R_3=Glc$
106. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl-(1 \rightarrow 4)- β -D-glucopyranosyl hederagenin 28-*O*- β -D-glucopyranosyl ester, $R_1=Glc(4\rightarrow 1)Glc(4\rightarrow 1)Glc$, $R_2=H$, $R_3=Glc$
107. 3,23-Bis(*O*- β -D-glucopyranosyloxy) olean-12-en-28-oic acid 28-*O*- α -L-arabinopyranosyl (1 \rightarrow 3)- β -D-glucopyranosyl ester, $R_1=Glc$, $R_2=Glc$, $R_3=Glc(3\rightarrow 1)Ara$
108. 3-*O*- β -D-Glucuronopyranosyl hederagenin 28-*O*- β -D-glucopyranosyl ester, $R_1=GlcA$, $R_2=H$, $R_3=Glc$
109. 3-*O*- β -D-Xylopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl hederagenin 28-*O*- β -D-glucopyranosyl ester, $R_1=GlcA(3\rightarrow 1)Xyl$, $R_2=H$, $R_3=Glc$

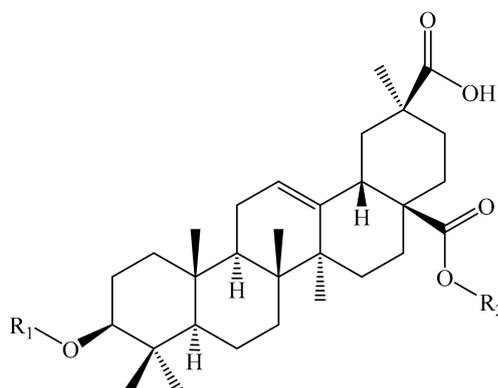
Figure 12. Structures of hederagenin and its glycosides isolated from quinoa.

4.3.3. Spergulagenic Acid Derivatives and Their Biological Activities or Functions

Spergulagenic acid (**84**), a pentacyclic triterpene used in medicine, was found in diverse plant families [258]. Until now, three spergulagenic acid glycosides (Table 11) were identified in quinoa [217,256], though spergulagenic acid as the aglycone has not been isolated from quinoa. Their structures are shown in Figure 13.

Table 11. Spergulagenic acid derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|---------------------------------|-------|
| 3- <i>O</i> - α -L-Arabinopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl spergulagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (110) | Seeds | - | [256] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl spergulagenic acid (111) | Bran | - | [217] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl spergulagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (112) | Seeds | - | [256] |



- 110.** 3-*O*- α -L-Arabinopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl spergulagenic acid
28-*O*- β -D-glucopyranosyl ester, R₁=GlcA(3 \rightarrow 1)Ara, R₂=Glc
- 111.** 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl spergulagenic acid,
R₁=Ara(3 \rightarrow 1)Glc(2 \rightarrow 1)Glc, R₂=H
- 112.** 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl spergulagenic acid,
28-*O*- β -D-glucopyranosyl ester, R₁=Ara(3 \rightarrow 1)Glc(2 \rightarrow 1)Glc, R₂=Glc

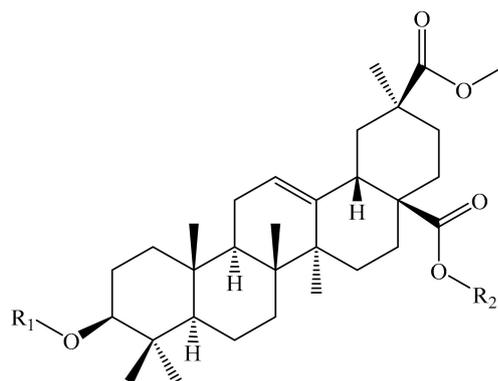
Figure 13. Structures of the spergulagenic acid glycosides isolated from quinoa.

4.3.4. Serjanic Acid Derivatives and Their Biological Activities or Functions

Serjanic acid (**85**) is the aglycone with only the bidesmosides to be found in quinoa [217]. About 5 serjanic acid analogues have been identified in quinoa (Table 12, Figure 14). Hemolysis tests showed that most monodesmoside saponins were active, and most bidesmoside saponins were inactive as the monodesmosides can reduce hydrophobic interactions with membrane lipids [208]. Similarly, both 3-*O*- α -L-arabinopyranosyl serjanic acid 28-*O*- β -D-glucopyranosyl ester (**113**) and 3-*O*- β -D-glucuronopyranosyl serjanic acid 28-*O*- β -D-glucopyranosyl ester (**116**) had weaker hemolytic activity (IC₅₀ > 100 μ g/mL) than their sapogenin (serjanic acid, IC₅₀ = 50 μ g/mL) [11].

Table 12. Serjanic acid derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|---------------------------------|--------------------------------------|------------------|
| Serjanic acid (85) | Flowers, fruits, seeds and bran | Cytotoxic activity on HeLa cell line | [11,217] |
| 3- <i>O</i> - α -L-Arabinopyranosyl serjanic acid 28- <i>O</i> - β -D-glucopyranosyl ester (113) | Flowers, fruits, seeds and bran | Cytotoxic activity on Hela cell line | [11] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl serjanic acid 28- <i>O</i> - β -D-glucopyranosyl ester = 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl-30- <i>O</i> -methyl spergulagenate 28- <i>O</i> - β -D-glucopyranosyl ester (114) | Flowers, fruits, seeds and bran | - | [11,236,238] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl serjanic acid 28- <i>O</i> - β -D-glucopyranosyl ester (?) = 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl-30- <i>O</i> -methyl spergulagenate 28- <i>O</i> - β -D-glucopyranosyl ester (115) | Flowers, fruits, seeds and bran | - | [11,217,238,256] |
| 3- <i>O</i> - β -D-Glucuronopyranosyl serjanic acid 28- <i>O</i> - β -D-glucopyranosyl ester (116) | Flowers, fruits, seeds and bran | Cytotoxic activity on HeLa cell line | [11] |



- 85.** Serjanic acid, $R_1 = H$, $R_2 = H$
113. 3-*O*- α -L-Arabinopyranosyl serjanic acid 28-*O*- β -D-glucopyranosyl ester, $R_1 = Ara$, $R_2 = Glc$
114. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl serjanic acid 28-*O*- β -D-glucopyranosyl ester, $R_1 = Ara(3\rightarrow 1)Glc$, $R_2 = Glc$
115. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl serjanic acid 28-*O*- β -D-glucopyranosyl ester, $R_1 = Ara(3\rightarrow 1)Glc(2\rightarrow 1)Glc$, $R_2 = Glc$
116. 3-*O*- β -D-Glucuronopyranosyl serjanic acid 28-*O*- β -D-glucopyranosyl ester, $R_1 = GlcA$, $R_2 = Glc$

Figure 14. Structures of serjanic acid and its glycosides isolated from quinoa.

4.3.5. Phytolaccagenic Acid Derivatives and Their Biological Activities or Functions

Phytolaccagenic acid (**86**) might be originated from serjanic acid (**85**) by subsequent oxidative enzymatic steps involving the formation of the corresponding alcohol substituted at C-23 *in planta* [11]. It is one of the main structures of quinoa saponins. About 10 phytolaccagenic acid analogues have been identified in quinoa. They are listed in Table 13, and the structures are shown in Figure 15.

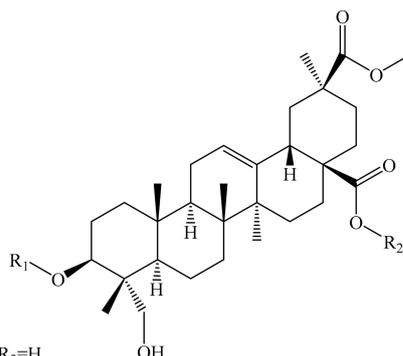
Phytolaccagenic acid saponins are highly concentrated in the bran (seed coats), which are more exposed to water during germination compared to oleanolic acid saponins [259]. It was suggested that a short saccharide chain (1 or 2 glycosyl residues) requires the presence of an additional longer one to make the saponin water-soluble [260]. Phytolaccagenic acid was employed as the anti-inflammatory drug of oral administration [234].

Table 13. Phytolaccagenic acid derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|---------------------------------|---------------------------------|------------------------------|
| Phytolaccagenic acid (86) | Bran | - | [216,217] |
| | Bran | Anti-inflammatory activity | [234] |
| 3- <i>O</i> - α -L-Arabinopyranosyl phytolaccagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (117) | Flowers, fruits, seeds and bran | - | [11,208,216,217,238] |
| 3- <i>O</i> - α -L-Arabinopyranosyl-(1 \rightarrow 3)- β -D-glucuronopyranosyl phytolaccagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (118) | Seeds | - | [235,236,256] |
| 3- <i>O</i> - β -D-Galactopyranosyl-(1 \rightarrow 3)- β -D-glucopyranosyl phytolaccagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (119) | Seeds | - | [238] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl phytolaccagenic acid (120) | Seeds | Antifungal activity | [208,238] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl phytolaccagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (121) | Flowers, fruits, seeds and bran | - | [11,208,216,217,235,236,238] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- β -D-galactopyranoside phytolaccagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (122) | Flowers, fruits, seeds and bran | - | [11,216,217,238] |
| 3- <i>O</i> - β -D-Glucopyranosyl(1 \rightarrow 2)- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranoside phytolaccagenic acid 28- <i>O</i> - β -D-glucopyranosyl ester (123) | Flowers, fruits, seeds and bran | - | [11,217,235,238] |

Table 13. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|---------------------------------|---------------------------------|----------|
| 3- <i>O</i> -β- <i>D</i> -Glucopyranosyl-(1→4)-β- <i>D</i> -glucopyranosyl-(1→4)-β- <i>D</i> -glucopyranosyl phytolaccagenic acid 28- <i>O</i> -β- <i>D</i> -glucopyranosyl ester (124) | Flowers, fruits, seeds and bran | - | [11,256] |
| 3- <i>O</i> -β- <i>D</i> -Glucopyranosyl-(1→3)-β- <i>D</i> -xylopyranosyl-(1→2)-β- <i>D</i> -glucopyranosyl phytolaccagenic acid 28- <i>O</i> -β- <i>D</i> -glucopyranosyl ester (125) | Seeds | - | [235] |



86. Phytolaccagenic acid, $R_1 = H$, $R_2 = H$
 117. 3-*O*-α-*L*-Arabinopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Ara$, $R_2 = Glc$
 118. 3-*O*-α-*L*-Arabinopyranosyl-(1→3)-β-*D*-glucuronopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = GlcA(3→1)Ara$, $R_2 = Glc$
 119. 3-*O*-β-*D*-Galactopyranosyl-(1→3)-β-*D*-glucopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Glc(3→1)Gal$, $R_2 = Glc$
 120. 3-*O*-β-*D*-Glucopyranosyl-(1→3)-α-*L*-arabinopyranosyl phytolaccagenic acid, $R_1 = Ara(3→1)Glc$, $R_2 = H$
 121. 3-*O*-β-*D*-Glucopyranosyl-(1→3)-α-*L*-arabinopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Ara(3→1)Glc$, $R_2 = Glc$
 122. 3-*O*-β-*D*-Glucopyranosyl-(1→3)-β-*D*-galactopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Gal(3→1)Glc$, $R_2 = Glc$
 123. 3-*O*-β-*D*-Glucopyranosyl-(1→2)-β-*D*-glucopyranosyl (1→3)-α-*L*-arabinopyranoside phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Ara(3→1)Glc(2→1)Glc$, $R_2 = Glc$
 124. 3-*O*-β-*D*-Glucopyranosyl-(1→4)-β-*D*-glucopyranosyl-(1→4)-β-*D*-glucopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Glc(4→1)Glc(4→1)Glc$, $R_2 = Glc$
 125. 3-*O*-β-*D*-Glucopyranosyl-(1→3)-β-*D*-xylopyranosyl-(1→2)-β-*D*-glucopyranosyl phytolaccagenic acid 28-*O*-β-*D*-glucopyranosyl ester, $R_1 = Glc(2→1)Xyl(3→1)Glc$, $R_2 = Glc$

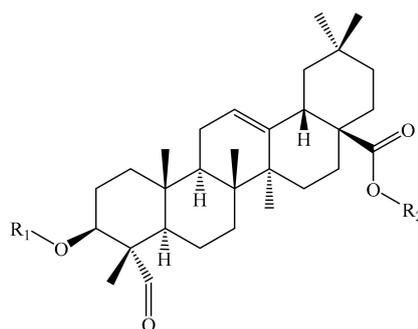
Figure 15. Structures of phytolaccagenic acid and its glycosides isolated from quinoa.

4.3.6. Gypsogenin Derivatives and Their Biological Activities or Functions

Gypsogenin (or named 3β-hydroxy-23-oxo-olean-12-en-28-oic acid) (**87**) and its glycoside 3-*O*-β-*D*-glucopyranosyl-(3)-α-*L*-arabinopyranosyl 23-oxo-olean-12-en-28-oic acid 28-*O*-β-*D*-gluco-pyranosyl ester (**126**) were isolated from quinoa (Table 14, Figure 16). They showed cytotoxic activity [11].

Table 14. Gypsogenin derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|---------------------------------|--------------------------------------|------|
| Gypsogenin = 3β-Hydroxy-23-oxo-olean-12-en-28-oic acid (87) | Flowers, fruits, seeds and bran | Cytotoxic activity on Hela cell line | [11] |
| 3- <i>O</i> -β- <i>D</i> -Glucopyranosyl-(1→3)-α- <i>L</i> -arabinopyranosyl 23-oxo-olean-12-en-28-oic acid 28- <i>O</i> -β- <i>D</i> -glucopyranosyl ester = 3β-[(<i>O</i> -β- <i>D</i> -Glucopyranosyl-(1→3)-α- <i>L</i> -arabinopyranosyl)oxy]-23-oxo-olean-12-en-28-oic acid 28- <i>O</i> -β- <i>D</i> -glucopyranosyl ester (126) | Flowers, fruits, seeds and bran | Cytotoxic activity on HeLa cell line | [11] |



87. Gypsogenin, $R_1=H$, $R_2=H$
 126. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 23-oxo-olean-12-en-28-oic acid
 28-*O*- β -D-glucopyranosyl ester, $R_1=Ara(3\rightarrow1)Glc$, $R_2=Glc$

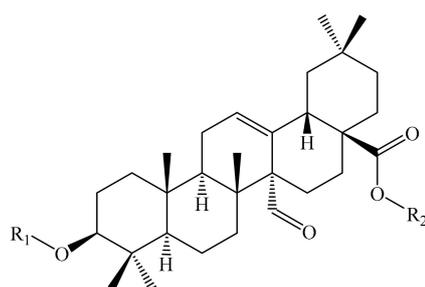
Figure 16. Structures of the gypsogenin derivatives isolated from quinoa.

4.3.7. 3 β -Hydroxy-27-oxo-olean-12-en-28-oic Acid Derivatives and Their Biological Activities or Functions

3 β -Hydroxy-27-oxo-olean-12-en-28-oic acid (**88**) and its glycoside 3-*O*- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 27-oxo-olean-12-en-28-oic acid 28-*O*- β -D-glucopyranosyl ester (**127**) were isolated from quinoa (Table 15, Figure 17). 3 β -Hydroxy-27-oxo-olean-12-en-28-oic acid (**88**) showed same cytotoxic effect as 3 β -hydroxy-23-oxo-olean-12-en-28-oic acid (**87**) with an IC_{50} value of 25.4 μ g/mL. This suggests that the CHO groups at C-23 or C-27 are correlated with the increased cytotoxicity [11].

Table 15. 3 β -Hydroxy-27-oxo-olean-12-en-28-oic acid derivatives and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|---------------------------------|--------------------------------------|------|
| 3 β -Hydroxy-27-oxo-olean-12-en-28-oic acid (88) | Flowers, fruits, seeds and bran | Cytotoxic activity on HeLa cell line | [11] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 27-oxo-olean-12-en-28-oic acid 28- <i>O</i> - β -D-glucopyranosyl ester = 3 β -[(<i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl)oxy]-27-oxo-olean-12-en-28-oic acid 28- <i>O</i> - β -D-glucopyranoside (127) | Flowers, fruits, seeds and bran | Cytotoxic activity on Hela cell line | [11] |



88. 3 β -Hydroxy-27-oxo-olean-12-en-28-oic acid, $R_1=H$, $R_2=H$
 127. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 27-oxo-olean-12-en-28-oic acid
 28-*O*- β -D-glucopyranosyl ester, $R_1=Ara(3\rightarrow1)Glc$, $R_2=Glc$

Figure 17. Structures of the 3 β -hydroxy-27-oxo-olean-12-en-28-oic acid triterpenoids isolated from quinoa.

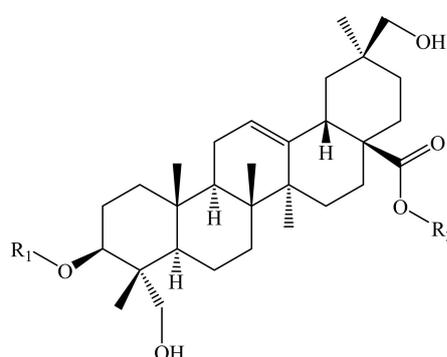
4.3.8. 3 β ,23,30-Trihydroxy-olean-12-en-28-oic Acid Triterpenoids and Their Biological Activities or Functions

3 β ,23,30-Trihydroxy-olean-12-en-28-oic acid (**89**) and 3-*O*- β -D-glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 3 β ,23,30-trihydroxy olean-12-en-28-oic acid 28-*O*- β -D-glucopyranosyl ester (**128**) have been isolated from quinoa (Table 16, Figure 18). Hederagenin (**83**) was considered as the substrate for the production of 3 β ,23,30-trihydroxyolean-12-en-28-oic acid (**89**), following a mechanism involving

a stereochemically specific enzyme able to insert one hydroxyl group into the C-30 position of the triterpene skeleton [11].

Table 16. 3 β ,23,30-Trihydroxy-olean-12-en-28-oic acid triterpenoids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|---------------------------------|---------------------------------|----------|
| 3 β ,23,30-Trihydroxy-olean-12-en-28-oic acid (89) | Flowers, fruits, seeds and bran | - | [11] |
| 3- <i>O</i> - β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 3 β ,23,30-trihydroxyolean-12-en-28-oic acid 28- <i>O</i> - β -D-glucopyranosyl ester (128) | Flowers, fruits, seeds and bran | - | [11,214] |



89. 3 β ,23,30-Trihydroxy-olean-12-en-28-oic acid. R₁=H, R₂=H
128. 3-*O*- β -D-Glucopyranosyl-(1 \rightarrow 3)- α -L-arabinopyranosyl 3 β ,23,30-trihydroxy olean-12-en-28-oic acid 28-*O*- β -D-glucopyranosyl ester, R₁= Ara (3 \rightarrow 1)Glc, R₂=Glc

Figure 18. Structures of the 3 β ,23,30-trihydroxy-olean-12-en-28-oic acid triterpenoids isolated from quinoa.

4.3.9. Other Triterpenoids and Their Biological Activities or Functions

Other triterpenoids include tetracyclic and pentacyclic triterpenoids. Their biological activities are shown in Table 17, and the structures are shown in Figures 19 and 20.

Four tetracyclic triterpenoids including two nortriterpenoids citrostadienol (**129**) and gramisterol (**130**) have been isolated from quinoa seeds [261]. Citrostadienol (**129**) showed anticomplementary activity [262], and gramisterol (**130**) showed anti-cancer activity [8].

Among the other pentacyclic triterpenoids, β -amyrin (**133**) was considered as the precursors of other triterpenoids in their biosynthetic pathways [23].

Table 17. Other triterpenoids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|--------------------------------|---|-------|
| Tetracyclic triterpenoids | | | |
| Citrostadienol (129) | Seeds | - | [261] |
| Gramisterol (130) | Seeds | Anticomplementary activity | [262] |
| | | - | [261] |
| | | Anti-cancer activity on mouse leukemic cell line WEHI-3 | [8] |
| 24-Methylene-cycloartenol (131) | Seeds | - | [261] |
| Parkeol (132) | Seeds | - | [261] |

Table 17. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|---|-----------|
| Pentacyclic triterpenoids | | | |
| α -Amyrin (133) | Seeds | - | [215] |
| | | Antibacterial activity | [263] |
| | | Antidiabetic effect | [264] |
| | | Antioxidant activity | [265] |
| | | Inhibitory activity against human oxidosqualene cyclase | [266] |
| β -Amyrin (134) | Seeds | - | [215] |
| | | Antibacterial activity | [263] |
| | | Antioxidant activity | [265] |
| | | Inhibitory activity against human oxidosqualene cyclase | [266] |
| | | Antifeedant and growth regulating activities | [267] |
| | | Insecticidal activity | [268] |
| Echinocystic acid (135) | Seeds | - | [215] |
| Erythrodiol (136) | Seeds | - | [215,261] |
| | | Antibacterial activity | [269] |
| | | Melanogenesis-inhibitory activity | [270] |
| | | Protecting the cardiovascular system | [271] |
| | | Antiproliferative and apoptotic activity | [272] |
| 3 β ,23-Dihydroxy-olean-12-ene-28,30-dioic acid (137) | Seeds | - | [208] |
| 2 β ,3 β ,23-Trihydroxy-olean-12-ene-28,30-dioic acid 30-methyl ester (138) | Bran | - | [216] |
| Queretaroic acid (139) | Seeds | - | [215] |
| Ursolic acid (140) | Seeds | - | [215] |
| | | Spasmolytic and antinociceptive activities | [85] |
| | | Cytotoxic activity | [270] |
| | | Anticancer activity | [273,274] |

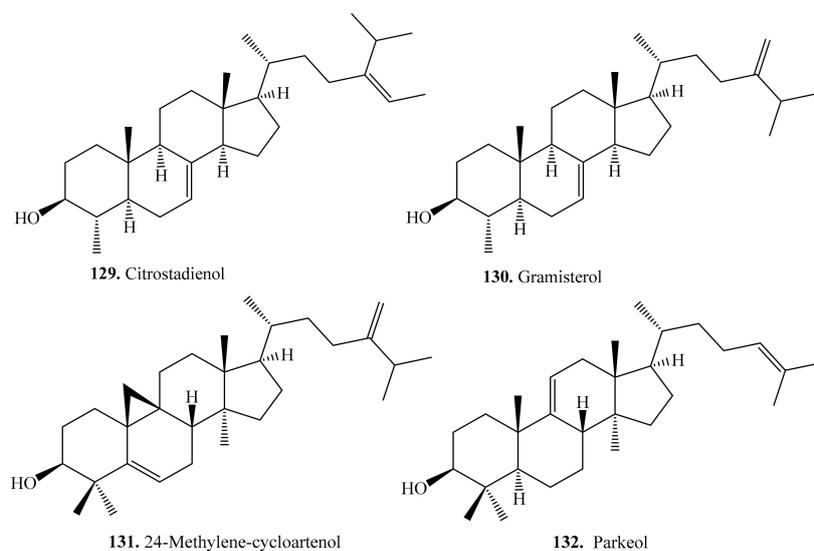


Figure 19. Structures of the tetracyclic triterpenoids isolated from quinoa.

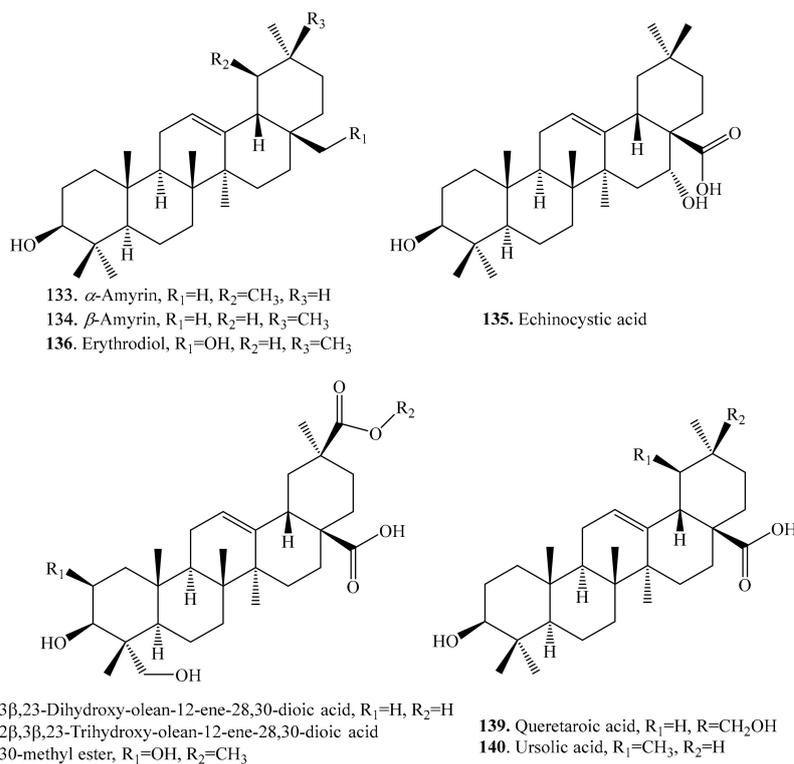


Figure 20. Structures of the other pentacyclic triterpenoids isolated from quinoa.

4.4. Meroterpenoids and Their Biological Activities or Functions

Mertoterpenoids are natural products of mixed biosynthetic origin which are partially derived from terpenoids. Meroterpenoids were also found in quinoa that include tocopherols (141–144) and tocotrienols (145,146). Their biological activities are listed in Table 18, and the structures are shown in Figure 21.

The total tocopherol content ranged from 37.49 to 59.82 $\mu\text{g/g}$ [275]. All four tocopherol isoforms (α , β , γ , and δ) have been detected in quinoa seeds, with γ -tocopherol (143) to be the most abundant followed by α -tocopherol (141), β -tocopherol (142) and δ -tocopherol (144) was the least [276]. Tocopherols acted as strong antioxidants and had many essential physiological functions such as anticoagulant, essential regulator of metabolic processes including inflammation and cancer in humans [277,278]. Among 4 tocopherols homologues, α -tocopherol (141) was considered a stronger antioxidant, whereas γ -tocopherol (143) was a stronger anti-inflammatory agent [279,280]. γ -Tocopherol (143) was the main lipophilic tocopherol in quinoa [281].

Both α -tocotrienol (145) and β -tocotrienol (146) were also identified in quinoa seeds [275]. They were the members of the vitamin E family to show antioxidant and anti-inflammatory properties [282].

Table 18. Meroterpenoids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|----------------------------|--------------------------------|---|-------|
| α -Tocopherol (141) | Seeds | - | [283] |
| | | Antioxidative, antihypercholesterolemic, anticancer, neuroprotective activities | [284] |
| β -Tocopherol (142) | Seeds | - | [37] |
| | | Antioxidative, antihypercholesterolemic, anticancer, neuroprotective activities | [284] |

Table 18. Cont.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|-----------------------------|--------------------------------|---|-------|
| γ -Tocopherol (143) | Seeds | - | [283] |
| | | Antioxidative, antihypercholesterolemic, anticancer, neuroprotective activities | [284] |
| δ -Tocopherol (144) | Seeds | - | [37] |
| | | Antioxidative, antihypercholesterolemic, anticancer, neuroprotective activities | [284] |
| α -Tocotrienol (145) | Seeds | - | [275] |
| | | Antioxidant and anti-inflammatory activities | [282] |
| β -Tocotrienol (146) | Seeds | - | [275] |
| | | Antioxidant and anti-inflammatory activities | [282] |

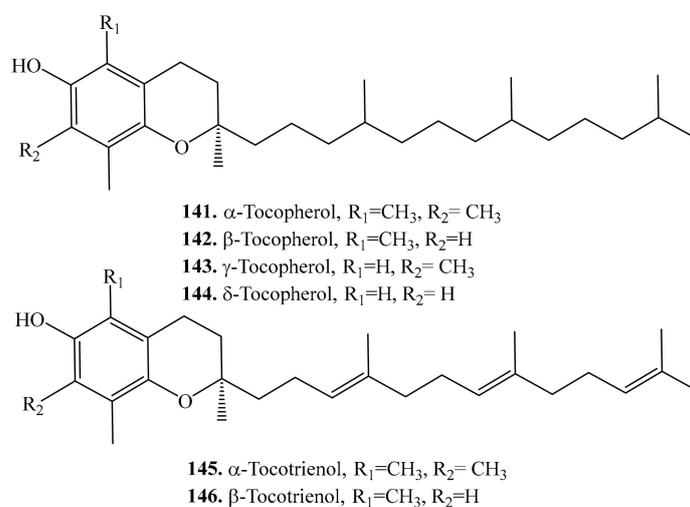


Figure 21. Structures of the meroterpenoids isolated from quinoa.

5. Steroids and Their Biological Activities or Functions

Quinoa contains a lot of biologically active phytoecdysteroids, which have been implicated in plant defense from insects, and have displayed potential pharmacologic and metabolic properties in mammals. According to the carbon skeletons, quinoa steroids can be classified as C_{27} -, C_{28} - and C_{29} -steroids.

About 36 steroids have been identified in quinoa. Seven sterols were identified among the quinoa lipids, namely cholesterol (**147**), campesterol (**160**), Δ^7 -campesterol (**161**), Δ^5 -avenasterol (**172**), β -sitosterol (**176**), stigmasterol (**181**), and Δ^7 -stigmasterol (**182**) [285]. Eleven 4,4-desmethylsterols were assigned, with Δ^7 -avenasterol (**173**), β -sitosterol (**176**), and Δ^7 -stigmasterol (**180**) being the most abundant (8.7, 27.2, and 51.3% of total sterols, respectively) [261].

5.1. C_{27} -Steroids and Their Biological Activities or Functions

Eleven C_{27} -steroids were identified in quinoa seeds which are listed in Table 19. Their structures are shown in Figure 22. Among them, ecdysteroids are main steroids which are insect moulting hormones and protect plants against non-adapted insects and nematodes [21]. Ecdysteroids are mainly present in the bran, the major component is 20-hydroxyecdysone (**148**) possessing a 14α -hydroxy-7-en-6-one chromophore and A/B-*cis* ring fusion (5β -H) [21].

Eating quinoa seeds or quinoa-derived products provides significant amounts of ecdysteroids that may be beneficial to animal or human health [22]. Quinoa extract enriched in 20-hydroxyecdysone has an antiobesity activity in vivo and could be used as a nutritional supplement for the prevention and treatment of obesity and obesity-associated disorders. The findings indicated that the extract

acted by reducing both fatty acid uptake and esterification in adipocyte [286]. It was found that 8 isolated ecdysteroids showed a stronger free-radical-scavenging activity, which was almost 3 to 8 times higher than that of the well-known antioxidant compound, BHA, and also possessed a strong ability to sequester ferrous ions. This observation supported that if the number of hydroxyl and methyl groups bearing the carbon skeleton of ecdysteroids is higher, the antioxidant activity becomes stronger. The ability of ecdysteroids to sequester ferrous ions is thought to be due to their carbonyl conjugated to a double bond attached to the C-7. Ecdysteroids are also able to inhibit skin collagenase, and could therefore also prevent skin ageing [287]. In addition, ecdysteroids have been reported to occur in Chenopodiaceae to show their possible chemotaxonomic and ecological implications [21].

Table 19. C₂₇-Steroids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|---|-----------|
| Cholesterol (147) | Seeds | - | [285,288] |
| 20-Hydroxyecdysone (148) | Seeds | Antioxidant activity | [289] |
| | | Inhibitory activity on collagenase | [287] |
| | | Insecticidal activity | [290] |
| 20,26-Dihydroxyecdysone (149) | Seeds | Antioxidant activity | [289] |
| | | Inhibitory activity on collagenase | [287] |
| 2-Deoxy-20-hydroxyecdysone (150) | Seeds | - | [22] |
| 3- <i>epi</i> -2-Deoxy-20-hydroxyecdysone (151) | Seeds | - | [22] |
| 2-Deoxy-20,26-dihydroxyecdysone (152) | Seeds | - | [22] |
| 20-Hydroxyecdysone 22-glycolate (153) | Seeds | Antioxidant activity and inhibitory activity on collagenase | [287] |
| 24,25-Dehydroinokosterone (154) | Seeds | - | [22] |
| 25,27-Dehydroinokosterone (155) | Seeds | - | [22] |
| Lathosterol (156) | Seeds | - | [261] |
| Polypodine B (157) | Seeds | - | [22] |

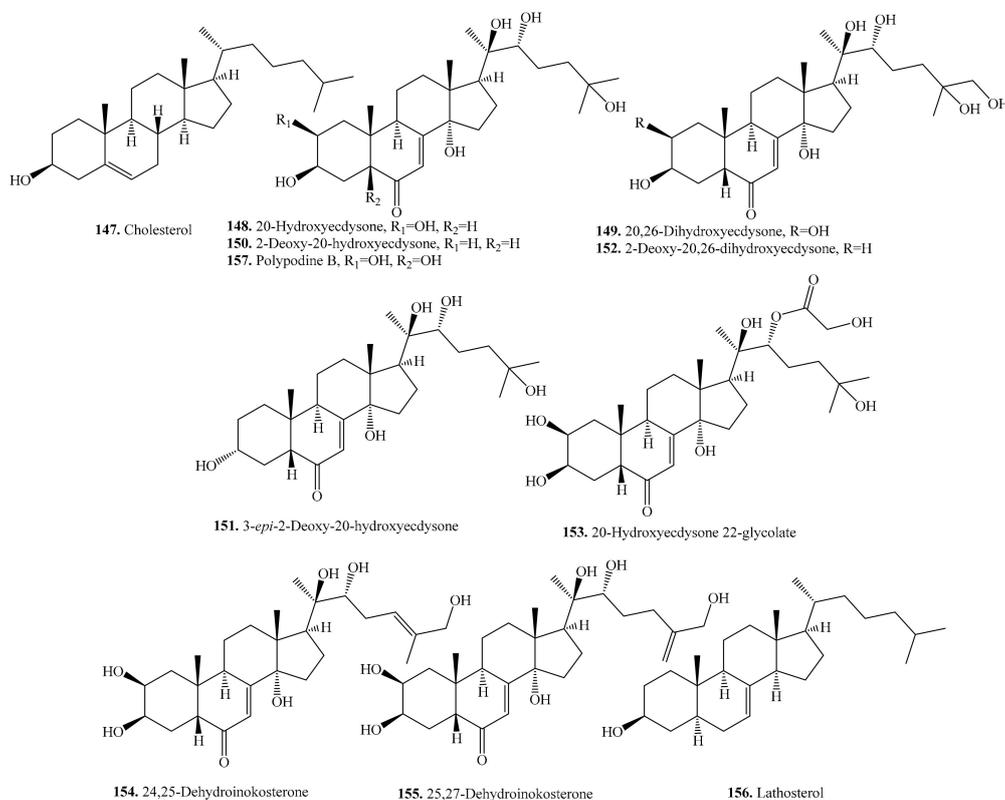


Figure 22. Structures of the C₂₇-steroids isolated from quinoa.

5.2. C₂₈-Steroids and Their Biological Activities or Functions

About 14 C₂₈-steroids such as campesterol (160), makisterone A (166), and their derivatives have been identified from quinoa seeds [34]. Their biological activities are listed in Table 20, and the structures are shown in Figure 23. The main biological activities include antioxidant activity [289], antiangiogenic activity [291], and inhibitory activity on collagenase [287].

Table 20. C₂₈-Steroids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|--|--------------------------------|--|---------------|
| Brassicasterol (158) | Seeds | - | [292] |
| Campestanol (159) | Seeds | - | [261] |
| Campesterol = Δ^5 -Campesterol (160) | Seeds | - | [261,285,288] |
| | | Antiangiogenic activity | [291] |
| Δ^7 -Campesterol (161) | Seeds | - | [285,288] |
| Dacrysterone (162) | Seeds | - | [22] |
| Episterol (163) | Seeds | - | [261] |
| Ergost-7-en-3 β -ol = Δ^7 -Ergosterol (164) | Seeds | - | [261] |
| Kancollosterone (165) | Seeds | - | [293] |
| Makisterone A (166) | Seeds | Antioxidant activity | [289] |
| | | Inhibitory activity on collagenase | [287] |
| 24- <i>epi</i> -Makisterone A (167) | Seeds | Antioxidant activity | [289] |
| | | Inhibitory activity on collagenase | [287] |
| 24(28)-Dehydromakisterone A (168) | Seeds | Antioxidant activity | [289] |
| | | Inhibitory activity on collagenase | [287] |
| 26-Hydroxy-24(28)-dehydromakisterone A (169) | Seeds | Antioxidant activity, inhibitory activity on collagenase | [287] |
| 5 β -Hydroxy-24(28)-dehydromakisterone A (170) | Seeds | - | [22] |
| 24-Methyl-20,26-dihydroxyecdysone (171) | Seeds | - | [22] |
| | Seeds | Antioxidant activity | [287] |

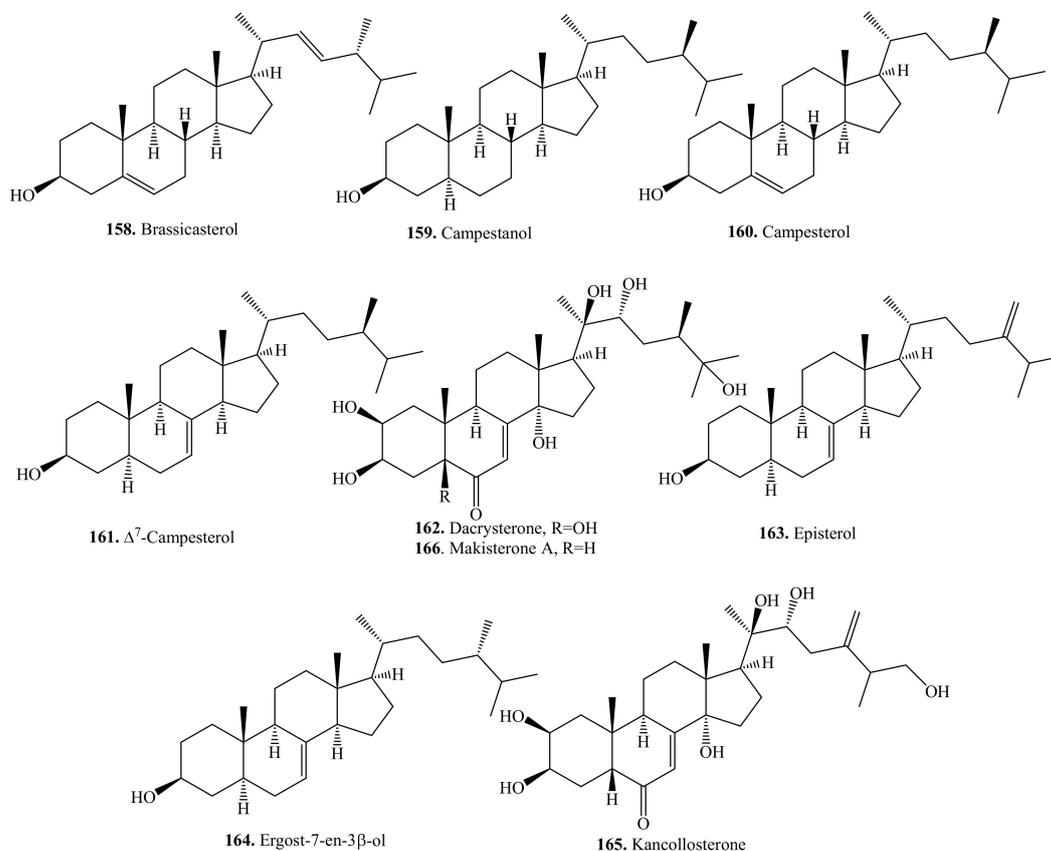


Figure 23. Cont.

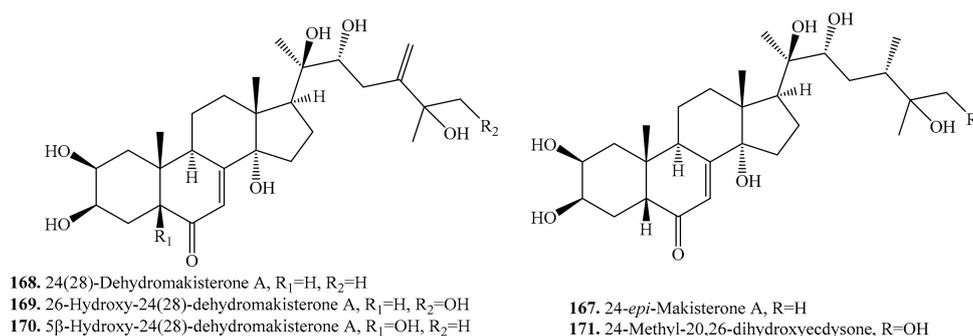


Figure 23. Structures of the C₂₈-steroids isolated from quinoa.

5.3. C₂₉-Steroids and Their Biological Activities or Functions

The main C₂₉-steroids in quinoa included avenasterol (172/173), sitosterol (176), stigmasterol (181), and their derivatives. They were all identified in the lipid extract of quinoa seeds [261]. Their biological activities are listed in Table 21, and the structures are shown in Figure 24.

β-Sitosterol (176) has been reported to have a variety of biological activities such as anti-inflammatory [294], antioxidant [295], and antidiabetic [296] activities. Stigmasterol (181) also exhibited various biological activities such as anti-inflammatory [297], anti-tumor [298], antifungal [299], anti-hypercholesterolemic [300], and cytotoxicity [301] activities.

Table 21. C₂₉-Steroids and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|------------------------------------|---------------|
| Δ ⁵ -Avenasterol = Δ ^{5,24(28)} -Avenasterol (172) | Seeds | - | [261,285,288] |
| Δ ⁷ -Avenasterol = Δ ^{7,24(28)} -Avenasterol (173) | Seeds | - | [261] |
| Makisterone C (174) | Seeds | - | [22] |
| Sitostanol (175) | Seeds | - | [261] |
| β-Sitosterol (176) | Seeds | - | [285,288] |
| | | Insecticidal activity | [268] |
| | | Anti-inflammatory activity | [294] |
| | | Anti-oxidant activity | [295] |
| | | Antidiabetic activity | [296] |
| | | Inducing apoptosis | [302] |
| | | Hypocholesterolemic activity | [303,304] |
| | | Angiogenic effect | [305] |
| | | Genotoxicity effect | [306] |
| | | Anthelminthic and | [307] |
| | | Anti-mutagenic activity | [307] |
| | | Immunomodulatory activity | [308] |
| | | Neuroprotection effect | [309] |
| Stigmast-4-en-3-one (177) | Seeds | | [292] |
| Stigmast-4,22-dien-3-one (178) | Seeds | | [292] |
| Stigmast-8-en-3-ol (179) | Seeds | - | [261] |
| Δ ⁷ -stigmasterol (180) | Seeds | - | [261] |
| Stigmasterol = Δ ⁵ -Stigmasterol (181) | Seeds | - | [261] |
| | | Anti-inflammatory activity | [297] |
| | | Anti-tumor activity | [298] |
| | | Antifungal activity | [299] |
| | | Anti-hypercholesterolemic activity | [300] |
| | | Cytotoxicity activity | [301] |
| | | Anti-osteoarthritic activity | [310] |
| Δ ⁷ -Stigmasterol (182) | Seeds | - | [285,288] |
| | Seeds | - | [261] |

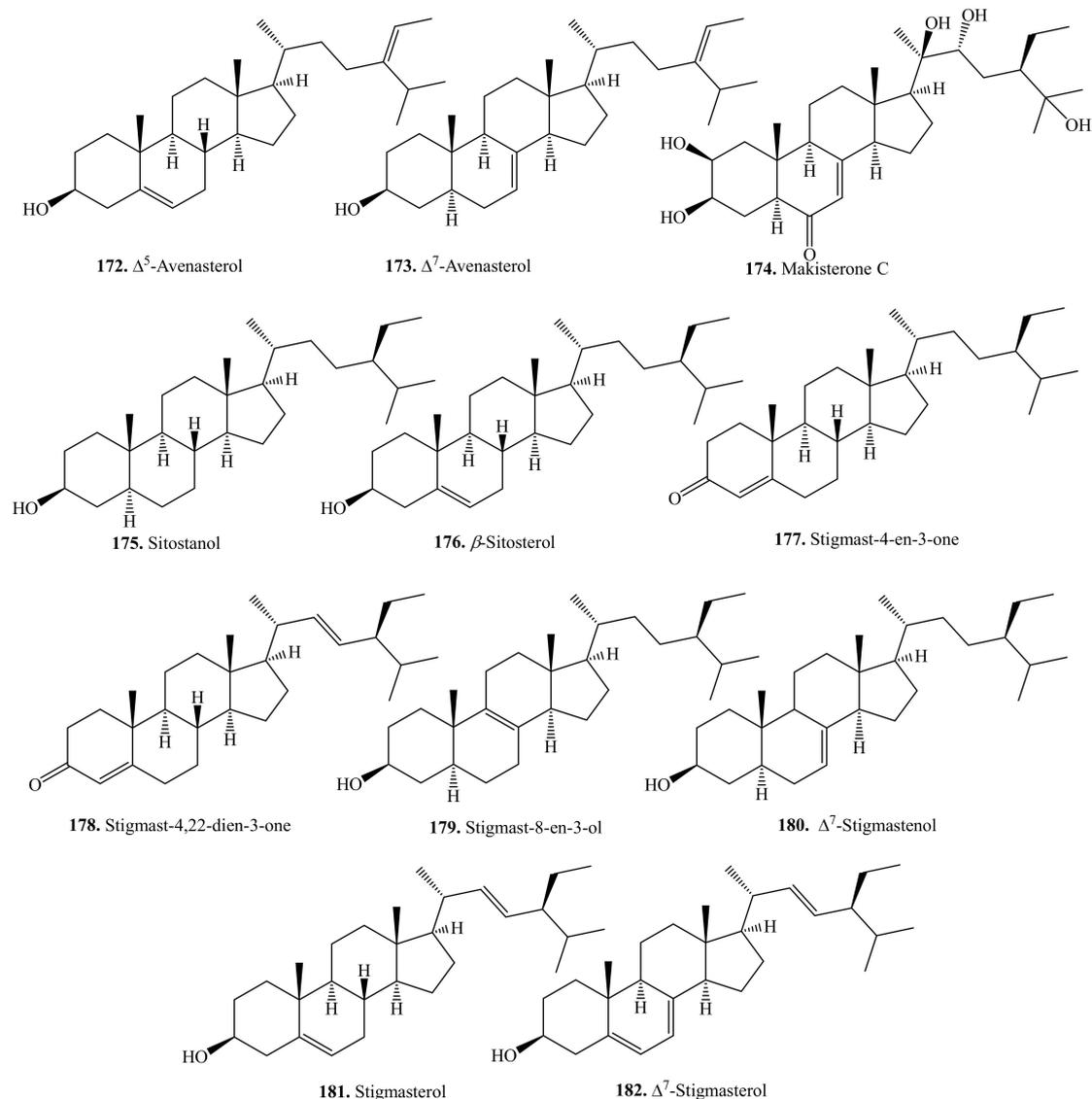


Figure 24. Structures of the C_{29} -steroids isolated from quinoa.

6. Nitrogen-Containing Metabolites and Their Biological Activities or Functions

About 12 nitrogen-containing metabolites have been identified in quinoa seeds. They belong to the derivatives of glycine and tyrosine. Their biological activities are listed in Table 22, and the structures are shown in Figure 25.

Betalains are tyrosine-derived red-violet and yellow pigments found in quinoa [311]. They are divided into two groups, betacyanins (red and purple) and betaxanthins (yellow and orange). Betacyanins are derivatives of betanidin, the conjugate of betalamic acid with *cyclo*-Dopa. Betacyanins, including amaranthin (183), betanin (184) and isobetanin (185), were confirmed in red and black quinoa seeds, instead of anthocyanins [35]. Betaxanthins are conjugates of betalamic acid with amino acids. Betaxanthins mainly include dopaxanthin (188), indicaxanthin (189), and miraxanthin V (190) in quinoa [312].

Betalains showed promising bioactive potential, such as high antioxidant and free radical scavenging activities [30]. Betacyanins and betaxanthins showed the highest antioxidant activity by comparing the white and black quinoa varieties [35]. These two varieties are characterized by a high content of dopaxanthin (188), whose dihydroxylated substructure demonstrated high antioxidant capacity [36].

Other nitrogen-containing metabolites in quinoa include betaine (186), trigonelline (191), and their derivatives. In mammals, betaine (186) acted as an osmolyte in the inner medulla of the kidney, preserving osmotic equilibrium, maintaining at the same time the tertiary structure of macromolecules [313]. Trigonelline (191) was considered to be an important multifunctional natural plant hormone with potential taxonomic value [314], and has been shown to stabilize enzyme activity in vitro [315].

Table 22. Nitrogen-containing metabolites and their biological activities or functions.

| Name | Quinoa Part Used for Isolation | Biological Activity or Function | Ref. |
|---|--------------------------------|---------------------------------|-------|
| Amaranthin (183) | Seeds | - | [316] |
| Betanin (184) | Seeds | - | [35] |
| | | Antioxidant activity | [317] |
| Isobetainin (185) | Seeds | - | [35] |
| Betaine (186) | Seeds | - | [35] |
| 3-Carboxy-1-(2-sulfoethyl)-pyridinium (187) | Seeds | - | [313] |
| Dopaxanthin (188) | Seeds | - | [316] |
| | | Antioxidant activity | [318] |
| Indicaxanthin (189) | Seeds | - | [316] |
| Miraxanthin V (190) | Seeds | - | [316] |
| Trigonelline (191) | Seeds | - | [313] |
| | | Anti-invasive activity | [319] |
| | | Hypoglycemic effect | [320] |
| Trigonelline glucosylester (192) | Seeds | - | [313] |
| Trigonelline methylester (193) | Seeds | - | [313] |

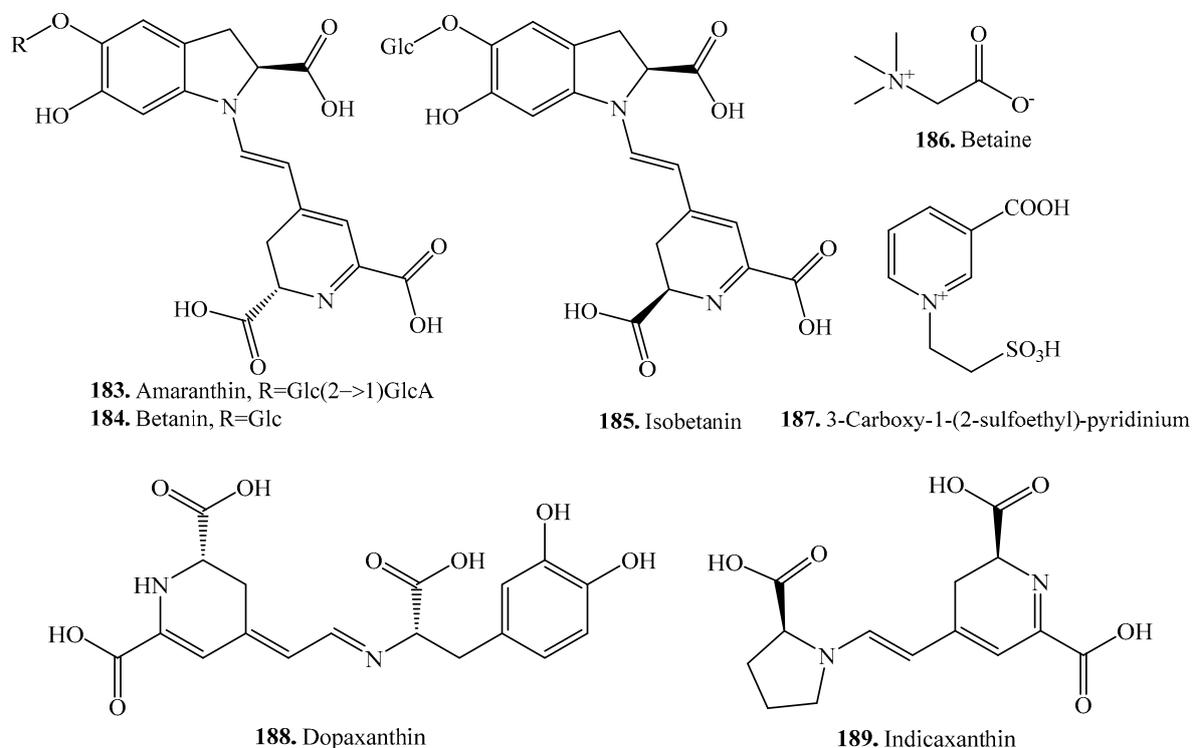


Figure 25. Cont.

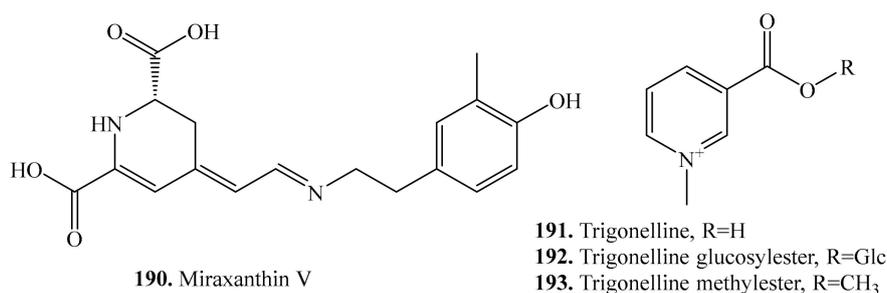


Figure 25. Structures of the nitrogen-containing metabolites isolated from quinoa.

7. Conclusions and Future Perspectives

This review focuses on the structures, isolation parts, biological activities or functions of quinoa secondary metabolites during the past 40 years. Flavonoids and phenolic acids were mostly derived from quinoa seeds. Steroids were mostly separated from quinoa bran. Triterpenoids were also mainly located in the bran. Their biological activities or functions have been reported but not comprehensive, and are needed to be systematically evaluated in the future.

The bitter taste associated with saponins (triterpenoids) greatly limits the use of quinoa as food [18]. Approximately 34% of quinoa saponins are present in the bran, indicating that dehulling could remove almost one half of the saponins. The seeds should be milled to remove the bran (seed coats) to make them edible [239]. Another method to remove saponins from the seeds is washing due to the high water solubility of saponins although this method can lead to the loss of some nutrients such as vitamins and minerals [18].

With the increased demand for quinoa, the problem that comes with it is that the bran is discarded as an industrial production waste. In order to increase the added value of quinoa, the bran (seed coat) should be fully exploited and utilized [321]. Quinoa saponins have shown their great potential applications. They can be used in the pharmaceutical industry as the saponins can induce changes in intestinal permeability which can be useful for the absorption of specific medicines and in hypocholesterolemia [15,322–324]. Quinoa saponins are also of interest as valuable adjuvants and the first saponin-based vaccines have been introduced commercially [203]. In addition, the saponins can be used as bitters, antibiotics to control pathogenic fungi and bacteria, or to protect crop against attack by birds and other pests [325]. Quinoa saponins have been successfully developed as a bioinsecticide in Bolivia [326]. They can also be used as emulsifiers and detergents due to surface active characteristics which saponins have [327]. Quinoa saponins might be developed into products like soaps, shampoos, and bitters in the future. As phenolic acids, flavonoids, and steroids are also abundant in the bran, they can be developed into antimicrobials, antioxidants, and insect moulting hormones, respectively [5,21]. It is worth mentioning that 20-hydroxyecdysone (148), mainly present in the bran, has potential for development as an insect moulting hormone [21]. After the above secondary metabolites are extracted from the bran, the remaining residues, which mainly contain cellulose, could be either used as feed, or fermented into biofuels and biofertilizer.

Biosynthesis research on quinoa secondary metabolites has rarely been reported. Methyl jasmonate was reported to induce accumulation of saponins in quinoa leaves and induce the expression of saponin biosynthetic genes in quinoa [328]. Knowledge of the saponin biosynthesis and its regulation in quinoa may aid the further development of sweet cultivars. Genome sequencing of quinoa revealed a diversity of biosynthetic core genes of secondary metabolites [329], indicating the great potential of this plant to produce various secondary metabolites with biological activities or functions which merit further investigation.

Author Contributions: Bibliographic research and original draft preparation, M.L.; manuscript discussion and corrections: P.H., Y.L. and W.W.; manuscript revision, D.L.; conception, design, supervision of the manuscript, L.Z. All authors read and approved the final manuscript.

Acknowledgments: This work was financed by the grant from the National Key R&D Program of China (2017YFD0201105).

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Jacobsen, S.E. The worldwide potential for quinoa (*Chenopodium quinoa* Willd.). *Food Rev. Int.* **2003**, *19*, 167–177.
2. Vega-Galvez, A.; Miranda, M.; Vergara, J.; Uribe, E.; Puente, L.; Martínez, E.A. Nutrition facts and functional potential of quinoa (*Chenopodium quinoa* Willd.), an ancient Andean grain: A review. *J. Sci. Food Agric.* **2010**, *90*, 2541–2547.
3. Repo-Carrasco, R.; Espinoza, C.; Jacobsen, S.E. Nutritional value and use of the Andean crops quinoa (*Chenopodium quinoa*) and kaniwa (*Chenopodium pallidicaule*). *Food Rev. Int.* **2003**, *19*, 179–189.
4. Ng, S.C.; Anderson, A.; Coker, J.; Ondrus, M. Characterization of lipid oxidation products in quinoa (*Chenopodium quinoa*). *Food Chem.* **2007**, *101*, 185–192.
5. Abugoch, L.E. Quinoa (*Chenopodium quinoa* Willd.): Composition, chemistry, nutritional and functional properties. *Adv. Food Nutr. Res.* **2009**, *58*, 1–31.
6. Jancurova, M.; Minarovicova, L.; Dandar, A. Quinoa—A review. *Czech J. Food Sci.* **2009**, *27*, 71–79.
7. Kim, S.J.; Pham, T.H.; Bak, Y.; Ryu, H.W.; Oh, S.R.; Yoon, D.Y. Orientin inhibits invasion by suppressing MMP-9 and IL-8 expression via the PKC α /ERK/AP-1/STAT3-mediated signaling pathways in TPA-treated MCF-7 breast cancer cells. *Phytomedicine* **2018**, *50*, 35–42.
8. Suttiarporn, P.; Chumpolsri, W.; Mahatheeranont, S.; Luangkamin, S.; Teepsawang, S.; Leardkamokkarn, V. Structures of phytosterols and triterpenoids with potential anti-cancer activity in bran of black non-glutinous rice. *Nutrients* **2015**, *7*, 1672–1687.
9. Graf, B.L.; Poulev, A.; Kuhn, P.; Grace, M.H.; Lila, M.A.; Raskin, I. Quinoa seeds leach phytoecdysteroids and other compounds with anti-diabetic properties. *Food Chem.* **2014**, *163*, 178–185.
10. Hu, Y.; Zhang, J.; Zou, L.; Fu, C.; Li, P.; Zhao, G. Chemical characterization, antioxidant, immune-regulating and anticancer activities of a novel bioactive polysaccharide from *Chenopodium quinoa* seeds. *Int. J. Biol. Macromol.* **2017**, *99*, 622–629.
11. Kuljanabhadgavad, T.; Thongphasuk, P.; Chamulitrat, W.; Wink, M. Triterpene saponins from *Chenopodium quinoa* Willd. *Phytochemistry* **2008**, *69*, 1919–1926.
12. Miranda, M.; Delatorre-Herrera, J.; Vega-Galvez, A.; Jorquera, E.; Quispe-Fuentes, I.; Martinez, E.A. Antimicrobial potential and phytochemical content of six diverse sources of quinoa seeds (*Chenopodium quinoa* Willd.). *Agric. Sci.* **2014**, *5*, 1015–1024.
13. Yao, Y.; Yang, X.; Shi, Z.; Ren, G. Anti-inflammatory activity of saponins from quinoa (*Chenopodium quinoa* Willd.) seeds in lipopolysaccharide-stimulated RAW 264.7 macrophages cells. *J. Food Sci.* **2014**, *79*, H1018–H1023.
14. Yao, Y.; Shi, Z.; Ren, G. Antioxidant and immunoregulatory activity of polysaccharides from quinoa (*Chenopodium quinoa* Willd.). *Int. J. Mol. Sci.* **2014**, *15*, 19307–19318.
15. Estrada, A.; Li, B.; Laarveld, B. Adjuvant action of *Chenopodium quinoa* saponins on the induction of antibody responses to intragastric and intranasal administered antigens in mice. *Comp. Immunol. Microbiol. Infect. Dis.* **1998**, *21*, 225–236.
16. Filho, A.M.M.; Pirozi, M.R.; Borge, J.T.D.S.; Sant’Ana, H.M.P.; Chaves, J.B.P.; Coimbra, S.D.R. Quinoa: Nutritional, functional, and antinutritional aspects. *Crit. Rev. Food Sci. Nutr.* **2017**, *57*, 1618–1630.
17. Tang, Y.; Tsao, R. Phytochemicals in quinoa and amaranth grains and their antioxidant, anti-inflammatory, and potential health beneficial effects: A review. *Mol. Nutr. Food Res.* **2017**, *61*, 1600767.
18. Suarez-Estrella, D.; Torri, L.; Pagani, M.A.; Marti, A. Quinoa bitterness: Causes and solutions for improving product acceptability. *J. Sci. Food Agric.* **2018**, *98*, 4033–4041.
19. Hinojosa, L.; Gonzalez, J.A.; Barrios-Masias, F.H.; Fuentes, F.; Murphy, K.M. Quinoa abiotic stress responses: A review. *Plants* **2018**, *7*, 106.
20. Ruiz, K.B.; Biondi, S.; Oses, R.; Acuna-Rodriguez, I.S.; Antognoni, F.; Martinez-Mosqueira, E.A.; Coulidaly, A.; Canahua-Murillo, A.; Pinto, M.; Zurita-Silva, A.; et al. Quinoa biodiversity and sustainability for food security under climate change. A review. *Agron. Sustain. Dev.* **2014**, *34*, 349–359.
21. Dinan, L. Phytoecdysteroids: Biological aspects. *Phytochemistry* **2001**, *57*, 325–353.

22. Kumpun, S.; Maria, A.; Crouzet, S.; Evrard-Todeschi, N.; Girault, J.P.; Lafont, R. Ecdysteroids from *Chenopodium quinoa* Willd., an ancient Andean crop of high nutritional value. *Food Chem.* **2011**, *125*, 1226–1234.
23. Kuljanabhadgavad, T.; Wink, M. Biological activities and chemistry of saponins from *Chenopodium quinoa* Willd. *Phytochem. Rev.* **2009**, *8*, 473–490.
24. Abd El-Mawla, A.M.A.; Beerhues, L. Benzoic acid biosynthesis in cell cultures of *Hypericum androsaemum*. *Planta* **2002**, *214*, 727–733.
25. Tang, Y.; Zhang, B.; Li, X.; Chen, P.X.; Zhang, H.; Tsao, R. Bound phenolics of quinoa seeds released by acid, alkaline, and enzymatic treatments and their antioxidant and α -glucosidase and pancreatic lipase inhibitory effects. *J. Agric. Food Chem.* **2016**, *64*, 1712–1719.
26. Gómez-Caravaca, A.M.; Iafelice, G.; Lavini, A.; Pulvento, C.; Caboni, M.F. Phenolic compounds and saponins in quinoa samples (*Chenopodium quinoa* Willd.) grown under different saline and nonsaline irrigation regimens. *J. Agric. Food Chem.* **2012**, *60*, 4620–4627.
27. Gawlik-Dziki, U.; Swieca, M.; Sułkowski, M.; Dziki, D.; Baraniak, B.; Czyz, J. Antioxidant and anticancer activities of *Chenopodium quinoa* leaves extracts—In vitro study. *Food Chem. Toxicol.* **2013**, *57*, 154–160.
28. Cho, J.Y.; Moon, J.H.; Seong, K.Y.; Park, K.H. Antimicrobial activity of 4-hydroxybenzoic acid and *trans* 4-hydroxycinnamic acid isolated and identified from rice hull. *Biosci. Biotech. Biochem.* **1998**, *62*, 2273–2276.
29. Tsou, M.F.; Hung, C.F.; Lu, H.F.; Wu, L.T.; Chang, S.H.; Chang, H.L.; Chen, G.W.; Chung, J.G. Effects of caffeic acid, chlorogenic acid and ferulic acid on growth and arylamine *N*-acetyltransferase activity in *Shigella sonnei* (group D). *Microbios* **2000**, *101*, 37–46.
30. Slimen, I.B.; Najjar, T.; Abderrabba, M. Chemical and antioxidant properties of betalains. *J. Agric. Food Chem.* **2017**, *65*, 675–689.
31. Ti, H.; Li, Q.; Zhang, R.; Zhang, M.; Deng, Y.; Wei, Z.; Chi, J.; Zhang, Y. Free and bound phenolic profiles and antioxidant activity of milled fractions of different indica rice varieties cultivated in Southern China. *Food Chem.* **2014**, *159*, 166–174.
32. Abou-Zaid, M.M.; Helson, B.V.; Nozzolillo, C.; Arnason, J.T. Ethyl *m*-digallate from red maple, *Acer rubrum* L., as the major resistance factor to forest tent caterpillar, *Malacosoma disstria* Hbn. *J. Chem. Ecol.* **2001**, *27*, 2517–2527.
33. Gomez-Caravaca, A.M.; Segura-Carretero, A.; Fernandez-Gutierrez, A.; Caboni, M.F. Simultaneous determination of phenolic compounds and saponins in quinoa (*Chenopodium quinoa* Willd) by a liquid chromatography-diode array detection-electrospray ionization-time-of-flight mass spectrometry methodology. *J. Agric. Food Chem.* **2011**, *59*, 10815–10825.
34. Pasko, P.; Sajewicz, M.; Gorinstein, S.; Zachwieja, Z. Analysis of selected phenolic acids and flavonoids in *Amaranthus cruentus* and *Chenopodium quinoa* seeds and sprouts by HPLC. *Acta Chromatogr.* **2008**, *20*, 661–672.
35. Tang, Y.; Li, X.; Zhang, B.; Chen, P.X.; Liu, R.; Tsao, R. Characterisation of phenolics, betanins and antioxidant activities in seeds of three *Chenopodium quinoa* Willd. genotypes. *Food Chem.* **2015**, *166*, 380–388.
36. Cai, L.; Wu, C.D. Compounds from *Syzygium aromaticum* possessing growth inhibitory activity against oral pathogens. *J. Nat. Prod.* **1996**, *59*, 987–990.
37. Alvarez-Jubete, L.; Wijngaard, H.; Arendt, E.K.; Gallagher, E. Polyphenol composition and in vitro antioxidant activity of amaranth, quinoa, buckwheat and wheat as affected by sprouting and baking. *Food Chem.* **2010**, *119*, 770–778.
38. Tanaka, T.; Tanaka, T.; Tanaka, M. Potential cancer chemopreventive activity of protocatechuic acid. *J. Exp. Clin. Med.* **2011**, *3*, 27–33.
39. Liu, K.S.; Tsao, S.M.; Yin, M.C. In vitro antibacterial activity of roselle calyx and protocatechuic acid. *Phytother. Res.* **2005**, *19*, 942–945.
40. Kore, K.J.; Bramhakule, P.P.; Rachhadiya, R.M.; Shete, R.V. Evaluation of antiulcer activity of protocatechuic acid ethyl ester in rats. *Int. J. Pharm. Life Sci.* **2011**, *2*, 909–915.
41. Shi, G.F.; An, L.J.; Jiang, B.; Guan, S.; Bao, Y.M. Alpinia protocatechuic acid protects against oxidative damage in vitro and reduces oxidative stress in vivo. *Neurosci. Lett.* **2006**, *403*, 206–210.
42. Kakkar, S.; Bais, S. A review on protocatechuic acid and its pharmacological potential. *ISRN Pharmacol.* **2014**, *2014*, 952943.
43. Zhou, Z.; Zhang, Y.; Ding, X.R.; Chen, S.H.; Yang, J.; Wang, X.J.; Wang, S.Q. Protocatechuic aldehyde inhibits hepatitis B virus replication both in vitro and in vivo. *Antivir. Res.* **2007**, *74*, 59–64.

44. Galano, A.; Francisco-Márquez, M.; Alvarez-Idaboy, J.R. Mechanism and kinetics studies on the antioxidant activity of sinapinic acid. *Phys. Chem. Chem. Phys.* **2011**, *13*, 11199–11205.
45. Chong, K.P.; Rossall, S.; Atong, M. In vitro antimicrobial activity and fungitoxicity of syringic acid, caffeic acid and 4-hydroxybenzoic acid against *Ganoderma boninense*. *J. Agric. Sci.* **2009**, *1*, 15.
46. Itoh, A.; Isoda, K.; Kondoh, M.; Kawase, M.; Kobayashi, M.; Tamesada, M.; Yagi, K. Hepatoprotective effect of syringic acid and vanillic acid on concanavalin a-induced liver injury. *Biol. Pharm. Bull.* **2009**, *32*, 1215–1219.
47. Fernandez, M.A.; Saenz, M.T.; Garcia, M.D. Natural Products: Anti-inflammatory activity in rats and mice of phenolic acids isolated from *Scrophularia frutescens*. *J. Pharm. Pharmacol.* **1998**, *50*, 1183–1186.
48. El-Hawary, S.; Mohammed, R.; AbouZid, S.; Ali, Z.Y.; El-Gendy, A.O.; Elwekeel, A. In-vitro cyclooxygenase inhibitory, antioxidant and antimicrobial activities of phytochemicals isolated from *Crassula arborescens* (Mill.) Willd. *Int. J. Appl. Res. Nat. Prod.* **2016**, *9*, 8–14.
49. Dini, I.; Tenore, G.C.; Dini, A. Phenolic constituents of *Kancolla* seeds. *Food Chem.* **2004**, *84*, 163–168.
50. Tai, A.; Sawano, T.; Yazama, F.; Ito, H. Evaluation of antioxidant activity of vanillin by using multiple antioxidant assays. *BBA Gen. Subjects* **2011**, *1810*, 170–177.
51. Cava-Roda, R.M.; Taboada-Rodríguez, A.; Valverde-Franco, M.T.; Marín-Iniesta, F. Antimicrobial activity of vanillin and mixtures with cinnamon and clove essential oils in controlling *Listeria monocytogenes* and *Escherichia coli* O157: H7 in milk. *Food Bioprocess Technol.* **2012**, *5*, 2120–2131.
52. Shoeb, A.; Chowta, M.; Pallemati, G.; Rai, A.; Singh, A. Evaluation of antidepressant activity of vanillin in mice. *Indian J. Pharmacol.* **2013**, *45*, 141.
53. Lim, E.J.; Kang, H.J.; Jung, H.J.; Song, Y.S.; Lim, C.J.; Park, E.H. Anti-angiogenic, anti-inflammatory and anti-nociceptive activities of vanillin in ICR mice. *Biomol. Ther.* **2008**, *16*, 132–136.
54. Gerig, T.M.; Blum, U. Effects of mixtures of four phenolic acids on leaf area expansion of cucumber seedlings grown in Portsmouth B 1 soil materials. *J. Chem. Ecol.* **1991**, *17*, 29–40.
55. Khanduja, K.L.; Avti, P.K.; Kumar, S.; Mittal, N.; Sohi, K.K.; Pathak, C.M. Anti-apoptotic activity of caffeic acid, ellagic acid and ferulic acid in normal human peripheral blood mononuclear cells: A Bcl-2 independent mechanism. *BBA Gen. Subjects* **2006**, *1760*, 283–289.
56. Hunyadi, A.; Martins, A.; Hsieh, T.J.; Seres, A.; Zupkó, I. Chlorogenic acid and rutin play a major role in the in vivo anti-diabetic activity of *Morus alba* leaf extract on type II diabetic rats. *PLoS ONE* **2012**, *7*, e50619.
57. Chang, W.S.; Chang, Y.H.; Lu, F.J.; Chiang, H.C. Inhibitory effects of phenolics on xanthine oxidase. *Anticancer Res.* **1994**, *14*, 501–506.
58. Ohnishi, M.; Morishita, H.; Iwahashi, H.; Toda, S.; Shirataki, Y.; Kimura, M.; Kido, R. Inhibitory effects of chlorogenic acids on linoleic acid peroxidation and haemolysis. *Phytochemistry* **1994**, *36*, 579–583.
59. Kwon, S.H.; Lee, H.K.; Kim, J.A.; Hong, S.I.; Kim, H.C.; Jo, T.H.; Jang, C.G. Neuroprotective effects of chlorogenic acid on scopolamine-induced amnesia via anti-acetylcholinesterase and anti-oxidative activities in mice. *Eur. J. Pharmacol.* **2010**, *649*, 210–217.
60. Cho, A.S.; Jeon, S.M.; Kim, M.J.; Yeo, J.; Seo, K.I.; Choi, M.S.; Lee, M.K. Chlorogenic acid exhibits anti-obesity property and improves lipid metabolism in high-fat diet-induced-obese mice. *Food Chem. Toxicol.* **2010**, *48*, 937–943.
61. Kapil, A.; Koul, I.B.; Suri, O.P. Antihepatotoxic effects of chlorogenic acid from *Anthocephalus cadamba*. *Phytother. Res* **1995**, *9*, 189–193.
62. Karunanidhi, A.; Thomas, R.; Van Belkum, A.; Neela, V. In vitro antibacterial and antibiofilm activities of chlorogenic acid against clinical isolates of *Stenotrophomonas maltophilia* including the trimethoprim/sulfamethoxazole resistant strain. *BioMed. Res. Int.* **2013**, *2013*, 392058.
63. Repo-Carrasco-Valencia, R.; Hellström, J.K.; Pihlava, J.M.; Mattila, P.H. Flavonoids and other phenolic compounds in Andean indigenous grains: Quinoa (*Chenopodium quinoa*), kaniwa (*Chenopodium pallidicaule*) and kiwicha (*Amaranthus caudatus*). *Food Chem.* **2010**, *120*, 128–133.
64. Wen, A.; Delaquis, P.; Stanich, K.; Toivonen, P. Antilisterial activity of selected phenolic acids. *Food Microbiol.* **2003**, *20*, 305–311.
65. Graf, E. Antioxidant potential of ferulic acid. *Free Radical Biol. Med.* **1992**, *13*, 435–448.
66. Kim, H.K.; Jeong, T.S.; Lee, M.K.; Park, Y.B.; Choi, M.S. Lipid-lowering efficacy of hesperetin metabolites in high-cholesterol fed rats. *Clin. Chim. Acta* **2003**, *327*, 129–137.
67. Ou, S.; Bao, H.; Lan, Z. Advances on pharmacological study of ferulic acid and its derivatives. *J. Chin. Med. Mater.* **2001**, *24*, 220–221.

68. Ou, S.; Kwok, K.C. Ferulic acid: Pharmaceutical functions, preparation and applications in foods. *J. Sci. Food Agric.* **2004**, *84*, 1261–1269.
69. Sakai, S.; Kawamata, H.; Kogure, T.; Mantani, N.; Terasawa, K.; Umatake, M.; Ochiai, H. Inhibitory effect of ferulic acid and isoferulic acid on the production of macrophage inflammatory protein-2 in response to respiratory syncytial virus infection in RAW264. 7 cells. *Mediat. Inflamm.* **1999**, *8*, 173–175.
70. Mori, H.; Kawabata, K.; Yoshimi, N.; Tanaka, T.; Murakami, T.; Okada, T.; Murai, H. Chemopreventive effects of ferulic acid on oral and rice germ on large bowel carcinogenesis. *Anticancer Res.* **1999**, *19*, 3775–3778.
71. Wang, X.; Li, X.; Chen, D. Evaluation of antioxidant activity of isoferulic acid in vitro. *Nat. Prod. Commun.* **2011**, *6*, 1285–1288.
72. Fiorito, S.; Preziuso, F.; Epifano, F.; Scotti, L.; Bucciarelli, T.; Taddeo, V.A.; Genovese, S. Novel biologically active principles from spinach, goji and quinoa. *Food Chem.* **2019**, *276*, 262–265.
73. Bais, H.P.; Walker, T.S.; Schweizer, H.P.; Vivanco, J.M. Root specific elicitation and antimicrobial activity of rosmarinic acid in hairy root cultures of *Ocimum basilicum*. *Plant Physiol. Biochem.* **2002**, *40*, 983–995.
74. Englberger, W.; Hadding, U.; Etschenberg, E.; Graf, E.; Leyck, S.; Winkelmann, J.; Parnham, M.J. Rosmarinic acid: A new inhibitor of complement C3-convertase with anti-inflammatory activity. *Int. J. Immunopharmacol.* **1988**, *10*, 729–737.
75. Cao, H.; Cheng, W.X.; Li, C.; Pan, X.L.; Xie, X.G.; Li, T.H. DFT study on the antioxidant activity of rosmarinic acid. *J. Mol. Struct.* **2005**, *719*, 177–183.
76. Furtado, M.A.; de Almeida, L.C.F.; Furtado, R.A.; Cunha, W.R.; Tavares, D.C. Antimutagenicity of rosmarinic acid in Swiss mice evaluated by the micronucleus assay. *Mutat. Res.-Genet. Toxicol. Environ. Mutagen.* **2008**, *657*, 150–154.
77. Swarup, V.; Ghosh, J.; Ghosh, S.; Saxena, A.; Basu, A. Antiviral and anti-inflammatory effects of rosmarinic acid in an experimental murine model of Japanese encephalitis. *Antimicrob. Agents Chemother.* **2007**, *51*, 3367–3370.
78. Yoon, B.H.; Jung, J.W.; Lee, J.J.; Cho, Y.W.; Jang, C.G.; Jin, C.; Ryu, J.H. Anxiolytic-like effects of sinapic acid in mice. *Life Sci.* **2007**, *81*, 234–240.
79. Karakida, F.; Ikeya, Y.; Tsunakawa, M.; Yamaguchi, T.; Ikarashi, Y.; Takeda, S.; Aburada, M. Cerebral protective and cognition-improving effects of sinapic acid in rodents. *Biol. Pharm. Bull.* **2007**, *30*, 514–519.
80. Kumar, S.; Pandey, A.K. Chemistry and biological activities of flavonoids: An overview. *Sci. World J.* **2013**, *2013*, 162750.
81. Harborne, J.B.; Williams, C.A. Advances in flavonoid research since 1992. *Phytochemistry* **2000**, *55*, 481–504.
82. Wuyts, N.; Swennen, R.; De Waele, D. Effects of plant phenylpropanoid pathway products and selected terpenoids and alkaloids on the behaviour of the plant-parasitic nematodes *Radopholus similis*, *Pratylenchus penetrans* and *Meloidogyne incognita*. *Nematology* **2006**, *8*, 89–101.
83. Kang, G.-H.; Chang, E.-J.; Choi, S.-W. Antioxidative activity of phenolic compounds in roasted safflower (*Carthamus tinctorius* L.) seeds. *J. Food Sci. Nutr.* **1999**, *4*, 221–225.
84. Lv, H.; Yu, Z.; Zheng, Y.; Wang, L.; Qin, X.; Cheng, G.; Ci, X. Isovitexin exerts anti-inflammatory and anti-oxidant activities on lipopolysaccharide-induced acute lung injury by inhibiting MAPK and NF- κ B and activating HO-1/Nrf2 pathways. *Int. J. Biol. Sci.* **2016**, *12*, 72.
85. González-Trujano, M.E.; Ventura-Martínez, R.; Chávez, M.; Díaz-Reval, I.; Pellicer, F. Spasmolytic and antinociceptive activities of ursolic acid and acacetin identified in *Agastache mexicana*. *Planta Med.* **2012**, *78*, 793–796.
86. Hsu, Y.L.; Kuo, P.L.; Liu, C.F.; Lin, C.C. Acacetin-induced cell cycle arrest and apoptosis in human non-small cell lung cancer A549 cells. *Cancer Lett.* **2004**, *212*, 53–60.
87. Hayashi, K.; Hayashi, T.; Arisawa, M.; Morita, N. Antiviral agents of plant origin. Antiherpetic activity of acacetin. *Antivir. Chem. Chemoth.* **1993**, *4*, 49–53.
88. Liu, L.Z.; Jing, Y.; Jiang, L.L.; Jiang, X.E.; Jiang, Y.; Rojanasakul, Y.; Jiang, B.H. Acacetin inhibits VEGF expression, tumor angiogenesis and growth through AKT/HIF-1 α pathway. *Biochem. Biophys. Res. Commun.* **2011**, *413*, 299–305.
89. Carballo-Villalobos, A.I.; Gonzalez-Trujano, M.E.; Lopez-Munoz, F.J. Evidence of mechanism of action of anti-inflammatory/antinociceptive activities of acacetin. *Eur. J. Pain.* **2014**, *18*, 396–405.

90. Nguyen, M.T.T.; Awale, S.; Tezuka, Y.; Shi, L.; Zaidi, S.F.H.; Ueda, J.Y.; Kadota, S. Hypouricemic effects of acacetin and 4,5-O-dicaffeoylquinic acid methyl ester on serum uric acid levels in potassium oxonate-pretreated rats. *Biol. Pharm. Bull.* **2005**, *28*, 2231–2234.
91. Shochet, G.E.; Drucker, L.; Pasmanik-Chor, M.; Pomeranz, M.; Fishman, A.; Matalon, S.T.; Lishner, M. First trimester human placental factors induce breast cancer cell autophagy. *Breast Cancer Res. Treat.* **2015**, *149*, 645–654.
92. Lee, C.Y.; Chien, Y.S.; Chiu, T.H.; Huang, W.W.; Lu, C.C.; Chiang, J.H.; Yang, J.S. Apoptosis triggered by vitexin in U937 human leukemia cells via a mitochondrial signaling pathway. *Oncol. Rep.* **2012**, *28*, 1883–1888.
93. De Oliveira, D.R.; Zamberlam, C.R.; Gaiardo, R.B.; Rêgo, G.M.; Cerutti, J.M.; Cavalheiro, A.J.; Cerutti, S.M. Flavones from *Erythrina falcata* are modulators of fear memory. *BMC Complement. Altern. Med.* **2014**, *14*, 288.
94. Soulimani, R.; Younos, C.; Jarmouni, S.; Bousta, D.; Misslin, R.; Mortier, F. Behavioural effects of *Passiflora incarnata* L. and its indole alkaloid and flavonoid derivatives and maltol in the mouse. *J. Ethnopharmacol.* **1997**, *57*, 11–20.
95. Choi, J.S.; Islam, M.N.; Ali, M.Y.; Kim, E.J.; Kim, Y.M.; Jung, H.A. Effects of C-glycosylation on anti-diabetic, anti-Alzheimer's disease and anti-inflammatory potential of apigenin. *Food Chem. Toxicol.* **2014**, *64*, 27–33.
96. Peng, X.; Zheng, Z.; Cheng, K.W.; Shan, F.; Ren, G.X.; Chen, F.; Wang, M. Inhibitory effect of mung bean extract and its constituents vitexin and isovitexin on the formation of advanced glycation endproducts. *Food Chem.* **2008**, *106*, 475–481.
97. Shibano, M.; Kakutani, K.; Taniguchi, M.; Yasuda, M.; Baba, K. Antioxidant constituents in the dayflower (*Commelina communis* L.) and their α -glucosidase-inhibitory activity. *J. Nat. Med.* **2008**, *62*, 349.
98. Li, H.M.; Hwang, S.H.; Kang, B.G.; Hong, J.S.; Lim, S.S. Inhibitory effects of *Colocasia esculenta* (L.) Schott constituents on aldose reductase. *Molecules* **2014**, *19*, 13212–13224.
99. Yoo, H.; Ku, S.-K.; Lee, T.; Bae, J.-S. Orientin inhibits HMGB1-induced inflammatory responses in HUVECs and in murine polymicrobial sepsis. *Inflammation* **2014**, *37*, 1705–1717.
100. An, F.; Yang, G.; Tian, J.; Wang, S. Antioxidant effects of the orientin and vitexin in *Trollius chinensis* Bunge in D-galactose-aged mice. *Neural Regen. Res.* **2012**, *7*, 2565.
101. Li, F.; Zong, J.; Zhang, H.; Zhang, P.; Xu, L.; Liang, K.; Qian, W. Orientin reduces myocardial infarction size via eNOS/NO signaling and thus mitigates adverse cardiac remodeling. *Front. Pharm.* **2017**, *8*, 926.
102. Lee, W.; Bae, J.S. Antithrombotic and antiplatelet activities of orientin in vitro and in vivo. *J. Funct. Foods* **2015**, *17*, 388–398.
103. Thangaraj, K.; Natesan, K.; Palani, M.; Vaiyapuri, M. Orientin, a flavanoid, mitigates 1,2-dimethylhydrazine-induced colorectal lesions in Wistar rats fed a high-fat diet. *Toxicol. Rep.* **2018**, *5*, 977–987.
104. Chen, J.; Zhong, J.; Liu, Y.; Huang, Y.; Luo, F.; Zhou, Y.; Wang, J. Purified vitexin compound 1, a new neolignan isolated compound, promotes PUMA-dependent apoptosis in colorectal cancer. *Cancer Med.* **2018**, *7*, 6158–6169.
105. Je, H.G.; Hong, S.M.; Je, H.D.; Sohn, U.D.; Choi, Y.S.; Seo, S.Y.; Park, E.S. The inhibitory effect of vitexin on the agonist-induced regulation of vascular contractility. *J. Pharm. Sci.* **2014**, *69*, 224–228.
106. Praveena, R.; Sadasivam, K.; Kumaresan, R.; Deepha, V.; Sivakumar, R. Experimental and DFT studies on the antioxidant activity of a C-glycoside from *Rhynchosia capitata*. *Spectrochim. Acta. A Mol. Biomol. Spectrosc.* **2013**, *103*, 442–452.
107. Borghi, S.M.; Carvalho, T.T.; Staurengo-Ferrari, L.; Hohmann, M.S.; Pinge-Filho, P.; Casagrande, R.; Verri, W.A., Jr. Vitexin inhibits inflammatory pain in mice by targeting TRPV1, oxidative stress, and cytokines. *J. Nat. Prod.* **2013**, *76*, 1141–1149.
108. Abbasi, E.; Nassiri-Asl, M.; Sheikhi, M.; Shafiee, M. Effects of vitexin on scopolamine-induced memory impairment in rats. *Chin. J. Physiol.* **2013**, *56*, 184–189.
109. Can, O.D.; Ozkay, U.D.; Ucel, U.I. Anti-depressant-like effect of vitexin in BALB/c mice and evidence for the involvement of monoaminergic mechanisms. *Eur. J. Pharm.* **2013**, *699*, 250–257.
110. Abbasi, E.; Nassiri-Asl, M.; Shafeei, M.; Sheikhi, M. Neuroprotective effects of vitexin, a flavonoid, on pentylenetetrazole-induced seizure in rats. *Chem. Biol. Drug Des.* **2012**, *80*, 274–278.
111. Aseervatham, G.S.B.; Suryakala, U.; Sundaram, S.; Bose, P.C.; Sivasudha, T. Expression pattern of NMDA receptors reveals antiepileptic potential of apigenin 8-C-glucoside and chlorogenic acid in pilocarpine induced epileptic mice. *Biomed. Pharm.* **2016**, *82*, 54–64.

112. Ozkay, U.D.; Can, O.D. Anti-nociceptive effect of vitexin mediated by the opioid system in mice. *Pharm. Biochem. Behav.* **2013**, *109*, 23–30.
113. Min, J.-W.; Hu, J.-J.; He, M.; Sanchez, R.M.; Huang, W.-X.; Liu, Y.-Q.; Bsoul, N.B.; Han, S.; Yin, J.; Liu, W.-H.; et al. Vitexin reduces hypoxia–ischemia neonatal brain injury by the inhibition of HIF-1 α in a rat pup model. *Neuropharmacology* **2015**, *99*, 38–50.
114. Wang, Y.; Zhen, Y.; Wu, X.; Jiang, Q.; Li, X.; Chen, Z.; Dong, L. Vitexin protects brain against ischemia/reperfusion injury via modulating mitogen-activated protein kinase and apoptosis signaling in mice. *Phytomedicine* **2015**, *22*, 379–384.
115. Brahmabhatt, S.; Brahmabhatt, R.M.; Boyages, S.C. Thyroid ultrasound is the best prevalence indicator for assessment of iodine deficiency disorders: A study in rural/tribal schoolchildren from Gujarat (Western India). *Eur. J. Endocrinol.* **2000**, *143*, 37–46.
116. Basile, A.; Giordano, S.; Lopez-Saez, J.A.; Cobiainchi, R.C. Antibacterial activity of pure flavonoids isolated from mosses. *Phytochemistry* **1999**, *52*, 1479–1482.
117. Knipping, K.; Garssen, J.; van't Land, B. An evaluation of the inhibitory effects against rotavirus infection of edible plant extracts. *Virology* **2012**, *9*, 137.
118. Arora, A.; Nair, M.G.; Strasburg, G.M. Structure-activity relationships for antioxidant activities of a series of flavonoids in a liposomal system. *Free Radical Biol. Med.* **1998**, *24*, 1355–1363.
119. Rice-evans, C.A.; Miller, N.J.; Bolwell, P.G.; Bramley, P.M.; Pridham, J.B. The relative antioxidant activities of plant-derived polyphenolic flavonoids. *Free Radical Res.* **1995**, *22*, 375–383.
120. Zhu, N.; Sheng, S.; Li, D.; LaVoie, E.J.; Karwe, M.V.; Rosen, R.T.; HO, C.T. Antioxidative flavonoid glycosides from quinoa seeds (*Chenopodium quinoa* Willd). *J. Food Lipids* **2001**, *8*, 37–44.
121. Bloor, S.J. An antimicrobial kaempferol-diacyl-rhamnoside from *Pentachondra pumila*. *Phytochemistry* **1995**, *38*, 1033–1035.
122. Martini, N.D.; Katerere, D.R.P.; Eloff, J.N. Biological activity of five antibacterial flavonoids from *Combretum erythrophyllum* (Combretaceae). *J. Ethnopharmacol.* **2004**, *93*, 207–212.
123. Treutter, D. Significance of flavonoids in plant resistance and enhancement of their biosynthesis. *Plant Biol.* **2005**, *7*, 581–591.
124. Parvez, M.M.; Tomita-Yokotani, K.; Fujii, Y.; Konishi, T.; Iwashina, T. Effects of quercetin and its seven derivatives on the growth of *Arabidopsis thaliana* and *Neurospora crassa*. *Biochem. Syst. Ecol.* **2004**, *32*, 631–635.
125. Bahrman, N.; Jay, M.; Gorenflot, R. Contribution to the chemosystematic knowledge of some species of the genus *Chenopodium*, L. *Lett. Bot.* **1985**, *2*, 107–113.
126. Saud, S.M.; Young, M.R.; Jones-Hall, Y.L.; Ileva, L.; Evbuomwan, M.O.; Wise, J.; Bobe, G. Chemopreventive activity of plant flavonoid isorhamnetin in colorectal cancer is mediated by oncogenic Src and β -catenin. *Cancer Res.* **2013**, *73*, 5473–5484.
127. Jnawali, H.N.; Jeon, D.; Jeong, M.-C.; Lee, E.; Jin, B.; Ryoo, S.; Yoo, J.; Jung, I.D.; Lee, S.J.; Park, Y.M.; et al. Antituberculosis activity of a naturally occurring flavonoid, isorhamnetin. *J. Nat. Prod.* **2016**, *79*, 961–969.
128. Torres, R.; Faini, F.; Modak, B.; Urbina, F.; Labbé, C.; Guerrero, J. Antioxidant activity of coumarins and flavonols from the resinous exudate of *Haplopappus multifolius*. *Phytochemistry* **2006**, *67*, 984–987.
129. Teng, B.S.; Lu, Y.H.; Wang, Z.T.; Tao, X.Y.; Wei, D.Z. In vitro anti-tumor activity of isorhamnetin isolated from *Hippophae rhamnoides* L. against BEL-7402 cells. *Pharm. Res.* **2006**, *54*, 186–194.
130. Li, C.; Yang, D.; Zhao, Y.; Qiu, Y.; Cao, X.; Yu, Y.; Yin, X. Inhibitory effects of isorhamnetin on the invasion of human breast carcinoma cells by downregulating the expression and activity of matrix metalloproteinase-2/9. *Nutr. Cancer* **2015**, *67*, 1191–1200.
131. Oh, H.M.; Kwon, B.M.; Baek, N.I.; Kim, S.H.; Chung, I.S.; Park, M.H.; Kim, D.K. Inhibitory activity of isorhamnetin from *Persicaria thunbergii* on farnesyl protein transferase. *Arch. Pharm. Res.* **2005**, *28*, 169–171.
132. Chirumbolo, S. Anti-inflammatory action of isorhamnetin. *Inflammation* **2014**, *37*, 1200.
133. Ku, S.K.; Kim, T.H.; Bae, J.S. Anticoagulant activities of persicarin and isorhamnetin. *Vasc. Pharm.* **2013**, *58*, 272–279.
134. Kim, S.K.; Kim, H.J.; Choi, S.E.; Park, K.H.; Choi, H.K.; Lee, M.W. Anti-oxidative and inhibitory activities on nitric oxide (NO) and prostaglandin E 2 (COX-2) production of flavonoids from seeds of *Prunus tomentosa* Thunberg. *Arch. Pharm. Res.* **2008**, *3*, 424.
135. Lee, K.M.; Lee, K.W.; Jung, S.K.; Lee, E.J.; Heo, Y.S.; Bode, A.M.; Dong, Z. Kaempferol inhibits UVB-induced COX-2 expression by suppressing Src kinase activity. *Biochem. Pharm.* **2010**, *80*, 2042–2049.

136. Rho, H.S.; Ghimeray, A.K.; Yoo, D.S.; Ahn, S.M.; Kwon, S.S.; Lee, K.H.; Cho, J.Y. Kaempferol and kaempferol rhamnosides with depigmenting and anti-inflammatory properties. *Molecules* **2011**, *16*, 3338–3344.
137. Prouillet, C.; Mazière, J.C.; Mazière, C.; Wattel, A.; Brazier, M.; Kamel, S. Stimulatory effect of naturally occurring flavonols quercetin and kaempferol on alkaline phosphatase activity in MG-63 human osteoblasts through ERK and estrogen receptor pathway. *Biochem. Pharm.* **2004**, *67*, 1307–1313.
138. De Simone, F.; Dini, A.; Pizza, C.; Saturnino, P.; Schettino, O. Two flavonol glycosides from *Chenopodium quinoa*. *Phytochemistry* **1990**, *29*, 3690–3692.
139. Hirose, Y.; Fujita, T.; Ishii, T.; Ueno, N. Antioxidative properties and flavonoid composition of *Chenopodium quinoa* seeds cultivated in Japan. *Food Chem.* **2010**, *119*, 1300–1306.
140. Chemmugil, P.; Lakshmi, P.T.V.; Annamalai, A. Exploring Morin as an anti-quorum sensing agent (anti-QSA) against resistant strains of *Staphylococcus aureus*. *Microb. Pathog.* **2019**, *127*, 304–315.
141. Qu, Y.; Wang, C.; Liu, N.; Gao, C.; Liu, F. Exhibits anti-inflammatory effects on IL-1 β -stimulated human osteoarthritis chondrocytes by activating the Nrf2 signaling pathway. *Cell. Physiol. Biochem.* **2018**, *51*, 1830–1838.
142. Ji, Y.; Jia, L.; Zhang, Y.; Xing, Y.; Wu, X.; Zhao, B.; Qiao, X. Antitumor activity of the plant extract morin in tongue squamous cell carcinoma cells. *Oncol. Rep.* **2018**, *40*, 3024–3032.
143. Yuan, W.; Ahmad, S.; Najar, A. Morin, a plant derived flavonoid, modulates the expression of peroxisome proliferator-activated receptor- γ coactivator-1 α mediated by AMPK pathway in hepatic stellate cells. *Am. J. Transl. Res.* **2017**, *9*, 5662.
144. Chen, X.; Deng, Z.; Zhang, C.; Zheng, S.; Pan, Y.; Wang, H.; Li, H. Is antioxidant activity of flavonoids mainly through the hydrogen-atom transfer mechanism? *Food Res. Int.* **2019**. [[CrossRef](#)]
145. Sithara, T.; Arun, K.B.; Syama, H.P.; Reshmitha, T.R.; Nisha, P. Morin Inhibits proliferation of SW480 colorectal cancer cells by inducing apoptosis mediated by reactive oxygen species formation and uncoupling of Warburg effect. *Front. Pharmacol.* **2017**, *8*, 640.
146. Wang, N.; Zhang, J.; Qin, M.; Yi, W.; Yu, S.; Chen, Y.; Zhang, R. Amelioration of streptozotocin-induced pancreatic β cell damage by morin: Involvement of the AMPK-FOXO3-catalase signaling pathway. *Int. J. Mol. Med.* **2018**, *41*, 1409–1418.
147. Carmona, V.; Martín-Aragon, S.; Goldberg, J.; Schubert, D.; Bermejo-Bescós, P. Several targets involved in Alzheimer's disease amyloidogenesis are affected by morin and isoquercitrin. *Nutr. Neurosci.* **2019**. [[CrossRef](#)]
148. Bhakuni, G.S.; Bedi, O.; Bariwal, J.; Kumar, P. Hepatoprotective activity of morin and its semi-synthetic derivatives against alcohol induced hepatotoxicity in rats. *Indian J. Physiol. Pharmacol.* **2017**, *61*, 175–190.
149. Fukumoto, L.R.; Mazza, G. Assessing antioxidant and prooxidant activities of phenolic compounds. *J. Agric. Food Chem.* **2000**, *48*, 3597–3604.
150. Lu, J.; Papp, L.V.; Fang, J.; Rodriguez-Nieto, S.; Zhivotovsky, B.; Holmgren, A. Inhibition of mammalian thioredoxin reductase by some flavonoids: Implications for myricetin and quercetin anticancer activity. *Cancer Res.* **2006**, *66*, 4410–4418.
151. Wang, S.J.; Tong, Y.; Lu, S.; Yang, R.; Liao, X.; Xu, Y.F.; Li, X. Anti-inflammatory activity of myricetin isolated from *Myrica rubra* Sieb. et Zucc. leaves. *Planta Med.* **2010**, *76*, 1492–1496.
152. Tong, Y.; Zhou, X.M.; Wang, S.J.; Yang, Y.; Cao, Y.L. Analgesic activity of myricetin isolated from *Myrica rubra* Sieb. et Zucc. leaves. *Arch. Pharm. Res.* **2009**, *32*, 527–533.
153. Comalada, M.; Ballester, I.; Bailon, E.; Sierra, S.; Xaus, J.; Galvez, J.; Zarzuelo, A. Inhibition of pro-inflammatory markers in primary bone marrow-derived mouse macrophages by naturally occurring flavonoids: Analysis of the structure–activity relationship. *Biochem. Pharmacol.* **2006**, *72*, 1010–1021.
154. Silva, T.B.; Costa, C.O.; Galvão, A.F.; Bomfim, L.M.; Rodrigues, A.C.; Mota, M.C.; Dantas, A.A.; Santos, T.R.; Soares, M.B.; Bezerra, D.P. Cytotoxic potential of selected medicinal plants in northeast Brazil. *BMC Complement. Altern. Med.* **2016**, *16*, 199.
155. Ng, T.B.; Liu, F.; Wang, Z.T. Antioxidative activity of natural products from plants. *Life Sci.* **2000**, *66*, 709–723.
156. Shimoyama, A.T.; Santin, J.R.; Machado, I.D.; de Silva, A.M.D.O.; De Melo, I.L.P.; Mancini-Filho, J.; Farsky, S.H. Antiulcerogenic activity of chlorogenic acid in different models of gastric ulcer. *N-S. Arch. Pharmacol.* **2013**, *386*, 5–14.
157. Al-Ashaal, H.A.; El-Sheltawy, S.T. Antioxidant capacity of hesperidin from citrus peel using electron spin resonance and cytotoxic activity against human carcinoma cell lines. *Pharm. Biol.* **2011**, *49*, 276–282.

158. Parhiz, H.; Roohbakhsh, A.; Soltani, F.; Rezaee, R.; Iranshahi, M. Antioxidant and anti-inflammatory properties of the citrus flavonoids hesperidin and hesperetin: An updated review of their molecular mechanisms and experimental models. *Phytother. Res.* **2015**, *29*, 323–331.
159. Salas, M.P.; Céliz, G.; Geronazzo, H.; Daz, M.; Resnik, S.L. Antifungal activity of natural and enzymatically-modified flavonoids isolated from citrus species. *Food Chem.* **2011**, *124*, 1411–1415.
160. Cincin, Z.B.; Unlu, M.; Kiran, B.; Bireller, E.S.; Baran, Y.; Cakmakoglu, B. Anti-proliferative, apoptotic and signal transduction effects of hesperidin in non-small cell lung cancer cells. *Cell. Oncol.* **2015**, *38*, 195–3204.
161. Mahmoud, A.M. Hesperidin protects against cyclophosphamide-induced hepatotoxicity by upregulation of PPAR γ and abrogation of oxidative stress and inflammation. *Can. J. Physiol. Pharm.* **2014**, *92*, 717–724.
162. Agrawal, Y.O.; Sharma, P.K.; Shrivastava, B.; Ojha, S.; Upadhyaya, H.M.; Arya, D.S.; Goyal, S.N. Hesperidin produces cardioprotective activity via PPAR- γ pathway in ischemic heart disease model in diabetic rats. *PLoS ONE* **2014**, *9*, e111212.
163. Lee, J.H.; Lee, S.H.; Kim, Y.S.; Jeong, C.S. Protective effects of neohesperidin and poncirin isolated from the fruits of *Poncirus trifoliata* on potential gastric disease. *Phytother. Res.* **2009**, *23*, 1748–1753.
164. Xu, F.; Zang, J.; Chen, D.; Zhang, T.; Zhan, H.; Lu, M.; Zhuge, H. Neohesperidin induces cellular apoptosis in human breast denocarcinoma MDA-MB-231 cells via activating the Bcl-2/Bax-mediated signaling pathway. *Nat. Prod. Commun.* **2012**, *7*, 1475–1478.
165. Jeon, S.M.; Bok, S.H.; Jang, M.K.; Lee, M.K.; Nam, K.T.; Park, Y.B.; Choi, M.S. Antioxidative activity of naringin and lovastatin in high cholesterol-fed rabbits. *Life Sci.* **2001**, *69*, 2855–2866.
166. Wei, M.; Yang, Z.; Li, P.; Zhang, Y.; Sse, W.C. Anti-osteoporosis activity of naringin in the retinoic acid-induced osteoporosis model. *Am. J. Chin. Med.* **2007**, *35*, 663–667.
167. Amaro, M.I.; Rocha, J.; Vila-Real, H.; Eduardo-Figueira, M.; Mota-Filipe, H.; Sepodes, B.; Ribeiro, M.H. Anti-inflammatory activity of naringin and the biosynthesised naringenin by naringinase immobilized in microstructured materials in a model of DSS-induced colitis in mice. *Food Res. Int.* **2009**, *42*, 1010–1017.
168. Huang, S.W.; Frankel, E.N. Antioxidant activity of tea catechins in different lipid systems. *J. Agric. Food Chem.* **1997**, *45*, 3033–3038.
169. Geetha, T.; Garg, A.; Chopra, K.; Kaur, I.P. Delineation of antimutagenic activity of catechin, epicatechin and green tea extract. *Mutat. Res.-Fund. Mol. Mech. Mutagen.* **2004**, *556*, 65–74.
170. Menon, L.G.; Kuttan, R.; Kuttan, G. Anti-metastatic activity of curcumin and catechin. *Cancer Lett.* **1999**, *141*, 159–165.
171. Hirasawa, M.; Takada, K. Multiple effects of green tea catechin on the antifungal activity of antimycotics against *Candida albicans*. *J. Antimicro. Chemoth.* **2004**, *53*, 225–229.
172. Saeki, K.; Hayakawa, S.; Isemura, M.; Miyase, T. Importance of a pyrogallol-type structure in catechin compounds for apoptosis-inducing activity. *Phytochemistry* **2000**, *53*, 391–394.
173. Iacopini, P.; Baldi, M.; Storchi, P.; Sebastiani, L. Catechin, epicatechin, quercetin, rutin and resveratrol in red grape: Content, in vitro antioxidant activity and interactions. *J. Food Compos. Anal.* **2008**, *21*, 589–598.
174. Kinjo, J.; Nagao, T.; Tanaka, T.; Nonaka, G.I.; Okawa, M.; Nohara, T.; Okabe, H. Activity-guided fractionation of green tea extract with antiproliferative activity against human stomach cancer cells. *Biol. Pharm. Bull.* **2002**, *2*, 1238–1240.
175. Han, Z.; Wang, G.; Yao, W.; Zhu, W. Isoflavonic phytoestrogens-new prebiotics for farm animals: A review on research in China. *Curr. Issues Intest. Microbiol.* **2006**, *7*, 53–60.
176. Lutz, M.; Martínez, A.; Martínez, E.A. Daidzein and genistein contents in seeds of quinoa (*Chenopodium quinoa* Willd.) from local ecotypes grown in arid Chile. *Ind. Crop. Prod.* **2013**, *49*, 117–121.
177. Foti, P.; Erba, D.; Riso, P.; Spadafranca, A.; Criscuoli, F.; Testolin, G. Comparison between daidzein and genistein antioxidant activity in primary and cancer lymphocytes. *Arch. Biochem. Biophys.* **2005**, *433*, 421–427.
178. Cho, K.W.; Lee, O.H.; Banz, W.J.; Moustaid-Moussa, N.; Shay, N.F.; Kim, Y.C. Daidzein and the daidzein metabolite, equol, enhance adipocyte differentiation and PPAR γ transcriptional activity. *J. Nutr. Biochem.* **2010**, *21*, 841–847.
179. Guo, J.M.; Kang, G.Z.; Xiao, B.X.; Liu, D.H.; Zhang, S. Effect of daidzein on cell growth, cell cycle, and telomerase activity of human cervical cancer in vitro. *Int. J. Gynecol. Cancer* **2004**, *14*, 882–888.
180. Picherit, C.; Dalle, M.; Neliat, G.; Lebecque, P.; Davicco, M.J.; Barlet, J.P.; Coxam, V. Genistein and daidzein modulate in vitro rat uterine contractile activity. *J. Steroid Biochem. Mol. Biol.* **2000**, *75*, 201–208.

181. Zeng, J.; Huang, Z.; Qiu, F.; Yao, X.; Ye, H. The anti-hypoxia activity of daidzein. *Chin. J. Mod. Appl. Pharm.* **2004**, *21*, 454–456.
182. Choo, M.K.; Park, E.K.; Yoon, H.K.; Kim, D.H. Antithrombotic and antiallergic activities of daidzein, a metabolite of puerarin and daidzin produced by human intestinal microflora. *Biol. Pharm. Bull.* **2002**, *25*, 1328–1332.
183. Lepri, S.R.; Luiz, R.C.; Zanelatto, L.C.; Da Silva, P.B.G.; Sartori, D.; Ribeiro, L.R.; Mantovani, M.S. Chemoprotective activity of the isoflavones, genistein and daidzein on mutagenicity induced by direct and indirect mutagens in cultured HTC cells. *Cytotechnology* **2013**, *65*, 213–222.
184. Fujioka, M.; Uehara, M.; Wu, J.; Adlercreutz, H.; Suzuki, K.; Kanazawa, K.; Ishimi, Y. Equol, a metabolite of daidzein, inhibits bone loss in ovariectomized mice. *J. Nutr.* **2004**, *134*, 2623–2627.
185. Choi, E.J.; Kim, G.H. Antiproliferative activity of daidzein and genistein may be related to ER α /c-erbB-2 expression in human breast cancer cells. *Mol. Med. Rep.* **2013**, *7*, 781–784.
186. Finking, G.; Wohlfrom, M.; Lenz, C.; Wolkenhauer, M.; Eberle, C.; Hanke, H. The phytoestrogens genistein and daidzein, and 17 beta-estradiol inhibit development of neointima in aortas from male and female rabbits in vitro after injury. *Coron. Artery Dis.* **1999**, *10*, 607–615.
187. Record, I.R.; Dreosti, I.E.; McInerney, J.K. The antioxidant activity of genistein in vitro. *J. Nutr. Biochem.* **1995**, *6*, 481–485.
188. Akiyama, T.; Ishida, J.; Nakagawa, S.; Ogawara, H.; Watanabe, S.I.; Itoh, N.; Fukami, Y. Genistein, a specific inhibitor of tyrosine-specific protein kinases. *J. Biol. Chem.* **1987**, *262*, 5592–5595.
189. Banerjee, S.; Zhang, Y.; Ali, S.; Bhuiyan, M.; Wang, Z.; Chiao, P.J.; Sarkar, F.H. Molecular evidence for increased antitumor activity of gemcitabine by genistein in vitro and in vivo using an orthotopic model of pancreatic cancer. *Cancer Res.* **2005**, *65*, 9064–9072.
190. Uckun, F.M.; Narla, R.K.; Jun, X.; Zeren, T.; Venkatachalam, T.; Waddick, K.G.; Rostostev, A.; Myers, D.E. Cytotoxic activity of epidermal growth factor-genistein against breast cancer cells. *Clin. Cancer Res.* **1998**, *4*, 901–912.
191. Büchler, P.; Reber, H.A.; Büchler, M.W.; Friess, H.; Lavey, R.S.; Hines, O.J. Antiangiogenic activity of genistein in pancreatic carcinoma cells is mediated by the inhibition of hypoxia-inducible factor-1 and the down-regulation of VEGF gene expression. *Cancer* **2004**, *10*, 201–210.
192. Farina, H.G.; Pomies, M.; Alonso, D.F.; Gomez, D.E. Antitumor and antiangiogenic activity of soy isoflavone genistein in mouse models of melanoma and breast cancer. *Oncol. Rep.* **2006**, *16*, 885–891.
193. Hong, H.; Landauer, M.R.; Foriska, M.A.; Ledney, G.D. Antibacterial activity of the soy isoflavone genistein. *J. Basic Microbiol.* **2006**, *46*, 329–335.
194. Ye, F.; Wu, J.; Dunn, T.; Yi, J.; Tong, X.; Zhang, D. Inhibition of cyclooxygenase-2 activity in head and neck cancer cells by genistein. *Cancer Lett.* **2004**, *211*, 39–46.
195. Aditya, N.P.; Shim, M.; Lee, I.; Lee, Y.; Im, M.H.; Ko, S. Curcumin and genistein coloaded nanostructured lipid carriers: In vitro digestion and antiprostata cancer activity. *J. Agric. Food Chem.* **2013**, *61*, 1878–1883.
196. Raynal, N.J.M.; Momparler, L.; Charbonneau, M.; Momparler, R.L. Antileukemic activity of genistein, a major isoflavone present in soy products. *J. Nat. Prod.* **2007**, *71*, 3–7.
197. Kim, J.S.; Nam, Y.J.; Kwon, T.W. Induction of quinone reductase activity by genistein, soybean isoflavone. *Food Sci. Biotechnol.* **1996**, *5*, 70–75.
198. Khaw, A.K.; Yong, J.W.Y.; Kalthur, G.; Hande, M.P. Genistein induces growth arrest and suppresses telomerase activity in brain tumor cells. *Gene Chromosome Canc.* **2012**, *51*, 961–974.
199. Yang, G.; Ham, I.; Choi, H.Y. Anti-inflammatory effect of prunetin via the suppression of NF- κ B pathway. *Food Chem. Toxicol.* **2013**, *58*, 124–132.
200. Xu, X.; Zhang, S.; Zhang, L.; Yan, W.; Zheng, X. The Neuroprotection of puerarin against cerebral ischemia is associated with the prevention of apoptosis in rats. *Planta Med.* **2005**, *71*, 585–591.
201. Cherdshewasart, W.; Sutjit, W. Correlation of antioxidant activity and major isoflavonoid contents of the phytoestrogen-rich *Pueraria mirifica* and *Pueraria lobata* tubers. *Phytomedicine* **2008**, *15*, 38–43.
202. Hsu, F.L.; Liu, I.M.; Kuo, D.H.; Chen, W.C.; Su, H.C.; Cheng, J.T. Antihyperglycemic effect of puerarin in streptozotocin-induced diabetic rats. *J. Nat. Prod.* **2003**, *66*, 788–792.
203. Sun, H.X.; Xie, Y.; Ye, Y.P. Advances in saponin-based adjuvants. *Vaccine* **2009**, *27*, 1787–1796.
204. Dembitsky, V.; Shkrob, I.; Hanus, L.O. Ascaridole and related peroxides from the genus *Chenopodium*. *Biomed. Pap. Med. Fac. Palacky. Olomouc. Czech. Repub.* **2008**, *152*, 209–215.

205. Yoshitomi, K.; Taniguchi, S.; Tanaka, K.; Uji, Y.; Akimitsu, K.; Gomi, K. Rice terpene synthase 24 (*PsTPS24*) encodes a jamonate-responsive monoterpene synthase that produces an antibacterial γ -terpinene against rice pathogen. *J. Plant Physiol.* **2016**, *191*, 120–126.
206. Ruiz, K.B.; Khakimov, B.; Engelsen, S.B.; Bak, S.; Biondi, S.; Jacobsen, S.-E. Quinoa seed coats as an expanding and sustainable source of bioactive compounds: An investigation of genotypic diversity in saponin profiles. *Ind. Crop. Prod.* **2017**, *104*, 156–163.
207. Mastebroek, H.D.; Limburg, H.; Gilles, T.; Marvin, H.J.P. Occurrence of saponins in leaves and seeds of quinoa (*Chenopodium quinoa* Willd). *J. Sci. Food Agric.* **2000**, *80*, 152–156.
208. Woldemichael, G.M.; Wink, M. Identification and biological activities of triterpenoid saponins from *Chenopodium quinoa*. *J. Agric. Food Chem.* **2001**, *49*, 2327–2332.
209. Sun, X.; Yang, X.; Xue, P.; Zhang, Z.; Ren, G. Improved antibacterial effects of alkali-transformed saponin from quinoa husks against halitosis-related bacteria. *BMC Complem. Altern. Med.* **2019**, *19*, 46.
210. Stuardo, M.; San Martín, R. Antifungal properties of quinoa (*Chenopodium quinoa* Willd) alkali treated saponins against *Botrytis cinerea*. *Ind. Crop. Prod.* **2008**, *27*, 296–302.
211. San Martín, R.; Ndjoko, K.; Hostettmann, K. Novel molluscicide against *Pomacea canaliculata* based on quinoa (*Chenopodium quinoa*) saponins. *Crop Prot.* **2008**, *27*, 310–319.
212. Castillo-Ruiz, M.; Cañon-Jones, H.; Schlotterbeck, T.; Lopez, M.A.; Tomas, A.; San Martin, R. Safety and efficacy of quinoa (*Chenopodium quinoa*) saponins derived molluscicide to control of *Pomacea maculata* in rice fields in the Ebro Delta, Spain. *Crop Prot.* **2018**, *111*, 42–49.
213. Joshi, R.C.; San Martin, R.; Saez-Navarrete, C.; Alarcon, J.; Sainz, J.; Antolin, M.M.; Martin, R.; Sebastian, L.S. Efficacy of quinoa (*Chenopodium quinoa*) saponins against golden apple snail (*Pomacea canaliculata*) in the Philippines under laboratory conditions. *Crop Prot.* **2008**, *27*, 553–557.
214. Madl, T.; Sterk, H.; Mittelbach, M.; Rechberger, G.N. Tandem mass spectrometric analysis of a complex triterpene saponin mixture of *Chenopodium quinoa*. *J. Am. Soc. Mass. Spectr.* **2006**, *17*, 795–806.
215. Burnouf-Radosevich, M.; Delfel, N.E.; England, R. Gas chromatography-mass spectrometry of oleanane-and ursane-type triterpenes—application to *Chenopodium quinoa* triterpenes. *Phytochemistry* **1985**, *24*, 2063–2066.
216. Mizui, F.; Kasai, R.; Ohtani, K.; Tanaka, O. Saponins from brans of quinoa, *Chenopodium quinoa* Willd. I. *Chem. Pharm. Bull.* **1988**, *36*, 1415–1418.
217. Mizui, F.; Kasai, R.; Ohtani, K.; Tanaka, O. Saponins from brans of quinoa, *Chenopodium quinoa* Willd. II. *Chem. Pharm. Bull.* **1990**, *38*, 375–377.
218. Burnouf-Radosevich, M.; Delfel, N.E. High-performance liquid chromatography of oleanane-type triterpenes. *J. Chromatogr. A* **1984**, *292*, 403–409.
219. Quispe-Fuentes, I.; Vega-Galvez, A.; Miranda, M.; Lemus-Mondaca, R.; Lozano, M.; Ah-Hen, K. A kinetic approach to saponin extraction during washing of quinoa (*Chenopodium quinoa* Willd.) seeds. *J. Food Process Eng.* **2013**, *36*, 202–210.
220. Horiuchi, K.; Shiota, S.; Hatano, T.; Yoshida, T.; Kuroda, T.; Tsuchiya, T. Antimicrobial activity of oleanolic acid from *Salvia officinalis* and related compounds on vancomycin-resistant enterococci (VRE). *Biol. Pharm. Bull.* **2007**, *30*, 1147–1149.
221. Wolska, K.I.; Grudniak, A.M.; Fiecek, B.; Kraczkiewicz-Dowjat, A.; Kurek, A. Antibacterial activity of oleanolic and ursolic acids and their derivatives. *Cent. Eur. J. Biol.* **2010**, *5*, 543–553.
222. Kashiwada, Y.; Wang, H.K.; Nagao, T.; Kitanaka, S.; Yasuda, I.; Fujioka, T.; Ikeshiro, Y. Anti-AIDS agents. 30. Anti-HIV activity of oleanolic acid, pomolic acid, and structurally related triterpenoids. *J. Nat. Prod.* **1998**, *61*, 1090–1095.
223. Singh, G.B.; Singh, S.; Bani, S.; Gupta, B.D.; Banerjee, S.K. Anti-inflammatory activity of oleanolic acid in rats and mice. *J. Pharm. Pharmacol.* **1992**, *44*, 456–458.
224. Ghosh, D.; Thejomoorthy, P. Anti-inflammatory and analgesic activities of oleanolic acid 3- β -glucoside (RDG-1) from *Randia dumetorum* (Rubiaceae). *Indian J. Pharmacol.* **1983**, *15*, 331.
225. Wang, X.; Ye, X.L.; Liu, R.; Chen, H.L.; Bai, H.; Liang, X.; Hai, C.X. Antioxidant activities of oleanolic acid in vitro: Possible role of Nrf2 and MAP kinases. *Chem.-Biol. Interact.* **2010**, *184*, 328–337.
226. Rajasekaran, M.; Bapna, J.S.; Lakshmanan, S.; Nair, A.R.; Veliath, A.J.; Panchanadam, M. Antifertility effect in male rats of oleanolic acid, a triterpene from *Eugenia jambolana* flowers. *J. Ethnopharmacol.* **1988**, *24*, 115–121.
227. Dzubak, P.; Hajduch, M.; Vydra, D.; Hustova, A.; Kvasnica, M.; Biedermann, D.; Sarek, J. Pharmacological activities of natural triterpenoids and their therapeutic implications. *Nat. Prod. Rep.* **2006**, *23*, 394–411.

228. Petronelli, A.; Pannitteri, G.; Testa, U. Triterpenoids as new promising anticancer drugs. *Anti-Cancer Drug*. **2009**, *20*, 880–892.
229. Zhu, Y.Y.; Huang, H.Y.; Wu, Y.L. Anticancer and apoptotic activities of oleanolic acid are mediated through cell cycle arrest and disruption of mitochondrial membrane potential in HepG2 human hepatocellular carcinoma cells. *Mol. Med. Rep.* **2015**, *12*, 5012–5018.
230. Yoshikawa, M.; Matsuda, H. Antidiabetogenic activity of oleanolic acid glycosides from medicinal foodstuffs. *BioFactors* **2000**, *13*, 231–237.
231. Park, S.H.; Oh, S.R.; Jung, K.Y.; Ahn, K.S.; Kim, J.G.; Lee, J.J.; Lee, H.K. Anticomplement activities of oleanolic acid monodesmosides and bisdesmosides isolated from *Tiarella polyphylla*. *Arch. Pharm. Res.* **1999**, *22*, 428–431.
232. Facino, R.M.; Carini, M.; Stefani, R.; Aldini, G.; Saibene, L. Anti-elastase and anti-hyaluronidase activities of saponins and sapogenins from *Hedera helix*, *Aesculus hippocastanum*, and *Ruscus aculeatus*: Factors contributing to their efficacy in the treatment of venous insufficiency. *Archiv. Der. Pharm.* **1995**, *328*, 720–724.
233. Ruiz, W.A.; Farfan, J.A. Determination of oleanolic acid in quinoa by gas-liquid chromatography (*Chenopodium quinoa*, Willd cv Kancolla). *Bol. Soc. Quim. Peru.* **1979**, *45*, 266–276.
234. Lozano, M.; Gonzales, E.; Flores, Y.; Almanza, G.R. Effect in acute inflammation of sapogenin extract and isolated sapogenins from quinoa waste (*Chenopodium quinoa* Willd). *Rev. Boliv. Quim.* **2013**, *30*, 115–121.
235. Dini, I.; Schettino, O.; Simioli, T.; Dini, A. Studies on the constituents of *Chenopodium quinoa* seeds: Isolation and characterization of new triterpene saponins. *J. Agric. Food Chem.* **2001**, *49*, 741–746.
236. Dini, I.; Tenore, G.C.; Dini, A. Oleanane saponins in “Kancolla”, a sweet variety of *Chenopodium quinoa*. *J. Nat. Prod.* **2002**, *65*, 1023–1026.
237. Ma, W.W.; Heinsteinst, P.F.; Mclaughlin, J.L. Additional toxic, bitter saponins from the seeds of *Chenopodium quinoa*. *J. Nat. Prod.* **1989**, *52*, 1132–1135.
238. Zhu, N.; Sheng, S.; Sang, S.; Jhoo, J.-W.; Bai, N.; Karwe, M.V.; Rosen, R.T.; Ho, C.-T. Triterpene saponins from debittered quinoa (*Chenopodium quinoa*) seeds. *J. Agric. Food Chem.* **2002**, *50*, 865–867.
239. Chauhan, G.S.; Eskin, N.A.M.; Tkachuk, R. Nutrients and antinutrients in quinoa seed. *Cereal Chem* **1992**, *69*, 85–88.
240. Hostettmann, K. Saponins with molluscicidal activity from *Hedera helix* L. *Helv. Chim. Acta* **1980**, *63*, 606–609.
241. Barthomeuf, C.; Debiton, E.; Mshvildadze, V.; Kemertelidze, E.; Balansard, G. In vitro activity of hederacolchisid A₁ compared with other saponins from *Hedera colchica* against proliferation of human carcinoma and melanoma cells. *Planta Med.* **2002**, *68*, 672–675.
242. Favel, A.; Steinmetz, M.D.; Regli, P.; Vidal-Ollivier, E.; Elias, R.; Balansard, G. In vitro antifungal activity of triterpenoid saponins. *Planta Med.* **1994**, *60*, 50–53.
243. Majester-Savornin, B.; Elias, R.; Diaz-Lanza, A.M.; Balansard, G.; Gasquet, M.; Delmas, F. Saponins of the ivy plant, *Hedera helix*, and their leishmanicidal activity. *Planta Med.* **1991**, *57*, 260–262.
244. Lee, S.J.; Shin, E.J.; Son, K.H.; Chang, H.W.; Kang, S.S.; Kim, H.P. Anti-inflammatory activity of the major constituents of *Lonicera japonica*. *Arch. Pharm. Res.* **1995**, *18*, 133.
245. Rodríguez-Hernández, D.; Demuner, A.J.; Barbosa, L.C.; Csuk, R.; Heller, L. Hederagenin as a triterpene template for the development of new antitumor compounds. *Eur. J. Med. Chem.* **2015**, *105*, 57–62.
246. Khalil, A.H.; El-Adawy, T.A. Isolation, identification and toxicity of saponin from different legumes. *Food Chem.* **1994**, *50*, 197–201.
247. Liu, B.-X.-Z.; Zhou, J.-Y.; Li, Y.; Zou, X.; Wu, J.; Gu, J.-F.; Yuan, J.-R.; Zhao, B.-J.; Feng, L.; Jia, X.-B.; et al. Hederagenin from the leaves of ivy (*Hedera helix* L.) induces apoptosis in human LoVo colon cells through the mitochondrial pathway. *BMC Complement. Altern. Med.* **2014**, *14*, 412.
248. Lee, K.-T.; Sohn, I.-C.; Park, H.-J.; Kim, D.-W.; Jung, G.-O.; Park, K.-Y. Essential moiety for antimutagenic and cytotoxic activity of hederagenin monodesmosides and bisdesmosides isolated from the stem bark of *Kalopanax pictus*. *Planta Med.* **2000**, *66*, 329–332.
249. Park, H.J.; Kwon, S.H.; Lee, J.H.; Lee, K.H.; Miyamoto, K.I.; Lee, K.T. Kalopanaxsaponin A is a basic saponin structure for the anti-tumor activity of hederagenin monodesmosides. *Planta Med.* **2001**, *67*, 118–121.
250. Voutquenne, L.; Lavaud, C.; Massiot, G.; Men-Olivier, L.L. Structure-activity relationships of haemolytic saponins. *Pharm. Biol.* **2002**, *40*, 253–262.
251. Houghton, P.; Patel, N.; Jurzysta, M.; Biely, Z.; Cheung, C. Antidermatophyte activity of medicago extracts and contained saponins and their structure–activity relationships. *Phytother. Res.* **2006**, *20*, 1061–1066.

252. He, W.; Van Puyvelde, L.; Maes, L.; Bosselaers, J.; De Kimpe, N. Antitrichomonas in vitro activity of *Cussonia holstii* Engl. *Nat. Prod. Res.* **2003**, *17*, 127–133.
253. Gopalsamy, N.; Gueho, J.; Julien, H.R.; Owadally, A.W.; Hostettmann, K. Molluscicidal saponins of *Polyscias dichroostachya*. *Phytochemistry* **1990**, *29*, 793–795.
254. Oh, S.R.; Jung, K.Y.; Son, K.H.; Park, S.H.; Ahn, K.S.; Lee, H.K. In vitro anticomplementary activity of hederagenin saponins isolated from roots of *Dipsacus asper*. *Arch. Pharm. Res.* **1999**, *22*, 317–319.
255. Jung, H.-J.; Lee, C.O.; Lee, K.-T.; Choi, J.; Park, H.-J. Structure–activity relationship of oleanane disaccharides isolated from *Akebia quinata* versus cytotoxicity against cancer cells and NO inhibition. *Biol. Pharm. Bull.* **2004**, *27*, 744–747.
256. Dini, I.; Tenore, G.C.; Schettino, O.; Dini, A. New oleanane saponins in *Chenopodium quinoa*. *J. Agric. Food Chem.* **2001**, *49*, 3976–3981.
257. Meyer, B.N.; Heinstein, P.F.; Burnoufradosevich, M.; Delfel, N.E.; Mclaughlin, J.L. Bioactivity-directed isolation and characterization of quinoside A: One of the toxic/bitter principles of quinoa seeds (*Chenopodium quinoa* Willd.). *J. Agric. Food Chem.* **1990**, *38*, 205–208.
258. Montoya, G.; Gutierrez, G.; D’vries, R.; Ellena, J.; Panay, A.J. Spergulagenic acid A: Isolation and single crystal structure elucidation. *J. Mol. Struct.* **2018**, *1173*, 937–941.
259. Lazo-Vélez, M.A.; Guajardo-Flores, D.; Mata-Ramírez, D.; Gutiérrez-Urbe, J.A.; Serna-Saldivar, S.O. Characterization and quantitation of triterpenoid saponins in raw and sprouted *Chenopodium berlandieri* spp. (Huauzontle) grains subjected to germination with or without selenium stress conditions. *J. Food Sci.* **2016**, *81*, C19–C26.
260. Vincken, J.P.; Heng, L.; de Groot, A.; Gruppen, H. Saponins, classification and occurrence in the plant kingdom. *Phytochemistry* **2007**, *68*, 275–297.
261. Fanali, C.; Beccaria, M.; Salivo, S.; Tranchida, P.; Tripodo, G.; Farnetti, S.; Dugo, L.; Dugo, P.; Mondello, L. Non-polar lipids characterization of quinoa (*Chenopodium quinoa*) seed by comprehensive two-dimensional gas chromatography with flame ionization/mass spectrometry detection and non-aqueous reversed-phase liquid chromatography with atmospheric pressure chemical ionization mass spectrometry detection. *J. Sep. Sci.* **2015**, *38*, 3151–3160.
262. Lee, I.S.; Oh, S.R.; Jung, K.Y.; Kim, D.S.; Kim, J.H.; Lee, H.K. Anticomplementary activity and complete ¹³C NMR assignment of citrostadienol from *Schizandra chinensis*. *Int. J. Pharmacogn.* **1997**, *35*, 358–363.
263. Saeed, M.A.; Sabir, A.W. Antibacterial activity of *Caesalpinia bonducella* seeds. *Fitoterapia* **2001**, *72*, 807–809.
264. Giacomán-Martínez, A.; Alarcón-Aguilar, F.J.; Zamilpa, A.; Hidalgo-Figueroa, S.; Navarrete-Vazquez, G.; García-Macedo, R.; Almanza-Perez, J.C. Triterpenoids from *Hibiscus sabdariffa* L. with PPAR δ/γ dual agonist action: In vivo, in vitro and *in silico* studies. *Planta Med.* **2019**, *85*, 412–423.
265. Cardoso, B.K.; de Oliveira, H.L.M.; Melo, U.Z.; Fernandez, C.M.M.; Campo, C.F.D.A.A.; Goncalves, J.E.; Gazim, Z.C. Antioxidant activity of α - and β -amyrin isolated from *Myrcianthes pungens* leaves. *Nat. Prod. Res.* **2019**, *33*. [[CrossRef](#)]
266. Chen, D.; Xu, F.; Zhang, P.; Deng, J.; Sun, H.; Wen, X.; Liu, J. Practical synthesis of α -amyrin, β -amyrin, and lupeol: The potential natural inhibitors of human oxidosqualene cyclase. *Arch. Pharm.* **2017**, *350*, 1700178.
267. Kannan, S.; Vijayakumar, B.; Sureshkumar, C.; Mohankumar, R.; Narasimhan, S. Insect antifeedant and growth regulating activities of β -amyrin from *Sarcostemma acidum*. *Asian J. Chem.* **2013**, *25*, 1167–1168.
268. Mhalla, D.; Ben Farhat-Touzri, D.; Tounsi, S.; Trigui, M. Combinational effect of *Rumex tingitanus* (Polygonaceae) hexane extract and *Bacillus thuringiensis* δ -endotoxin against *Spodoptera littoralis* (Lepidoptera: Noctuidae). *BioMed. Res. Int.* **2018**, *2018*, 3895834.
269. Kemboi, D. Phytochemistry and antimicrobial activity of extracts from medicinal plant *Olea africana* and *Olea europea*. *Int. J. Biochem. Res. Rev.* **2016**, *12*, 25863.
270. Zhang, J.; Yamada, S.; Ogihara, E.; Kurita, M.; Banno, N.; Qu, W.; Akihisa, T. Biological activities of triterpenoids and phenolic compounds from *Myrica cerifera* bark. *Chem. Biodivers.* **2016**, *13*, 1601–1609.
271. Ntchapda, F.; Talla, E.; Sakava, P.; Tanzi, F.; Fohouo, F.N.T.; Tanyi, J.M.; Dimo, T. Nitric oxide-dependent vasodilation and Ca²⁺ signalling induced by erythrodiol in rat aorta. *Asian Pac. J. Trop. Dis.* **2015**, *5*, S214–S223.
272. Juan, M.E.; Wenzel, U.; Daniel, H.; Planas, J.M. Erythrodiol, a natural triterpenoid from olives, has antiproliferative and apoptotic activity in HT-29 human adenocarcinoma cells. *Mol. Nutr. Food. Res.* **2008**, *52*, 595–599.

273. Zheng, Q.; Li, P.; Jin, F.; Yao, C.; Zhang, G.; Zang, T.; Ai, X. Ursolic acid induces ER stress response to activate ASK1-JNK signaling and induce apoptosis in human bladder cancer T24 cells. *Cell. Signal.* **2013**, *25*, 206–213.
274. Wang, C.M.; Tsai, S.J.; Jhan, Y.L.; Yeh, K.L.; Chou, C.H. Anti-proliferative activity of triterpenoids and sterols isolated from *Alstonia scholaris* against non-small-cell lung carcinoma cells. *Molecules* **2017**, *22*, 2119.
275. Tang, Y.; Li, X.; Chen, P.X.; Zhang, B.; Hernandez, M.; Zhang, H.; Marccone, M.F.; Liu, R.; Tsao, R. Characterisation of fatty acid, carotenoid, tocopherol/tocotrienol compositions and antioxidant activities in seeds of three *Chenopodium quinoa* Willd. genotypes. *Food Chem.* **2015**, *174*, 502–508.
276. Alvarez-Jubete, L.; Hulse, M.; Hansen, A.; Arendt, E.K.; Gallagher, E. Impact of baking on vitamin E content of pseudocereals amaranth, quinoa, and buckwheat. *Cereal Chem.* **2009**, *86*, 511–515.
277. Ju, J.; Picinich, S.C.; Yang, Z.; Zhao, Y.; Suh, N.; Kong, A.N.; Yang, C.S. Cancer-preventive activities of tocopherols and tocotrienols. *Carcinogenesis* **2009**, *31*, 533–542.
278. Gil-Chávez, G.J.; Villa, J.A.; Ayala-Zavala, J.F.; Heredia, J.B.; Sepulveda, D.; Yahia, E.M.; González-Aguilar, G.A. Technologies for extraction and production of bioactive compounds to be used as nutraceuticals and food ingredients: An overview. *Compr. Rev. Food Sci. Food Saf.* **2013**, *12*, 5–23.
279. Sen, C.K.; Khanna, S.; Roy, S. Tocotrienols in health and disease: The other half of the natural vitamin E family. *Mol. Asp. Med.* **2007**, *28*, 692–728.
280. Zingg, J.-M. Vitamin E: An overview of major research directions. *Mol. Asp. Med.* **2007**, *28*, 400–422.
281. Pereira, E.; Encina-Zelada, C.; Barros, L.; Gonzales-Barron, U.; Cadavez, V.; Ferreira, I.C. Chemical and nutritional characterization of *Chenopodium quinoa* Willd (quinoa) grains: A good alternative to nutritious food. *Food Chem.* **2019**, *280*, 110–114.
282. Ahsan, H.; Ahad, A.; Iqbal, J.; Siddiqui, W.A. Pharmacological potential of tocotrienols: A review. *Nutr. Metab.* **2014**, *11*, 52.
283. Koziol, M.J. Chemical composition and nutritional evaluation of quinoa (*Chenopodium quinoa* Willd.). *J. Food Compos. Anal.* **1992**, *5*, 35–68.
284. Sookwong, P.; Murata, K.; Nakagawa, K.; Shibata, A.; Kimura, T.; Yamaguchi, M.; Kojima, Y.; Miyazawa, T. Cross-fertilization for enhancing tocotrienol biosynthesis in rice plants and QTL analysis of their F2 progenies. *J. Agric. Food Chem.* **2009**, *57*, 4620–4625.
285. Ahamed, N.T.; Singhal, R.S.; Kulkarni, P.R.; Pal, M. A lesser-known grain, *Chenopodium quinoa*: Review of the chemical composition of its edible parts. *Food Nutr. Bull.* **1998**, *19*, 61–70.
286. Foucault, A.S.; Mathé, V.; Lafont, R.; Even, P.; Dioh, W.; Veillet, S.; Tomé, D.; Huneau, J.F.; Hermier, D.; Quignard-Boulangé, A. Quinoa extract enriched in 20-hydroxyecdysone protects mice from diet-induced obesity and modulates adipokines expression. *Obesity* **2012**, *20*, 270–277.
287. Nsimba, R.Y.; Kikuzaki, H.; Konishi, Y. Ecdysteroids act as inhibitors of calf skin collagenase and oxidative stress. *J. Biochem. Mol. Toxic.* **2008**, *22*, 240–250.
288. Dini, A.; Rastrelli, L.; Saturnino, P.; Schettino, O. A compositional study of *Chenopodium quinoa* seeds. *Nahrung* **1992**, *36*, 400–404.
289. Zhu, N.; Kikuzaki, H.; Vastano, B.C.; Nakatani, N.; Karwe, M.V.; Rosen, R.T.; Ho, C.T. Ecdysteroids of quinoa seeds (*Chenopodium quinoa* Willd.). *J. Agric. Food Chem.* **2001**, *49*, 2576–2578.
290. Xu, D.; Ali, S.; Huang, Z. Insecticidal activity influence of 20-hydroxyecdysone on the pathogenicity of *Isaria fumosorosea* against *Plutella xylostella*. *Biol. Control* **2011**, *56*, 239–244.
291. Choi, J.M.; Lee, E.O.; Lee, H.J.; Kim, K.H.; Ahn, K.S.; Shim, B.S.; Kim, N.I.; Song, M.C.; Baek, N.I.; Kim, S.H. Identification of campesterol from *Chrysanthemum coronarium* L. and its antiangiogenic activities. *Phytother. Res.* **2007**, *21*, 954–959.
292. Villacrés, E.; Pástor, G.; Quelal, M.B.; Zambrano, I.; Morales, S.H. Effect of processing on the content of fatty acids, tocopherols and sterols in the oils of quinoa (*Chenopodium quinoa* Willd), lupine (*Lupinus mutabilis* Sweet), amaranth (*Amaranthus caudatus* L.) and sangorache (*Amaranthus quitensis* L.). *Glob. Adv. Res. J. Food Sci. Technol.* **2013**, *2*, 44–53.
293. Dini, I.; Tenore, G.C.; Dini, A. Nutritional and antinutritional composition of Kancolla seeds: An interesting and underexploited andine food plant. *Food Chem.* **2005**, *92*, 125–132.
294. Prieto, J.M.; Recio, M.C.; Giner, R.M. Anti-inflammatory activity of β -sitosterol in a model of oxazolone-induced contact-delayed-type hypersensitivity. *Bol. Lat. Am. Caribb. Bull. Med. Plants* **2006**, *5*, 57–62.
295. Vivancos, M.; Moreno, J.J. β -Sitosterol modulates antioxidant enzyme response in RAW 264.7 macrophages. *Free Radical Bio. Med.* **2005**, *39*, 91–97.

296. Radika, M.K.; Viswanathan, P.; Anuradha, C.V. Nitric oxide mediates the insulin sensitizing effects of β -sitosterol in high fat diet-fed rats. *Nitric Oxide Biol. Chem.* **2013**, *32*, 43–53.
297. Garcia, M.D.; Saenz, M.T.; Gomez, M.A.; Fernandez, M.A. Topical antiinflammatory activity of phytosterols isolated from *Eryngium foetidum* on chronic and acute inflammation models. *Phytother. Res.* **1999**, *13*, 78–80.
298. Ghosh, T.; Maity, T.K.; Singh, J. Evaluation of antitumor activity of stigmasterol, a constituent isolated from *Bacopa monnieri* Linn aerial parts against Ehrlich Ascites Carcinoma in mice. *Orient. Pharm. Exp. Med.* **2011**, *11*, 41–49.
299. Mbambo, B.; Odhav, B.; Mohanlall, V. Antifungal activity of stigmasterol, sitosterol and ergosterol from *Bulbine natalensis* Baker. (Asphodelaceae). *J. Med. Plants Res.* **2012**, *6*, 5135–5141.
300. Batta, A.K.; Xu, G.; Honda, A.; Miyazaki, T.; Salen, G. Stigmasterol reduces plasma cholesterol levels and inhibits hepatic synthesis and intestinal absorption in the rat. *Metabolism* **2006**, *55*, 292–299.
301. Huang, J.G.; Zhou, L.J.; Xu, H.H.; Li, W.O. Insecticidal and cytotoxic activities of extracts of *Cacalia tangutica* and its two active ingredients against *Musca domestica* and *Aedes albopictus*. *J. Econ. Entomol.* **2009**, *102*, 1444–1447.
302. Chai, J.W.; Kuppasamy, U.R.; Kanthimathi, M.S. Beta-sitosterol induces apoptosis in MCF-7 cells. *Malay. J. Biochem. Mol. Biol.* **2008**, *16*, 28–30.
303. Saeidnia, S.; Manayi, A.; Gohari, A.R.; Abdollahi, M. The story of beta-sitosterol—a review. *Eur. J. Med. Plants* **2014**, *4*, 590.
304. Sugano, M.; Morioka, H.; Ikeda, I. A comparison of hypocholesterolemic activity of β -sitosterol and β -sitostanol in rats. *J. Nutr.* **1977**, *107*, 2011–2019.
305. Moon, E.J.; Lee, Y.M.; Lee, O.H.; Lee, M.J.; Lee, S.K.; Chung, M.H.; Kim, K.W. A novel angiogenic factor derived from *Aloe vera* gel: β -sitosterol, a plant sterol. *Angiogenesis* **1999**, *3*, 117–123.
306. Paniagua-Pérez, R.; Madrigal-Bujaidar, E.; Reyes-Cadena, S.; Molina-Jasso, D.; Gallaga, J.P.; Silva-Miranda, A.; Chamorro, G. Genotoxic and cytotoxic studies of beta-sitosterol and pteropodine in mouse. *Biomed. Res. Int.* **2005**, *2005*, 242–247.
307. Villasenor, I.M.; Angelada, J.; Canlas, A.P.; Echegoyen, D. Bioactivity studies on β -sitosterol and its glucoside. *Phytother. Res.* **2002**, *16*, 417–421.
308. Bouic, P.J.D.; Etsebeth, S.; Liebenberg, R.W.; Albrecht, C.F.; Pegel, K.; Van Jaarsveld, P.P. Beta-sitosterol and beta-sitosterol glucoside stimulate human peripheral blood lymphocyte proliferation: Implications for their use as an immunomodulatory vitamin combination. *Int. J. Immunopharmacol.* **1996**, *18*, 693–700.
309. Shi, C.; Wu, F.; Zhu, X.; Xu, J. Incorporation of β -sitosterol into the membrane increases resistance to oxidative stress and lipid peroxidation via estrogen receptor-mediated PI3K/GSK3 β signaling. *BBA Gen. Subj.* **2013**, *1830*, 2538–2544.
310. Gabay, O.; Sanchez, C.; Salvat, C.; Chevy, F.; Breton, M.; Nourissat, G.; Wolf, C.; Jacques, C.; Berenbaum, F. Stigmasterol: A phytosterol with potential anti-osteoarthritic properties. *Osteoarthr. Cartil.* **2010**, *18*, 106–116.
311. Imamura, T.; Takagi, H.; Miyazato, A.; Ohki, S.; Mizukoshi, H.; Mori, M. Isolation and characterization of the betalain biosynthesis gene involved in hypocotyl pigmentation of the allotetraploid *Chenopodium quinoa*. *Biochem. Biophys. Res. Commun.* **2018**, *496*, 280–286.
312. Kobayashi, N.; Schmidt, J.; Wray, V.; Schliemann, W. Formation and occurrence of dopamine-derived betacyanins. *Phytochemistry* **2001**, *56*, 429–436.
313. Dini, I.; Tenore, G.C.; Trimarco, E.; Dini, A. Two novel betaine derivatives from *Kancolla* seeds (Chenopodiaceae). *Food Chem.* **2006**, *98*, 209–213.
314. Tramontano, W.A.; Jouve, D. Trigonelline accumulation in salt-stressed legumes and the role of other osmoregulators as cell cycle control agents. *Phytochemistry* **1997**, *44*, 1037–1040.
315. Jones, G.P.; Paleg, L.G. In vitro thermal and salt stability of pyruvate kinase are increased by proline analogues and trigonelline. *Funct. Plant. Biol.* **1991**, *18*, 279–286.
316. Escribano, J.; Cabanes, J.; Jimenez-Atienzar, M.; Ibañez-Tremolada, M.; Gomez-Pando, L.R.; García-Carmona, F.; Gandía-Herrero, F. Characterization of betalains, saponins and antioxidant power in differently colored quinoa (*Chenopodium quinoa*) varieties. *Food Chem.* **2017**, *234*, 285–294.
317. Esatbeyoglu, T.; Wagner, A.E.; Motafakkerzad, R.; Nakajima, Y.; Matsugo, S.; Rimbach, G. Free radical scavenging and antioxidant activity of betanin: Electron spin resonance spectroscopy studies and studies in cultured cells. *Food Chem. Toxicol.* **2014**, *73*, 119–126.

318. Cai, Y.; Sun, M.; Corke, H. Antioxidant activity of betalains from plants of the Amaranthaceae. *J. Agric. Food Chem.* **2003**, *51*, 2288–2294.
319. Hirakawa, N.; Okauchi, R.; Miura, Y.; Yagasaki, K. Anti-invasive activity of niacin and trigonelline against cancer cells. *Biosci. Biotech. Biochem.* **2005**, *69*, 653–658.
320. Shah, S.N.; Bodhankar, S.L.; Bhonde, R.; Mohan, V. Hypoglycemic activity of the combination of active ingredients isolated from *Trigonella foenumgraecum* in alloxan induced diabetic mice. *Pharmacologyonline* **2006**, *1*, 65–82.
321. Letelier, M.E.; Rodríguez-Rojas, C.; Sánchez-Jofré, S.; Aracena-Parks, P. Surfactant and antioxidant properties of an extract from *Chenopodium quinoa* Willd seed coats. *J. Cereal Sci.* **2011**, *53*, 239–243.
322. Johnson, I.T.; Gee, J.M.; Price, K.; Curl, C.; Fenwick, G.R. Influence of saponins on gut permeability and active nutrient transport in vitro. *J. Nutr.* **1986**, *116*, 2270–2277.
323. Gee, J.M.; Price, K.R.; Ridout, C.L.; Wortley, G.M.; Hurrell, R.F.; Johnson, I.T. Saponins of quinoa (*Chenopodium quinoa*): Effects of processing on their abundance in quinoa products and their biological effects on intestinal mucosal tissue. *J. Sci. Food Agric.* **1993**, *63*, 201–209.
324. Sharma, V.; Chandra, S.; Dwivedi, P.; Parturkar, M. Quinoa (*Chenopodium quinoa* Willd.): A nutritional healthy grain. *Int. J. Adv. Res.* **2015**, *3*, 725–736.
325. Risi, J.C. The *Chenopodium* grains of the Andes: Inca crops for modern agriculture. *Adv. Appl. Biol.* **1984**, *10*, 145–216.
326. Jiang, X.; Hansen, H.C.B.; Strobel, B.W.; Cedergreen, N. What is the aquatic toxicity of saponin-rich plant extracts used as biopesticides? *Environ. Pollut.* **2018**, *236*, 416–424.
327. Juneja, V.K.; Dwivedi, H.P.; Yan, X. Novel natural food antimicrobials. *Annu. Rev. Food Sci. Technol.* **2012**, *3*, 381–403.
328. Fiallos-Jurado, J.; Pollier, J.; Moses, T.; Arendt, P.; Barriga-Medina, N.; Morillo, E.; Arahana, V.; de Lourdes Torres, M.; Goossens, A.; Leon-Reyes, A. Saponin determination, expression analysis and functional characterization of saponin biosynthetic genes in *Chenopodium quinoa* leaves. *Plant Sci.* **2016**, *250*, 188–197.
329. Jarvis, D.E.; Ho, Y.S.; Lightfoot, D.J.; Schmockel, S.M.; Li, B.; Borm, T.J.A.; Ohyanagi, H.; Mineta, K.; Michell, C.T.; Saber, N.; et al. The genome of *Chenopodium quinoa*. *Nature* **2017**, *542*, 307–312.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).