## SUPPLEMENTARY MATERIAL FOR

# Biofilm Inhibitory Abscisic Acid Derivatives from the Plant-Associated Dothideomycete Fungus, *Roussoella* sp.

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- S1. Morphological features and molecular phylogenetic
- S1.1 Identification of Roussoella sp. MFLUCC 17-2059



**Figure S1.1** *Roussoella sp.* MFLUCC 17-2059 **a** Appearance of ascomata on host surface **b-c** Asci **d** Ascospores e-f Culture characters on YMG agar. **Scale bars:**  $a = 500 \ \mu m$ ,  $b-c = 50 \ \mu m$ ,  $d = 10 \ \mu m$ .

Material examined: Dried branched *Clematis subumbellata* (Ranunculaceae), Thailand, living culture = MFLUCC 17-2059.

#### S1.2 Phylogenetic study

The aligned ITS and TEF-1 $\alpha$  sequences were concatenated to generate multigenes alignment phylogenetic tree. Maximum Likelihood were constructed by using CIPRES webportal online tools (Miller et al. 2010), including 1,000 bootstrap replicates. According to the data set combining the morphological features and phylogenetic analysis, the fungus was identified as *Roussoellaceae* member.







**S2.** <sup>1</sup>H NMR spectrum (MeOH- $d_4$ , 700 MHz) of roussoellenic acid (1).



**S3.** <sup>13</sup>C NMR spectrum (MeOH- $d_4$ , 175 MHz) of roussoellenic acid (1).



**S5.** COSY spectrum of I roussoellenic acid (1).





\* NOE correlations are indicated by blue off-diagonal peaks.



**S8.** LC-DAD-HR-ESIMS spectrum of roussoellenic acid (1).



**S9.** <sup>1</sup>H NMR spectrum (CDCl<sub>3</sub>, 700 MHz) of pestabacillin B (**2**).



**S10.** <sup>13</sup>C NMR spectrum (CDCl<sub>3</sub>, 175 MHz) of pestabacillin B (2).

Liu, S.; Dai, H.; Heering, C.; Jania, C.; Lin, W.; Liu, Z.; Proksch, P. Tetrahedron Lett. 2017, 58, 257-261.



**S11.** <sup>1</sup>H NMR spectrum (MeOH-*d*<sub>4</sub>, 700 MHz) of *cyclo*(*S*-Pro-*S*-IIe) (**3**).



**S12.** <sup>13</sup>C NMR spectrum (MeOH-*d*<sub>4</sub>, 175 MHz) of *cyclo*(*S*-Pro-*S*-Ile) (**3**).

Pedras, M.; Soledade C.; Yu, Y.; Liu, J.; Tandron-Moya, Y.A. *Zeitschrift fuer Naturforschung, C: J. Biosci.* **2005**, *60*, 717–722.

## S13. Antimicrobial assay results of compounds 1-3.

Strain	Compound 1	Compound 2	Compound 3	Positive control (µg/mL)	
Bacteria	MIC (µg/mL)				
Bacillus subtilis DSM 10	66.7	-	-	4.1	Oxytetracyclin
Chromobacterium violaceum DSM 30191	-	-	-	0.4	Oxytetracyclin
Escherichia coli DSM 1116	-	-	-	3.3	Oxytetracyclin
Micrococcus luteus DSM 1790	66.7	-	-	0.2	Oxytetracyclin
<i>Mycobacterium smegmatis</i> ATCC 700084	-	-	-	3.3	Kanamycin
Pseudomonas aeruginosa PA14	-	-	-	1.7	Gentamycin
Staphylococcus aureus DSM 346	66.7	-	-	3.3	Oxytetracyclin
Fungi					
Candida albicans DSM 1665	-	-	-	66.7	Nystatin
Mucor hiemalis DSM 2656	66.7	-	-	33.3	Nystatin
Pichia anomala DSM 6766	-	-	-	33.3	Nystatin
Rhodoturula glutinis DSM 10134	-	-	-	33.3	Nystatin
Schizosaccharomyces pombe DSM 70572	-	-	-	33.3	Nystatin
Cell lines	Cytotoxicity IC <sub>50</sub> (μg/mL)				
KB 3.1	14	-		5 × 10 <sup>-4</sup>	epothilone B
L929	27	-		1.1 × 10 <sup>-3</sup>	epothilone B

### Table S13. Antimicrobial activities of compounds 1-3.

(-) no activity

#### S14. Cytotoxicity assay results of compounds 1.







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**Figure S14.** Proliferation assay. Graph of MTT assay after 5 days of incubation. *In vitro* cytotoxicity  $(IC_{50})$  of compounds **1** was determined against HeLa (KB-3.1) (A) and mouse fibroblast L929 (B) cell lines. Epothilone B was used as the positive control while methanol was used as the negative control.



#### **S15.** Biofilm inhibitory activity of compounds 1-3.

**Figure S15.** Inhibition of the biofilm formation from *Staphylococcus aureus*. CASO medium containing 4% glucose was used as the negative control and tetracycline (100 µg/mL) was used as the positive control.