

# Effects of In Vitro Digestion on the Content and Biological Activity of Polyphenols from *Acacia mearnsii* Bark

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**Abstract:** The stability and bioaccessibility of polyphenol from *Acacia mearnsii* bark were measured at various stages during in vitro simulated digestion. Subsequently, the changes in the total polyphenol content (TPC) and biological activity were studied. The results showed that the phenolic compounds from *A. mearnsii* remained stable, and TPC underwent few changes during gastric digestion. Nonetheless, intestinal digestion led to the degradation of proanthocyanidins (PAs) and a significant decrease in TPC (26%). Degradation was determined by normal-phase HPLC and gel permeation chromatography. Only monomers, dimers, and trimers of flavan-3-ols were identified in the serum-accessible fraction for characterization of their bioaccessibility. The results also indicated the obvious antioxidant capacity of PAs from *A. mearnsii* bark, and ~53% of the  $\alpha$ -glucosidase-inhibitory effect was preserved. All these findings show that PAs from *A. mearnsii* bark as a native plant source may be particularly beneficial for human health as a natural nutritional supplement.

**Keywords:** *Acacia mearnsii*; proanthocyanidins; in vitro digestion; antioxidant; antidiabetic; HPLC/MS

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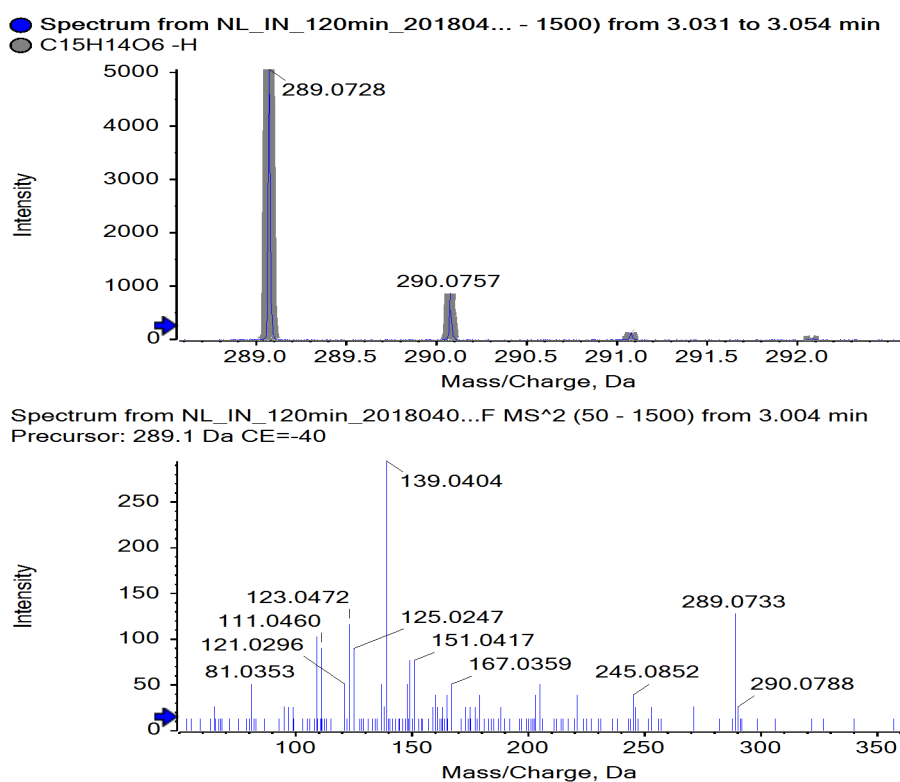
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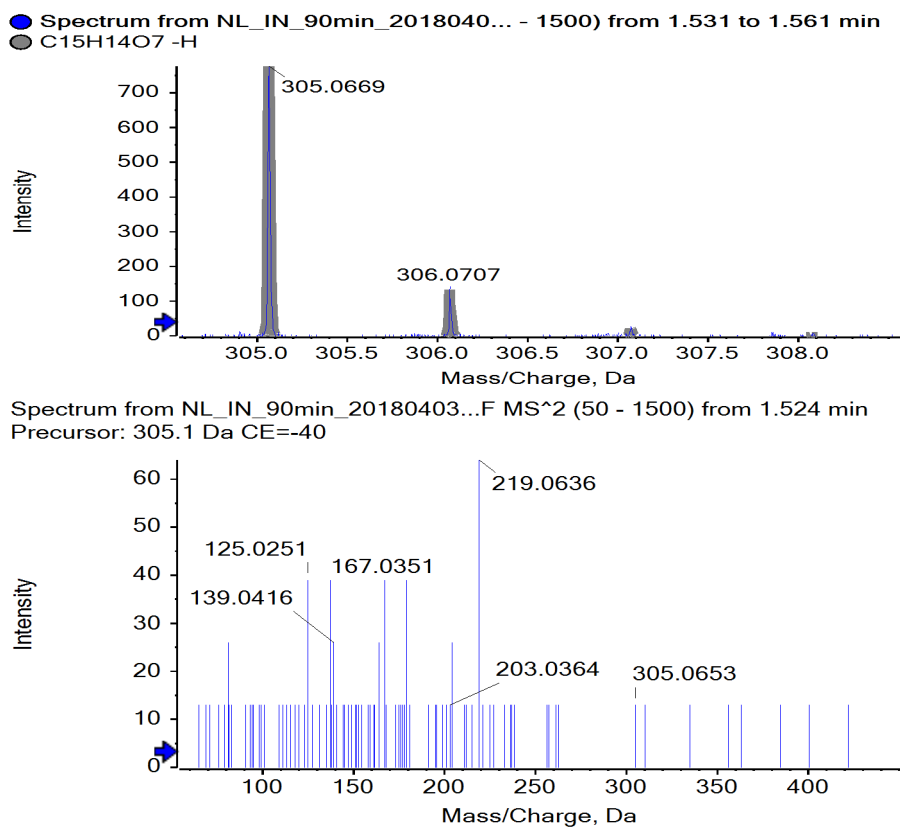
**Table S1.** Total polyphenol contents (TPC) of samples from *A. mearnsii* bark at different times during the simulated gastric-intestinal digestion without enzymes.

| Digestion Time<br>(min) | TPC of Digestion Samples (mg/mL) |             |             |
|-------------------------|----------------------------------|-------------|-------------|
|                         | Gastric Digestion                | OUT Sample  | IN Sample   |
| 0                       | 5.97 ± 0.05                      | 5.09 ± 0.02 | 0           |
| 30                      | 5.83 ± 0.06                      | 5.06 ± 0.03 | 0.25 ± 0.02 |
| 60                      | 5.88 ± 0.04                      | 5.01 ± 0.04 | 0.68 ± 0.02 |
| 90                      | 5.91 ± 0.03                      | 4.97 ± 0.02 | 1.23 ± 0.03 |
| 120                     | 5.89 ± 0.04                      | 4.99 ± 0.04 | 1.74 ± 0.04 |

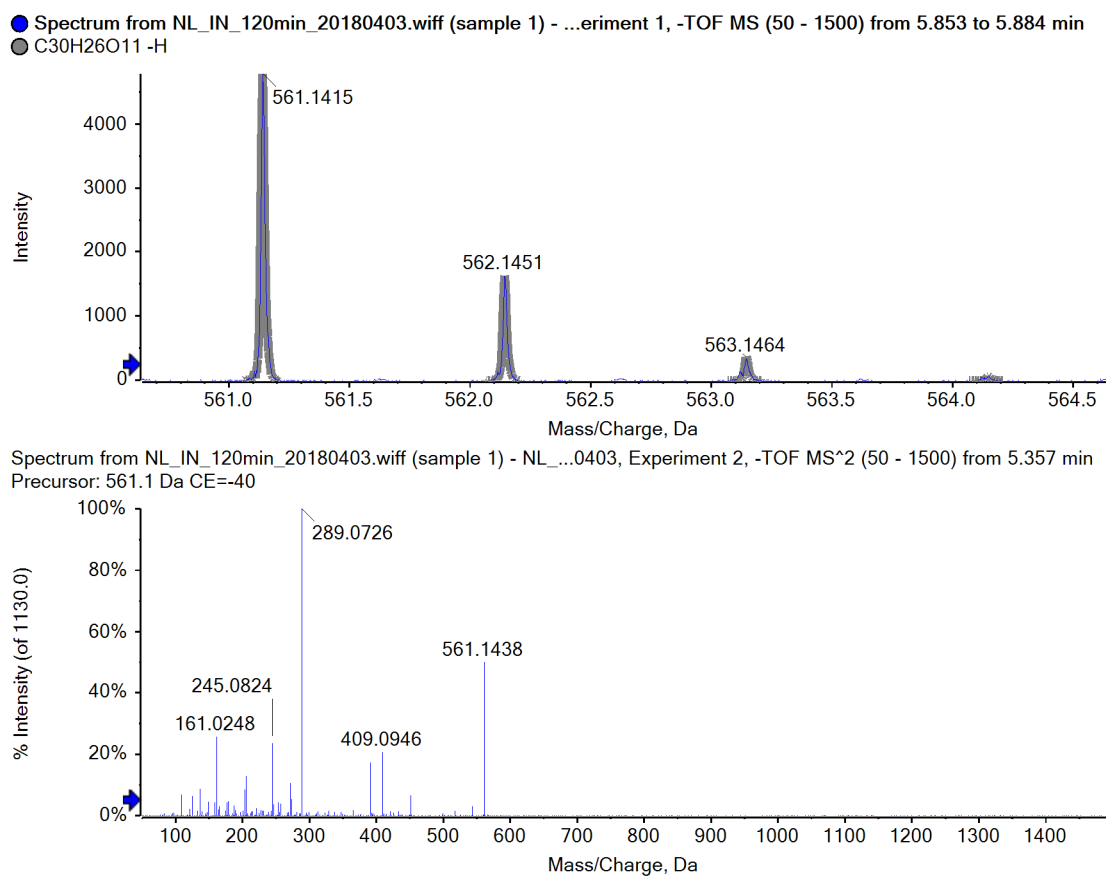
The IN sample represents the solution that diffused into the dialysis tubing, the OUT sample represents the solution outside of the tubing. Data represent the means of three independent determinations ± SD.



**Figure S1.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 289 in the IN sample.

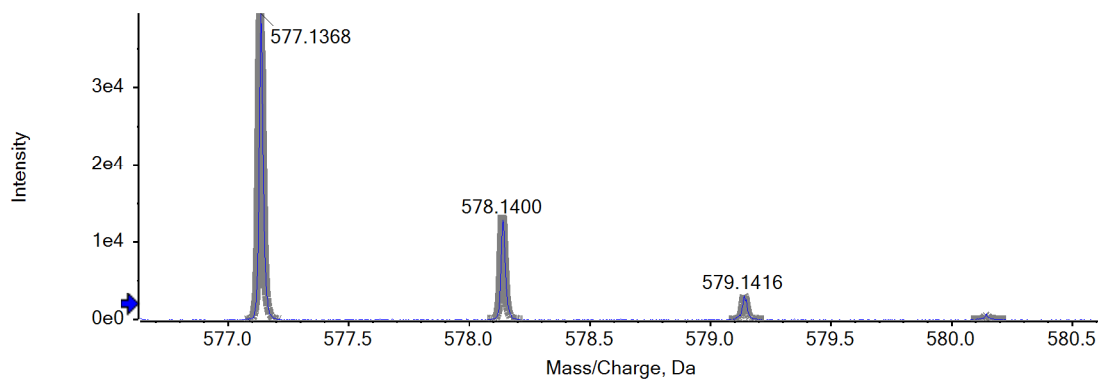


**Figure S2.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 305 fragment in the IN sample.

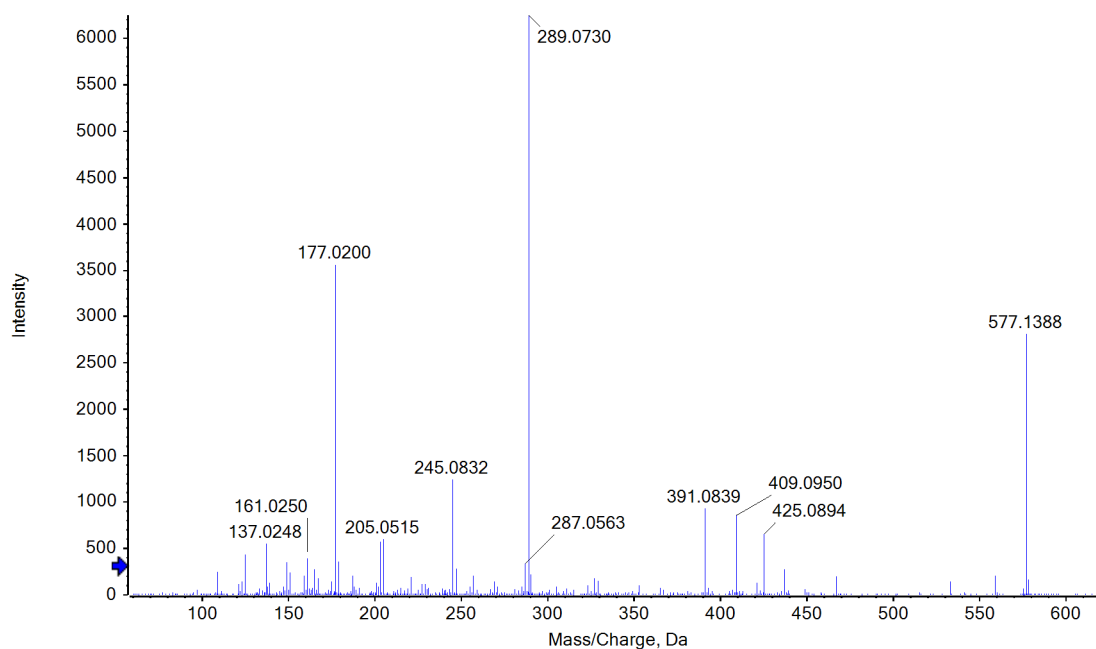


**Figure S3.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 561 in the IN sample.

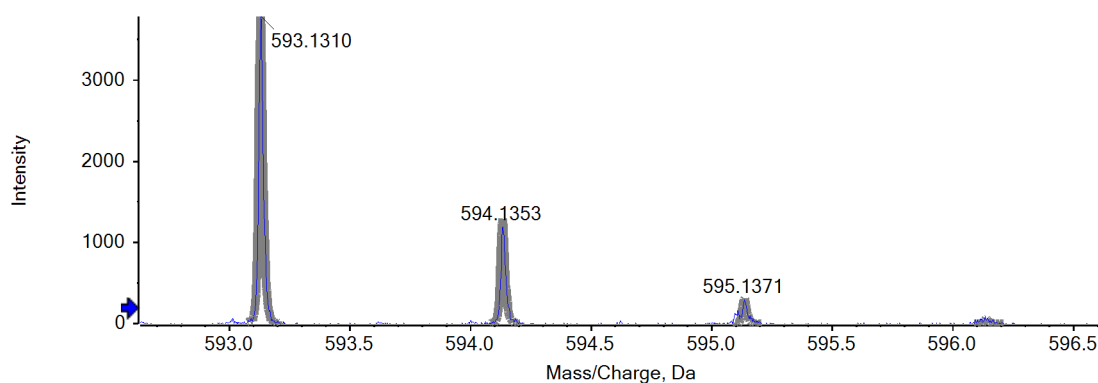
● Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - Experiment 1, -TOF MS (50 - 1500) from 4.232 to 4.262 min  
● C<sub>30</sub>H<sub>26</sub>O<sub>12</sub>-H



Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - NL\_IN\_1...\_20180403, Experiment 7, -TOF MS<sup>2</sup> (50 - 1500) from 4.307 min  
Precursor: 577.2 Da

**Figure S4.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 577 in the IN sample.

● Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - Experiment 1, -TOF MS (50 - 1500) from 2.580 to 2.610 min  
● C<sub>30</sub>H<sub>26</sub>O<sub>13</sub>-H



Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - NL\_...0403, Experiment 5, -TOF MS<sup>2</sup> (50 - 1500) from 2.345 min  
Precursor: 593.1 Da CE=-40

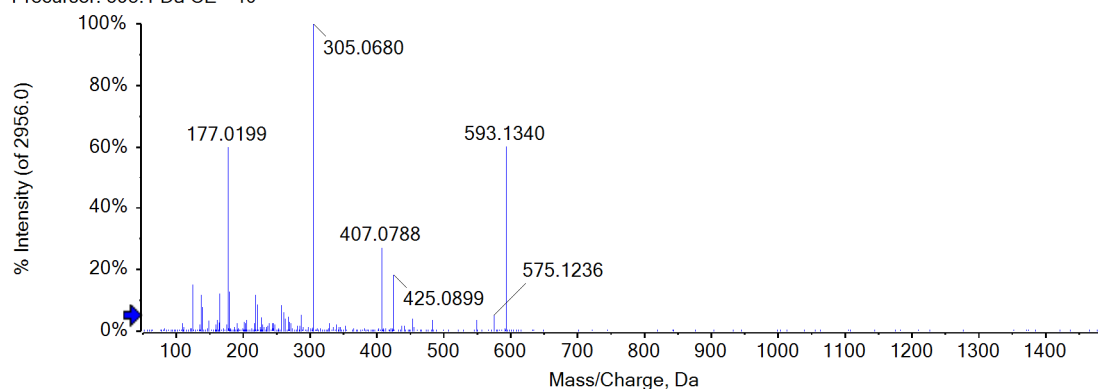
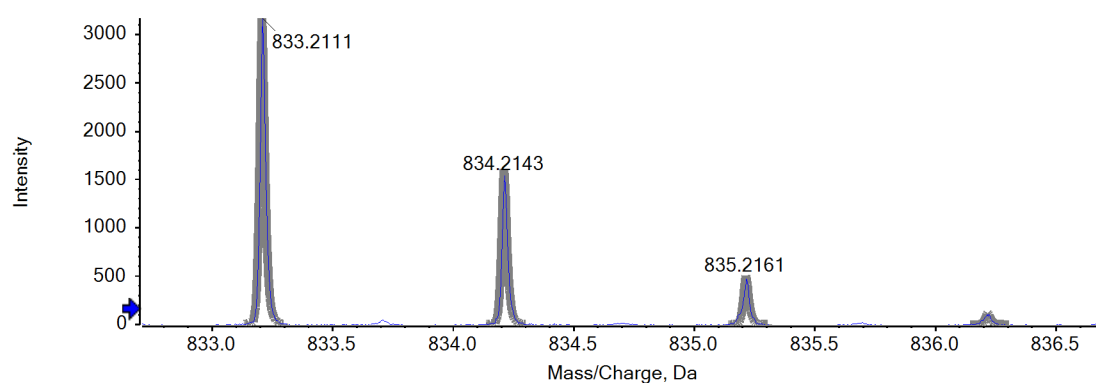


Figure S5. Product ion scan (MS and MS<sup>2</sup>) of the m/z 593 in the IN sample.

● Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - ...iment 1, -TOF MS (50 - 1500) from 10.948 to 10.980 min  
● C45H38O16 -H



Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - NL\_...403, Experiment 3, -TOF MS<sup>2</sup> (50 - 1500) from 10.942 min  
Precursor: 833.2 Da CE=-40

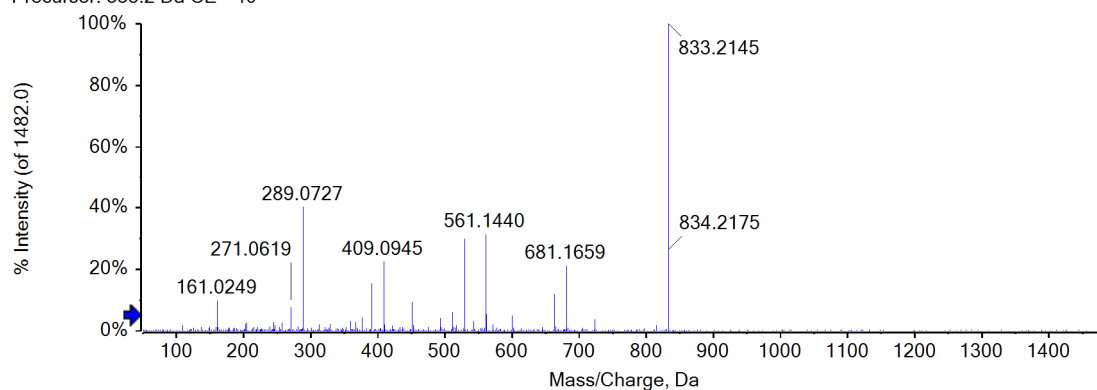
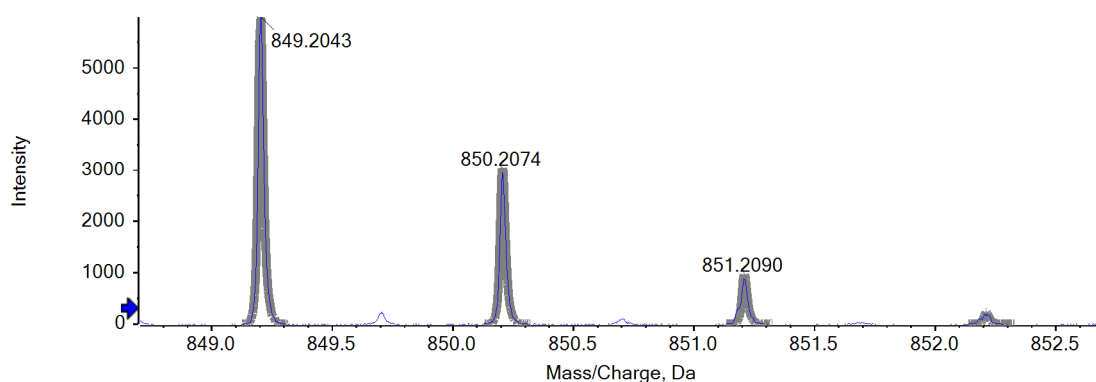
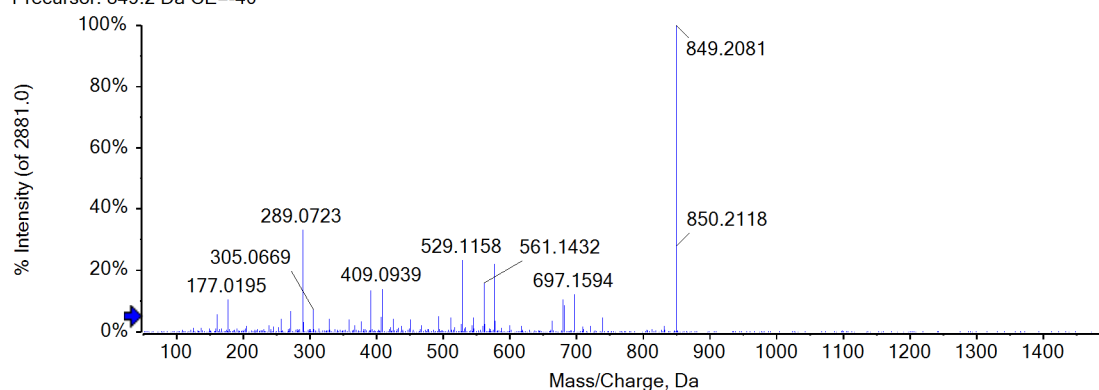


Figure S6. Product ion scan (MS and MS<sup>2</sup>) of the m/z 833 in the IN sample.

● Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - Experiment 1, -TOF MS (50 - 1500) from 9.672 to 9.703 min  
● C<sub>45</sub>H<sub>38</sub>O<sub>17</sub> -H

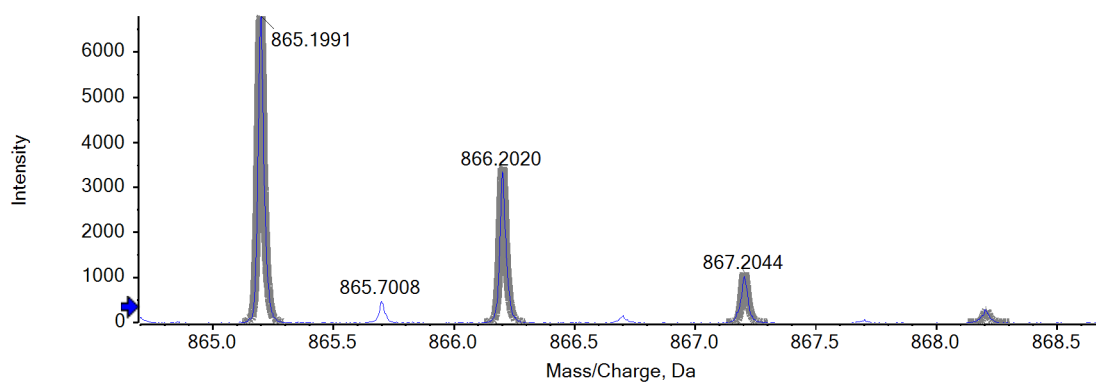


Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - NL\_...0403, Experiment 4, -TOF MS<sup>2</sup> (50 - 1500) from 9.593 min  
Precursor: 849.2 Da CE=40

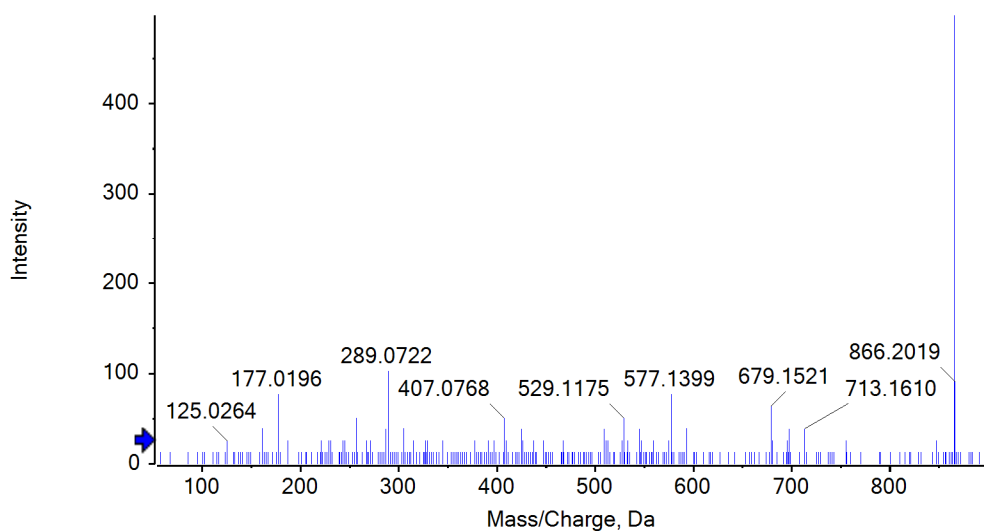


**Figure S7.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 849 in the IN sample.

● Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - Experiment 1, -TOF MS (50 - 1500) from 7.981 to 8.012 min  
● C<sub>45</sub>H<sub>38</sub>O<sub>18</sub> -H

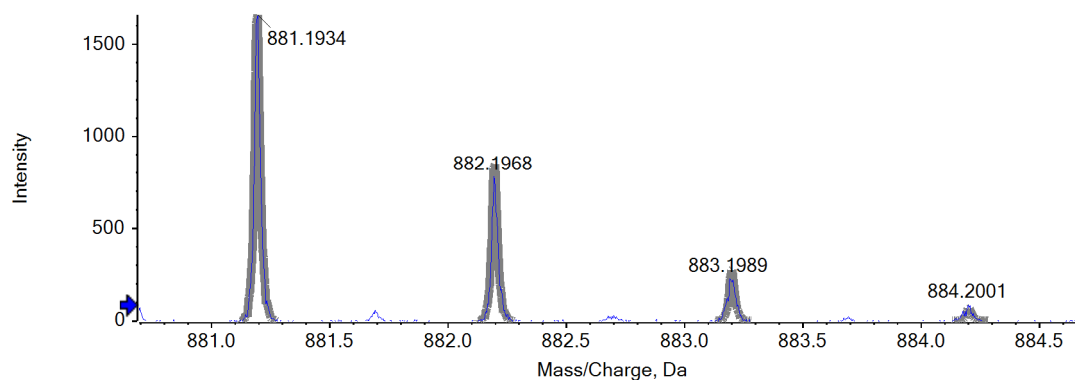


Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - Experiment 7, -TOF MS<sup>2</sup> (50 - 1500) from 6.468 min  
Precursor: 865.2 Da

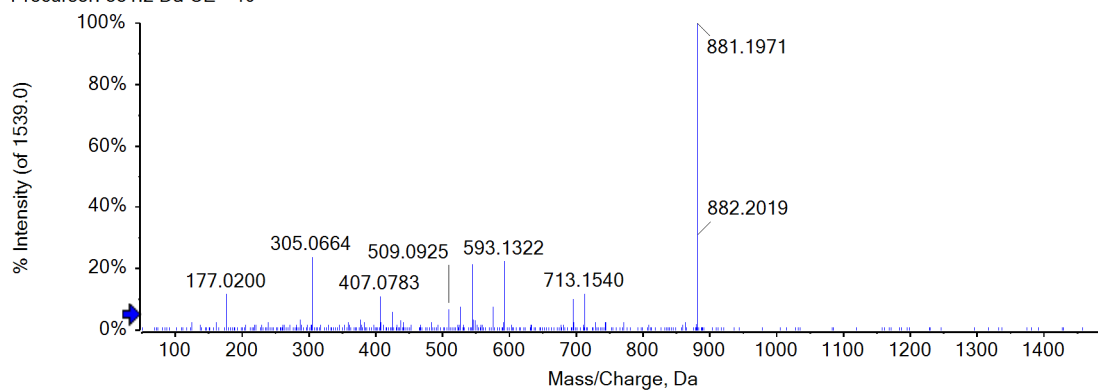


**Figure S8.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 865 in the IN sample.

● Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - Experiment 1, -TOF MS (50 - 1500) from 5.990 to 6.021 min  
● C45H38O19 -H



Spectrum from NL\_IN\_120min\_20180403.wiff (sample 1) - NL\_...0403, Experiment 7, -TOF MS<sup>2</sup> (50 - 1500) from 5.877 min  
Precursor: 881.2 Da CE=-40



**Figure S9.** Product ion scan (MS and MS<sup>2</sup>) of the m/z 881 in the IN sample.