Supplementary Material

Dynamic Allostery Modulates Catalytic Activity by Modifying the Hydrogen-Bonding Network in the Catalytic Site of Human Pin1

Jing Wang^{1,†}, Ryosuke Kawasaki^{1,†}, Jun-ichi Uewaki², Arif U.R. Rashid¹, Naoya Tochio² and Shin-ichi Tate^{1,2,*}

¹Department of Mathematical and Life Sciences, School of Science, Hiroshima University, 1-3-1 Kagamiyama, Higashi-Hiroshima 739-8526, Japan
²Research Center for the Mathematics on Chromatin Live Dynamics (RcMcD), Hiroshima University, 1-3-1 Kagamiyama, Higashi-Hiroshima 739-8526, Japan

[†]These authors contributed equally to this work.

*Correspondence:

Shin-ichi Tate

Department of Mathematical and Life Sciences,

School of Science, Hiroshima University

1-3-1 Kagamiyama, Higashi-Hiroshima 739-8526, Japan.

Tel: +81-82-424-7387

e-mail: tate@hiroshima-u.ac.jp

Figure Captions

Figure S1. Structural and backbone chemical shift changes induced by the S138A mutation. (A) Structural superposition of the wild-type (grey) [1] and S138A mutant (red). The basic triad and W73 are shown as stick models. The S138A mutation site is indicated as a sphere in red. In the inset, the NOEs related to W73 in the S138A mutant are shown as dashed yellow lines: the NOE connectivities from W73 are listed in Table S3. (B) Normalized ¹H_N and ¹⁵N backbone chemical shift differences between the wild-type [2] and the S138A mutant residues. The normalized chemical shift difference for each residue is defined as $\Delta \delta = [(\Delta \delta^1 H)^2 + (\Delta \delta^{15} N/5)^2]^{1/2}$, where $\Delta \delta^1 H$ and $\Delta \delta^{15} N$ are the chemical shift differences in ¹H_N and ¹⁵N dimensions, respectively [3]. (C) The NOE signals observed between C113 H^N and W73 H^{c1}. Signals with asterisks come from the other residues at different ¹⁵N planes in a 3D ¹⁵N-editted NOESY spectrum.

Figure S2. Comparison of phosphorylated peptide binding ability between the wild-type protein and the S138A mutant. Isothermal calorimetric titration data at 298 K for the wild-type Pin1-PPIase (left) and the S138A mutant (right) are shown. The red solid lines drawn in the Δ H plot (bottom panels) represent the best-fit model function assuming a 1:1 stoichiometry.

Figure S3. Comparison of heteronuclear NOEs between the S138A mutant and the wild-type Pin1 PPIase domain. (A) The heteronuclear NOEs for the S138A mutant (red) and those for the wild-type Pin1 PPIase domain (black) [2]. (B) Difference between the ¹⁵N-¹H heteronuclear NOEs (hNOEs) for the S138A mutant and those for the wild-type Pin1 PPIase domain [2]. The difference, Δ hNOE, is defined as Δ hNOE = hNOE^{S138A} – hNOE^{wild-type}. The region F103–A116 exhibited reduced $\Delta J(\omega_h)$ values relative to other residues are orange-boxed. **Figure S4.** Graphical representation of the correlation between J(0) and $J(\omega_N)$ [4]. The simple correlation between J(0) and $J(\omega_N)$ described in the main text eq. (1) is shown as the blue curve. Point *a* represents a residue with the N–H bond motion completely dominated by τ_c ; *b* represents a residue with significant rapid internal motion due to the local correlation time, τ_c ; *c* represents a residue with the N–H bond motion mainly dominated by rapid internal motion; and *d* represents a residue with slow conformational fluctuations.

Figure S5. A 2D ¹H–¹⁵N multi-bond HSQC spectrum [5] for the imidazole rings of the S138A Pin1 PPIase domain mutant. The set of signals marked with an asterisk represent the histidine located within the N-terminal GSHM segment, which is present because of the expression construct.

Completeness of resonance assignments (%) ^a	
Backbone	97.8
Side chain	99.3
Aromatic	100
Stereospecific methyl	100
Conformationally restricting restraints	
Distance restraints	
Total	2,691
Intraresidue $(i = j)$	543
Sequential $(i - j = 1)$	609
Medium range $(1 < i - j < 5)$	528
Long range $(i-j \ge 5)$	1,011
Dihedral angle restraints ^b	102
Hydrogen-bond restraints	0
Disulfide restraints	0
No. of restraints per residue	23.0
No. of long-range restraints per residue	8.6
Residual restraint violations	
Average no. of distance violations per structure	
0.1–0.3 Å	2.4
0.3–0.5 Å	0.1
> 0.5 Å	0
Average no. of dihedral angle violations per structure	
> 5°	0
Model quality ^c	
RMSD backbone atoms (Å) ^d	0.7
RMSD heavy atoms (Å) ^d	1.1
RMSD bond lengths (Å)	0.007
RMSD bond angles (°)	0.6
MolProbity Ramachandran statistics ^{c,d}	
Most favored regions (%)	94.7
Allowed regions (%)	5.3
Disallowed regions (%)	0
Global quality scores (raw / Z score) ^c	
Verify3D	0.40 / -0.96
Prosall	0.72 / 0.29
PROCHECK $(\phi - \psi)^d$	-0.28 / -0.79
PROCHECK (all) ^d	-0.19 / -1.12
MolProbity clash score	16.68 / -1.34
Model contents	
Total no. of residues	117
BMRB accession number	36014
PDB ID code	5GPH

Table S1. Structural statistics of the final 10 structures of S138A Pin1 PPIase mutant.

^aThe number excluding highly exchangeable protons, nitrogens bound with highly exchangeable protons, nonprotonated carbons and nitrogens.

^bThe angle restraints were derived from TALOS+ [6] with the angle ranges $\pm 30^{\circ}$.

^cCalculated using PSVS version 1.5 [7]. ^dFor residues P52-S111, S114-S126, Q129-M146, and P149-T162 as the ordered residues estimated by PSVS.

	<i>J</i> ₍ 0) [ns]	$J(\omega_{ m N})$ [ns]	$J(\omega_{\rm h})$ [ps]	$R_1 \\ [s^{-1}]$	R_2 [s ⁻¹]	hNOE [ratio]
Wild-type ^a	7.72 ± 0.03	0.190 ± 0.002	3.34 ± 0.18	0.92 ± 0.01	24.52 ± 0.08	0.77 ± 0.01
S138A	6.31 ± 0.03	0.232 ± 0.002	3.97 ± 0.26	1.12 ± 0.01	20.23 ± 0.09	0.77 ± 0.02
C113D ^a	6.62 ± 0.05	0.218 ± 0.003	3.51 ± 0.20	1.05 ± 0.01	21.15 ± 0.16	0.79 ± 0.01
C113A ^a	9.95 ± 0.18	0.204 ± 0.007	3.51 ± 0.32	0.98 ± 0.03	31.49 ± 0.58	0.77 ± 0.02
C113S ^a	6.67 ± 0.06	0.196 ± 0.006	2.97 ± 0.27	0.94 ± 0.03	21.24 ± 0.19	0.80 ± 0.02

Table S2. The relaxation parameters for the side chain of W73 (N ϵ).

Side chain relaxation for W73 was analyzed in a manner identical to that for the backbone amide group except using -107.7 ppm as the ¹⁵N CSA [8]. ^aThese values were determined using data in our previous studies [1,9].

Residue1	Atom1	Residue2	Atom2	
W73	ΗN	S72	H ^ℕ	
W73	ΗN	S72	Η ^{β#}	
W73	H ^{ε1}	S72	Η ^{β#}	
W73	ΗN	R74	ΗN	
W73	H^{δ_1}	R74	Η ^N	
W73	H^{δ_1}	R74	Η ^{γ#}	
W73	Η ^{ε1}	R74	Η ^{γ#}	
W73	$H^{\epsilon 3}$	R74	Hα	
W73	H^{ζ_2}	R74	H ^{δ#}	
W73	H^{ζ_2}	R74	$H^{\delta \#}$	
W73	$H^{\zeta 2}$	R74	Η ^{γ#}	
W73	$H^{\zeta 3}$	R74	Hα	
W73	$H^{\eta 2}$	Q109	Hα	
W73	$H^{\eta 2}$	Q109	Η ^{γ#}	
W73	$H^{\zeta 2}$	Q109	Hα	
W73	H ^{ε1}	D112	Hα	
W73	H ^{ε1}	C113	Η ^N	
W73	$H^{\delta 1}$	S114	H ^{β#}	
W73	$H^{\delta 1}$	A116	H ^{β#}	
W73	H ^{ε1}	A116	H ^{β#}	
W73	$H^{\eta 2}$	A116	H ^{β#}	
W73	$H^{\zeta 2}$	A116	H ^{β#}	

Table S3. Observed NOEs related to W73 (See Figure S1A inset).

represents ambiguous methylene protons.

Supplementary References

- Xu, N.; Tochio, N.; Wang, J.; Tamari, Y.; Uewaki, J.-i.; Utsunomiya-Tate, N.; Igarashi, K.; Shiraki, T.; Kobayashi, N.; Tate, S.-i. The c113d mutation in human pin1 causes allosteric structural changes in the phosphate binding pocket of the ppiase domain through the tug of war in the dualhistidine motif. *Biochemistry* 2014, *53*, 5568-5578.
- Xu, N.; Tochio, N.; Wang, J.; Tamari, Y.; Uewaki, J.; Utsunomiya-Tate, N.; Igarashi, K.; Shiraki, T.; Kobayashi, N.; Tate, S. The c113d mutation in human pin1 causes allosteric structural changes in the phosphate binding pocket of the ppiase domain through the tug of war in the dual-histidine motif. *Biochemistry* 2014, *53*, 5568-5578.
- Hall, D.A.; Vander Kooi, C.W.; Stasik, C.N.; Stevens, S.Y.; Zuiderweg, E.R.; Matthews, R.G. Mapping the interactions between flavodoxin and its physiological partners flavodoxin reductase and cobalamin-dependent methionine synthase. *Proc Natl Acad Sci U S A* 2001, *98*, 9521-9526.
- Krizova, H.; Zidek, L.; Stone, M.J.; Novotny, M.V.; Sklenar, V. Temperature-dependent spectral density analysis applied to monitoring backbone dynamics of major urinary protein-i complexed with the pheromone 2- sec-butyl-4,5-dihydrothiazole. *Journal of biomolecular NMR* 2004, 28, 369-384.
- 5. Pelton, J.G.; Torchia, D.A.; Meadow, N.D.; Roseman, S. Tautomeric states of the active-site histidines of phosphorylated and unphosphorylated iiigle, a signal-transducing protein from escherichia coli, using two-dimensional heteronuclear nmr techniques. *Protein science : a publication of the Protein Society* **1993**, *2*, 543-558.
- Shen, Y.; Delaglio, F.; Cornilescu, G.; Bax, A. Talos+: A hybrid method for predicting protein backbone torsion angles from nmr chemical shifts. *J Biomol NMR* 2009, 44, 213-223.
- Bhattacharya, A.; Tejero, R.; Montelione, G.T. Evaluating protein structures determined by structural genomics consortia. *Proteins: Structure, Function, and Bioinformatics* 2007, 66, 778-795.
- Miloushev, V.Z.; Bahna, F.; Ciatto, C.; Ahlsen, G.; Honig, B.; Shapiro, L.; Palmer, A.G., 3rd. Dynamic properties of a type ii cadherin adhesive domain: Implications for the mechanism of strand-swapping of classical cadherins. *Structure (London, England : 1993)* 2008, *16*, 1195-1205.
- Wang, J.; Tochio, N.; Kawasaki, R.; Tamari, Y.; Xu, N.; Uewaki, J.-i.; Utsunomiya-Tate, N.; Tate, S.-i. Allosteric breakage of the hydrogen bond within the dual-histidine motif in the active site of human pin1 ppiase. *Biochemistry* 2015, *54*, 5242-5253.





B



Figure S1

С











