

Article

Systemic Induction of the Defensin and Phytoalexin Pisatin Pathways in Pea (*Pisum sativum*) against *Aphanomyces euteiches* by Acetylated and Nonacetylated Oligogalacturonides

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Abstract: Oligogalacturonides (OGs) are known for their powerful ability to stimulate the plant immune system but little is known about their mode of action in pea (*Pisum sativum*). In the present study, we investigated the elicitor activity of two fractions of OGs, with polymerization degrees (DPs) of 2–25, in pea against *Aphanomyces euteiches*. One fraction was nonacetylated (OGs – Ac) whereas the second one was 30% acetylated (OGs + Ac). OGs were applied by injecting the upper two rachises of the plants at three- and/or four-weeks-old. Five-week-old roots were inoculated with 10⁵ zoospores of *A. euteiches*. The root infection level was determined at 7, 10 and 14 days after inoculation using the quantitative real-time polymerase chain reaction (qPCR). Results showed significant root infection reductions namely 58, 45 and 48% in the plants treated with 80 µg OGs + Ac and 59, 56 and 65% with 200 µg of OGs – Ac. Gene expression results showed the upregulation of genes involved in the antifungal defensins, lignans and the phytoalexin pisatin pathways and a priming effect in the basal defense, SA and ROS gene markers as a response to OGs. The reduction of the efficient dose in OGs + Ac is suggesting that acetylation is necessary for some specific responses. Our work provides the first evidence for the potential of OGs in the defense induction in pea against *Aphanomyces* root rot.

Keywords: pea root rot; *Aphanomyces euteiches*; oligogalacturonides; real-time qPCR; gene expression; pea defense pathways; defensins; pisatins; phytoalexins

1. Introduction

Plants need to be able to recognize pathogen attacks in a timely manner in order to activate their defenses that provide protection against the infection process. The plant cell wall is the site of initial interaction with microbial pathogens. Pectin is one of the most accessible components of the cell wall and, therefore, is among the first structures to be altered upon pathogen invasion. The oligogalacturonides (OGs) are produced upon partial degradation of the pectin homogalacturon by pathogen pectinases and polygalacturonases [1]. However, OGs have been indicated as damage-associated molecular patterns (DAMPs) which may trigger plant defenses against pathogens. OGs endogenous elicitors consist of linear chains of α -(1-4)-linked D-galacturonic acid [2,3] and those with high biological activity have often a degree of polymerization (DP) between 10 and 15 [4]. This size

is optimal for the formation of Ca^{2+} mediated inter molecular cross links resulting in structures called “egg boxes” that are thought to be necessary for OGs activity [5,6]. The short OGs with a DP of 2–6 have been reported in few cases to exhibit elicitor activity in tomato [7,8]; however, they appear to suppress defense responses in wheat [9]. It has been shown that the degree of OGs methylation clearly influences plant defense responses [10–12]. In wild strawberry (*Fragaria vesca*), partial demethylation of OGs in transgenic fruit enhanced resistance to *Botrytis cinerea* [13]. Furthermore, in wheat, Wietholter et al. [14] found a significant difference in the methyl ester distribution in OGs from cultivars susceptible or resistant to stem rust. Recently, we reported more than 57% protection in wheat against *Blumeria graminis* f. sp. *tritici* using 30% chemically acetylated or nonacetylated citrus OGs with DPs of 2–25 [15]. We found that only the acetylated OGs led to an increase in papilla-associated fluorescence and a reduction in the fungal haustoria formation, suggesting that acetylation is necessary for some specific responses. However, OGs elicit in several plant species [16–18] a wide range of defense responses, including induction of polygalacturonase-inhibiting protein (PGIP) [19,20], accumulation of phytoalexins [21], glucanase, and chitinase [22,23], deposition of callose, production of reactive oxygen species [15], and nitric oxide [24]. Root rot caused by the oomycete *Aphanomyces euteiches* is the major destructive soil-borne fungal disease of pea (*Pisum sativum*) with up to 80% yield loss per year. It is widespread in North America, Europe, Japan, Australia and New Zealand [25]. Oospores released from infected roots into the rhizosphere constitute the primary source of inoculum. *A. euteiches* invades the root system leading to a complete arrest of root growth and ultimately plant death. To date, disease control measures are limited to crop rotation and no resistant pea lines are available. The fact that oospores are able to remain dormant in the soil for up to 10 years reduces the effectiveness of crop rotation in decreasing the propagation of this pathogen [26]. Moreover, oomycetes are distantly related to true fungi and their particular physiology makes them insensitive to most fungicides [27]. Therefore, the development of alternative control methods against oomycetes is becoming urgent. In the present work, the efficiency of two distinct biochemical fractions of OGs to protect pea against *A. euteiches* was studied. These fractions consisted of OGs with DPs ranging from 2 to 25 (OGs – Ac) and one fraction that was chemically 30% acetylated (OGs + Ac). The systemic defense mechanisms elicited in pea roots as a response to rachis injections with OGs are discussed.

2. Results

2.1. Elicitor Effect of OG Compounds

A. euteiches oospores were observed within necrotic tissues between 7 and 14 days after inoculation (dai). However, at ten dai, high *P. sativum*-*A. euteiches* compatibility was observed with the disease severity index (DSI) 3.5 and 76% of root fragments with more than 50 oospores (Figure 1).

No significant difference was observed between controls injected with water at one and/or two weeks before inoculation (wbi). At 10 dai, OGs at the dose of 20 $\mu\text{g}/\text{plant}$ and all other tested elicitors did not lead to any significant protection compared to the control injected with water (Figure 2). The protection level conferred on pea against *A. euteiches* was significant and increased to 43.5% and 47.8% as a response to the increase in the injected dose of OGs – Ac and OGs + Ac, respectively, to 40 $\mu\text{g}/\text{plant}$ at two wbi (Figure 3a). This protection level was associated with a significant reduction (50.7% and 60%, respectively) in the percentages of root fragments containing more than 50 oospores (Figure 3b).

As the classic methods using the DSI and the counting of oospores are time-consuming and tedious, especially with the large number of root samples, we developed primers and probes to evaluate the disease severity using qPCR. Figure 4 shows the relation between the *A. euteiches* specific gene and the qPCR threshold cycles. The efficiency of the qPCR was 99.58% with high sensitivity to detecting one copy of the *A. euteiches* specific gene (Figure 4).

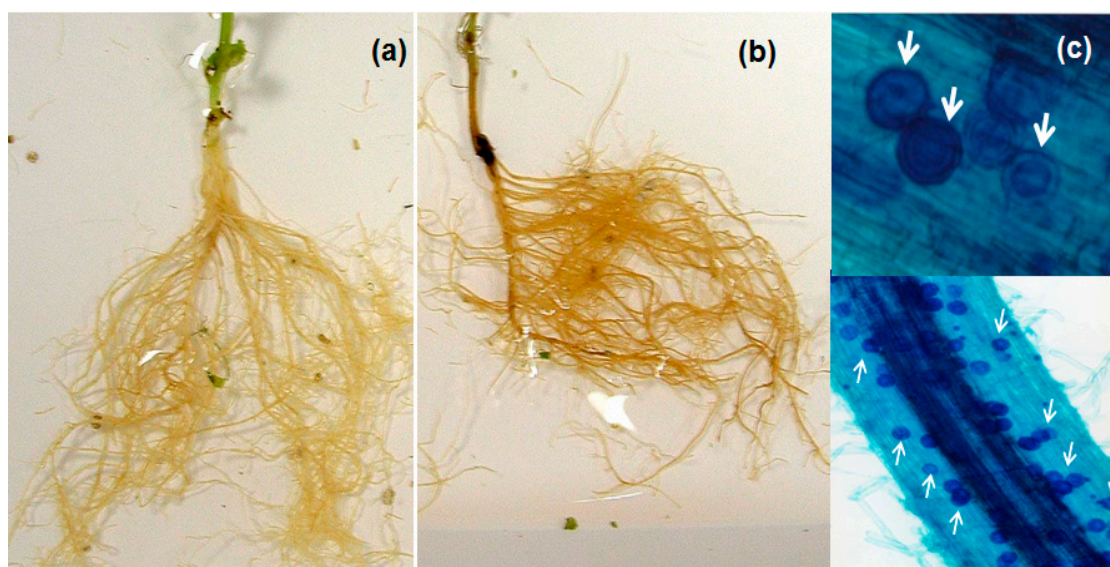


Figure 1. Pea stem basal part and root necrosis at 10 days after inoculation with 10^5 zoospores of *Aphanomyces euteiches* at 5-weeks-old; (a) Roots of plant injected with a solution of nonacetylated oligogalacturonides (OGs – Ac) elicitor in the upper two rachises ($20 \mu\text{g/rachis} = 40 \mu\text{g/plant}$) at two weeks before inoculation; (b) Controls injected with water; (c) Plant roots colored with lactophenol cotton blue. *A. euteiches* oospores indicated with white arrows.

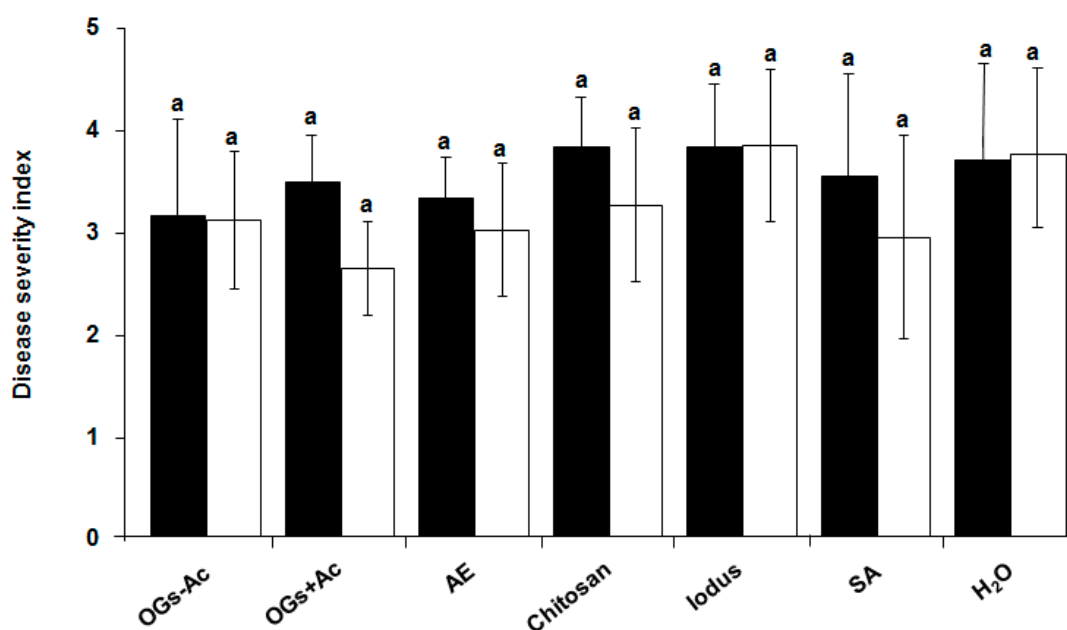


Figure 2. Pea root rot disease severity index at 10 days after inoculation with *Aphanomyces euteiches* at 10^5 zoospores/plant at 5-week-old. The upper plant rachis was injected with elicitors ($20 \mu\text{g/rachis}$) at two (white) or one (black) weeks before inoculation. OGs – Ac; nonacetylated oligogalacturonides, OGs + Ac; acetylated oligogalacturonides, AE; an inoculum of *A. euteiches* zoospores (10^5 zoospores. mL^{-1}) heated at 100°C for 10 min, Chitosan, Iodine, SA; salicylic acid. Controls were injected with water. The values shown are means with SD ($n = 5$). Different lower-case letters indicate significant differences between treatments according to the Tukey test ($p \leq 0.05$).

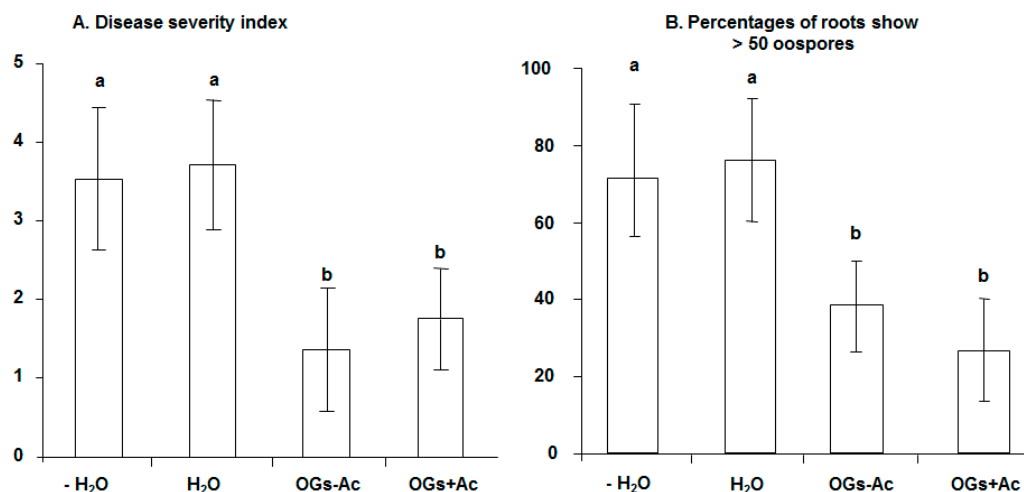


Figure 3. (A) Pea root rot disease severity index; (B) Percentage of roots showing >50% of oospores, at 10 days after inoculation with *Aphanomyces euteiches* at 10^5 zoospores/plant at 5-weeks-old. The upper two rachises of plants were injected with $20 \mu\text{g}/\text{rachis}$ ($=40 \mu\text{g}/\text{plant}$) of acetylated oligogalacturonides (OGs + Ac) or nonacetylated OGs (OGs – Ac) two weeks before inoculation. H₂O; controls injected with water, –H₂O; controls without water injection. The values shown are means with SD ($n = 5$). Different lower-case letters indicate significant differences between treatments according to the Tukey test ($p \leq 0.05$).

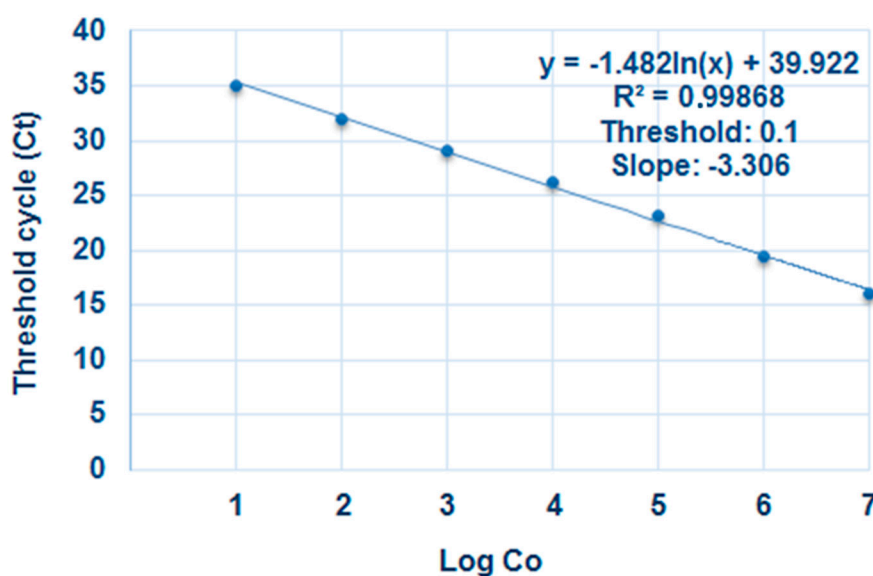


Figure 4. Standard curve using known copies (10^1 to 10^7) of the appropriate cloned target sequence of the *Aphanomyces euteiches* specific gene (GenBank accession No. AF228037.1). For each reaction, the cycle threshold (Ct), the initial cycle number at which an increase in fluorescence above a baseline can be detected, is plotted against the log₁₀ (Log Co) of the *A. euteiches* specific gene copies. Three technical PCR replicates were performed for each concentration.

The results in Figure 5 show the *A. euteiches* specific gene copy numbers in 100 ng of total DNA (AESG_{100ng}) extracted from root samples collected at 7, 10 and 14 dai. Elicitor efficiency was calculated by comparing the levels of AESG_{100ng} in the inoculated plants after the elicitor pretreatment with those in the inoculated control plants without elicitor pretreatment. The well-known endogenous elicitor salicylic acid (SA) was used as a reference. No significant differences in AESG_{100ng} were observed between all the inoculated control modalities, injected or non-injected with water, at the three

observation dates. These controls were grouped and used as repetitions for non-treated inoculated controls. Plants treated with the high doses (200 and 400 $\mu\text{g}/\text{plant}$) of SA showed phytotoxicity symptoms of brown necrotic lesions on the leaves and the injected rachises were dead. On the other hand, unstable protection efficiencies (50, 38 and 4%) and (48, 32 and 7%) were recorded in root samples collected at 7, 10 and 14 dai, respectively, as a response to the injection with SA (40 $\mu\text{g}/\text{plant}$) once at two wbi or twice (= 80 $\mu\text{g}/\text{plant}$) at two and one wbi (Figure 5). The same phytotoxicity symptoms, but less severe than those seen with SA, were observed in plants injected with the high dose (400 $\mu\text{g}/\text{plant}$) of OGs – Ac and OGs + Ac. Unstable protection was observed with the one-date injection modalities (two wbi) with 40 $\mu\text{g}/\text{plant}$ of OGs – Ac (74, 33 and 6%) and OGs + Ac (38, 35 and 7%) at 7, 10 and 14 dai, respectively (Figure 5). However, significant and stable protection efficiencies of 58, 45 and 55% were recorded at 7, 10 and 14 dai, respectively, in the plants treated twice, at two and one wbi, with 40 $\mu\text{g}/\text{plant}$ OGs + Ac at each date (=80 $\mu\text{g}/\text{plant}$) and of 59, 56 and 65% in the plants treated at only one date (two wbi) with 200 $\mu\text{g}/\text{plant}$ OGs – Ac (=2.5 folds more than OGs + Ac) (Figure 5). The differences between these two treatments were not significant. The root samples of these two efficient treatments were used for gene expression studies.

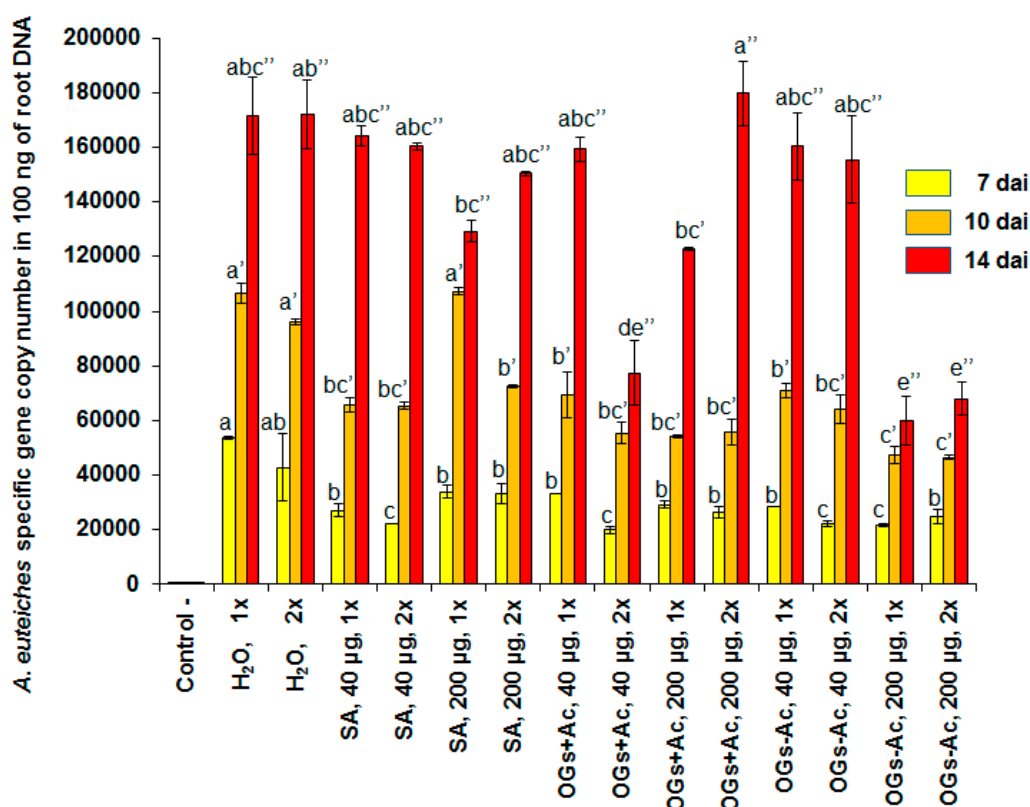


Figure 5. *Aphanomyces euteiches* specific gene copy numbers in 100 ng of the total DNA extracted from root samples collected at 7, 10 and 14 dai (days after inoculation). Pea plants injected in the upper two rachises with 20 $\mu\text{L}/\text{rachis}$ of salicylic acid (SA), acetylated oligogalacturonides (OGs + Ac) or nonacetylated OGs (OGs – Ac). The elicitor injections were done once (1 \times) two weeks before inoculation (wbi) or twice (2 \times) two and one wbi. The final elicitor concentrations were 40, 80, 200 or 400 $\mu\text{g}/\text{plant}$. Controls were injected with water. The values shown are means with SD ($n = 5$). Different lower-case letters indicate significant differences between treatments according to the Tukey test ($p \leq 0.05$).

2.2. Gene Expression

The expression of Pathogenesis Related protein 1 (PR1), 1,3 β glucanase and phenylalanine ammonia-lyase (PAL) genes was followed at 3, 6, 12, 24, 48, 96, 168 and 336 hat and using the

same time course after inoculation (hai) in root samples harvested from plants injected twice at two and one wbi with 80 µg/plant of OGs + Ac (40 µg/plant/date) or once at two wbi with OGs – Ac (200 µg/plant). No upregulation of the three tested genes was detected in the controls injected with water in comparison with the control non-injected and non-inoculated. Neither elicitor treatments nor inoculation with *Aphanomyces* showed any expression changes for the *PR1* and *1,3 β glucanase* genes over the tested time course. However, significant upregulation (≥ 2 folds) of the *PAL* gene was recorded at 6, 12, 48, 168 and 336 hai with *A. euteiches* (Figure 6). On the other hand, high and early induction (3 hat) of *PAL* was observed in roots pretreated with both OG compounds. This active response of *PAL* over the tested time course in non-inoculated plants was higher and more stable with OGs – Ac (until 360 hat) than with OGs + Ac (until 168 hat) (Figure 6). The highest recorded *PAL* expression value was found at 168 hat in the plant roots pretreated with OGs + Ac (12.9-fold) and 31-fold at 24 hat in the plant roots pretreated with OGs – Ac. At the inoculation time (336 hat), *PAL* expression in OGs – Ac samples was upregulated 2.8-fold and this value increased strongly after inoculation to reach 13.8, 56.7, 8.0, 10.8 and 8.4-fold at 3, 6, 12, 24, and 96 hai, respectively (Figure 6). These values were significantly higher than those in the inoculated non-treated control which were up-regulated 1.4, 3.6, 2.7, 1.5 and 1.3-fold, respectively. In the case of OGs + Ac, *PAL* expression was 0.3-fold at the time of inoculation and upregulated after inoculation to reach 5.7-, 18.2-, 2.9- and 5.9-fold at 3, 6, 24 and 48 hai, respectively (Figure 6). These values were significantly higher than those in the inoculated non-treated plants which were up-regulated 1.4, 3.6, 1.5 and 2.5-fold, respectively.

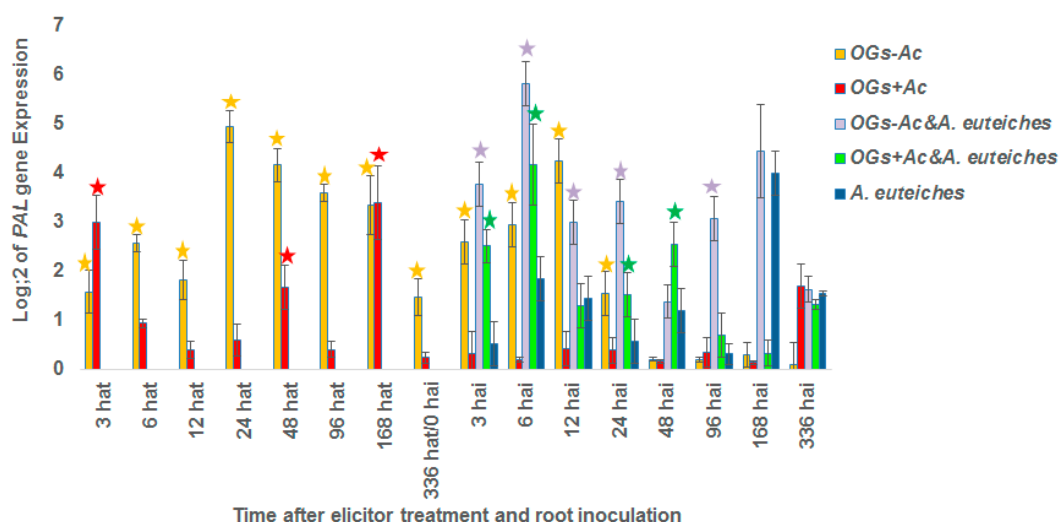


Figure 6. *PAL* gene expression ratio at 3, 6, 12, 24, 48, 96, 168 and 336 hours after treatment (hat) with water, 80 µg/plant of acetylated oligogalacturonides (OGs + Ac) or 200 µg/plant of nonacetylated OGs (OGs – Ac). The elicitor injections were done two weeks before inoculation (wbi) and two and one wbi for OGs – Ac and OGs + Ac, respectively. After inoculation with 10^5 *Aphanomyces euteiches* zoospores/plant, *PAL* gene expression was followed over the same time course (hours after inoculation (hai)). The values shown are means of five repetitions. ★ Stars indicate gene induction ≥ 2 -folds and significant differences between elicitor treatments and inoculated non-treated control according to the Tukey test ($p \leq 0.05$).

The expression changes in twenty-two pea defense genes as a response to OG pretreatments were followed using RT-PCR in root samples at 0 (just at the time of root inoculation), 7, 10 and 14 dai with *A. euteiches* in three modalities: inoculated without OG elicitor pretreatment; inoculated and pretreated with 80 µg/plant OGs + Ac; and inoculated and pretreated with 200 µg/plant OGs – Ac. These modalities were compared with the inoculated untreated controls. The results in Figure 7 show that at the inoculation time (0 dai = 14 dat), except for *GST* with OGs – Ac pretreatment, no significant induction of the genes involved in the ROS pathway (*SOD*, *POX*, *Catalase*, *NOS*, *Metallothionein*) was

observed. After inoculation, only the *Catalase* and *NOS* genes were significantly upregulated at 10 dai in plants pretreated with OGs – Ac compared to their levels in the inoculated non-treated control. In addition, no significant induction was observed for the genes coding for *MAPK* and *PRP* (Figure 8). For the genes coding for pathogenesis-related proteins, *PR1*, β 1,3 glucanase, *chitinase*, *DRR230*, *DRR276*, *DRR206* and *DRR49*, significant inductions of *chitinase* expression, at 0 and 10 dai, were recorded in the plant roots pretreated with OGs – Ac and OGs + Ac, respectively. At 7 dai, *DRR276* and *DRR49* were recorded in the plant roots pretreated with OGs – Ac (Figures 7 and 8). The *DRR206* and *DRR230* genes showed significant upregulations at 0 dai and at 0 and 7 dai respectively, in the plants pretreated with the two OG compounds (Figure 8). However, no significant activation was observed for *PR1* and β 1,3 glucanase genes with either of the two OG elicitors. The *PGIP* gene, coding for polygalacturinase inhibitor protein enzyme (marker gene for the basal defense), and *LOX*, coding for lipoxygenase enzyme (marker gene of the jasmonic acid pathway), showed no significant upregulation except for *PGIP* at 10 dai and only with OGs – Ac.

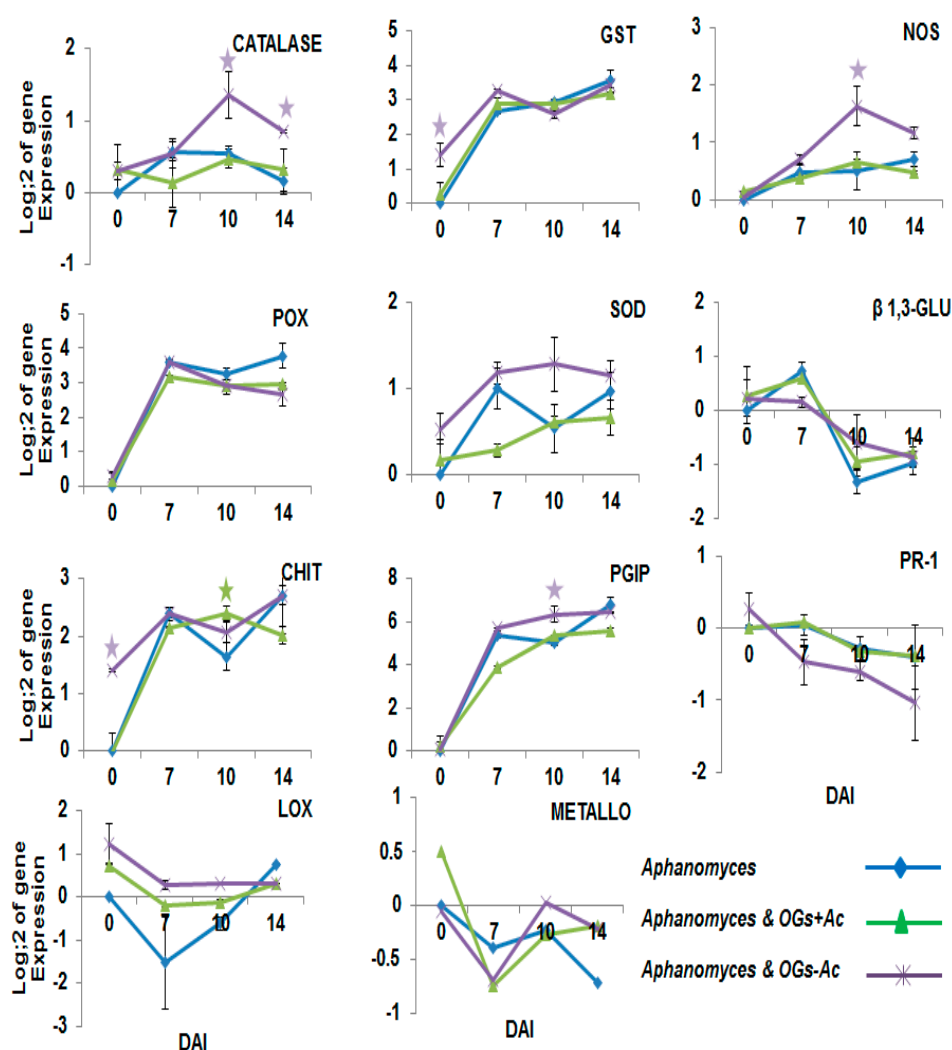


Figure 7. Pea gene expression ratio at 0 (just at the time of root inoculation), 7, 10 and 14 days after inoculation with 10^5 *Aphanomyces euteiches* zoospores/plant. Pea plants were injected with acetylated oligogalacturonides (OGs + Ac) one and two weeks before inoculation (final concentration = 80 μ g/plant) or with nonacetylated OGs (OGs – Ac) two (wbi) (final concentration = 200 μ g/plant). Controls were injected with water. The values shown are means of 5 repetitions. ★ Stars indicate gene induction ≥ 2 -folds and significant differences between elicitor treatments and inoculated non-treated control according to the Tukey test ($p \leq 0.05$).

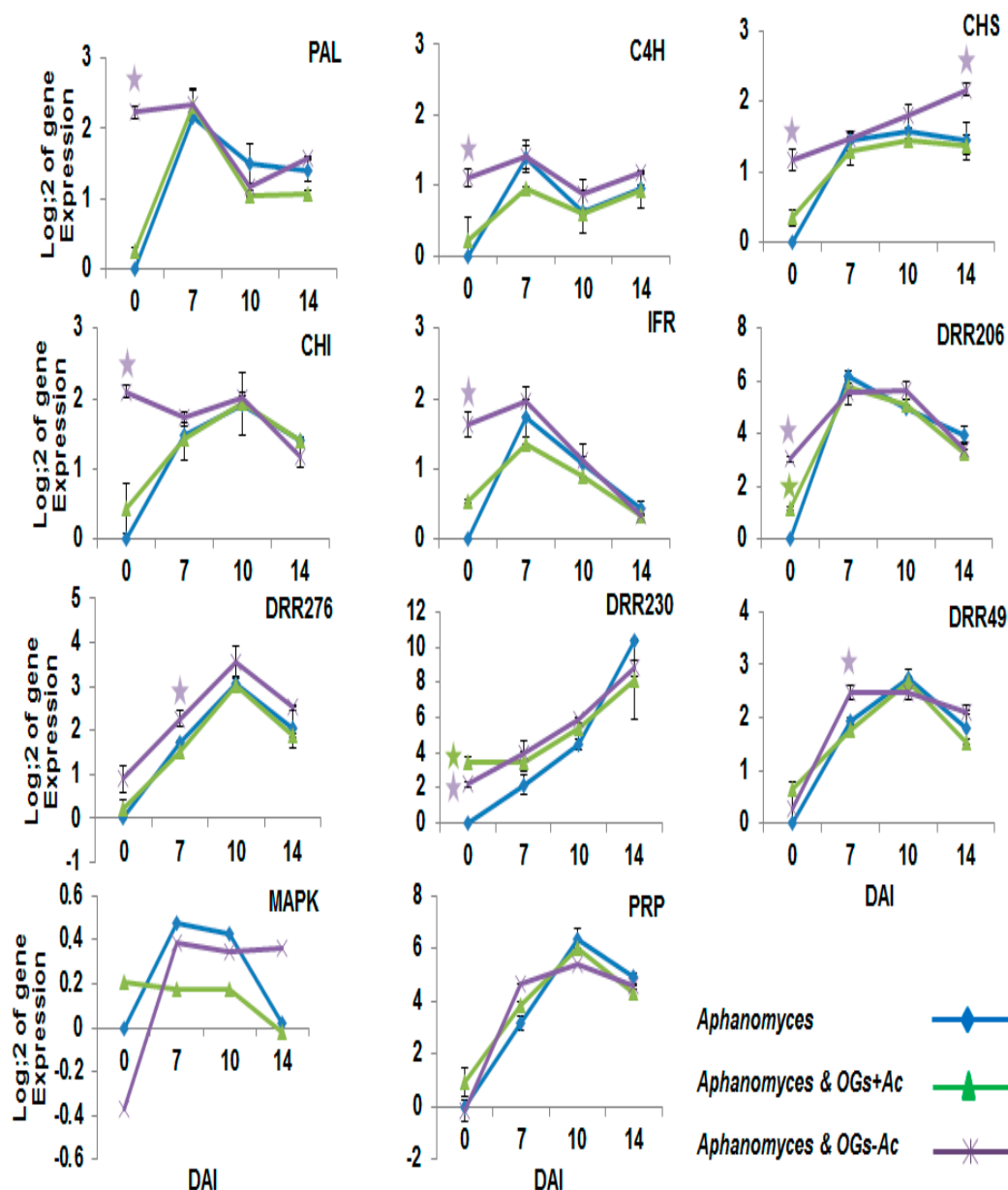


Figure 8. Pea gene expression ratio at 0 (at the time of root inoculation), 7, 10 and 14 days after inoculation with 10^5 *Aphanomyces euteiches* zoospores/plant. Pea plants were injected with acetylated oligogalacturonides (OGs + Ac) one and two weeks before inoculation (final concentration = 80 $\mu\text{g}/\text{plant}$) or with nonacetylated OGs (OGs – Ac) two (wbi) (final concentration = 200 $\mu\text{g}/\text{plant}$). Controls were injected with water. The values shown are means of 5 repetitions. ★ Stars indicate gene induction ≥ 2 -folds and significant differences between elicitor treatments and inoculated non-treated control according to the Tukey test ($p \leq 0.05$).

For the genes involved in the phenylpropanoid and phytoalexin pathways (*PAL*, cinnamate-4-hydroxylase (*C4H*), chalcone synthase (*CHS*), chalcone isomerase (*CHI*), and isoflavone reductase (*IFR*)), and known as SA gene markers, these were all significantly upregulated at the time just before inoculation (0 dai) in the plants pretreated with OGs – Ac. Only the *CHS* gene was significantly upregulated at 14 dai in roots from plants pretreated with OGs – Ac compared to those untreated and inoculated with *A. euteiches*. None of these genes showed upregulation at the time of inoculation in the plants pretreated with OGs + Ac.

3. Discussion

The oomycete *Aphanomyces euteiches* causes up to 80% crop loss in pea (*P. sativum*). To date, disease control measures are limited to crop rotation and no resistant pea lines or efficient fungicides are available. The present study aimed to investigate the potential of citrus-derived OGs to stimulate defense mechanisms in pea roots against *A. euteiches*. We tested two OGs fractions with DPs ranging from 2 to 25, nonacetylated (OGs – Ac) and acetylated at 30% (OGs + Ac).

Our results revealed no protection efficiency against *A. euteiches* root rot as a response to the commercial elicitors tested, Chitosan® and Iodus®, as well as the heated inoculum of *A. euteiches* zoospores. In parallel, SA showed unstable protection efficiencies and phytotoxicity symptoms with the higher doses tested.

In contrast, plants injected with the OGs showed a significant disease reduction. High and stable protection efficiencies (>45%) were recorded in plants pretreated with OGs. On the other hand, the acetylation of OGs reduced the efficient dose more than two times compared to the nonacetylated OGs. It has been shown that the degree of OGs methylation clearly influences plant defense responses in wild strawberry against *B. cinerea* [13], in tomato against *Ralstonia solanacearum* [11], in wheat against *Puccinia graminis* f. sp. *tritici* [14] and against *B. graminis* f. sp. *tritici* [15]. Indeed, the ability of bacterial or fungal necrotrophs to produce pectin methylesterases (PME) is often related to a successful initiation of the infective process. Pectin is synthesized in a highly methylesterified form and is subsequently de-esterified in muro by PME. De-esterification makes pectin more susceptible to the degradation by pectic enzymes such as endopolygalacturonases and pectate lyases [28]. Wayra and Bari [11] observed in their immunohistochemical studies constitutive differences between tomato genotypes susceptibility to *R. solanacearum* which manifested in methyl-ester distribution of homogalacturonan (HG), arabinan and galactan side chain composition of rhamnogalacturonan I (RG I) and arabinogalactan-protein (AGP) in the xylem parenchyma and in vessel cell walls. They suggested that *R. solanacearum* PME may act on HGs of the susceptible plant in a non-blockwise deesterification pattern, while in the resistant genotype the constitutive, more blockwise methyl-ester distribution and the increased AGP content and higher side chain branching of RG I in vessel cell walls may inhibit easy degradation. However, it is not yet clear how esterification affects OGs biological activity [29].

Gene expression analysis showed no upregulation of the genes involved in the ROS pathway as a response to OGs except for the *GST* gene with OGs – Ac. However, a priming effect in the expression of the *catalase* and *NOS* genes were observed after challenging the OGs-treated plants with *A. euteiches*. Most interestingly, it is known that elicitors such as jasmonic acid, salicylic acid, nitric oxide and superoxides or their precursors do not significantly enhance the resistance induction of pea, and at high concentrations can negatively affect resistance [30]. The induction of the *GST* gene could be explained by its broad spectrum of functions in plants, such as transport and storage of reduced sulfur, detoxification, and antioxidation, as well as a role as cofactor in enzymatic processes, protein reduction, and in phytochelatin by complex-binding heavy metals that have thiol affinity [31,32]. The observed priming induction of the *PGIP* as a response to OGs – Ac indicates the importance of this gene against *A. euteiches*. PGIP are plant extracellular leucine-rich repeat proteins that effectively and specifically bind and inhibit fungal [33] and bacterial [19,20] polygalacturonases and inhibit further invasion of these pathogens. The down-regulation of *PGIP* at early time points after inoculation could be attributed to virulence factors released by the pathogen to suppress the host resistance and facilitate host colonization [34].

Looking at the genes coding for pathogenesis-related proteins, only *DRR276* and *DDR49* genes were upregulated as a response to OGs – Ac. Both the *PR10 DRR49*, *DRR276* pea genes code for a protein homologous with RNase [35,36]. Transgenic ‘Shepody’ potatoes possessing the pea gene *DRR49* displayed resistance to potato early dying disease (*Verticillium dahliae*) [37]. However, the utility of this gene may be related to the close homology of its product to other plant allergens [38].

In parallel, the *chitinase*, *DDR206* and *DDR230* genes were significantly overexpressed as a response to both OGs. The stimulation of *chitinase*, which codes for an enzyme that digests chitin

in the fungus cell wall, could be an important mechanism against *A. euteiches*, which contains 10% of chitin in its wall structure [39]. It has been reported that the pea defense gene *DRR206* confers resistance to black leg (*Leptosphaeria maculans*) disease in transgenic canola (*Brassica napus*) by inhibiting fungal germination and decreasing hyphal growth at inoculation sites [40]. Recently, Seneviratne et al. [41] investigated *in planta* the biochemical function of the *DRR206* and reported that the metabolite associated with its gene induction is the pinorensinol monoglucoside. The pinorensinol is a member of a large, structurally diverse, class of lignans, which have a wide range of physiological and pharmacologically important properties [42,43]. Because of their pronounced biological (antimicrobial, antifungal, antiviral, antioxidant and anti-feedant) properties, a major role of lignans in vascular plants is to apparently help confer resistance against various opportunistic pathogens and predators [41]. The *DDR230* gene that codes for proteins with a high cysteine content, called defensins [44], has been found previously actively expressed in the pea endocarp during the resistance response to *Fusarium solani* f. sp. *phaseoli* [45]. Defensins are antifungal products of some PR genes and one of the first defensin genes cloned, *DRR230*, was isolated from pea [44]. When it was overexpressed in canola, extracts of these plants inhibited the *in vitro* germination of *L. maculans* [40]. Pea defensins have also been used in the biological control of blue mold in apple [46]. The mode of action of pea defensins corresponds to that of similar highly-conserved antimicrobial peptides present in a broad range of biological organisms [47] and could consist of inducing the membrane destabilization/permeabilization required for fungal growth, inhibiting protein synthesis, enzyme activity, and ion channels [48].

The earlier induction of the *PAL* gene, marker of the phenylpropanoid and phytoalexin pathway, in pea roots infected with *A. euteiches* indicates the importance of this pathway in pea defense reactions. The preventive induction of this pathway has previously been reported as one of the important strategies to control root legume diseases [49]. Before inoculation with *A. euteiches*, a high and early induction (3 hat) of *PAL* was observed in pea roots as a response to both OGs elicitors, with higher levels with OGs – Ac. After root inoculation with *A. euteiches*, the induction of *PAL* as a response to both OGs was primed to be significantly higher than its levels in inoculated non-treated plants. This priming effect was observed during the first 4 dai. In fact, *PAL* enzyme catalyzes the first step in the phenylpropanoid pathway toward the biosynthesis of a large variety of products, including antimicrobial phytoalexin compounds such as pisatins, antioxidant protectants such as flavonoid compounds, and precursors of lignin [50–52]. Pisatin production is dependent on *PAL* and a series of other secondary enzymes, such as *TCAH*, *CHS*, *CHI*, and *IFR* [49,53]. At the inoculation time, all of these genes were significantly upregulated as a response to OGs – Ac pretreatment. Previous investigations reported that multiple genes control the pathogenicity of fungal isolates on pea and of these, the gene for pisatin demethylase enzyme (*PDA*) [54], which detoxifies the phytoalexin pisatin [55], is considered the most important. It has also been reported that all isolates without the *PDA* gene were essentially non-pathogenic on peas [30,56] showing the importance of the pisatin pathway in the pea defense against pathogen attack. Interestingly, both pinorensinol monoglucoside and pisatin were found co-localized in pea pod endocarp epidermal cells and associated with *CHS* and *DRR206* gene expression, indicating that both pisatin and pinorensinol monoglucoside function in the overall phytoalexin responses [41].

In conclusion, acetylated and nonacetylated oligogalacturonides confer protection in pea against *A. euteiches* root rot. Acetylation allows a significant reduction of the efficient elicitor dose of OGs, suggesting that acetylation is necessary for some specific responses. The induction of the antifungal defensins, lignans and the phytoalexin pisatin pathways and their priming effect in the expression of the basal defense, *SA* and *ROS* gene markers could explain their stable and synergetic protection efficiency. Taken together, acetylated OGs are interesting elicitors to stimulate defense mechanisms in pea.

4. Materials and Methods

4.1. Elicitor Compounds

OGs were produced at the Laboratoire des Polysaccharides Microbiens et Végétaux (Université Jules Verne, Amiens, France). A mixture of OGs was obtained by thermal degradation of polygalacturonic acid from citrus fruit following the same methods described previously in Randoux et al. (2010). OGs with polymerization degrees (DPs) of 2 to 25 were selected by sequences of purification using acetic acid and isopropanol and then mixed together. This mixture of OGs is hereafter referred to as the nonacetylated OGs (OGs – Ac). Dried OGs – Ac were acetylated using acetic anhydride. After addition of H₂O, the preparation was dialyzed and the acetylated galacturonides were freeze-dried. Samples were then dissolved in D₂O, 99.96% D; the final concentration was 15 g L^{−1}. The degree of acetylation was calculated by integration of the signals in the downfield, upfield, and acetyl regions as described in the literature [57] and OGs with a degree of substitution (DS) of 30% (OGs + Ac) were used in further experiments. OGs + Ac were characterized by the presence of acetyl groups linked on either the C2 or C3 of galacturonan residues, as described in [15]. For all the esterification procedures applied, the DS and the distribution of acetyl groups on the galacturonan residue were always the same.

4.2. Plant Material and Growth Conditions

Pisum sativum L. commercial cv. Alezan, highly susceptible to pea root rot caused by *Aphanomyces euteiches*, was used in the experiments. The seeds were sterilized by immersing them in 70% ethanol alcohol for 5 min and then in a solution of 1% NaOCl for 15 min with three intervals of washing in sterilized distilled water. Plants were grown from seeds in 0.52 L pots (9 × 9 × 8 cm) filled with autoclaved vermiculite in a growth chamber at 20 ± 2 °C and under a photoperiod of 16 h daylight with a light intensity of 150 μmol m^{−2} s^{−1} photon flux density supplied by high-output white fluorescent tubes (Philips Master Cool White 80 W//865, Lamotte Beuvron, France). Plants were irrigated daily and once a week with 25% Murashige and Skoog nutritive solution (Murashige and Skoog medium, Sigma-Aldrich, Saint Louis, Mo, USA).

4.3. Inoculum Preparation and Inoculation

A. euteiches was cultured on corn meal agar medium (Sigma-Aldrich, St. Quentin Fallavier, France) at 18 °C in the dark for three days. Then, zoospores of *A. euteiches* were produced in a mineral salt solution as described by Carman and Lockwood [58]. At 5 weeks post-germination, 25 mL of water containing 10⁵ zoospores was added to each pot on top of the vermiculite. Control plants were irrigated with the same amount of water without zoospores. Three, seven, ten and fourteen days after inoculation, plant roots were harvested and colored with lactophenol cotton blue stain for microscopic observations, or conserved at −80 °C until DNA extraction to follow the disease progression using qPCR. *P. sativum*-*A. euteiches* compatibility was evaluated at 7, 10 and 14 dai using the DSI ratings from 1 to 5 as follows: 1 = no necrosis of roots and hypocotyls; 2 = slight necrosis of roots and hypocotyls; 3 = necrosis of roots and lower hypocotyls, slight chlorosis of cotyledons, and moderate stunting of stem; 4 = extensive necrosis of roots, hypocotyls, cotyledons, and severe stunting of stem; 5 = plant death. In the same samples, the percentage of roots containing more than 50 oospores was determined in samples of one hundred 1-cm root fragments per condition.

4.4. Protection Assay

All tested elicitors were dissolved in water. OGs – Ac and OGs + Ac elicitor solutions were prepared at 1 and 5 g L^{−1}. Solutions of SA (Sigma-Aldrich, St. Quentin Fallavier, France), Chitosan® (Sigma-Aldrich, St. Quentin Fallavier, France), Iodus® (Goëmare, Saint Malo, France) at 1 g L^{−1} were tested. An inoculum of *A. euteiches* (AE) zoospores (10⁵ zoospores mL^{−1}), heated at 100 °C for 10 min to kill the zoospores, was also tested for its elicitor activity. Plants were injected with each

elicitor compound or with water for the control plants on the upper one or two proximal rachises (20 µL/rachis). Different elicitor doses were used: 20, 40, 80, 200 and 400 µg/plant. Depending on the date of elicitor injection, three timing modalities were carried out: at 3-weeks, at 4-weeks, or twice at 3- and 4-weeks post-germination. The plants were then inoculated at 5-weeks-old as mentioned above. Three plant control modalities were used: injected with water and inoculated; non-injected and inoculated; non-injected and non-inoculated. The roots from controls and treated plants were harvested at 7, 10 and 14 dai for disease observation and at 3, 6, 12, 24, 48, 96, 168 and 336 hours after treatment (hat) and at the same hours after inoculation (hai) for gene expression studies.

4.5. DNA and RNA Extraction

For *A. euteiches* DNA quantification and gene expression experiments, roots were harvested and stored immediately in liquid nitrogen and subsequently used for DNA and RNA extraction. Total DNA and RNA were isolated from *P. sativum* roots with the DNeasy and RNeasy plant mini kits (Qiagen, Les Ulis, France), respectively, in accordance with the manufacturer's recommendations. DNA and RNA concentrations and qualities were evaluated using absorption values at 260 and 280 nm, and RNA quality was also checked by gel electrophoresis.

4.6. Real-Time PCR

4.6.1. Real-Time Quantitative PCR (qPCR)

To quantify infection levels of *A. euteiches*, primers and TaqMan minor groove binder probes (Table 1) were designed, using the Primer Express 3 software (Applied Biosystems, Foster, CA, USA), to target a 61-bp fragment of the *A. euteiches* specific gene (GenBank accession No. AF228037.1 [59]). A TaqMan assay was carried out in 25 µL of a reaction mixture containing the following: 12.5 µL of universal TaqMan PCR Master Mix (Applied Biosystems, Foster, CA, USA), 0.3 µM of each primer, 0.2 µM of probe, 200 ng of DNA and water up to a volume of 25 µL. The conditions of qPCR amplification were the following: 10 min at 95 °C, followed by 40 cycles of 15 seconds at 95 °C and 1 min at 60 °C. qPCR analysis of the *A. euteiches* specific gene was calibrated from 10² to 10⁷ copies by serial dilution of the appropriate cloned target sequence.

Table 1. Oligonucleotide primer sequences of wheat defense genes

Gene Name	GenBank Accession N°	Forward, Reverse Primers and Probes (5'–3')	T _m (°C)	Amplicon Length
<i>Aphanomyces euteiches</i>	AF228037	TTTTGGAACACCCAAACGTACTG AGTCCAAGAGGCATTGACAA ACGCTGAGCTTGAC	58 58 68	61
Housekeeping genes				
GAPDH (Mtgap1) ¹	X73150	GTCTTTGCACACAGGAACCCA GGCACCACCTTCAAATGAG CCCATGGGCCAGCAC	59 59 70	123
Defense and cell rescue				
Pathogenesis protein 1 (PR1)	AJ586324	CCTTCCCCTCATGGCTATCC TGTGGTGAGTTTGTGACATATGAGA AGTACTATCCACATCAACAC	59 59 68	69
Proline-rich protein (PRP)	AJ233399	TGGCTTCCTTAACCTTCTACTGT TTGGCAAACCTTGAGGAAT ACTCCTTCTTGCTCTTAT	58 58 68	64
Mitogen-activated protein kinase (MAPK)	X70703	CATTCCGCGAATGTTTTC TTGGCGTTCAGGAGAAGGTT AGGGACTTAAACCC	58 58 70	59

Table 1. Cont.

Gene Name	GenBank Accession N°	Forward, Reverse Primers and Probes (5'–3')	T _m (°C)	Amplicon Length
Reactive oxygen species (ROS)				
Superoxide dismutase (SOD)	AB087845	CCATCATAGGAAGGGCTGTTGT CGTGACCACCTTTCCAAGA CCATGCCGATCCTGAT	59 59 70	63
Peroxidase (POX)	AB193816	ATGCAAGAACAGCAAGCCAAA GGGTGCAAGGTCAGATGATG AACAGTCAAATCCC	59 59 70	69
Nitric oxide synthase (NOS)	AY672712	GGCGGTGGTCAGGGTCTT CCCTTTGGGACACGCTTTT TGGAAGAATGGATCTATT	59 59 68	63
Glutathione S-transferase (GST)	AB087837	GAGAATGCCCTTGGTAAATTTGA ACGCAATATCCACCAAACTGAAT CCCTTCCTTCTTGGTC	58 58 69	70
Catalase (Cat)	X60169	CCAAGTGGTCTCACCACAACAAT TGACCTCCTCATCCTGTGAA CCATGAGGGTTTCATG	59 59 69	69
Metallothionein (Metal)	AB176564	TCCGGCGAAGATCCAGTTT CCACACTTGCAGCCACCAT TGGTGCTGAAATGAGTG	59 59 69	69
Cell wall proteins & Basal defense				
Chitinase (Chit)	L37876	CCTTCAAGACCGCTTATGGTT ACGTCGTGGCAGGATGGTT ACGCCTCAGTCACCT	58 60 68	64
Beta-1,3-glucanase (β 1,3-Glu)	S51479	TGGAATTGGTTGGGTGAATGT TTGCAGAGCCTCCATCTGAA TTGTTTCTGAGAGTGTTG	58 58 68	65
Polygalacturonase inhibiting protein (PGIP)	AB087839	CAGTGCTTTTCGGGAGCAA CAAACGACAGCAAGTTCCTTGA AAAGGACACAGATACTTGAT	59 59 69	66
JA signaling pathways				
Lipoxygenase (LOX)	X17061	TGATCCGCGGTCTTCAAGAG CACCGTATCTGCGGGATCT TTCTCCGAAAAGC	59 59 69	60
Phenylpropanoid & Phytoalexin pathway				
Phenylalanine ammonia lyase (PAL)	D10001	GCACTTAGAACTTACCAGCAATG GAAAGTTTCCACCATGCAAAGC CCCTTTGATTGATGTTTC	60 60 69	
Cinnamate-4-hydroxylase (C4H)	U29243	GCCATAACCGCCATCACAAT GGGCCAGGAGGGAGTTTGAA AACTCCGCGGCAAA	59 59 68	61
Chalcone isomerase (CHI)	U03433	GCTGCAGCATCCTCCATCA CACCGCTGGGAATCATGT CGCAATCCAGTCGAG	58 58 67	56
Chalcone synthase (CHS)	D10662	GACATGGTGGTCTCGAGGTA GCCCCATCTTTTATAGCTTTC AGACTAGGGAAAGAGGCT	58 58 70	70
Isoflavone reductase (CHR)	S72472	CTTTTGGCGTTGTACCATTCG TCTTTGGCAGGTCAATCTCA AACAAATAAAGGGAGATGCAG	59 59 70	68
Disease resistance response (DRR)				
DRR230	AJ308155	TTGCAGGAACAACGAGCACTT GCACCAGCAGCGAAAAATCAT CTCAGTGGGAGGTGCA	60 60 69	61
DRR276	M18249	TGCTGACACTCTTACTCCAAAGGT CCGTTTCCTTCAACAATTTCG TTGATGCCATCAAAAGTA	58 58 69	66
Disease resistance response protein (DRR)				
DRR49	X13383	GGTGATGCTGCTCCTAGTGAAGA CTTGAAAAGACCATCCCCCTTA CAACTCAAGACTGACAAAAG	58 58 68	66
DRR206	M18250	GCTGGAGCTGACCCAATTGT AAGAAATCTCCAGTACCACAGTGA CCAAAAGTAGAGATATTCT	59 59 69	68

¹ The GAPDH (glyceraldehyde-3-phosphate dehydrogenase) genes were used as an internal reference control for equivalent amplification in the PCR.

4.6.2. Real-Time Reverse Transcription PCR (RT-PCR)

The cDNAs were prepared as follows; 1 µg of total RNA was added to 1.5 µg of oligo(dT)₁₅-dNTP (2.5 mM each) and made up to a final volume of 11.5 µL with sterile distilled water. RNA was denatured for 5 min at 70 °C and placed on ice, and then 5 µL of Moloney murine leukemia virus (MMLV) 5× reaction buffer, 300 U of MMLV reverse transcriptase, and 80 U of RNase inhibitor were added. First-strand cDNA was synthesized at 25 °C for 15 min, followed by incubation for 50 min at 42 °C and 2 min at 96 °C. Then, gene-specific fragments were amplified by real-time PCR using the defense gene specific primers and probes listed in Table 1, which were designed using the Primer Express 3 software. The TaqMan assays were carried out as mentioned above. All PCR experiments were carried out using an ABI PRISM 7300 sequence detection system (Applied Biosystems, Foster, CA, USA). The *Mtgap1* gene was used as an internal reference control for equivalent reverse transcription to cDNA and equivalent amplification in the PCR. Expression ratio for each cDNA was calculated relatively to corresponding controls, injected with water, using the $2^{-\Delta\Delta C_t}$ method as described by Livak et al., 2001 [60].

4.7. Statistical Analyses

Five technical repetitions were used for each experimental condition and four separate experiments were carried out. For all experiments, significant differences were evaluated using the Tukey test at $p \leq 0.05$.

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References

1. Cervone, F.; Hahn, M.G.; DeLorenzo, G.; Darvill, A.; Albersheim, P. Host–pathogen interactions: XXXIII. A plant protein converts a fungal pathogenesis factor into an elicitor of plant defense responses. *Plant Physiol.* **1989**, *90*, 542–548. [[CrossRef](#)] [[PubMed](#)]
2. Albersheim, P.; Darvill, A.; Augur, C.; Cheong, J.J.; Eberhard, S.; Hahn, M.G.; Marfa, V.; Mohnen, D.; O'Neill, M.A.; Spiro, M.D.; et al. Oligosaccharins: Oligosaccharide regulatory molecules. *Acc. Chem. Res.* **1992**, *25*, 77–83. [[CrossRef](#)]
3. Côté, F.; Ham, K.S.; Hahn, M.; Bergmann, C.W.; Côté, F.; Ham, K.S.; Hahn, M.G.; Bergmann, C.W. Oligosaccharide elicitors in host–pathogen interactions. Generation, perception and signal transduction. *Subcell. Biochem.* **1998**, *29*, 385–432. [[PubMed](#)]
4. Côté, F.; Hahn, M.G. Oligosaccharins: Structures and signal transduction. *Plant Mol. Biol.* **1994**, *26*, 1379–1411. [[CrossRef](#)] [[PubMed](#)]
5. Braccini, I.; Perez, S. Molecular basis of Ca(2+)-induced gelation in alginates and pectins: The egg-box model revisited. *Biomacromolecules* **2001**, *2*, 1089–1096. [[CrossRef](#)] [[PubMed](#)]
6. Cabrera, J.C.; Boland, A.; Messiaen, J.; Cambier, P.; Van Cutsem, P. Egg box conformation of oligogalacturonides: The time-dependent stabilization of the elicitor-active conformation increases its biological activity. *Glycobiology* **2008**, *18*, 473–482. [[CrossRef](#)] [[PubMed](#)]
7. Farmer, E.E.; Ryan, C.A. Inter plant communication: Airborne methyl jasmonate induces expression of protease inhibitor genes in plant leaves. *Proc. Natl. Acad. Sci. USA* **1990**, *87*, 7713–7716. [[CrossRef](#)] [[PubMed](#)]
8. Moloshok, T.; Pearce, G.; Ryan, C.A. Oligouronide signaling of proteinase inhibitor genes in plants: Structure-activity relationships of di- and trigalacturonic acids and their derivatives. *Arch. Biochem. Biophys.* **1992**, *294*, 731–734. [[CrossRef](#)]
9. Moerschbacher, B.M.; Mierau, M.; Graefner, B.; Noll, U.; Mor, A.J. Small oligomers of galacturonic acid are endogenous suppressors of disease resistance reactions in wheat leaves. *J. Exp. Bot.* **1999**, *50*, 605–612. [[CrossRef](#)]

10. Wydra, K.; Beri, H. Structural changes of homogalacturonan, rhamnogalacturonan I and arabinogalactan protein in xylem cell walls of tomato genotypes in reaction to *Ralstonia solanacearum*. *Physiol. Mol. Plant Pathol.* **2006**, *68*, 41–50. [[CrossRef](#)]
11. Wydra, K.; Beri, H. Immunohistochemical changes in methyl-ester distribution of homogalacturonan and side chain composition of rhamnogalacturonan I as possible components of basal resistance in tomato inoculated with *Ralstonia solanacearum*. *Physiol. Mol. Plant Pathol.* **2007**, *70*, 13–24. [[CrossRef](#)]
12. Diogo, R.; Wydra, K. Silicon-induced basal resistance in tomato against *Ralstonia solanacearum* is related to modification of pectic cell wall polysaccharide structure. *Physiol. Mol. Plant Pathol.* **2007**, *70*, 120–129. [[CrossRef](#)]
13. Osorio, S.; Castillejo, C.; Quesada, M.A.; Medina-Escobar, N.; Brownsey, G.J.; Suau, R.; Heredia, A.; Botella, M.A.; Valpuesta, V. Partial demethylation of oligogalacturonides by pectin methyl esterase 1 is required for eliciting defence responses in wild strawberry (*Fragaria vesca*). *Plant J.* **2008**, *54*, 43–55. [[CrossRef](#)] [[PubMed](#)]
14. Wiethölter, N.; Graessener, B.; Mierau, M.; Mort, A.J.; Moerschbacher, B.M. Differences in the methyl ester distribution of homogalacturonans from near-isogenic wheat lines resistant and susceptible to the wheat stem rust fungus. *Mol. Plant. Microbe Interact.* **2003**, *16*, 945–952. [[CrossRef](#)] [[PubMed](#)]
15. Randoux, B.; Renard-Merlier, D.; Mulard, G.; Rossard, S.; Duyme, F.; Sanssené, J.; Courtois, J.; Durand, R.; Reignault, P. Distinct defenses induced in wheat against powdery mildew by acetylated and nonacetylated oligogalacturonides. *Phytopathology* **2010**, *100*, 1352–1363. [[CrossRef](#)] [[PubMed](#)]
16. Boller, T.; Felix, G. A renaissance of elicitors: Perception of microbe-associated molecular patterns and danger signals by pattern-recognition receptors. *Annu. Rev. Plant Biol.* **2009**, *60*, 379–406. [[CrossRef](#)] [[PubMed](#)]
17. De Lorenzo, G.; Brutus, A.; Savatin, D.; Sicilia, F.; Gervone, F. Engineering plant resistance by constructing chimeric receptors that recognize damage-associated molecular patterns (DAMPs). *FEBS Lett.* **2011**, *585*, 1521–1528. [[CrossRef](#)] [[PubMed](#)]
18. Ranf, S.; Eschen-Lippold, L.; Pecher, P.; Lee, J.; Scheel, D. Interplay between calcium signalling and early signalling elements during defence responses to microbe- or damage-associated molecular patterns. *Plant J.* **2011**, *68*, 100–113. [[CrossRef](#)] [[PubMed](#)]
19. Wydra, K.; Beri, H.; Schacht, H. Polygalacturonase-inhibiting protein (PGIP) and structure and composition of cell wall polysaccharides of tomato in relation to resistance to *Ralstonia solanacearum*. In *Emerging Trends in Plant-Microbe Interactions*; Gananamanickam, S., Balasubramanian, R., Anand, N., Eds.; Centre for Advanced Studies in Botany: Chennai, India, 2005; pp. 217–223.
20. Schacht, T.; Unger, C.; Pich, A.; Wydra, K. Endo- and exopolygalacturonases of *Ralstonia solanacearum* are inhibited by polygalacturonase-inhibiting protein (PGIP) activity in tomato stem extracts. *Plant Physiol. Biochem.* **2011**, *49*, 377–387. [[CrossRef](#)] [[PubMed](#)]
21. Davis, K.R.; Darvill, A.G.; Alber-sheim, P.; Dell, A. Host-pathogen interactions: XXIX. Oligogalacturonides released from sodium polypectate by endopolygalacturonic acid lyase are elicitors of phytoalexins in soybean. *Plant Physiol.* **1986**, *80*, 568–577. [[CrossRef](#)] [[PubMed](#)]
22. Davis, K.R.; Hahlbrock, K. Induction of defense responses in cultured parsley cells by plant cell wall fragments. *Plant Physiol.* **1987**, 1286–1290. [[CrossRef](#)]
23. Broekaert, W.F.; Pneumas, W.J. Pectic polysaccharides elicit chitinase accumulation in tobacco. *Plant Physiol.* **1988**, *74*, 740–744. [[CrossRef](#)]
24. Rasul, S.; Dubreuil-Maurizi, C.; Lamotte, O.; Koen, E.; Poinssot, B.; Alcaraz, G.; Wendehenne, D.; Jeandroz, S. Nitric oxide production mediates oligogalacturonide-triggered immunity and resistance to *Botrytis cinerea* in *Arabidopsis thaliana*. *Plant Cell Environ.* **2012**, *35*, 1483–1499. [[CrossRef](#)] [[PubMed](#)]
25. Cannesan, M.A.; Gangneux, C.; Lanoue, A.; Giron, D.; Laval, K.; Hawes, M.; Driouich, A.; Vire-Gibouin, M. Association between border cell responses and localized root infection by pathogenic *Aphanomyces euteiches*. *Ann. Bot.* **2011**, *108*, 459–469. [[CrossRef](#)] [[PubMed](#)]
26. Shang, H.; Grau, C.R.; Peters, R.D. Oospore germination of *Aphanomyces euteiches* in root exudates and on the rhizoplanes of crop plants. *Plant Dis.* **2000**, *84*, 994–998. [[CrossRef](#)]
27. Baldauf, S.L.; Roger, A.J.; Wenk-Siefert, J.; Doolittle, W.F. A kingdom level phylogeny of eukaryotes based on combined protein data. *Science* **2000**, *290*, 972–977. [[CrossRef](#)] [[PubMed](#)]

28. Limberg, G.; Korner, R.; Buchholt, H.C.; Christensen, T.M.; Roepstorff, P.; Mikkelsen, J.D. Analysis of different de-esterification mechanisms for pectin by enzymatic fingerprinting using endopolygalacturonase II from *A. niger*. *Carbohydr. Res.* **2000**, *327*, 293–307. [[CrossRef](#)]
29. Ferrari, S. Oligogalacturonides: Plant damage-associated molecular patterns and regulators of growth and development. *Front. Plant Sci.* **2013**, *4*, 49. [[CrossRef](#)] [[PubMed](#)]
30. Hadwiger, L.A. Pea-*Fusarium solani* interactions contributions of a system toward understanding disease resistance. *Phytopathology* **2008**, *98*, 372–379. [[CrossRef](#)] [[PubMed](#)]
31. Rennenberg, H. Glutathione metabolism and possible biological roles in the higher plant. *Phytochemistry* **1982**, *21*, 2771–2781. [[CrossRef](#)]
32. Smith, I.K.; Polle, A.; Rennemerg, H. Glutathione. In *Stress Responses in Plants: Adaptation and Acclimation Mechanisms*; Alschér, R.G., Cumming, J.R., Eds.; Wiley & Liss: New York, NY, USA, 1990; pp. 201–215.
33. Liu, N.; Zhang, X.; Sun, Y.; Wang, P.; Li, X.; Pei, Y.; Li, F.; Hou, Y. Molecular evidence for the involvement of a polygalacturonase-inhibiting protein, GhPGIP1, in enhanced resistance to Verticillium and Fusarium wilts in cotton. *Sci. Rep.* **2017**, *7*, 39840. [[CrossRef](#)] [[PubMed](#)]
34. Ghareeb, H.; Bozsó, Z.; Ott, P.G.; Repenning, C.; Stahl, F.; Wydra, K. Transcriptome of silicon-induced resistance against *Ralstonia solanacearum* in the silicon non accumulator tomato implicates priming effect. *Physiol. Mol. Plant Pathol.* **2011**, *75*, 83–89. [[CrossRef](#)]
35. Moiseyev, G.P.; Beintema, J.J.; Fedoreyeva, L.I.; Yakovlev, J.I. High sequence similarity between a ribonuclease from Ginseng calluses and fungus-elicited proteins from parsley indicates that intracellular pathogenesis-related proteins are ribonucleases. *Planta* **1994**, *193*, 470–472. [[CrossRef](#)] [[PubMed](#)]
36. Riggelman, R.D.; Fristensky, B.; Hadwiger, L.A. The disease resistance response in pea is associated with increased levels of specific mRNAs. *Plant Mol. Biol.* **1985**, *4*, 81–86. [[CrossRef](#)] [[PubMed](#)]
37. Chang, M.M.; Chiang, C.C.; Martin, M.W.; Hadwiger, L.A. Expression of a pea disease resistance response gene in the potato cultivar shepody. *Am. Potato J.* **1993**, *70*, 635–647. [[CrossRef](#)]
38. Breiteneder, H.; Pettenburger, K.; Bito, A.; Valenta, R.; Kraft, D.; Rumpold, H.; Scheiner, O.; Breitenbach, M. The gene coding for the major birch pollen allergen Betv 1 is highly homologous to a pea disease resistance response gene. *EMBO J.* **1989**, *8*, 1935–1938. [[PubMed](#)]
39. Badreddine, I.; Lafitte, C.; Heux, L.; Skandalis, N.; Spanou, Z.; Martinez, Y.; Esquerre-Tugaye, M.-T.; Bulone, V.; Dumas, B.; Bottin, A. Cell wall chitosaccharides are essential components and exposed patterns of the phytopathogenic oomycete *Aphanomyces euteiches*. *Eukaryot. Cell* **2008**, *7*, 1980–1993. [[CrossRef](#)] [[PubMed](#)]
40. Wang, Y.; Nowak, G.; Culley, D.; Hadwiger, L.A.; Fristensky, B. Constitutive expression of pea defense gene DRR206 confers resistance to blackleg (*Leptosphaeria maculans*) disease in transgenic canola (*Brassica napus*). *Mol. Plant. Microbe Interact.* **1999**, *12*, 410–418. [[CrossRef](#)]
41. Seneviratne, H.K.; Dalisay, D.S.; Kim, K.-W.; Moinuddin, S.G.A.; Yang, H.; Hartshorn, C.M.; Davin, L.B.; Lewis, N.G. Non-host disease resistance response in pea (*Pisum sativum*) pods: Biochemical function of DRR206 and phytoalexin pathway localization. *Phytochemistry* **2015**, *113*, 140–148. [[CrossRef](#)] [[PubMed](#)]
42. Chu, H.D.; Le, Q.N.; Nguyen, H.Q.; Le, D.T. Genome-wide analysis of genes encoding methionine-rich proteins in *Arabidopsis* and Soybean suggesting their roles in the adaptation of plants to abiotic stress. *Int. J. Genomics* **2016**, *2016*, 1–8. [[CrossRef](#)] [[PubMed](#)]
43. Vassão, D.G.; Kim, K.W.; Davin, L.B.; Lewis, N.G. Lignans (neolignans) and allyl/propenyl phenols: Biogenesis, structural biology, and biological/human health considerations. In *Comprehensive Natural Products Chemistry II, Structural Diversity I*; Townsend, C., Ebizuka, Y., Eds.; Elsevier: Oxford, UK, 2010; Volume 1, pp. 815–928.
44. Chiang, C.C.; Hadwiger, L.A. The *Fusarium solani*-induced expression of a pea gene family encoding high cysteine content proteins. *Mol. Plant. Microbe Interact.* **1991**, *4*, 324–331. [[CrossRef](#)] [[PubMed](#)]
45. Fristensky, B.; Riggelman, C.R.; Wagoner, W.; Hadwiger, L.A. Gene expression in susceptible and disease resistant interactions of peas induced with *Fusarium solani* pathogens and chitosan. *Physiol. Plant Pathol.* **1985**, *27*, 15–28. [[CrossRef](#)]
46. Janisiewicz, W.J.; Pereira, I.B.; Almeida, M.S.; Roberts, D.P.; Wisniewski, M.E.; Kurtenbach, E. Biocontrol activity of recombinant *Pichia pastoris* constitutively expressing pea defensin against blue mold of apple. (Abstr.). *Phytopathology* **2006**, *96*, S53.
47. Selitrennikoff, C.P. Antifungal Proteins. *Appl. Environ. Microbiol.* **2001**, *63*, 834–839. [[CrossRef](#)] [[PubMed](#)]

48. Lay, F.T.; Anderson, M.A. Defensins—Components of the innate immune system in plants. *Curr. Protein Pept. Sci.* **2005**, *5*, 85–101. [[CrossRef](#)]
49. Selim, S.; Negrel, J.; Wendehenne, D.; Ochatt, S.; Gianinazzi, S.; van Tuinen, D. Stimulation of defense reactions in *Medicago truncatula* by antagonistic lipopeptides from *Paenibacillus* sp. strain B2. *Appl. Environ. Microbiol.* **2010**, *76*, 7420–7428. [[CrossRef](#)] [[PubMed](#)]
50. Lamb, C.; Dixon, R.A. The oxidative burst in plant disease resistance. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **1997**, *48*, 251–275. [[CrossRef](#)] [[PubMed](#)]
51. Jeandet, P.; Clément, C.; Courot, E.; Cordelier, S. Modulation of phytoalexin biosynthesis in engineered plants for disease resistance. *Int. J. Mol. Sci.* **2013**, *14*, 14136–14170. [[CrossRef](#)] [[PubMed](#)]
52. Jeandet, P.; Hébrard, C.; Deville, M.A.; Crouzet, J. Deciphering the role of phytoalexins in plant-microorganism interactions and human health. *Molecules* **2014**, *19*, 18033–18056. [[CrossRef](#)] [[PubMed](#)]
53. Wu, Q.; Van Etten, H.D. Introduction of plant and fungal genes into pea (*Pisum sativum* L.) hairy roots reduces their ability to produce pisatin and affects their response to a fungal pathogen. *Mol. Plant. Microbe Interact.* **2004**, *17*, 798–804. [[CrossRef](#)] [[PubMed](#)]
54. Maloney, A.P.; Van Etten, H.D. A gene from the fungal pathogen *Nectria haematococca* that encodes the phytoalexin-detoxifying enzyme pisatin demethylase defines a new cytochrome P450 family. *Mol. Genet. Genomics* **1994**, *243*, 506–514. [[CrossRef](#)]
55. Delserone, L.M.; McCluskey, K.D.; Mathews, E.; Van Etten, H.D. Pisatin demethylation by fungal pathogens and nonpathogens of pea: Association with pisatin tolerance and virulence. *Physiol. Mol. Plant Pathol.* **1999**, *55*, 317–326. [[CrossRef](#)]
56. Mackintosh, S.F.; Matthews, D.E.; Van Etten, H.D. Two additional genes for pisatin demethylation and their relationship to the pathogenicity of *Nectria haematococca* on pea. *Mol. Plant. Microbe Interact.* **1989**, *2*, 354–362. [[CrossRef](#)]
57. Courtois, J.; Seguin, J.P.; Roblot, C.; Heyraud, A.; Gey, C.; Dantas, L.; Barbotin, J.N.; Courtois, B. Exopolysaccharide production by the *Rhizobium meliloti* M5N1CS strain. Location and quantification of the sites of *O*-acetylation. *Carbohydr. Polym.* **1994**, *25*, 7–12. [[CrossRef](#)]
58. Carman, L.M.; Lockwood, J.L. Factors affecting zoospore production by *Aphanomyces euteiches*. *Phytopathology* **1959**, *49*, 535.
59. Vandemark, G.J.; Kraft, J.M.; Larsen, R.C.; Gritsenko, M.A.; Boge, W.L. A PCR-based assay by sequence-characterized DNA markers for the identification and detection of *Aphanomyces euteiches*. *Phytopathology* **2000**, *90*, 1137–1144. [[CrossRef](#)] [[PubMed](#)]
60. Livak, K.J.; Schmittgen, T.D. Analysis of relative gene expression data using real-time quantitative PCR and the $2^{-\Delta\Delta CT}$ method. *Methods* **2001**, *25*, 402–408. [[CrossRef](#)] [[PubMed](#)]

Sample Availability: Samples of the compounds are available from the authors.



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