



Article Facile Chemical Access to Biologically Active Norcantharidin Derivatives from Biomass

Konstantin I. Galkin¹, Fedor A. Kucherov¹, Oleg N. Markov¹, Ksenia S. Egorova¹, Alexandra V. Posvyatenko^{2,3} and Valentine P. Ananikov^{1,*}

- ¹ Zelinsky Institute of Organic Chemistry, Russian Academy of Sciences, Leninsky Prospect 47, 119991 Moscow, Russia; glkn.ioc@gmail.com (K.I.G.); fkucherov@gmail.com (F.A.K.); markoff.o@yandex.ru (O.N.M.); egorova-ks@ioc.ac.ru (K.S.E.)
- ² Institute of Gene Biology, Russian Academy of Sciences, Vavilova Str. 34/5, 119334 Moscow, Russia; sandra.posvjatenko@gmail.com
- ³ Dmitry Rogachev National Medical Research Center of Pediatric Hematology, Oncology and Immunology, Ministry of Health of Russian Federation, Samory Mashela Str., 117198 Moscow, Russia
- * Correspondence: val@ioc.ac.ru; Tel.: +7-499-137-29-44

Received: 9 November 2017; Accepted: 7 December 2017; Published: 12 December 2017

Abstract: Reductive amination of 2,5-diformylfuran (DFF) was used to implement the transition from bio-derived 5-hydroxymethylfurfural (HMF) to pharmaceuticals. The synthesized bis(aminomethyl)furans were utilized as building blocks for the construction of new derivatives with structural cores of naturally occurring biologically active compounds. Using the one-pot procedure, which included the Diels–Alder reaction followed by hydrogenation of the double bond, bio-derived analogues of the anticancer drug norcantharidin were obtained. The cyclization process was diastereoselective, and resulted in the formation of tricyclic products with the endo configuration. Analysis of cytotoxycity for the resulting tricyclic amine-containing compounds showed an increase of anticancer activity as compared with the unsubstituted norcantharimide.

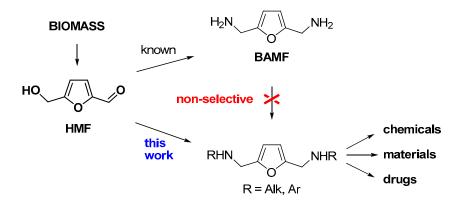
Keywords: biomass; 2,5-diformylfuran; reductive amination; norcantharidin; diastereoselectivity; Diels–Alder reaction

1. Introduction

Plant biomass is the largest renewable source of carbohydrates, with a mass of about 10¹¹ tons of carbon per year [1]. To date, the most promising approach to the synthetic utilization of biomass involves the catalytic conversion of carbohydrates to molecular building blocks, which are often defined as platform chemicals [2,3]. 5-Hydroxymethylfurfural (HMF) is one of the most important platform chemicals, as it has many possibilities, especially in the fields of biofuels [4–8], biomaterials production [9–12], and fine chemical applications [13–15]. In contrast to several other classes of organic compounds, obtaining substituted amines from HMF is quite rare [13–15]. The most important applications are in the fields of medicine and material science. Indeed, the amine functional group is represented in many bioactive compounds, so the functionalization by an amine-containing fragment can improve the pharmacological properties of potential drugs. In addition, primary and secondary HMF-based amines can be used as monomers in the synthesis of bio-derived polyamides [9,16–19].

The synthesis of 2,5-bis(aminomethyl)furan (BAMF), a useful diamine compound, from HMF is well known [16,20,21], but examples of the synthesis of *N*-substituted BAMF derivatives are rarely presented in the literature [22,23]. Possible ways to obtaining such derivatives are the alkylation of BAMF or the alkylation of primary amines by halogenated derivatives of HMF, especially by 5-(chloromethyl)furfural. Such transformations are not practical, since it is impossible to control the

degree of alkylation and the formation of quaternary byproducts. Overall, these limitations lead to non-selective transformations (Scheme 1).



Scheme 1. 2,5-Bis(aminomethyl)furan (BAMF) synthesis from 5-hydroxymethylfurfural (HMF) and route to *N*-substituted BAMF derivatives.

In the present work, we report the reductive amination of 2,5-diformylfuran (DFF) to connect biomass with pharmaceuticals containing an amino function (Scheme 1). For this purpose, we synthesized di- and tri-substituted BAMF derivatives as building blocks for further synthetic applications. The Diels–Alder protocol was used in order to obtain bio-derived analogues of the anticancer drug norcantharidin. Additionally, disubstituted amines were of considerable interest as monomers for the production of new biomaterials [19].

2. Results and Discussion

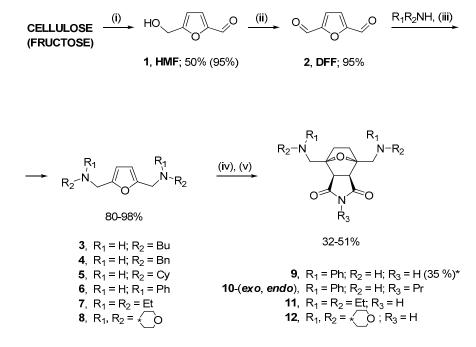
2.1. Synthesis of Norcantharidin Derivatives from HMF

HMF was obtained from natural carbohydrates (cellulose or fructose), and was isolated in a pure crystalline form, as described previously (Scheme 2) [24,25]. DFF was synthesized by oxidation of HMF using a recyclable catalyst [Pip'(O)][BF₄] (4-acetamido-2,2,6,6-tetramethyl-1-oxopiperidinium tetrafluoroborate) in ionic liquid media, according to the previously developed method [26]. In order to obtain amine-based building blocks, the reductive amination of DFF with mono- and disubstituted amines was implemented (Scheme 2). The reaction of DFF with two equivalents of amines, followed by the treatment with sodium triacetoxyborohydride resulted in amines **3–8**, which were obtained with high yields and very good purity of >95%.

The prepared 2,5-bis(aminomethyl)furan derivatives can be further modified at the amine or furan functional groups. The modification of substituents on the furan ring in HMF-based compounds has been well studied [13–15], whereas the chemistry of the furan ring is a challenging problem. For evaluation of the synthetic potential of the furan ring in the prepared amines, we synthesized norcantharimides **9–12**, which were bio-derived analogues of the anticancer drug norcantharidin. We used a modified one-pot protocol based on the Diels–Alder reaction of compounds **6–8** with alkenes, followed by hydrogenation of the formed double bond, as reported recently [27]. The Diels–Alder reactions were performed in THF media at room temperature and monitored by NMR. The second stage was performed by hydrogen addition over 10% Pd/C. Final products were isolated by column chromatography on silica with moderate yields and high purity.

Evaluation of the molecular structure of the formed tricyclic products was carried out using standard 1D-, 2D-NMR, and NOE experiments along with LC-MS data. In comparison with the previous results, the endo selectivity of the Diels–Alder reaction was observed for most of the isolated products. Introduction of the *N*-propyl moiety into maleimide led to the loss of diastereoselectivity of the cycloaddition, possibly due to the steric effects of the *N*-alkyl

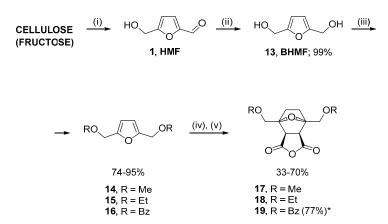
substituent, and the formation of the mixture of **10**-endo and **10**-exo products with a diastereomeric ratio of 4:1, respectively. Both diastereomers were isolated by column chromatography and characterized spectroscopically. The diastereomeric configuration was determined by ¹H-NMR and NOE experiments. After considering the NMR data confirmed by the X-ray analysis of HMF-derived norcantharimide analogues obtained in the previous study [27], the spatial structure of the Diels–Alder adducts was confirmed (see the Supplementary Materials). Chemical shifts of the diastereomeric protons at the articulated position were characterized by a singlet at ~3.5–3.7 ppm in the ¹H-NMR spectra, while the signals of the appropriate protons in the exo products (compounds norcantharimide [28] and **10**-exo) were observed in the stronger magnetic fields at ~2.9–3.1 ppm. Additional confirmation of the stereo configuration of compounds **10**-exo and **10**-endo was accomplished by nuclear Overhauser effect spectroscopy (NOESY) and analysis of the obtained spectral data (see the Supplementary Materials).



Scheme 2. Reductive amination of 2,5-diformylfuran (DFF) for the synthesis of bio-based norcantharimides. (i) $CrCl_3 \cdot 6H_2O$, [BMIM]Cl, 120 °C (H_2SO_4 , [BMIM]Cl, 65 °C); (ii) [Pip'(O)][BF₄], [BMIM]Cl; (iii) NaBH(OAc)₃, CHCl₃; (iv) maleimide, THF; (v) H_2 , 1 atm, 10% Pd/C, THF. *—yield using step-by-step procedure.

Norcantharidins and norcantharimides are synthetic analogues of the natural product cantharidin, which possesses anticancer activity and is of considerable practical interest [29–34]. For comparison of the cytotoxic activity of the prepared amine-containing tricyclic products, we prepared a number of their oxo analogues by the reaction of 2,5-(alkoxymethyl)furans **14–16** with maleic anhydride, followed by the hydrogenation reaction (Scheme 3). The initial furans were prepared by alkylation or acylation of BHMF using known methods (see the Section 3). The preparation of final products **17–19** was performed under the reaction conditions used for the synthesis of the furan–maleimide adducts, and endo selectivity of the cyclization reaction was observed.

The one-pot protocol for the synthesis of the tricyclic products was used because we were unable to isolate the intermediates of the Diels–Alder reaction due to their high polarity and the reversibility of the cycloaddition process. However, two unsaturated products were obtained using Et_2O instead of THF, and were precipitated from the reaction mixture in pure form. The hydrogenation of these products in THF media under standard reaction conditions led to the formation of products **9** and **19**, with an overall yield that was higher than that when using the one-pot procedure (Scheme 3).



Scheme 3. Preparation of norcantharidin analogues. (i) $CrCl_3 \cdot 6H_2O$, [BMIM]Cl, 120 °C (H₂SO₄, [BMIM]Cl, 65 °C); (ii) NaBH₄, water; (iii) alkylation or acylation; (iv) maleic anhydride, THF; (v) H₂, 1 atm, 10% Pd/C, THF. *—yield using step-by-step procedure.

2.2. Biological Activity of Synthesized Compounds

Preliminary testing of the biological activity of selected substances was carried out (Figure 1). According to a cytotoxicity assay in human colorectal adenocarcinoma cell line HT-29, the activity of norcantharidin was in accordance with the published data [35]. Despite a recently reported improvement of anticancer activity of norcantharidin by introduction of *O*-benzoylated hydroxymethyl groups [29], isomeric product **19** and methylated analogue **17** did not show high cytotoxicity towards HT-29 cells. The cytotoxic activity of norcantharimide and its derivatives was low [36], but we found for the first time that it could be improved by introduction of the phenylaminomethyl moiety into the bicyclic core. The activity of aminomethylated norcantharimide **9** towards HT-29 cells was significantly higher than that of norcantharimide (see Figure 1 and the Supplementary Materials for details), but lower than the norcantharidin activity, indicating the necessity of further structural tuning to improve the biological activity.

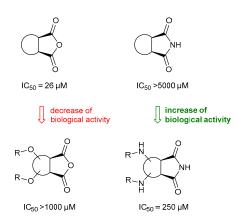


Figure 1. Increasing biological activity of cantharimide by introduction of an amino function (IC₅₀ in HT-29 cells is given in μ M).

3. Materials and Methods

Commercially available reagents and solvents were of analytical grade or were purified prior to use by standard methods. NMR spectra were recorded using a Bruker Fourier 300 HD, Bruker Avance III 400 and Bruker DRX-500 spectrometers (Fällanden, Switzerland) at the following frequencies: 300.1/400.1/500.1 MHz (¹H), 75.5/100.6/125.8 MHz (¹³C). NMR chemical shifts were measured relative to residual protio solvent peaks. Mass spectrometric detection was performed on a high

resolution Bruker maXis instrument equipped with an electrospray ionization (ESI) ion source. Chromatographic separations were performed on silica gel (Kieselgel, 230–400 mesh, Merck Schuchardt, Darmstadt, Germany) with analytical grade solvents. Analytical TLC was performed on Merck silica gel plates with a QF-254 indicator. Visualization was accomplished with UV light or using KMnO₄ in aqueous NaOH or (NH₄)₆Mo₇O₂₄ in ethanolic H₂SO₄, and the TLC was heated until the development of color. Norcantharidin and norcantharimide were obtained from commercial sources.

3.1. Synthesis of Substrates

5-Hydroxymethylfurfural (HMF, 1) was prepared from cellulose or fructose with 50% (45% after recrystallization) or 95% (90% after recrystallization) yields, respectively [24].

- 2,5-Diformylfuran (DFF, 2) was prepared from 1 with 95% yield [26].
- 2,5-Bis(hydroxymethyl)furan (BHMF, 13) was prepared from 1 with 99% yield [37].
- 2,5-Bis(methoxymethyl)furan (14) was prepared from 13 with 74% yield [38].

2,5-Bis(ethoxymethyl)furan (15) was prepared from 13 with 76% yield [27].

Furan-2,5-*diylbis(methylene)dibenzoate* (**16**) was prepared from **13** with 95% yield [**39**]. ¹H-NMR (500 MHz, CDCl₃) δ = 8.06 (d, *J* = 7.8 Hz, 4H), 7.55 (t, *J* = 7.1 Hz, 2H), 7.42 (t, *J* = 7.5 Hz, 4H), 6.49 (s, 2H), 5.31 (s, 4H). ¹³C-NMR (126 MHz, CDCl₃) δ 166.3, 150.4, 133.2, 129.9, 128.5, 111.9, 58.6. HRMS (ESI) Calcd. for C₂₀H₁₆O₅ [M + Na]: 359.0890. Found 359.0875.

3.2. General Method for Synthesis of 2,5-Bis(aminomethyl)furans

A solution of 2,5-diformylfuran **2** (0.5 g, 4 mmol) and the corresponding amine (8.4 mmol) in chloroform (15 mL) was stirred for 20 min, and sodium triacetoxyborohydride (3.4 g, 16 mmol) was added portion-wise. The reaction mixture was stirred overnight, poured into a saturated solution of sodium bicarbonate, and extracted with chloroform (3×7 mL). The combined organic phases were washed with water, dried with sodium sulfate, evaporated, and dried under high vacuum. Target compounds **3–8** were obtained as pale yellow oils.

2,5-*Bis*(*N*-*butylaminomethyl*)*furan* (**3**). Yield 0.89 g, 93%. ¹H-NMR (500 MHz, CDCl₃) δ = 6.05 (s, 2H), 3.71 (s, 4H), 2.58 (t, *J* = 7.2 Hz, 4H), 1.45 (dt, *J* = 14.7, 7.3 Hz, 4H), 1.30 (m, 4H), 0.88 (t, *J* = 7.3 Hz, 6H). ¹³C-NMR (126 MHz, CDCl₃) δ = 153.4, 107.4, 49.0, 46.5, 32.2, 20.5, 14.1. HRMS (ESI) Calcd. for C₁₄H₂₆N₂O [M + H]: 239.2118. Found 239.2129.

2,5-Bis(*N*-benzylaminomethyl)furan (4). Yield 0.99 g, 80%. ¹H-NMR (500 MHz, CDCl₃) δ = 7.34 (d, *J* = 4.4 Hz, 8H), 7.27 (dt, *J* = 6.8, 4.4 Hz, 2H), 6.13 (s, 2H), 3.80 (d, *J* = 13.5 Hz, 8H), 1.73 (s, 2H). ¹³C-NMR (126 MHz, CDCl₃) δ = 153.2, 140.0, 128.5, 128.3, 127.1, 107.7, 52.9, 45.6. HRMS (ESI) Calcd. for C₂₀H₂₂N₂O [M + H]: 307.1805. Found 307.1815.

2,5-*Bis*(*N*-*cyclohexylaminomethyl*)*furan* (5). Yield 1.10 g, 95%. ¹H-NMR (500 MHz, CDCl₃) δ = 6.04 (s, 2H), 3.75 (s, 4H), 2.42 (m, 2H), 1.85 (d, *J* = 10.6 Hz, 4H), 1.71 (d, *J* = 12.9 Hz, 4H), 1.59 (d, *J* = 12.0 Hz, 2H), 1.40 (br. s, 2H), 1.30–1.00 (m, 10H). ¹³C-NMR (126 MHz, CDCl₃) δ = 153.6, 107.2, 55.9, 43.6, 33.5, 26.3, 25.1. HRMS (ESI) Calcd. for C₁₈H₃₀N₂O [M + H]: 291.2431. Found 291.2442.

2,5-Bis(*N*-phenylaminomethyl)furan (6). Yield 1.02 g, 92%. ¹H-NMR (500 MHz, CDCl₃) δ = 7.20 (t, *J* = 7.9 Hz, 4H), 6.76 (t, *J* = 7.3 Hz, 2H), 6.69 (d, *J* = 7.7 Hz, 4H), 6.17 (s, 2H), 4.29 (s, 4H). ¹³C-NMR (126 MHz, CDCl₃) δ = 152.1, 147.5, 129.4, 118.4, 113.5, 108.1, 41.8. HRMS (ESI) Calcd. for C₁₈H₁₈N₂O [M + Na]: 301.1311. Found 301.1322.

2,5-Bis(N,N-diethylaminomethyl)furan (7). Yield 760 mg, 80% [27]. ¹H-NMR (500 MHz, CDCl₃) δ = 6.10 (s, 2H), 3.67 (s, 4H), 2.52 (q, 8H, *J* = 7.2 Hz), 1.08 (t, 12H, *J* = 7.2 Hz). ¹³C-NMR (126 MHz, CDCl₃) δ = 151.4, 109.1, 48.6, 46.9, 12.0. HRMS (ESI) Calcd. for C₁₄H₂₆N₂O [M + H]: 239.2118. Found 239.2118.

2,5-*Bis(morpholinomethyl)furan* (8). Yield 1.04 g, 98% [27]. ¹H-NMR (500 MHz, CDCl₃) δ = 6.13 (s, 2H), 3.70 (m, 8H), 3.51 (s, 4H), 2.45 (m, 8H). ¹³C-NMR (126 MHz, CDCl₃) δ = 150.9, 109.7, 66.8, 55.3, 53.2. HRMS (ESI) Calcd. for C₁₄H₂₂N₂O₃ [M + H]: 267.1703. Found 267.1703.

3.3. General Method for One-Pot Synthesis of Norcantharimide Derivatives

To a solution of 2,5-bis(aminomethyl)furans 6–8 (0.6 mmol) in 2 mL of THF, maleimide (0.12 g, 1.2 mmol, 2 eq.) was added. The reaction mixture was stirred for 24 h at 24 °C. Approximately 20 mg of 10% Pd/C was added, and the reaction mixture was placed under a hydrogen atmosphere (1 atm) for 8 h at 24 °C. The catalyst was filtered off and washed thoroughly with hot THF (3 × 4 mL). The filtrate was evaporated under reduced pressure. The residue was purified by flash chromatography (ethyl acetate as eluent). Target compounds **10–12** were obtained as pale-yellow oils. Diastereomeric products **10**-endo and **10**-exo were formed in a ratio of 4:1 and separated using column chromatography.

endo-4,7-Bis((phenylamino)methyl)-2-propylhexahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione(endo-4,7-bis-(phe nylamino)-2-propyl-norcantharimide) (**10**-endo). Yield 133 mg, 53%. ¹H-NMR (500 MHz, CDCl₃) δ = 7.22 (t, J = 7.8 Hz, 8H), 6.77 (t, J = 7.4 Hz, 12H), 4.37 (br. s, 2H), 3.70 (d, J = 13.3 Hz, 2H), 3.59 (d, J = 13.3 Hz, 2H), 3.53–3.41 (m, 4H), 1.89 (d, J = 7.6 Hz, 2H), 1.76 (d, J = 7.7 Hz, 2H), 1.63 (m, 1H), 0.97 (t, J = 7.4 Hz, 1H). ¹³C-NMR (126 MHz, CDCl₃) δ = 176.2, 148.0, 129.4, 118.1, 113.2, 88.6, 53.4, 47.3, 40.7, 30.0, 21.1, 11.5. HRMS (ESI) Calcd. for C₂₅H₂₉N₃O₃ [M + Na]: 442.2101. Found 442.2114.

exo-4,7-Bis((phenylamino)methyl)-2-propylhexahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione(exo-4,7-bis(phe-nylamino)-2-propyl-norcantharimide) (**10**-exo). Yield 33 mg, 13%. ¹H-NMR (400 MHz, CDCl₃) δ = 7.19 (t, J = 7.9 Hz, 4H), 6.72 (dd, J = 12.5, 7.6 Hz, 6H), 4.21 (s, 2H), 3.69 (q, J = 14.0 Hz, 4H), 3.49 (t, J = 7.1 Hz, 2H), 3.10 (s, 2H), 2.06 (d, J = 7.2 Hz, 2H), 1.76 (d, J = 7.2 Hz, 2H), 1.61 (m, 2H), 0.88 (t, J = 7.4 Hz, 3H). ¹³C-NMR (126 MHz, CDCl₃) δ = 176.1, 148.2, 129.4, 117.9, 113.4, 88.2, 52.1, 45.4, 40.7, 34.2, 21.1, 11.3. HRMS (ESI) Calcd. for C₂₅H₂₉N₃O₃ [M + Na]: 442.2101. Found 442.2111.

endo-4,7-Bis((diethylamino)methyl)hexahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione(endo-4,7-bis(diethylamino)norcantharimide) (11). Yield 103 mg, 51% [27]. ¹H-NMR (500 MHz, CD₃OD) δ = 3.66 (s, 2H), 2.99 (m, 4H), 2.76–2.89 (m, 8H), 1.76–1.85 (m, 4H), 1.12 (t, 12H, J = 7.1 Hz). ¹³C-NMR (126 MHz, CD₃OD) δ = 177.8, 88.6, 54.5, 53.6, 48.4, 29.1, 10.3.

endo-4,7-Bis(morpholinomethyl)hexahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione(endo-4,7-bis(morpholinomethyl)norcantharimide) (12). Yield 93 mg, 42% [27]. ¹H-NMR (500 MHz, CD₃OD) δ = 3.58–3.65 (m, 10H), 2.72 (m, 4H), 2.48–2.66 (m, 8H), 1.69–1.83 (m, 4H). ¹³C-NMR (126 MHz, CD₃OD) δ = 178.2, 89.1, 66.7, 59.8, 54.6, 53.7, 28.9.

3.4. General Method for One-Pot Synthesis of Norcantharidin Derivatives

To a solution of compounds **14–16** (1 mmol) in 2 mL of THF, maleic anhydride (196 mg, 2 mmol, 2 eq.) was added, and the reaction mixture was stirred for 24 h at 24 °C. Approximately 30 mg of 10% Pd/C was added, and the reaction mixture was placed under a hydrogen atmosphere (1 atm) for 12 h at 24 °C. The catalyst was filtered off and washed thoroughly with hot THF (3×4 mL). The filtrate was evaporated under reduced pressure, and the resulting mixture was treated by column chromatography. Target compounds **17–19** were obtained as white solids or light yellow oils.

endo-4,7-Bis(methoxymethyl)hexahydro-4,7-epoxyisobenzofuran-1,3-dione(endo-4,7-bis(methoxymethyl)-nor cantharidin) (17). Yield 43 mg, 17% (33% based on reacted substrate). ¹H-NMR (500 MHz, CDCl₃) δ = 3.86 (d, *J* = 10.6 Hz, 2H), 3.76 (d, *J* = 10.6 Hz, 2H), 3.42 (s, 6H), 3.31 (s, 2H), 2.06 (d, *J* = 7.4 Hz, 2H), 1.71 (d, *J* = 7.5 Hz, 2H). ¹³C-NMR (126 MHz, CDCl₃) δ = 169.3, 88.1, 70.8, 59.7, 52.5, 32.0. HRMS (ESI) Calcd. for C₁₂H₁₆O₆ [M + Na]: 279.0839. Found 279.0836.

endo-4,7-*Bis*(*ethoxymethyl*)*hexahydro*-4,7-*epoxyisobenzofuran*-1,3-*dione*(*endo*-4,7-*bis*(*ethoxymethyl*)*nor*-*canth aridin*) (**18**). Yield 77 mg, 27% (59% based on reacted substrate). ¹H-NMR (500 MHz, CDCl₃) δ = 3.92

(d, *J* = 10.6 Hz, 2H), 3.81 (d, *J* = 10.6 Hz, 2H), 3.61 (m, 4H), 3.31 (s, 2H), 2.08 (q, *J* = 3.5 Hz, 2H), 1.73 (q, *J* = 3.4 Hz, 2H), 1.22 (t, *J* = 7.0 Hz, 6H). ¹³C-NMR (126 MHz, CDCl₃) δ = 169.4, 88.3, 68.8, 67.5, 52.6, 32.2, 15.1. HRMS (ESI) Calcd. for C₁₄H₂₀O₆ [M + H]: 307.1152. Found 307.1148.

endo-1,3-Dioxohexahydro-4,7-epoxyisobenzofuran-4,7-diyl)bis(methylene)dibenzoate(endo-4,7-bis ((O-ben-zoyl)-oxymethyl)-norcantharidin) (**19**). Yield 112 mg, 26% (70% based on reacted substrate). ¹H-NMR (500 MHz, DMSO-d₆) δ = 7.97 (d, J = 7.4 Hz, 4H), 7.68 (t, J = 7.4 Hz, 2H), 7.54 (t, J = 7.6 Hz, 4H), 4.72 (m, 4H), 3.80 (s, 2H), 2.01 (s, 4H). ¹³C-NMR (126 MHz, DMSO-d₆) δ = 170.6, 165.1, 133.6, 129.3, 128.8, 86.6, 62.4, 52.9, 31.2. HRMS (ESI) Calcd. for C₂₄H₂₀O₈ [M + H]: 459.1050. Found 459.1040.

3.5. Synthesis of Compounds 9 and 19 Using Step-by-Step Procedure

3.5.1. Preparation of Compound 9

To a solution of compound **6** (100 mg, 0.36 mmol) in 0.2 mL of Et₂O, maleimide (105 mg, 1.1 mmol, 3 eq.) was added, and the reaction mixture was stirred for 24 h at 24 °C. The precipitate that formed was filtered, washed with cold Et₂O, and dried in vacuo. Compound **9a** was obtained as white crystals (49 mg, 36%). Approximately 10 mg of 10% Pd/C was added to a solution of compound **9a** (49 mg, 0.13 mmol) in THF (1 mL), and the reaction mixture was placed under a hydrogen atmosphere (1 atm) for 8 h at 24 °C. The catalyst was filtered off and washed thoroughly with hot THF (3 × 4 mL). The filtrate was evaporated under reduced pressure. The residue was purified by flash chromatography (ethyl acetate as eluent). Compound **9** was obtained as a pale yellow oil (47 mg, 35% over two steps).

endo-4,7-Bis((*phenylamino*)*methyl*)-*3a,4,7,7a-tetrahydro-1H-4,7-epoxyisoindole-1,3*(2*H*)-*dione* (**9a**). ¹H·NMR (500 MHz, Acetone-*d*₆) δ = 9.80 (s, 1H), 7.14 (t, *J* = 7.8 Hz, 4H), 6.78 (d, *J* = 7.9 Hz, 4H), 6.64 (t, *J* = 7.3 Hz, 2H), 6.52 (s, 2H), 5.00 (s, 2H), 3.80 (qd, *J* = 13.7, 6.0 Hz, 4H), 3.67 (s, 2H). ¹³C-NMR (126 MHz, Acetone-*d*₆) δ 176.8, 149.8, 137.5, 129.9, 117.9, 113.7, 92.1, 52.2, 45.8. HRMS (ESI) Calcd. for C₂₂H₁₁N₃O₃ [M + H]: 376.1656. Found 376.1648.

endo-4,7-Bis((phenylamino)methyl)hexahydro-1H-4,7-epoxyisoindole-1,3(2H)-dione(endo-4,7-bis(phenylamino)norcantharimide) (9). ¹H-NMR (500 MHz, CDCl₃) δ = 8.26 (s, 1H), 7.21 (t, *J* = 7.6 Hz, 4H), 6.75 (t, *J* = 8.9 Hz, 6H), 4.19 (s, 2H), 3.68 (d, *J* = 13.6 Hz, 2H), 3.58 (d, *J* = 10.3 Hz, 4H), 1.93 (s, 4H). ¹³C-NMR (126 MHz, CDCl₃) δ = 176.1, 148.1, 129.5, 118.2, 113.2, 88.9, 54.6, 47.1, 30.1. HRMS (ESI) Calcd. for C₂₂H₂₃N₃O₃ [M + H]: 378.1812. Found 378.1818.

3.5.2. Preparation of Compound 19

To a solution of compound **16** (100 mg, 0.3 mmol) in 2 mL of Et₂O, maleic anhydride (87 mg, 0.9 mmol, 3 eq.) was added, and the reaction mixture was stirred for 24 h at 24 °C. The precipitate that formed was filtered, washed with cold Et₂O, and dried in vacuo. Compound **19a** was obtained as white crystals (110 mg, 84%). Approximately 10 mg of 10% Pd/C was added to a solution of compound **19a** (110 mg, 0.25 mmol) in THF (1 mL), and the reaction mixture was placed under a hydrogen atmosphere (1 atm) for 8 h at 24 °C. The catalyst was filtered off and washed thoroughly with hot THF (3 × 4 mL). The filtrate was evaporated under reduced pressure. The residue was purified by flash chromatography (ethyl acetate as eluent). Compound **19** was obtained as a pale yellow oil (100 mg, 77% over two steps).

endo-1,3-Dioxo-1,3,3a,7a-tetrahydro-4,7-epoxyisobenzofuran-4,7-diylbis(methylene)dibenzoate (**19a**). ¹H-NMR (500 MHz, Acetone- d_6) δ = 8.02 (d, *J* = 7.6 Hz, 4H), 7.65 (t, *J* = 7.3 Hz, 2H), 7.50 (t, *J* = 7.6 Hz, 4H), 6.84 (s, 2H), 5.13 (d, *J* = 12.6 Hz, 2H), 4.85 (d, *J* = 12.6 Hz, 2H), 3.90 (s, 2H). ¹³C-NMR (75 MHz, Acetone- d_6) δ 169.8, 166.2, 139.2, 134.2, 130.7, 130.4, 129.5, 91.4, 62.3, 53.8. HRMS (ESI) Calcd. for C₂₄H₁₈O₈ [M + NH₄]: 452.1340. Found 452.1342.

3.6.1. Cell Culture

The human colorectal adenocarcinoma cell line HT-29 (The Russian Cell Culture Collection (RCCC), the Institute of Cytology RAS, St. Petersburg, Russia) was cultured in clear plastic tissue culture-treated dishes or multiple-well plates (Corning Inc., Corning, NY, USA) in a HeraCell 150 incubator (Thermo Electron Corp., Waltham, MA, USA) at 37 °C, 95% humidity, and 5% CO₂. The DMEM (Dulbecco's modified Eagle's medium)/F-12 (1:1) medium with 2.5 mM L-glutamine and 1.5 mM HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid) (HyClone, Logan, UT, USA) was used. The medium was supplemented with 10% fetal bovine serum (HyClone, Logan, UT, USA), 100 units·mL⁻¹ penicillin (OAO Sintez, Kurgan, Russia), and 100 μ g*mL⁻¹ streptomycin (OAO Biokhimik, Saransk, Russia).

3.6.2. MTS Assay

The MTS assay was used to evaluate the cytotoxicity of the target compounds, as previously described [40]. Before the test, cells were placed into 96-well flat-bottomed plates, with 10,000 cells per well. The utmost wells were filled with 200 μ L of phosphate-buffered saline (HyClone, Logan, UT, USA). Cells were cultivated until reaching a 70% monolayer, and then were incubated for 48 h or 72 h with the target compounds in concentrations ranging from 1 μ M to 5000 μ M, depending on the compound, with 10 points in total. The same substance concentrations were applied to an empty plate to allow an adjustment for the substance influence on optical density. All of the test points were measured in triplicate. Triton-100 (1%, Sigma-Aldrich, St. Louis, MO, USA) in the culture medium was used as a positive control, and the medium was used as a negative control. After incubation, 20 μ L of the MTS (3-(4,5-dimethylthiazol-2-yl)-5-(3-carboxymethoxyphenyl)-2-(4-sulfophenyl)-2H-tetrazolium) reagent (CellTiter 96[®] AQueous One Solution Cell Proliferation Assay, Promega, Fitchburg, WI, USA) were added into each well, and the plates were incubated for additional 4 h. Optical density was measured at 492 nm and 650 nm were subtracted from those obtained at 492 nm to exclude background absorption.

4. Conclusions

In summary, we have developed the reductive amination of DFF in order to obtain new amine-containing bio-derived building blocks **3–6** and norcantharidin derivatives **9**, **10**, and **17–19**, which can be used in the production of drugs and materials. The prepared derivatives extend applications of HMF as a platform chemical in organic synthesis, particularly connecting biomass processing with amine-containing building blocks. An efficient one-pot procedure was implemented, and the Diels–Alder reaction of substituted BAMF derivatives with maleimide and maleic anhydride was carried out with high endo selectivity; the influence of substituents in maleimide on diastereoselectivity of the cycloaddition was observed. It was shown that the introduction of amines in the side-chains resulted in the enhanced bioactivity of cantharimide.

Supplementary Materials: The spectra of synthesized compounds and Table S1: Cytotoxicity of selected substances towards HT-29 cells, are available online.

Acknowledgments: This study was supported by Russian Science Foundation (Grant RSF 17-13-01176).

Author Contributions: K.I.G., F.A.K. and V.P.A. conceived and designed the experiments; K.I.G., F.A.K., O.N.M. and A.V.P. performed the experiments; K.I.G., K.S.E. and V.P.A. analyzed the data and wrote the paper.

Conflicts of Interest: The authors declare no conflict of interest.

References

 Field, C.B. Primary Production of the Biosphere: Integrating Terrestrial and Oceanic Components. *Science* 1998, 281, 237–240. [CrossRef] [PubMed]

- Werpy, T.; Petersen, G.; Aden, A.; Bozell, J.; Holladay, J.; White, J.; Manheim, A.; Eliot, D.; Lasure, L.; Jones, S. Top Value Added Chemicals from Biomass. Volume 1—Results of Screening for Potential Candidates from Sugars and Synthesis Gas; Department of Energy: Washington, DC, USA, 2004.
- 3. Bozell, J.J.; Petersen, G.R. Technology development for the production of biobased products from biorefinery carbohydrates—The US Department of Energy's "Top 10" revisited. *Green Chem.* **2010**, *12*, 539–554. [CrossRef]
- 4. Román-Leshkov, Y.; Barrett, C.J.; Liu, Z.Y.; Dumesic, J.A. Production of dimethylfuran for liquid fuels from biomass-derived carbohydrates. *Nature* **2007**, *447*, 982–985. [CrossRef] [PubMed]
- Sutton, A.D.; Waldie, F.D.; Wu, R.; Schlaf, M.; 'Pete' Silks, L.A., III; Gordon, J.C. The hydrodeoxygenation of bioderived furans into alkanes. *Nat. Chem.* 2013, *5*, 428–432. [CrossRef] [PubMed]
- Hu, L.; Lin, L.; Wu, Z.; Zhou, S.; Liu, S. Recent advances in catalytic transformation of biomass-derived 5-hydroxymethylfurfural into the innovative fuels and chemicals. *Renew. Sustain. Energy Rev.* 2017, 74, 230–257. [CrossRef]
- 7. Wu, K.; Wu, Y.; Chen, Y.; Chen, H.; Wang, J.; Yang, M. Heterogeneous Catalytic Conversion of Biobased Chemicals into Liquid Fuels in the Aqueous Phase. *ChemSusChem* **2016**, *9*, 1355–1385. [CrossRef] [PubMed]
- 8. Thananatthanachon, T.; Rauchfuss, T.B. Efficient Production of the Liquid Fuel 2,5-Dimethylfuran from Fructose Using Formic Acid as a Reagent. *Angew. Chem. Int. Ed.* **2010**, *49*, 6616–6618. [CrossRef] [PubMed]
- 9. Amarasekara, A.S. *Renewable Polymers: Synthesis, Processing, and Technology*; Mittal, V., Ed.; John Wiley & Sons, Scrivener Pub.: Hoboken, NJ, USA, 2012; pp. 398–428.
- 10. Liu, W.-J.; Jiang, H.; Yu, H.-Q. Development of Biochar-Based Functional Materials: Toward a Sustainable Platform Carbon Material. *Chem. Rev.* **2015**, *115*, 12251–12285. [CrossRef] [PubMed]
- 11. Dutta, S.; De, S.; Saha, B. A Brief Summary of the Synthesis of Polyester Building-Block Chemicals and Biofuels from 5-Hydroxymethylfurfural. *ChemPlusChem* **2012**, *77*, 259–272. [CrossRef]
- 12. Zhang, X.; Fevre, M.; Jones, G.O.; Waymouth, R.M. Catalysis as an Enabling Science for Sustainable Polymers. *Chem. Rev.* **2017**. [CrossRef] [PubMed]
- Van Putten, R.-J.; van der Waal, J.C.; de Jong, E.; Rasrendra, C.B.; Heeres, H.J.; de Vries, J.G. Hydroxymethylfurfural, A Versatile Platform Chemical Made from Renewable Resources. *Chem. Rev.* 2013, 113, 1499–1597. [CrossRef] [PubMed]
- 14. Rosatella, A.A.; Simeonov, S.P.; Frade, R.F.M.; Afonso, C.A.M. 5-Hydroxymethylfurfural (HMF) as a building block platform: Biological properties, synthesis and synthetic applications. *Green Chem.* **2011**, *13*, 754. [CrossRef]
- 15. Lewkowski, J. Synthesis, chemistry and applications of 5-hydroxymethyl-furfural and its derivatives. *ARKIVOC* **2005**, 2001, 17–54. [CrossRef]
- Delidovich, I.; Hausoul, P.J.C.; Deng, L.; Pfützenreuter, R.; Rose, M.; Palkovits, R. Alternative Monomers Based on Lignocellulose and Their Use for Polymer Production. *Chem. Rev.* 2016, 116, 1540–1599. [CrossRef] [PubMed]
- 17. Galbis, J.A.; García-Martín, M.d.G.; de Paz, M.V.; Galbis, E. Synthetic Polymers from Sugar-Based Monomers. *Chem. Rev.* **2016**, *116*, 1600–1636. [CrossRef] [PubMed]
- 18. Gandini, A.; Lacerda, T.M.; Carvalho, A.J.F.; Trovatti, E. Progress of Polymers from Renewable Resources: Furans, Vegetable Oils, and Polysaccharides. *Chem. Rev.* **2016**, *116*, 1637–1669. [CrossRef] [PubMed]
- 19. Froidevaux, V.; Negrell, C.; Caillol, S.; Pascault, J.-P.; Boutevin, B. Biobased Amines: From Synthesis to Polymers; Present and Future. *Chem. Rev.* **2016**, *116*, 14181–14224. [CrossRef] [PubMed]
- 20. Dunbabin, A.; Subrizi, F.; Ward, J.M.; Sheppard, T.D.; Hailes, H.C. Furfurylamines from biomass: transaminase catalysed upgrading of furfurals. *Green Chem.* **2017**, *19*, 397–404. [CrossRef]
- Komanoya, T.; Kinemura, T.; Kita, Y.; Kamata, K.; Hara, M. Electronic Effect of Ruthenium Nanoparticles on Efficient Reductive Amination of Carbonyl Compounds. *J. Am. Chem. Soc.* 2017, 139, 11493–11499. [CrossRef] [PubMed]
- 22. Roylance, J.J.; Choi, K.-S. Electrochemical reductive amination of furfural-based biomass intermediates. *Green Chem.* **2016**, *18*, 5412–5417. [CrossRef]
- Sánchez, M.I.; Vázquez, O.; Martínez-Costas, J.; Vázquez, M.E.; Mascareñas, J.L. Straightforward access to bisbenzamidine DNA binders and their use as versatile adaptors for DNA-promoted processes. *Chem. Sci.* 2012, 3, 2383. [CrossRef]
- 24. Galkin, K.I.; Krivodaeva, E.A.; Romashov, L.V.; Zalesskiy, S.S.; Kachala, V.V.; Burykina, J.V.; Ananikov, V.P. Critical Influence of 5-Hydroxymethylfurfural Aging and Decomposition on the Utility of Biomass Conversion in Organic Synthesis. *Angew. Chem. Int. Ed.* **2016**, *55*, 8338–8342. [CrossRef] [PubMed]

- 25. Kashin, A.S.; Galkin, K.I.; Khokhlova, E.A.; Ananikov, V.P. Direct Observation of Self-Organized Water-Containing Structures in the Liquid Phase and Their Influence on 5-(Hydroxymethyl)furfural Formation in Ionic Liquids. *Angew. Chem. Int. Ed.* **2016**, *55*, 2161–2166. [CrossRef] [PubMed]
- Kashparova, V.P.; Khokhlova, E.A.; Galkin, K.I.; Chernyshev, V.M.; Ananikov, V.P. The "one-pot" synthesis of 2,5-diformylfuran, a promising synthon for organic materials in the conversion of biomass. *Russ. Chem. Bull.* 2015, 64, 1069–1073. [CrossRef]
- 27. Kucherov, F.A.; Galkin, K.I.; Gordeev, E.G.; Ananikov, V.P. Efficient route for the construction of polycyclic systems from bioderived HMF. *Green Chem.* **2017**, *19*, 4858–4864. [CrossRef]
- 28. De Graaff, C.; Bensch, L.; van Lint, M.J.; Ruijter, E.; Orru, R.V.A. IBX-mediated oxidation of unactivated cyclic amines: application in highly diastereoselective oxidative Ugi-type and aza-Friedel–Crafts reactions. *Org. Biomol. Chem.* **2015**, *13*, 10108–10112. [CrossRef] [PubMed]
- 29. Baba, Y.; Hirukawa, N.; Tanohira, N.; Sodeoka, M. Structure-Based Design of a Highly Selective Catalytic Site-Directed Inhibitor of Ser/Thr Protein Phosphatase 2B (Calcineurin). *J. Am. Chem. Soc.* **2003**, 125, 9740–9749. [CrossRef] [PubMed]
- Hirai, G.; Sodeoka, M. Focused Library with a Core Structure Extracted from Natural Products and Modified: Application to Phosphatase Inhibitors and Several Biochemical Findings. *Acc. Chem. Res.* 2015, 48, 1464–1473. [CrossRef] [PubMed]
- Shen, M.; Wu, M.-Y.; Chen, L.-P.; Zhi, Q.; Gong, F.-R.; Chen, K.; Li, D.-M.; Wu, Y.; Tao, M.; Li, W. Cantharidin represses invasion of pancreatic cancer cells through accelerated degradation of MMP2 mRNA. *Sci. Rep.* 2015, 5, 11836–11848. [CrossRef] [PubMed]
- 32. Puerto Galvis, C.E.; Vargas Méndez, L.Y.; Kouznetsov, V.V. Cantharidin-Based Small Molecules as Potential Therapeutic Agents. *Chem. Biol. Drug Des.* **2013**, *82*, 477–499. [CrossRef] [PubMed]
- 33. Hill, T.A.; Stewart, S.G.; Gordon, C.P.; Ackland, S.P.; Gilbert, J.; Sauer, B.; Sakoff, J.A.; McCluskey, A. Norcantharidin Analogues: Synthesis, Anticancer Activity and Protein Phosphatase 1 and 2A Inhibition. *ChemMedChem* 2008, *3*, 1878–1892. [CrossRef] [PubMed]
- 34. Tarleton, M.; Gilbert, J.; Sakoff, J.A.; McCluskey, A. Synthesis and anticancer activity of a series of norcantharidin analogues. *Eur. J. Med. Chem.* **2012**, *54*, 573–581. [CrossRef] [PubMed]
- 35. Robertson, M.J.; Gordon, C.P.; Gilbert, J.; McCluskey, A.; Sakoff, J.A. Norcantharimide analogues possessing terminal phosphate esters and their anti-cancer activity. *Bioorg. Med. Chem.* **2011**, *19*, 5734–5741. [CrossRef] [PubMed]
- McCluskey, A.; Walkom, C.; Bowyer, M.C.; Ackland, S.P.; Gardiner, E.; Sakoff, J.A. Cantharimides: A new class of modified cantharidin analogues inhibiting protein phosphatases 1 and 2A. *Bioorg. Med. Chem. Lett.* 2001, *11*, 2941–2946. [CrossRef]
- 37. De, S.; Kumar, T.; Bohre, A.; Singh, L.R.; Saha, B. Furan-based acetylating agent for the chemical modification of proteins. *Bioorg. Med. Chem.* 2015, 23, 791–796. [CrossRef] [PubMed]
- Cao, Q.; Liang, W.; Guan, J.; Wang, L.; Qu, Q.; Zhang, X.; Wang, X.; Mu, X. Catalytic synthesis of 2,5-bis-methoxymethylfuran: A promising cetane number improver for diesel. *Appl. Catal. A* 2014, 481, 49–53. [CrossRef]
- 39. Kuo, Y.-H.; Shieh, C.-J. Sensitized Photooxidation of 2,5-Furanyldimethyl Dibenzoate. *Heterocycles* **1986**, *24*, 1271–1274. [CrossRef]
- 40. Egorova, K.S.; Seitkalieva, M.M.; Posvyatenko, A.V.; Ananikov, V.P. An unexpected increase of toxicity of amino acid-containing ionic liquids. *Toxicol. Res.* **2015**, *4*, 152–159. [CrossRef]

Sample Availability: Samples of the compounds are available from the authors.



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).