



Article

Targeting Cancer Stem Cells with Novel 4-(4-Substituted phenyl)-5-(3,4,5-trimethoxy/3,4dimethoxy)-benzoyl-3,4-dihydropyrimidine-2(1*H*)-one/thiones

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Abstract: Novel 4-(4-substituted phenyl)-5-(3,4,5-trimethoxy/3,4-dimethoxy)-benzoyl-3,4-dihydropyrimidine-2(1*H*)-one/thione derivatives (**DHP 1–9**) were designed, synthesized, characterized and evaluated for antitumor activity against cancer stem cells. The compounds were synthesized in one pot. Enaminones **E1** and **E2** were reacted with substituted benzaldehydes and urea/thiourea in the presence of glacial acetic acid. The synthesized compounds were characterized by spectral analysis. The compounds were screened in vitro against colon cancer cell line (LOVO) colon cancer stem cells. Most of the compounds were found to be active against side population cancer stem cells with an inhibition of >50% at a 10 μ M concentration. Compounds **DHP-1**, **DHP-7** and **DHP-9** were found to be inactive. Compound **DHP-5** exhibited an in vitro anti-proliferative effect and arrested cancer cells at the Gap 2 phase (G2) checkpoint and demonstrated an inhibitory effect on tumor growth for a LOVO xenograft in a nude mouse experiment.

Keywords: cancer stem cells; antitumor activity; dihydropyrimidine; enaminones

1. Introduction

Compounds, which have potential anticancer activity, are often screened out in drug discovery programs for cancer research [1] due to the presence of cells which have the capability to regrow in vivo, called cancer stem cells (CSCs). Thus, the antitumor activity of the compounds in vivo is not adequate for the treatment of cancer in preclinical models. Tumors are maintained by a self-growing CSC population [2]. Research has confirmed the presence of cancer stem cells in leukemia [3], as well as in tumors of the breast [4], brain [5], lung [6] and colon [7]. In cancer relapse, CSCs must have resisted the primary drug action [8]. Literature has reported that aldehyde dehydrogenase 1 (ALDH-1) is a more potent marker of breast CSCs [9–11] and ALDH-1–positive cells are resistant to Epirubicin and Paclitaxel [12]. Adult stem cells can be predicted by a side population (SP) phenotype. A SP is confined to the tumorigenic part of the breast cancer cell line MCF-7 [13,14]. Normal chemotherapy could lead to augmentation of CSCs in treated patients [15,16]. Thus, there remains an urgent need to discover new drugs to effectively eliminate both proliferating cells and CSCs in order to treat cancer [17].

Multicomponent reactions (MCR) are important in the discovery of new lead compounds. The acid-catalyzed cyclocondensation reaction of a diketone with benzaldehyde and urea was reported in 1893 by Pietro Biginelli. The product obtained was identified as a dihydropyridimidine-2-one. Dihydropyrimidines presented a varied range of biological activities, e.g., calcium channel blockers, α -adrenoceptor selective antagonists and anti-mitotics [18]. Furthermore, (S)-Monastrol (1) has been identified as a novel molecule for the development of potentially new anticancer drugs [19]. Monastrol causes specific and reversible inhibition of kinesin Eg5. Oxo-Monastrol and its thio-analogues have been investigated for their anti-proliferative activity. The 4-methoxy derivative 2 and 3-methoxy-4-hydoxy derivative 3 of Monastrol have been synthesized as anticancer agents [20]. The 3,4-methylenedioxy derivative of Monastrol, Piperastrol (4), was found to be three times more potent than Monastrol [21]. Pyrimidinone peptoid hybrids have been reported as active against SKBr-3 breast cancer cells [22]. Improved efficiency was reported in cell-based assays by optimization of the Monastrol-based dihydropyrimidine (DHPM) analogue R-Monastrol-97 (5) [23]. The 3,4-difluoro derivative *R*-fluorestrol (6) was also reported to be a potent anticancer compound. Compound 7, derived from Monastrol-97, has been reported to be active in anticancer screens. Deaths of over 80% of cancer cells were observed after 72 h of treatment with the Biginelli adducts Enastron (8) and dimethyl Enastron (9) [24]. These compounds showed minute toxic effects against healthy fibroblast cells. Amide-derived Biginelli adducts exhibited moderate anti-proliferative activity against HepG2 epithelial carcinoma. Compounds 10 and 11 showed IC₅₀ values of (190 μ g/mL) against HeLa hepatocellular carcinoma cells [25]. Additionally, cinnamoyl derivatives of dihydropyrimidine have been reported as potent anticancer agents [26]. Examples of dihydropyrimidines demonstrating anticancer activities are presented in Figure 1.



Figure 1. Dihydropyrimidine derivatives demonstrating anticancer activity.

There is a need for structural optimization of dihydropyrimidine derivatives with the aim of modifying the profile of current lead molecules. In an effort to discover novel dihydropyrimidine derivatives with potent anticancer activity against cancer stem cells, modulation of the Monastrol-97 structure was carried out as illustrated in Figure 2.



Figure 2. Lead compound Monstrol-97 and newly synthesized compounds (DHP 1-9).

These dihydropyrimidine derivatives were then evaluated for antitumor activity.

2. Results and Discussion

Enaminones E1 and E2 were reacted with substituted benzaldehydes and urea/thiourea in the presence of acetic acid to yield dihydropyrimidinone/thione derivatives (DHP 1–9). The purity of the compounds was confirmed by elemental analysis and thin-layer chromatography. The compounds were characterized using spectroscopic methods. In the ¹H-NMR spectra, the signals of the individual protons of the compounds were verified on the basis of multiplicity, chemical shifts and the coupling constant. All the compounds showed the D₂O exchangeable broad singlet at 8.8–9.8 ppm and 9.5–10.5 ppm corresponding to the two NH protons. Analytical and spectral data for the compounds were in good agreement with the expected structures of the synthesized derivatives. The physicochemical properties of all compounds are given in Table 1.

Table 1. Physical data of the synthesized dihydropyrimidinone/thione compounds (DHP 1-9).



Compounds	R	R ¹	R ²	(Yield %)	m.p. (°C)
DHP-1	Phenyl	OCH ₃	0	70	153–155
DHP-2	4-Chlorophenyl	OCH ₃	0	75	138-140
DHP-3	4-Nitrophenyl	OCH ₃	0	65	158-160
DHP-4	3,4-Dimethoxyphenyl	OCH ₃	0	72	165–167
DHP-5	4-Ethoxyphenyl	OCH ₃	0	60	168-170
DHP-6	Phenyl	Н	S	65	248-250
DHP-7	4-Chlorophenyl	Н	S	65	243-245
DHP-8	4-Nitrophenyl	Н	S	68	258-260
DHP-9	3,4-Dimethoxyphenyl	Н	S	70	228-230

The newly synthesized compounds (**DHP 1–9**) were evaluated for side population percent inhibition on colon cancer cell line (LOVO) at a 10 μ M concentration (Figure 3, Table 2).

Compounds	* Side Population (%) at 10 μM	$^{\#}$ Side Population Inhibition (%) at 10 μM
DHP-1	4.90 ± 0.2	0
DHP-2	1.72 ± 0.1	64.7
DHP-3	1.76 ± 0.3	64
DHP-4	1.44 ± 0.5	70.5
DHP-5	2.01 ± 0.7	58.82
DHP-6	1.47 ± 0.6	70
DHP-7	4.90 ± 0.3	0
DHP-8	2.4 ± 0.1	50
DHP-9	4.90 ± 0.1	0

Table 2. Side population inhibition on LOVO colon cancer cells (%) at 10 μ M concentration.

* Side population% as mean \pm SD of three independent experiments; [#] Inhibition% = 100 - (SP% of treated cells/SP% of untreated cells) \times 100.



Figure 3. Scatter plot showing results of side population analyses of tumor-derived cells of LOVO untreated, treated with **DHP-1**, **DHP-4**, **DHP-5** and **DHP-6**. Furthermore, compound **DHP-5** exhibit an in vitro anti-proliferative effect and arrested cancer cells at the G2 checkpoint (Figure 4). Blue color represents the percentage of cancer stem cells and red color represents the percentage of remaining cells other than cancer stem cells.



Figure 4. Compound DHP-5 arrested cancer cells at G2 checkpoint.

The structure-activity relationships of the compounds were studied. From the compounds (**DHP 1–9**), four compounds were found to be very effective, namely **DHP-4**, **DHP-6**, **DHP-2** and **DHP-3**, when the side population inhibition percentage was measured at a 10 µM concentration. Compounds **DHP-5** and **DHP-8** were moderately active as indicated by a low value of the side population inhibition percentage. Most of the dihydropyrimidine compounds (**DHP 1–9**) presented significant activity against side population inhibition percentage. It was noted that most of the compounds having a methoxy group at R¹ were active. Compounds with an oxygen atom at R² were also active. Compound **DHP-6**, with a hydrogen at R¹ and a sulfur atom at R², displayed significant activity. Compound **DHP-4** was found to be the most active compound of the series.

A side population analysis of tumor-derived cells of LOVO xenografts that were untreated, treated with the side population inhibitor reference drug Verapamil 200 μ M, and with compound **DHP-5** (50 μ M) confirmed that **DHP-5** had a more potent inhibitory effect on the side population cancer stem cells than the reference drug Verapamil (Figure 5).



Figure 5. Scatter plot showing results of side population analyses of tumor-derived cells of the LOVO xenograft that were untreated, treated with side population inhibitor reference drug Verapamil (200 μ M) and with **DHP-5** (50 μ M).

The tumor growth of LOVO (colon cancer xenografts) was recorded in untreated mice groups and in **DHP-5**–treated (50 mg/kg) mice groups. A potent anti-tumor effect was demonstrated by a shrinking of tumors in the animals which were treated by compound **DHP-5**. A remarkable anti-tumor effect of compound **DHP-5** was demonstrated on tumors of colon cancer xenografts (Figure 6).



Figure 6. Graph showing tumor growth record of LOVO (colon cancer xenograft) in untreated mice group (**red line**) and **DHP-5**–treated (50 mg/kg) mice group (**blue line**).

3. Material and Methods

3.1. Experimental

All solvents were obtained from Merck (Kenilworth, NJ, USA). The homogeneity of the compounds was checked by TLC performed on silica gel G; An iodine chamber was used for visualization of TLC spots. The FT-IR spectra were recorded in KBr pellets on a Spectrum BX Perkin Elmer FT-IR spectrophotometer (Perkin Elmer, Hopkinton, MA, USA). Melting points were determined on a Gallenkamp melting point apparatus (Gallenkamp, Loughborough, UK), and are uncorrected. NMR spectra were scanned in DMSO- d_6 on a Bruker NMR spectrophotometer (Bruker, Billerica, MA, USA) operating at 500 MHz for ¹H and 125.76 MHz for ¹³C at the Research Center, College of Pharmacy, King Saud University, Saudi Arabia. Chemical shifts δ are expressed in parts per million (ppm) relative to TMS as an internal standard and D₂O was added to confirm the exchangeable protons. Coupling constants (*J*) are in Hertz. The molecular masses of compounds were determined by UPLC/TQMS and all tested compounds yielded data consistent with a purity of \geq 95%, as measured by HPLC (Agilent 1260 affinity). The elemental analyses (C, H, N (\pm 0.4%); and S (\pm 0.3%)) were performed on a CHN Elementar (Analysensysteme GmbH, Langenselbold, Germany).

The synthesis of dihydropyrimidine derivatives was carried out in single step as shown in Scheme 1.



Scheme 1. Synthetic route of compounds (DHP 1-9).

3.2. General Synthesis of 4-(Substituted phenyl)-5-(3,4,5-trimethoxybenzoyl/3,4-dimethoxybenzoyl)-3,4dihydropyrimidin-2(1H)-ones **DHP 1–9**

A solution of enaminone **E1/E2** (0.01 mol), substituted benzaldehyde (0.01 mol), urea/thiourea (0.01 mol) and glacial acetic acid (10 mL) was heated under reflux for 3 h. The precipitates (**DHP 1–9**) thus formed were collected by filtration, washed with water and recrystallized from acetic acid.

4-Phenyl-5-(3,4,5-trimethoxybenzoyl)-3,4-dihydropyrimidin-2(1H)-one (**DPH-1**): Yield: 70%; m.p.: 153–155 °C; IR (KBr): 3412 (N-H), 2938 (ArC-H), 1700 (C=O), 1636 (C=O), 1618 (C=C), 1126 (C-O); ¹H-NMR (500 MHz, DMSO- d_6); δ = 3.80 (9H, s, 3× -OCH₃), 5.40 (1H, d, *J* = 2.5 Hz, H-4), 6.73–7.36 (7H, m, Ar-H), 7.88 (1H, d, *J* = 2.5 Hz, =CH), 9.50 (1H, bs, NH, D₂O exchg.), 10.00 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO- d_6): δ = 54.0, 56.4, 56.5, 56.7, 60.5, 106.2, 112.4, 126.9, 127.8, 128.9, 134.4, 140.3, 142.3, 144.6, 151.8, 152.9, 153.0, 191.0; MS: *m*/*z* = 368.46 [M]⁺; Analysis: C₂₀H₂₀N₂O₅ for, calcd. C 65.21, H 5.47, N 7.60%; found C 65.45, H 5.48, N 7.62%.

4-(4-Chlorophenyl)-5-(3,4,5-trimethoxybenzoyl)-3,4-dihydropyrimidin-2(1H)-one (**DPH-2**): Yield: 75%; m.p.: 138–140 °C; IR (KBr): 3412 (N-H), 2938 (ArC-H), 1686 (C=O), 1654 (C=O), 1618 (C=C), 1123 (C-O); ¹H-NMR (500 MHz, DMSO-*d*₆); δ = 3.79 (9H, s, 3× -OCH₃), 5.39 (1H, d, *J* = 3.0 Hz, H-4), 6.74–7.43 (6H, m, Ar-H), 7.91 (1H, d, *J* = 2.5 Hz, =CH), 9.50 (1H, bs, NH, D₂O exchg.), 10.00 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO-*d*₆): δ = 53.6, 56.4, 56.5, 56.7, 60.5, 60.7, 106.2, 108.2, 112.0, 128.4, 128.9, 128.9, 129.8, 131.6, 152.3, 134.2, 139.8, 140.2, 140.3, 142.5, 143.5, 151.6, 152.9, 153.0, 153.2, 191.0,

192.5, 193.0; MS: $m/z = 402.8 \text{ [M]}^+$, 403.8 [M + 1]⁺; Analysis: C₂₀H₁₉N₂O₅Cl for, calcd. C 59.63, H 4.75, N 6.95%; found C 59.45, H 4.73, N 6.97%.

4-(4-Nitrophenyl)-5-(3,4,5-trimethoxybenzoyl)-3,4-dihydropyrimidin-2(1H)-one (**DPH-3**): Yield: 65%; m.p.: 158–160 °C; IR (KBr): 3421 (N-H), 2936 (ArC-H), 1685 (C=O), 1654 (C=O), 1618 (C=C), 1125 (C-O); ¹H-NMR (500 MHz, DMSO- d_6); δ = 3.77 (9H, s, 3× -OCH₃), 5.53 (1H, d, *J* = 2.5 Hz, H-4), 6.74–7.40 (6H, m, Ar-H), 8.20 (1H, d, *J* = 2.5 Hz, =CH), 9.47 (1H, bs, NH, D₂O exchg.), 10.20 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO- d_6): δ = 53.8, 56.5, 56.5, 56.7, 60.2, 60.5, 60.7, 65.3, 106.2, 124.7, 128.4, 131.0, 134.1, 138.2, 140.4, 143.0, 147.2, 151.4, 151.7, 153.0, 153.0, 153.2, 190.9; MS: *m*/*z* = 413.47 [M]⁺; Analysis: C₂₀H₁₉N₃O₇ for, calcd. C 58.11, H 4.63, N 10.16%; found C 58.32, H 4.62, N 10.19%.

4-(3,4-Dimethoxyphenyl)-5-(3,4,5-trimethoxybenzoyl)-3,4-dihydropyrimidin-2(1H)-one (**DPH-4**): Yield: 72%; m.p.: 165–167 °C; IR (KBr): 3367 (N-H), 2937 (ArC-H), 1700 (C=O), 1624 (C=O), 1578 (C=C), 1123 (C-O); ¹H-NMR (500 MHz, DMSO- d_6); δ = 3.81 (15H, s, 5× -OCH₃), 5.36 (1H, d, *J* = 2.5 Hz, H-4), 6.75–7.28 (5H, m, Ar-H), 7.81 (1H, d, *J* = 2.5 Hz, =CH), 9.24 (1H, bs, NH, D₂O exchg.), 9.84 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO- d_6): δ = 53.5, 55.9, 56.0, 56.3, 56.4, 56.5, 56.7, 60.5, 60.7, 65.3, 106.1, 106.2, 108.2, 109.9, 111.0, 112.2, 116.4, 118.8, 120.3, 126.5, 130.1, 131.8, 134.4, 135.1, 136.9, 139.3, 140.1, 142.1, 147.6, 148.6, 149.6, 151.7, 152.9, 153.0, 154.6, 193.3; MS: *m*/*z* = 428.26 [M]⁺; Analysis: C₂₂H₂₄N₂O₇ for, calcd. C 61.67, H 5.65, N 6.54%; found C 61.45, H 5.66, N 6.56%.

4-(4-*Ethoxyphenyl*)-5-(3,4,5-*trimethoxybenzoyl*)-3,4-*dihydropyrimidin*-2(1*H*)-*one* (**DPH-5**): Yield: 60%; m.p.: 168–170 °C; IR (KBr): 3411 (N-H), 2938 (ArC-H), 1696 (C=O), 1648 (C=O), 1618 (C=C), 1126 (C-O); ¹H-NMR (500 MHz, DMSO-*d*₆); δ = 1.31 (3H, t, *J* = 7.0 Hz, -CH₃), 3.80 (9H, s, 3× -OCH₃), 4.20 (2H, q, *J* = 2.0 Hz, -OCH₂), 5.32 (1H, d, *J* = 2.5 Hz, H-4), 6.75–7.25 (6H, m, Ar-H), 7.79 (1H, d, *J* = 2.5 Hz, =CH), 8.81 (1H, bs, NH, D₂O exchg.), 9.50 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO-*d*₆): δ = 15.1, 53.3, 56.5, 60.5, 63.4, 106.1, 112.7, 114.7, 128.1, 140.2, 153.0, 192.0; MS: *m*/*z* = 412.28 [M]⁺; Analysis: C₂₂H₂₄N₂O₆ for, calcd. C 64.07, H 5.87, N 6.79%; found C 64.25, H 5.88, N 6.76%.

(3,4-Dimethoxyphenyl)(4-phenyl-2-thioxo-1,2,3,4-tetrahydropyrimidin-5-yl)methanone (**DHP-6**): Yield: 65%; m.p.: 248–250 °C; IR (KBr): 3413 (N-H), 2955 (ArC-H), 1653 (C=O), 1636 (C=O), 1595 (C=C), 1199 (C-O); ¹H-NMR (500 MHz, DMSO- d_6); δ = 3.81 (6H, s, 2× -OCH₃), 5.45 (1H, d, *J* = 3.0 Hz, H-4), 6.97–7.28 (7H, m, Ar-H), 7.34 (1H, d, *J* = 3.0 Hz, =CH), 9.70 (1H, bs, NH, D₂O exchg.), 10.40 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO- d_6): δ = 54.2, 56.6, 56.1, 56.3, 111.1, 111.2, 111.7, 112.2, 113.6, 122.7, 127.1, 128.2, 129.1, 130.8, 136.7, 143.4, 149.1, 149.3, 152.2, 153.8, 162.7, 174.3, 191.0, 193.5; MS: *m*/*z* = 355.0 [M + 1]⁺; Analysis: C₁₉H₁₈N₂O₃S for, calcd. C 64.39, H 5.12, N 7.90, S 9.05%; found C 64.54, H 5.11, N 7.92, S 9.04%.

(3,4-Dimethoxyphenyl)(4-chlorophenyl-2-thioxo-1,2,3,4-tetrahydropyrimidin-5-yl)methanone (**DHP-7**): Yield: 65%; m.p.: 243–245 °C; IR (KBr): 3413 (N-H), 2933 (ArC-H), 1670 (C=O), 1647 (C=O), 1616 (C=C), 1195 (C-O); ¹H-NMR (500 MHz, DMSO-*d*₆); δ = 3.78 (6H, s, 2× -OCH₃), 5.45 (1H, d, *J* = 3.0 Hz, H-4), 6.98–7.45 (7H, m, Ar-H), 7.96 (1H, d, *J* = 3.0 Hz, =CH), 9.76 (1H, bs, NH, D₂O exchg.); 10.49 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO-*d*₆): δ = 53.7, 56.0, 56.1, 111.1, 111.7, 113.2, 122.7, 129.0, 129.1, 130.7, 132.8, 137.0, 142.3, 149.1, 152.2, 162.7, 174.3, 190.9; MS: *m*/*z* = 387.99 [M]⁺; Analysis: C₁₉H₁₇N₂O₃ClS for, calcd. C 58.68, H 4.41, N 7.20, S 8.25%; found C 58.85, H 4.43, N 7.23, S 8.24%.

(3,4-Dimethoxyphenyl)(4-nitrophenyl-2-thioxo-1,2,3,4-tetrahydropyrimidin-5-yl)methanone (**DHP-8**): Yield: 68%; m.p.: 258–260 °C; IR (KBr): 3412 (N-H), 2933 (ArC-H), 1676 (C=O), 1654 (C=O), 1615 (C=C), 1141 (C-O); ¹H-NMR (500 MHz, DMSO-*d*₆); δ = 3.82 (6H, s, 2× -OCH₃), 5.58 (1H, d, *J* = 3.0 Hz, H-4), 7.0–7.95 (7H, m, Ar-H), 8.26 (1H, d, *J* = 2.5 Hz, =CH), 9.85 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO-*d*₆): δ = 53.9, 56.0, 56.1, 111.1, 111.7, 112.6, 122.8, 124.4, 128.5, 130.6, 137.5, 147.4, 149.1, 150.3, 152.3, 162.7, 174.5, 190.8; MS: *m*/*z* = 402.23 [M + 3]⁺; Analysis: C₁₉H₁₇N₃O₅S for, calcd. C 57.13, H 4.29, N 10.52, S 8.03%; found C 57.23, H 4.28, N 10.55, S 8.01%.

(3,4-Dimethoxyphenyl)(3,4-dimethoxyphenyl-2-thioxo-1,2,3,4-tetrahydropyrimidin-5-yl)methanone (**DHP-9**): Yield: 70%; m.p.: 228–230 °C; IR (KBr): 3410 (N-H), 2932 (ArC-H), 1684 (C=O), 1654 (C=O), 1611 (C=C), 1134 (C-O); ¹H-NMR (500 MHz, DMSO- d_6); δ = 3.81 (12H, s, 4× -OCH₃), 5.40 (1H, d, *J* = 3.0 Hz, H-4), 6.82–7.21 (6H, m, Ar-H), 7.96 (1H, d, *J* = 2.5 Hz, =CH), 9.68 (1H, bs, NH, D₂O exchg.), 10.38 (1H, bs, NH, D₂O exchg.); ¹³C-NMR (125.76 MHz, DMSO- d_6): δ = 43.7, 55.9, 56.0, 56.0, 56.1, 111.1, 111.7, 112.3, 113.5, 119.0, 122.7, 130.8, 135.7, 136.6, 148.9, 149.1, 149.1, 152.2, 162.7, 174.1, 191.0; MS: *m*/*z* = 413.6 [M – 1]⁺; Analysis: C₂₁H₂₂N₂O₅S for, calcd. C 60.85, H 5.35, N 6.76, S 7.74%; found C 61.05, H 5.36, N 6.78, S 7.73%.

3.3. Cell line and Tissue Culture

LOVO colon cancer cells were purchased from the American Type Culture Collection. LOVO cells were cultured in RPMI. The medium was supplemented with 10% FBS (Cambrex Bio Science, Franklin Lakes, NJ, USA), 100 IU/mL of Penicillin and 100 mg/mL of Streptomycin. Cell viability was assessed by trypan blue exclusion analysis. Cell numbers were determined by using a hemacytometer.

3.4. Flow Cytometric Analysis of Cellular DNA Content

Cells (2×10^6) were fixed in 1 mL of ethanol (70%) for 60 min at room temperature. Harvested cells were resuspended in 1 mL of sodium citrate (50 mM) containing 250 µg RNase A and incubated at 50 °C for 60 min Next, cells were resuspended in the same buffer containing 4 µg of propidium iodide (PI) and incubated for 30 min before being analyzed by flow cytometry (Becton Dickinson, San Jose, CA, USA). The percentage of cells in various cell cycle phases was determined by using Cell Quest Pro software (version 5.1, Becton Dickinson, East Rutherford, NJ, USA).

3.5. Side Population Staining by DYECYCLE Violet Stain

For DCV staining, cells were pelleted and suspended in DMEM cell culture medium at a concentration of 1×10^6 cells/mL. DCV (Invitrogen Molecular Probes[®], Eugene, OR, USA) was added at a final staining concentration of 10 μ M, as this concentration gave optimal separation between SP and non SP cells. PI staining was used to exclude dead cells. Functionally, to gate only side population cells, Verapamil 200 μ M or Emtricitabine (FTC, 10 μ g/mL) was used. All analyses were performed on a FACS LSRII (BD Biosciences, San Jose, CA, USA). Debris and cell clusters were excluded during side-scatter and forward-scatter analyses.

3.6. Antitumor Activity in Mice

Nude mice (Jackson Laboratories, Bar Harbor, ME, USA), six to seven weeks old, weighing 20 g, were obtained from the Animal Care and Use Committee of the King Faisal Specialist Hospital and Research Centre, Riyadh, KSA. All of the animals were acclimatized to laboratory conditions for one week before experiments. The animals were maintained under standard conditions, housed in a pathogen-free environment, and fed adequately. Each treatment and vehicle group consisted of six animals. The breeding, care and sacrifice of the animals were performed in accordance with the protocols approved by the Animal Care and Use Committee of the King Faisal Specialist Hospital and Research Centre. The mice were injected with 4×10^6 cells of LOVO subcutaneously in the right flank, and tumor size was measured weekly using a caliper. When the tumor reached approximately 400 mm³ diameter, the mice were divided into control, treated groups, the treatment including administration of **DHP-5** (50 mg/kg) via intraperitoneal injection daily for 14 days. The general toxicity of the treatment was determined by measuring the total body weight of the treated and control mice.

4. Conclusions

In conclusion, we focused on the synthesis of dihydropyrimidine derivatives (DHP 1–9). The synthesized compounds were screened in vitro against LOVO colon cancer cells. DHP-4 was

found to be the most active compound of the series in its side population inhibition percentage at 10 µM. The anti-tumor effect of compound **DHP-5** was demonstrated on tumors of colon cancer xenografts. Compound **DHP-5** was found to be a more potent inhibitor of side population cells than the reference drug Verapamil. Compound **DHP-5** exhibited an in vitro anti-proliferative effect and arrested cancer cells at the G2 checkpoint. Furthermore, treatment with compound **DHP-5** enabled blocking of the self-renewal ability of breast cancer cells in a dose-dependent manner. Compound **DHP-5** induced apoptosis and blocked cell proliferation in vitro and presented superior efficacy compared to the reference drug Doxorubicin in advanced animal models of colon cancer without any sign of general toxicity.

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Sample Availability: Samples of the compounds (DHP 1–9) in pure form are available from the authors.



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