

## Supplementary Materials

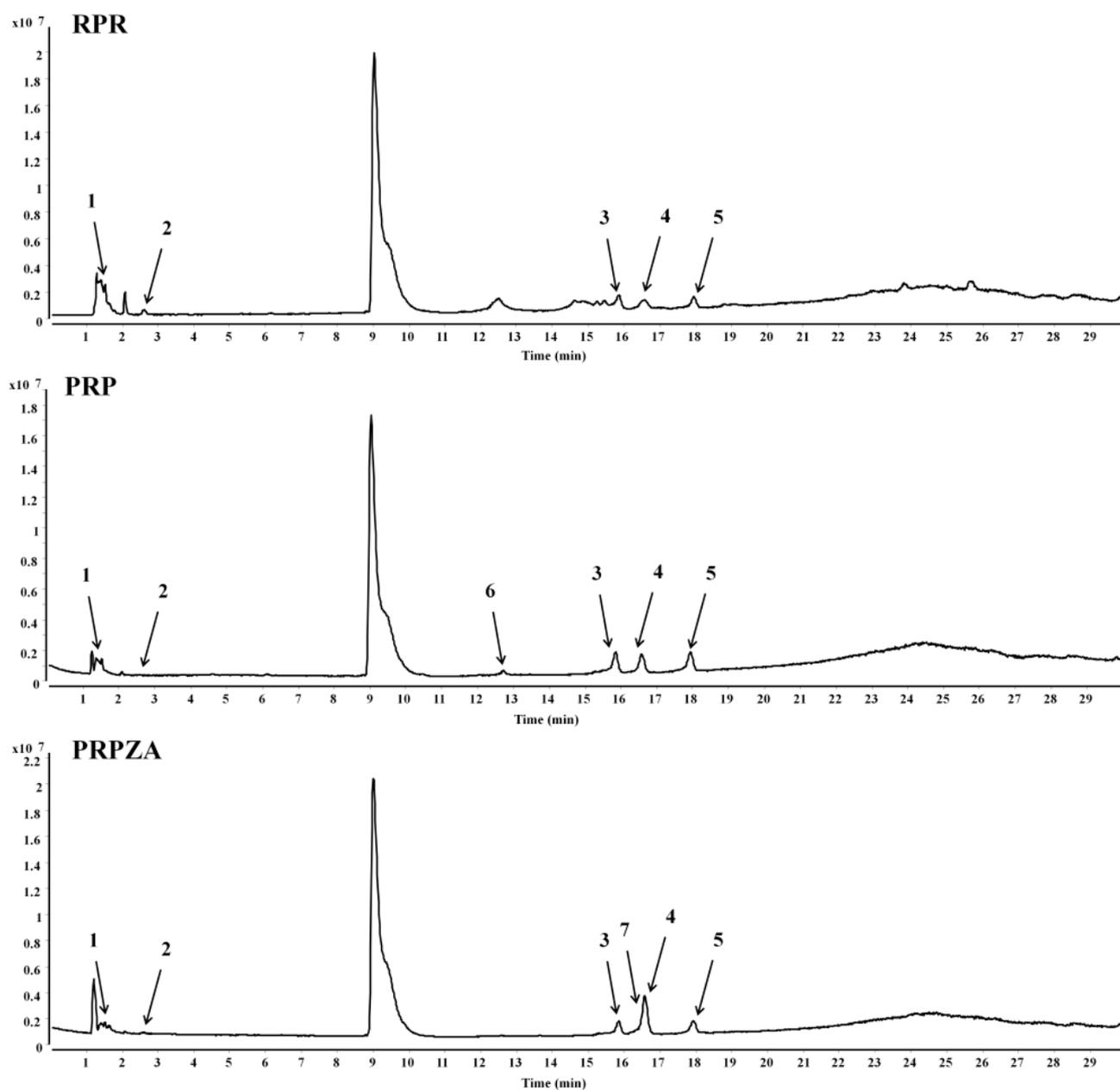
### Identification of the Possible Ingredients in RPR, PRP and PRPZA Water Extracts by UHPLC-MS-Q-TOF

The prepared water extracts (10  $\mu$ L) were diluted 100 times with methanol. The mixture was subsequently vortexed and then centrifuged at 18,000 g for 30 min, and the supernatant was subjected to UHPLC-MS-Q-TOF for analysis.

Chromatographic analysis was performed on an Agilent 1290 Infinity UHPLC system (Agilent, Santa Clara, CA, USA) equipped with a binary solvent delivery system, an auto-sampler and a MS-Q-TOF detector. Chromatographic separation was carried out on an Agilent ZORBAX Eclipse plus C18 column (1.7  $\mu$ m 100  $\times$  2.1 mm) at 30°C. The mobile phase consisted of (A) 0.1% acetic acid in water, and (B) acetonitrile. The eluting conditions were optimized as follows: isocratic at 10% B (0–3 min), linear gradient from 5% to 40% B (3–10 min), 40% to 90% B (10–25 min), 90% to 90% B (25–28 min) and 90% to 10% B (28–30 min). The flow rate was set at 0.3 mL/min, the auto-sampler was maintained at 10 °C, and the sample injection volume was 5  $\mu$ L.

The mass spectra were acquired using a 6540 Q-TOF mass spectrometer (Agilent Technologies, Santa Clara, CA, USA) equipped with an ESI interface in the positive ion mode (ESI+). The optimized mass spectrometric parameters were as follows: capillary voltage 4000 V, nebulizer gas pressure 50 psig, drying gas flow rate 11 L/min, gas temperature 350 °C, fragmentor and skimmer voltages 130 V and 50 V. The mass scale was calibrated using the calibration solution provided by the manufacturer between  $m/z$  50 and 1500. A second orthogonal sprayer with a reference solution was used for continuous calibration using the following reference masses:  $m/z$  121.0509 and 922.0098 in the positive ion mode. Masses were determined at the center of the mass peak using the width of the peak at half-peak height measured by computer-generated algorithms thus avoiding human bias.

A good chromatographic separation of constituents in RPR, PRP and PRPZA were achieved on a reversed-phase column using gradient elution with 0.1% formic acid and acetonitrile (Figure S1). And the information of possible ingredients in RPR, PRP and PRPZA were shown in Table S1.



**Figure S1.** The chromatographic profile of RPR, PRP and PRPZA water extracts.

**Table S1.** Retention time (RT), HRMS data and molecular formula in RPR, PRP and PRPZA determined by UHPLC-MS-Q-TOF.

No.	RT (min)	<i>m/z</i> experimental	Formula	<i>m/z</i> calculated	Error (ppm)	Proposed Compound
1	1.44	138.0590	C <sub>7</sub> H <sub>15</sub> NO <sub>2</sub>	138.0550	-0.57	Trigonelline
2	2.6	166.0862	C <sub>10</sub> H <sub>15</sub> NO	166.1226	3.34	Ephedrine
3	15.86	387.1806	C <sub>22</sub> H <sub>26</sub> O <sub>6</sub>	387.1802	-0.87	Pinoresinol dimethyl ether
4	16.58	149.0256	C <sub>8</sub> H <sub>4</sub> O <sub>3</sub>	149.0233	-0.98	Phthalic anhydride
5	17.91	437.1931	C <sub>24</sub> H <sub>30</sub> O <sub>6</sub>	437.1935	1.02	Magnoshinin
6	12.72	823.4166	C <sub>42</sub> H <sub>62</sub> O <sub>16</sub>	823.4111	-1.91	Glycyrrhizic acid
7	16.66	177.0553	C <sub>17</sub> H <sub>18</sub> O <sub>5</sub>	177.0546	-0.36	Herniarin