



Newiew On the Future Perspectives of Some Medicinal Plants within *Lamiaceae* Botanic Family Regarding Their Comprehensive Properties and Resistance against Biotic and Abiotic Stresses

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Abstract: Lamiaceae is one of the largest botanical families, encompassing over 6000 species that include a variety of aromatic and medicinal spices. The current study is focused on three plants within this botanical family: basil (Ocimum basilicum L.), thyme (Thymus vulgaris L.), and summer savory (Satureja hortensis L.). These three species contain primary and secondary metabolites such as phenolic and flavonoid compounds, fatty acids, antioxidants, and essential oils and have traditionally been used for flavoring, food preservation, and medicinal purposes. The goal of this study is to provide an overview of the nutraceutical, therapeutic, antioxidant, and antibacterial key features of these three aromatics to explore new breeding challenges and opportunities for varietal development. In this context, a literature search has been performed to describe the phytochemical profile of both primary and secondary metabolites and their pharmacological uses, as well as to further explore accession availability in the medicine industry and also to emphasize their bioactive roles in plant ecology and biotic and abiotic stress adaptability. The aim of this review is to explore future perspectives on the development of new, highly valuable basil, summer savory, and thyme cultivars. The findings of the current review emphasize the importance of identifying the key compounds and genes involved in stress resistance that can also provide valuable insights for further improvement of these important medicinal plants.

Keywords: aromatics; nutrient content; essential oils; salinity stress; drought stress; breeding prospects

1. Introduction

The economic importance of medicinal and aromatic plants within the context of agro-alimentary, pharmaceutical, natural cosmetics, and perfume development uses is of paramount significance. In addition to providing food flavoring and pleasing aromas, their secondary metabolites and antioxidants provide additional nutritional value and make them an invaluable part of the human diet, alongside cereals, fruits, and vegetables [1,2].

The *Lamiaceae* botanical family encompasses around 236 genera and over 6000 species of herbs and shrubs that have a global distribution. Within this family, basil (*O. basilicum* L.), thyme (*T. vulgaris* L.), and summer savory (*S. hortensis* L.) are species of particular importance, both for their specialized metabolites, such as essential oils and various non-volatile constituents with multiple applications in the food industry, cosmetics, and medicine, and their amazing adaptability to a whole range of biotic and abiotic stress factors. They are highly valued for their nutritional, medicinal, and industrial properties, providing flavor and fragrance to food, promoting health and well-being in traditional medicine due to the anti-inflammatory, antiseptic,



Citation: Avasiloaiei, D.I.; Calara, M.; Brezeanu, P.M.; Murariu, O.C.; Brezeanu, C. On the Future Perspectives of Some Medicinal Plants within *Lamiaceae* Botanic Family Regarding Their Comprehensive Properties and Resistance against Biotic and Abiotic Stresses. *Genes* **2023**, *14*, 955. https:// doi.org/10.3390/genes14050955

Academic Editors: Qingyi Yu, Wajid Zaman and Hakim Manghwar

Received: 17 March 2023 Revised: 7 April 2023 Accepted: 20 April 2023 Published: 22 April 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). and analgesic properties of their essential oils, and serving as valuable ingredients in various industries (perfumes, cosmetics, and cleaning agents) [1,2].

With regards to the main abiotic stress factors, the climate change evidence from the past decade indicates that salinity and drought will likely be the highest causes of significant concern. Salinity has emerged as a major environmental factor that has impacted over 20% of cultivated land worldwide, with the affected regions continuing to expand every year [3,4]. In fact, there is a significant risk that salinization may affect more than 50% of arable land by the mid-21st century [5], posing a serious challenge to global food security. Salinity can have a negative impact on Lamiaceae aromatic plants, affecting their growth, yield, and quality. High levels of salt in the soil decrease water availability and impose osmotic stress conditions on plants, leading to reduced photosynthesis and yield limitation [3]. In response to such conditions, the uptake and transport of the major essential ions (such as Ca, K, Mg, and nitrate) is transiently decreasing, and as a consequence, it impacts the amount and quality of secondary metabolites, such as essential oils. There is a significant increase in the reactive oxygen species (ROS) production under osmotic stress conditions that can exceed the scavenging ability of the plants. The accumulation of ROS has harmful cellular effects, such as DNA damage, membrane lipid peroxidation, and enzymatic activity impairment.

Drought stress is another significant environmental concern, representing a serious threat to agriculture worldwide by reducing the yield and quality for principal cash crops. It may have a negative impact on *Lamiaceae* aromatic plants as well affecting their growth, yield, and quality. However, most of these plants possess the ability to respond to drought stress at morphological, anatomical, physiological, biochemical, and molecular levels with a series of adjustments, allowing the plant to avoid the stress or to increase its tolerance [6,7].

2. Methods

Plenty of scientific literature discusses the benefits of these three aromatic species, and there is an abundance of data regarding their adaptation to key stressors and breeding perspectives. As a result, we searched SCOPUS and the Google Academic database for topics relating to "*O. basilicum*", "*T. vulgaris*", and "*S. hortensis*" and performed an extensive keyword search for "(Aromatic plant species name) composition and properties" and "(Aromatic plant species name) (a) biotic stress resistance." Our search generated around 350 documents published over a period of 35 years (1989–2022). The returned results highlighted a multitude of studies from countries with a long-standing traditional cultivation of these plants, especially from the Middle and Far East, focusing on a variety of complex topics.

This review aims to present the progress that has been made in studying the resistance potential of these three species on a series of stressors and to emphasize the need to focus on breeding and developing new cultivars with higher resistance capacity.

3. Discussion

3.1. Aromatic Plant Composition and Accessions Availability

A synthetic characterization of the main biocomponents of the three species studied, including the volatile oils, is presented in Table 1.

Aromatic Plant	Chemical	Fresh Leaves	References
Species	Composition	Volatile Oil	
O. basilicum L.		Dry matter (909.1 g kg ⁻¹), cude ash (89.84 g kg ⁻¹), crude protein (208.8 g kg ⁻¹), ether extract (11.21 g kg ⁻¹), crude fiber (45.91 g kg ⁻¹), NFI (sugars readily hydrolyzed) (553.3 g kg ⁻¹), Mg (79.8 μ g g ⁻¹), Ca (1278 μ g g ⁻¹), K (2135 μ g g ⁻¹), Na (218.5 μ g g ⁻¹), Fe (26.31 μ g g ⁻¹), Cu (1.95 μ g g ⁻¹), Mn (8.56 μ g g ⁻¹) and Zn (45.14 μ g g ⁻¹) Alkaloids, tannins, flavonoids, cholesterol, terpernoids, glycosides, cardiac glycosides, phenols, carbohydrates, and phlobatannins	[8,9]
		$\frac{(\sim 6.20 \text{ mg/g})}{\text{Linalool (56.7-60.6\%), epi-α-cadinol (8.6-11.4\%), α-bergamotene} (7.4-9.2\%) and γ-cadinene (3.2-5.4\%, germacrene D (1.13.3\%), camphor (1.13.1\%)$	[10,11]
T. vulgaris L.		 Oxygen terpene derivatives (1,8-cineole, linalool, followed by camphor, endo-borneol, α-terpineol and linalyl acetate), terpene hydrocarbons (α-pinene, camphene and β-pinene, trans-caryophylle, four flavonoids (two flavanones and two flavones)—sakuranetin, 6,7-dimethylcarthamidin, respectively 5-desmethylsinensetin and -hydroxy-3,7,8,2',4'-pentamethoxy-flavone 	[12]
		$(12 \text{ mL/kg} \leq)$ Thymol (~47.59%), γ -Terpinene (~30.90%), para-Cymene (~8.41%), Carene< δ -2-> (~3.76%), Caryophyllene (2.68%), α -Thujene, α -Pinene, β -Pinene, β -Myrcene, α -Phellandrene, D-Limonene, β -Phellandrene, Terpineol, Terpinen-4-ol, Cyclohexene, 1-methyl-4-(5-methyl-1-methylene-4-hexenyl)	[13]
S. hortensis L.		$\begin{array}{l} \mbox{Moisture (72\%), protein (4.2\%), fat (1.65\%), sugar (4.45\%), \\ \mbox{fibre (8.60\%), ash (2.11\%)} \\ \mbox{Minerals: K (1.68-3.38 mg\cdot kg^{-1} DM), P (0.31-0.72 mg\cdot kg^{-1} DM), \\ \mbox{Ca (1.08-2.84 mg\cdot kg^{-1} DM), Mg (0.25-0.61 mg\cdot kg^{-1} DM), \\ \mbox{Fe (242-726 mg\cdot kg^{-1} DM), and Na (0.007-0.013 mg\cdot kg^{-1} DM) \end{array}$	[14,15] [16]
		Carvacrol (11–67%, Thymol ($\underline{\geq}$ 5%) p-cymene (3.5–19.6%), <i>α</i> -phellandrene, <i>α</i> - and β-pinene, Sabinene, terpineol, <i>α</i> -thujene	[15,17], [18–22]

Table 1. Main biocomponent profile of some *Lamiaceae* family aromatic plants presented by original research papers.

3.2. O. basilicum L. Row Plant and Essential Oil Composition and Accession Availability

Ocimum is one of the largest genera in the *Lamiaceae* family, which consists of 65 species native to Africa, South America, and Asia [23]. Among the species, sweet basil (*O. basilicum* Linn.), which originated from the warm tropical climates of India, Africa, and southern Asia, is probably the most important crop, being cultivated as a culinary herb worldwide under various ecological circumstances [24]. Both the raw plant and essential oil of basil have marked culinary, pharmaceutical, and cosmetic purposes [23]. It is used frequently in traditional medicine, having antispasmodic, stomachic, carminative, anti-ulcerogenic, anti-inflammatory, anti-carcinogenic, analgesic, stimulant radioprotective, and febrifuge properties [25]. Basil's leafy components exhibit antimicrobial properties and antioxidant activity [26,27] that can be used for alleviating pain and otitis [28]. The essential oil extracted from European genotype basil is recognized for its superior aroma, primarily comprising linalool and methyl chavicol.

Due to their vast diversity, various plant species and cultivars exhibit unique levels of resistance and ability to withstand physiological functions and produce yields in varying environmental conditions and stressful situations.

Furthermore, at the European level, based on the EURSICO National Inventory Report Taxonomy (ipk-gatersleben.de), a total number of 834 *O. basilicum* L. accessions are available for multiplication and future breeding perspectives (Figure 1). The countries that have uploaded the largest number of accessions are Germany (268 acc.), Croatia (119), the Czech Republic (64), and Romania (38).

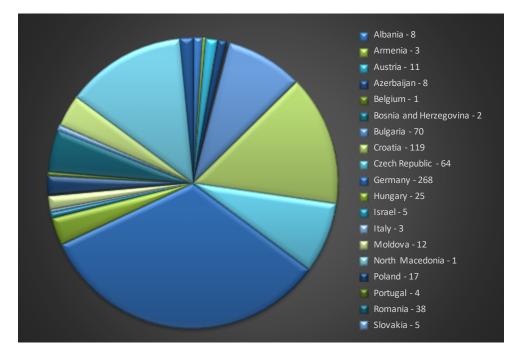


Figure 1. O. basilicum L. accession availability at the European level (https://eurisco.ipk-gatersleben.de/ apex/eurisco_ws/r/eurisco/taxon-search-results?p26_genus=OCIMUM&p26_species=BASILICUM, accessed on 14 February 2023).

3.3. T. vulgaris L. Composition and Accession Availability

The *Thymus* genera of the *Lamiaceae* family is highly significant due to its large number of species [29]. Thyme (*T. vulgaris* L.), commonly known as garden thyme or common thyme, appears in many different areas worldwide, including the drier Mediterranean regions [30], and is the most commercially cultivated species in the genus *Thymus* [31] due to the dietary trends in the recent decades [32]. It has several aromatic and medicinal properties. The leaves can be used either fresh or dried as a flavoring component in various culinary preparations, containing a high ratio of minerals (K, Ca, Mg, Fe, Mn, and Se), antioxidants (flavonoids, phenolic compounds such as pigenin, naringenin, luteolin, thymonin, lutein, and zeaxanthin), and vitamins (A, B6, B9, C, E, and K) [7,33,34]. In addition, the essential oil of common thyme is extracted by the distillation of the fresh leaves and flowering tops, and it contains 20–54% thymol, which is a monoterpene known as the main active ingredient with a wide range of pharmacological properties [7,35,36]. It has demonstrated anti-pathogenic and antioxidant effects, being intensively utilized in various industries, particularly in medication, agriculture, and food production.

At the European level, the total number of *T. vulgaris* L. accessions is 202, with Spain (77), Albania (51), Poland (15), and Ukraine (14) having uploaded the highest numbers of accessions to the EURISCO database (Figure 2).

3.4. S. hortensis L. Composition and Accession Availability

Summer savory (*S. hortensis* L.) is an annual herbaceous plant that is among the principal *Satureja* species grown in southern Europe, as well as central and southwestern Asia [37]. It contains numerous vitamins, including B-complex vitamins, vitamin A, vitamin C, niacin, thiamine, and pyridoxine, as well as carvacrol, terpinene, cymene, and caryophyllene, which make it an excellent choice for medicinal purposes [19,38]. The aerial parts of the *Satureja* species, such as *S. hortensis*, contain essential oil that is widely used

in the medicine, food, and health industries for therapeutic purposes. Scientific studies have highlighted several pharmacological properties of *Satureja*, such as antispasmodic, antioxidant, antimicrobial, antidiarrheal, and sedative properties [37,39–41]. Its beneficial effects on hypertension have also been discussed [42].

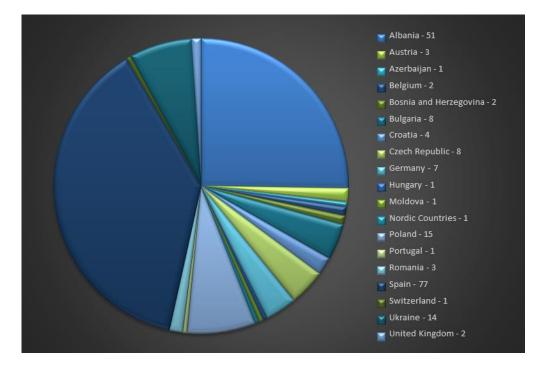


Figure 2. *T. vulgaris* L. accession availability at the European level (https://eurisco.ipk-gatersleben.de/ apex/eurisco_ws/r/eurisco/taxon-search-results?p26_genus=THYMUS&p26_species=VULGARIS, accessed on 14 February 2023).

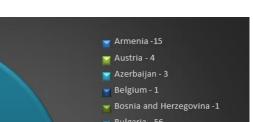
The savory essential oil contains two major compounds, thyme and carvacrol, that have antiseptic, antifungal, and antibacterial properties [3]. The concentration and composition of secondary metabolites in savory oils, such as y-terpinene, p-cymene, carvacrol methyl ether, and caryophyllene, play significant ecological roles as they possess insecticidal, antifungal, and antibacterial properties [43–45].

At the European level, the total number of *S. hortensis* L. accessions is 269, with Bulgaria (56), Romania (54), Germany (38), and Hungary (32) being the countries with the highest number of accessions uploaded to the EURISCO database (Figure 3).

3.5. Aromatic Plant Biological Activities and Stress Resistance

Various elements, such as genetic and ecological factors, have a notable impact on the chemical components of medicinal plants and their physiological and morphological characteristics [46]. Diverse environmental circumstances can be the primary cause of variability in morphological characteristics, which may induce alterations in the phenotype in the short term and in its genotype in the long term [47]. The presence of abiotic environmental stressors, such as salinity and drought, can inhibit the plant growth and development [48,49].

Essential oils perform a crucial function in plant defense by serving as antiviral, antibacterial, antimycotic, and insecticidal agents and deterring herbivores [50]. Due to these properties, diverse plant essential oils may serve as remedial or auxiliary agents in the pharmaceutical sector [51] and function as fragrances, seasonings, and natural preservatives in the food industry [52]. Finally, these oils may also serve as eco-friendly and biodegradable substances for protecting plants in agriculture [53].



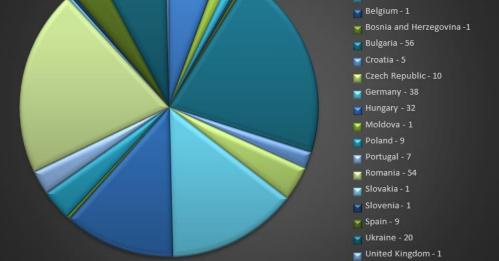


Figure 3. *S. hortensis* L. accession availability at the European level (https://eurisco.ipk-gatersleben.de/ apex/eurisco_ws/r/eurisco/taxon-search-results?p26_genus=SATUREJA&p26_species=HORTENSIS accessed on 14 February 2023).

3.6. O. basilicum L.—Biological Activities and Stress Resistance

Basil is regarded as a functional food plant due to its abundant secondary metabolites and antioxidant traits that are believed to enable oxidative stress disease prevention. The constituents of basil's essential oil (EO) are known to differ significantly based on the genetic factors (cultivar, origin, season, chemotype, and phonological stage), environmental factors (climatic conditions, agricultural practices, and postharvest processes), and the combined effects of these factors [4,54]. Oxygenated monoterpenes and phenylpropanoids are the primary chemical compounds present in the *Ocimum* genus, while linalool, eugenol, methyl chavicol, methyl cinnamate, methyl eugenol, and geraniol are some of the significant constituents identified in various *O. basilicum* cultivars and chemotypes [55]. These constituents act as potent antioxidants by scavenging free radicals and functioning as electron donors, making them effective in safeguarding the plants against pathogens and predators. At the cellular level, they can also protect cells from the adverse effects of ROS arising from different abiotic stressors [56].

Plants that have a high concentration of antioxidants can resist damage caused by ROS. These antioxidants can also function as protective substances [57,58]. The existence of phenolic compounds relies on various factors, such as the type of soil, plant species, genetics, growth stage, and location [59]. The primary phenolic compounds responsible for the antioxidant effects in basil are caffeic acid (CA) and rosmarinic acid (RA), which is an ester of CA. These compounds are mostly produced in roots and leaves [60]. CA is an active participant in plant physiology and stress tolerance mechanisms [56]. RA has been utilized as an anti-inflammatory, anti-proliferative, and chemoprotective agent [61]. The synthesis of RA in plants occurs through the phenylpropanoid and tyrosine-derived pathways. Phenylalanine ammonia-lyase (PAL) is the principal enzyme in the phenylpropanoid pathway, catalyzing the transformation of l-phenylalanine to trans-cinnamic acid and ammonia. Research has confirmed that PAL plays a role in RA biosynthesis [62]. An increase in PAL activity induced by stress could be the starting point for the cells to adapt to drought conditions [63]. Phosphorus (P) is an essential element to produce secondary metabolites in plants, and its availability can affect the amount and composition of phenolic

compounds. A competition exists between the production of phenolic compounds and the required proteins for growth.

Regarding the response of *O. basilicum* L. to different stressors, Table 2 presents the main anatomical, physiological, and molecular changes, as well as some contributing factors that could occur.

Туре	of Stress	Anatomical, Physiological, and Molecular Changes	Contributing Factors	References
		II' has NL+	Increased MDA accumulation	[64]
	of Stress Salinity Drought Twospotted spider mite (<i>Tetranychus</i>	Higher Na ⁺ concentrations	Enhanced proline content	[4]
		5	Photosynthetic pigments decrease	Chlorophyllase enzyme activity enhancement
		Induces essential oil production	Higher oil gland density	[4,65,66]
		Plant growth process is inhibited	Constrained cell elongation and differentiation	[67,68]
Abiotic			Chlorophyll reduction	[69]
Abloue	Disruption of main metabolic processes	Photosynthesis inhibation	[69]	
			Cell division suppresion	[69]
	Drought	Protein complexes imbalance	Chlorophyll a and b depletion	[70,71]
		Chlorophyllase activity enhancement		
		Photosynthesis inhibition	Stomatal blockage	[72]
			RubisCO enzyme activity cut	[73]
			Proline accumulation	[69]
	Cell osmotic adjustment	CO_2 assimilation	[74]	
		Small chlorotic spots	Lower concentrations of nitrogen, phosphorous, and protein	[75]
Biotic spider mite	Biotic Spider mite	Cell physiology disruption	Photosynthesis reduction and phytotoxic compounds injection	[75,76]
Diotic		Plasma membrane potential change	Cytosolic free Ca ²⁺ changes	[75]
		· -	Oxidative damage	Increase in cellular concentration of reactive oxygen species and, subsequently, of H ₂ O ₂

Table 2. O. basilicum L. response to different stressors.

Salinity stress has various negative impacts on plants, including reduced growth and water content [77]. Additionally, salinity stress results in reduced soil moisture content and limited water absorption from the soil, leading to osmotic stress in plants [66]. However, the osmotic potential significantly improves. Moreover, in many plants, salt stress increases the levels of cell free radicals to a point where it can damage the membrane, further intensifying the effects of the stress. The sensitivity of plants to salt stress can be measured by malondialdehyde (MDA), which is a widely used parameter for estimating lipid peroxidation in plant tissue that increases under oxidative stress. At the cellular level, osmotic stress modifies the properties and composition of the membrane lipids. MDA accumulation increased in the severity of NaCl stress in the leaves of summer savory (*S. hortensis* L.) and *O. basilicum* L. The MDA content serves as an indicator of oxidative

stress resulting from membrane lipid peroxidation, and it can be reduced by lowering lipid peroxidation and increasing the activity of antioxidant enzymes in salt-affected plants [64].

The composition and amount of essential oils can be influenced by environmental factors, including salinity stress, which can have a negative impact on plant growth and osmotic balance [78]. Under salinity stress, proline plays a crucial role in osmotic adjustment, helping to maintain the osmotic balance of the plant and enhance its tolerance to salt stress [79]. Proline also acts as an ROS scavenger, protein stabilizer, and osmoprotectant [80]. The accumulation of proline during drought stress has been shown to be related to improved plant performance, probably due to its antioxidant properties and its ability to stabilize macromolecules [81,82].

Regarding the drought stress, the accumulation of secondary metabolites is a defensive mechanism employed by plants to cope with stress by altering their cellular metabolism to overcome various challenges [83].

The concentration of leaf chlorophyll (Ch) is a crucial physiological characteristic that directly impacts a plant's photosynthetic ability. Both chlorophyll-a and chlorophyll-b levels in sweet basil plants decreased when they were deprived of water. The amount of chlorophyll in leaves, which is an indicator of plant vigor, is influenced by a variety of environmental factors. The reduction in chlorophyll content due to the water scarcity could be linked to the generation of ROS in cells [84], which have negative effects on plants in stressful situations. The plant water status is an indicator of their response to water scarcity, with higher relative water contents indicating a healthy plant condition [85]. In experiments involving various water supplies, the accurate evaluation of the plant water status is crucial. The water content of plants can be expressed per unit of fresh or dry weight (or, less commonly, per unit of leaf area) [86]. Particularly, the leaf relative water content (RWC) is utilized as a dependable measure of a plant's susceptibility to dehydration [87]. To reduce water loss through the leaves, the transpiration rate can be adjusted, and the leaf area can be limited. Typically, plants restrict water loss through their foliage by closing stomata, which reduces the rate of transpiration from the leaves. Nevertheless, under conditions of water scarcity, the root water uptake may be more critical in mitigating the damage caused by drought stress than controlling water loss through the leaves [88].

Employing plant growth regulators (PGRs) represents a viable means of enhancing plant resilience to stress, alongside other techniques such as selective breeding and genetic modification. There are numerous compounds that can alleviate drought-related stress in plants [89]. Salicylic acid, or 2-hydroxybenzoic acid, is a phenolic compound with hormone-like properties that can disrupt plant growth regulation, particularly when confronted with diverse stresses [90]. Additionally, it may trigger various physiological and biochemical functions in plants.

Moreover, salicylic acid (SA) has the ability to hinder catalase (CAT) function, which could result in the accumulation of hydrogen peroxide (H_2O_2) . This, in turn, may stimulate the operation of ROS-detoxifying enzymes and the production of antioxidant metabolites [91].

The ability of plants to adapt to environmental pressures, such as drought or salinity, requires temporary and long-lasting reductions in transpiration water flow. This physiological reaction is influenced by both inherent and induced genetic factors [92]. Stomata are of paramount importance in transpiration management, regulating plant water loss through density on the leaf surface and the mechanism of closure in response to environmental stimuli.

On the biotic nature stressors, mites prevent harm to the epidermis, thus reducing the chances of the leaf surface detecting the attack and postponing the plant's reaction. This pressure results in a modification of the plasma membrane potential and consequent alterations in the concentration of free cytosolic Ca^{2+} , which set off a signal that initiates a series of reactions [93]. One of the primary effects of biotic stress encountered by the plant is a rise in the cellular levels of ROS, which are then transformed into H₂O₂.

3.7. T. vulgaris L.—Biological Activities and Stress Resistance

T. vulgaris L. displays antimicrobial, anti-inflammatory, antioxidant, and immunomodulatory properties, being effective against a variety of ailments related to the respiratory, cardiovascular, and nervous systems, among others [94,95]. These effects are ascribed to phenolic acids, other phenols, and particularly the plant's essential oil. The essential oil is composed mainly of thymol, carvacrol, geraniol, α -terpineol, 4-thujanol, linalool, 1,8-cineole, myrcene, γ -terpinene, and p-cymene. The key components' abundance varies greatly depending on the plant chemotype, with the thymol chemotype being the most widespread [96,97]. This versatile plant finds widespread use in the food and pharmaceutical industries [51,52]. It is also a promising agent for crop protection and storage preservation [52]. From an economic perspective, the plant's biomass yield and essential oil quality are crucial, and various factors can affect them, including crop nutrition, manure application, water stress, seasonal variations, or processing [98,99].

When using the DPPH free radical scavenging method, *T. vulgaris* demonstrated a robust antioxidant activity of approximately 85% [100–102]. A similar outcome was observed regarding the antioxidant activity of the thyme methanol extract [103].

The disk diffusion method revealed that the extract exhibited very strong inhibition (20 mm inhibition zone) against various bacteria, including *Bacillus subtilis*, *Staphylococcus aureus*, *Pseudomonas aeruginosa*, and *Candida albicans* [104]. However, its inhibitory activity against *Candida tropicalis* was found to be moderate [13].

The antimicrobial activity of essential oils (EOs) is dependent on their chemical constituents. The antimicrobial activity of the analyzed EO is likely associated with the presence of phenolic compounds (thymol) and terpene hydrocarbons (γ -terpinene) [105,106]. It is believed that p-Cymene, the third major component, has synergistic effects with thymol and γ -terpinene [107], which may contribute to the observed antimicrobial activity. Furthermore, several studies have indicated that EOs exhibit stronger antimicrobial activity than their major constituents or mixtures [108,109], suggesting that minor components may have synergistic effects and emphasizing the importance of all components in relation to the biological activity of EOs [13].

The *T. vulgaris* essential oil demonstrated potent antimicrobial and antibiofilm effects, with MIC values ranging from 0.0625% to 2% v/v [110]. It also exhibited lower minimum inhibitory concentration values compared with the antibiotics tested on eradicating *Candida* genus biofilm [111]. When utilized in the vapor phase, it could serve as a viable alternative to antimicrobials in the food industry due to the lower concentration of EO required compared with the liquid phase contact effect [112].

The thyme essential oil contains elevated levels of TPC along with strong radical scavenging ability against DPPH, ABTS, and linoleic acid radicals, in addition to iron chelating capabilities. This positive correlation highlights the antioxidant potential of TEO to combat various oxidation systems and prevent oxidative damage [113].

At a concentration of 0.1 mg/mL, thyme oil and a CNC-based formulation of thyme white oil demonstrated complete larvicidal activity against *Aedes albopictus* [114].

The composition of bioactive secondary metabolites synthesized by medicinal plants varies widely depending on the species, having a profound impact on their relationship with endophytic microorganisms [115]. *Bacillus* spp. connected with *T. vulgaris*, such as *Bacillus sonorensis* (EGY05), *Bacillus tequilensis* (EGY21), and *Bacillus mojavensis* (EGY25), generated plant-growth-stimulating substances such as auxin, fixed nitrogen, soluble phosphate and iron, and lytic enzymes such as chitinase, cellulase, protease, and lipase. These bacteria may provide novel tactics to alleviate salt stress [116].

Phenolic substances and their corresponding enzymes (PAL and PPO) could potentially serve as protective factors against drought stress [6] that impacts the water status of leaves, pigments, and stomatal conductance, leading to the inhibition of photosynthesis [6]. The PPOs, in particular, play a role in various processes such as the Mehler reaction, the photoreduction of molecular oxygen by PSI, the regulation of oxygen levels in plastids, and the generation of the phenylpropanoid pathway [117]. The decrease in chlorophyll-a, chlorophyll-b, and total chlorophyll concentrations on thyme plants experiencing water stress may be viewed as an important regulatory measure to prevent excessive light absorption and to restrict the over-reduction of the photosynthetic electron transport chain, thereby limiting the generation of ROS [118]. ROS have been shown to be produced by both biotic and abiotic stresses [119], and these molecules are responsible for most of the oxidative damage to biological structures, including DNA, RNA, amino acids, proteins, and lipids [120].

Hydrogen peroxide (H_2O_2) is one of the most stable ROS which is produced in plant cells during different physiological processes, including photosynthesis, photorespiration, and, to a relatively lesser extent, respiration; it plays an important role as a signaling molecule under stressful conditions [121].

Malondialdehyde (MDA) is also widely known as a biochemical marker to increase the activity of ROS and the oxidative stress in plant tissues under adverse conditions. It is considered the most final product of lipid peroxidation and an important indicator of the oxidative damage that could occur in the cellular membrane under different stress conditions [7].

In plants, mutations on the epigenetic regulator histone deacetylase-6 (HDA-6) appear to improve survival in drought conditions [122]. This response is associated with the expression of genes involved in acetic acid biosynthesis. Therefore, in conditions of water stress, there would be a relationship between HAD-6 and the regulation of genes involved in acetic acid synthesis [123].

Preserving the integrity of cellular membranes during stressful circumstances is deemed a crucial aspect of any salinity adaptation mechanisms. The percentage of electrolyte leakage (ELP) indicates the level of injury to cell membranes. The application of salicylic acid (SA) amplified the ion leakage in thyme seedlings exposed to salt stress at greater concentrations, indicating that SA concentrations play a crucial role in saline environments.

The potential of ascorbic acid in mitigating and modifying the effects of salt stress on plants is well known. As a rule, its concentration is higher in leaves compared with other plant parts and is 5–10 times higher than that of glutathione [124]. In addition, the ascorbic acid's antioxidant role has been confirmed [125]. Therefore, plants require high endogenous levels of ascorbic acid to regulate various processes of plant metabolism in addition to countering oxidative stress. Endogenous levels of ascorbic acid can be elevated by exogenously administering ascorbic acid via the rooting medium, as a foliar spray or as seed priming. It plays a significant role in photosynthesis, specifically by regulating the redox state of photosynthetic electron carriers through the Mehler peroxidase reaction with ascorbate peroxidase and acting as a co-factor for violaxanthin deep oxidase, which is involved in xanthophyll cycle-mediated photoprotection [126]. As a result, in plants treated with ascorbic acid, high levels of pigments can work synergistically with the ascorbic acid to provide an efficient barrier against oxidation under salinity stress. Ascorbic acid can mitigate the detrimental effects of salinity by increasing the auxin and gibberellin content while reducing abscisic acid levels [127], which may help protect the photosynthetic apparatus and subsequently increase photosynthetic pigments.

During stressful periods, the accumulation of compatible osmolytes, such as proline, can serve as an appropriate marker for heavy metal contamination. In addition, proline may exhibit antioxidative properties that safeguard the cells from the detrimental effects of ROSs induced by Cd contamination due to a conducive environment for Cd sequestration and phytochelatin synthesis [128].

The main anatomical, physiological, and molecular changes of *T. vulgaris* related to different stressors are presented in Table 3.

Type of Stress		Anatomical, Physiological, and Molecular Changes	Contributing Factors	References
	Salinity	Nutritional imbalance in plant tissues	Electrolyte leakage increase	[103]
		Reduction of the photosynthetic capacity	CO ₂ assimilation reduction	[129]
	Drought	Mitigate cell division, elongation, and differentiation	Decreases cell turgor	[7,130]
			Minimizes enzyme activities	[7,130]
			Decreases energy supply	[7,130]
Abiotic		Photosynthetic processes reduction	Lower level of relative water content (RWC)	[131,132]
		Affects the level of endogenous phytohormones	Alters relations between ABA, ethylene, GA3, cytokinins, and auxins	[7,133–135]
		Reduces concentration of chlorophyll a and b and total chlorophyll	Increases ROS production	[7,135–137]
		Carotenoids concentration reduction	Enhancement of ABA hormone	[7,133]
		H ₂ O ₂ and lipid peroxidation enhancement	MDA concentration increase	[7]
		Adjustment of osmotic potential	Increased soluble sugars, proline, and free amino acid concentrations	[7]
		Considerable synthesis of total soluble phenols and phenylalanine ammonia-lyase (PAL)	Enhanced specific activity of PPO (polyphenol oxidase)	[7]
		Plant gene expression adjustment	Increased HDA-6 levels	[138–140]
		Secondary metabolites boost	Phe, Trp, and Asn amino acids	[141]
	Cd contamination (seeds)	Phytochelatin synthesis	Osmolytes (proline) accumulation	[142]
		ROS production enhancement	Increase in MDA content	[142]
Biotic	Aphis serpylli Koch	Cell physiology disruption	Carvacrol, Geraniol, and Thymol monoterpenes mitigate attack	[143,144]
			Linalol enhances attack	[143,144]

Table 3. T. vulgaris L. response to different stressors.

3.8. S. hortensis L.—Biological Activities and Stress Resistance

The EO extracted from summer savory contains a significant amount of carvacrol, which plays a crucial role in various biological activities, such as antimicrobial, antioxidant, antidiabetic, antihyperlipidemic, antispasmodic, antinociceptive, anti-inflammatory, antiproliferative, sedative, and reproduction stimulatory effects [145]. The EO content in different species of this genus is more than 5%, the major oil constituents being carvacrol, thymol, γ -terpinene, and borneol [146]. The chemical composition of the plant extracts is influenced by several factors, including the plant part used, harvest time, extraction method, plant cultivar or genotype, geographical location, storage, and climatic conditions [39].

Regarding its antimicrobial properties, summer savory volatile oils exhibit actions on cell membranes, causing interference, destabilization, and consequent effects on the phospholipid bilayer and enzyme activity [39]. The inhibitory effect of volatile oil against bacteria and fungi can be attributed to the higher content of biologically active compounds from the monoterpenes group, particularly terpinene, thymol, and carvacrol, where thymol has significant inhibitory activity against *S. aureus*, carvacrol and p-cymene against *Escherichia coli*, and γ -terpinene against *C. albicans* and *S. aureus* [17]. Thymol and carvacrol have increased activity against bacterial strains, while γ -terpinene and p-cymene are active against fungal strains [21,147]. The volatile oil extracted from *S. hortensis* L. has a broad antimicrobial spectrum, exhibiting inhibitory effects against 25 bacterial, eight fungal, and one yeast species [20]. Its activity against *E. coli*, *Salmonella typhimurium*, *S. aureus*, *Listeria monocytogenes*, and *Pseudomonas putida* isolated strains was also demonstrated [19]. The volatile oil has a higher concentration of antimicrobial compounds compared with the extracts [20,148].

The antioxidant activity of summer savory essential oil (SHEO) could be ascribed to the abundant content of carvacrol, γ -terpinene, p-cymene, and thymol compounds, which are known for their antioxidant properties [149,150]. Meanwhile, the components of extracts from *S. hortensis* (rosmarinic acid, caffeic acid, naringenin, quercetin, apigenin, kaempferol, luteolin, chlorogenic acid, rutin, and apigenin-glycoside) are also recognized for their antioxidant potential [151,152]. Due to their antioxidant activity, natural extracts derived from *S. hortensis* are being considered for use in the meat industry, with water leaf extract found to increase the shelf life of ground beef [153]. They can also be utilized as an antioxidant in mayonnaise formulations [154]. The presence of monoterpenes, such as carvacrol, cymene, and thymol, in the essential oil of *S. hortensis* suggests its potential for antimicrobial activities against food, plant, and human pathogens [155]. The antimicrobial mechanism involves damage to the integrity of the cell membrane, leading to the leakage of ions and other cell components and eventual death. At the same time, the antimicrobial properties of individual components of the essential oil are being evaluated [15].

The potential use of *S. hortensis* essential oil (SHEO) as a natural herbicide against two widely spread weeds, *Amaranthus retroflexus* and *Chenopodium album*, was also assessed [156]. The aerial parts of the plant were used during the fruit stage by hydrodistillation, and it was found to be rich in carvacrol and γ -terpinene (determined by GC-MS to be 55.66% and 31.98%, respectively). The essential oil was formulated as a nanoemulsion with a concentration of 5 mL/L, with an observed herbicidal activity both in laboratory conditions (at a nanoemulsion concentration of 1 mL/L) and in greenhouse conditions (at a nanoemulsion concentration of 4 mL/L).

There is a lack of data on the potential applications of *S. hortensis* essential oil in cancer treatment, although several other *Satureja* species have exhibited anticancer properties. For example, *S. intermedia* essential oil has shown potential against oesophageal squamous cell carcinoma and human bladder carcinoma cell lines, while *S. spicigera* has shown promise against Rectosigmoid adenocarcinoma cells, human epithelial colorectal adenocarcinoma cells, mouse embryo fibroblast cells, and ductal carcinoma cells. *S. sahendica* essential oil has demonstrated anticancer properties against breast cancer cells, fibroblast-like kidney cells, colon adenocarcinoma cells, and choriocarcinoma cells, while *S. montana* essential oil has shown potential against colon adenocarcinoma cells. These findings have been reported in various studies [17,155].

Some of the most important anatomical, physiological, and molecular changes of *S. hortensis* L. related to different stressors are presented in Table 4.

Type of Stress		Anatomical, Physiological and Molecular Changes	Contributing Factors	Reference
		Growth decrese	High osmotic proficiency	[49,66]
			Salt ions toxicity	[66]
			Cytokinin cutoff	[66]
			Enhanced inhibitor production	[66]
			Decreasing water and nutrient uptake	[49]
	-	Higher Na ⁺ concentrations	Lipid peroxidation increase	[157]
			Enhanced membrane damage	[157]
Abiotic	Salinity - -		Electrolyte leakage	[158]
			Increased MDA accumulation	[158]
			TPC, TSC, proline, and essential oil enhancement	[49]
		Chlorophyll content	Free oxygen radicals exposure/peroxidation	[49,159]
		Decreased transpiration rate	Gas exchange mittigation	[159]
		Imbalance in plant tissues	Reduced Ca and K	[49]
			Increased Cl and Na concentration	[49]
	Drought	Mitigate cell division, elongation, and differentiation	Decreases cell turgor	[160]
			Decreased relative water content (RWC)	
			Minimizes enzyme activities	[160]
			Decreases energy supply	[160]
			Reduced the plant height and the number of subsidiary branches	[43]
			Intensified malondialdehyde (MDA), H2O2, and proline contents	[43]
			Improved total chlorophyll, chlorophyll a and b, and carotenoid contents	[43]
Biotic	Botrytis	Necrosis and narrowing tissues s.	Endopolygalacturonase content enhancement	[161–163]
	<i>cinerea</i> Pers.		Pectin degradation	

During stress periods, terpene emissions and related attracting mechanisms can indirectly contribute to plant defense mechanisms [164]. Furthermore, certain volatile compounds may act as airborne signals that can either directly or indirectly trigger systemic resistance and defense responses in neighboring plants [165,166].

Methyl jasmonate (MJ) treatment has been found to up-regulate genes involved in Jasmonate biosynthesis, secondary metabolism, and cell wall formation, as well as genes that encode stress-protective and defense proteins. Conversely, genes that are involved in photosynthesis, such as ribulose bisphosphate carboxylase/oxygenase, chlorophyll a/b-binding protein, and light-harvesting complex II, are down-regulated [167].

Gibberellin may have a potential role in aiding plants to adapt to stress [168,169]. Thus, the external application of it may reduce the negative impacts of salinity, while also enhancing growth under saline conditions, as evidenced by increased nutrient uptake, dry weights, plant height, leaf area, and yield, mitigating NaCl-induced growth inhibition [170,171]. Specifically, gibberellin application increased transpiration rate, relative water content, chlorophyll b, total chlorophyll, and xanthophyll content under salinity stress conditions for savory plants. The external application of gibberellin may enhance plant growth by elevating endogenous gibberellin levels [168]. Overall, gibberellin plays a crucial role in boosting plant growth, pigment synthesis, and photosynthesis rate under salt stress conditions [169].

In plants, the contents of hydrogen peroxide (H_2O_2) and malondialdehyde (MDA) reduced when inoculated with bacteria under well-watered and water stress conditions due to a significant increase in the expression of antioxidant enzymes, leading to a decline in MDA levels and electrolyte leakage [172]. Within plant cells, certain compounds such as lipid-soluble antioxidants (e.g., α -tocopherol and carotenoids), water-soluble reductants (e.g., glutathione and ascorbate), and antioxidant enzymes can provide protection against the harmful effects of ROS [173]. Moreover, the accumulation of certain osmolytes (e.g., proline) in plant cells can aid in scavenging free radicals and safeguarding enzymes [174].

3.9. Basil, Thyme, and Summer Savory Morphological Response to Different Biotic and Abiotic Stressors

All the three *Lamiaceae* herbs have developed several morphological adaptations that enable them to cope with biotic and abiotic stresses, such as leathery leaves, pubescence, and deep root systems.

Leathery leaves, one of the main adaptations, are characterized by a thick cuticle and more sclerenchyma tissue than other types of leaves, with a pronounced retaining water role, reducing water loss through transpiration, while conferring resistance to drought stress [175]. This adaptation allows these herbs to survive in arid conditions by maintaining hydration. Leathery leaves also help the plants to cope with high temperatures, which can cause dehydration and damage to the plant tissues. Moreover, the amount of epicuticular wax, a waxy substance found on the surface of leaves, was positively correlated with drought tolerance in these herbs [175]. The ability to tolerate water stress is linked to minor modifications in cellular biochemistry due to the buildup of compatible solutes and particular proteins that can be swiftly triggered by osmotic stress [176]. Water scarcity impacts plant development at diverse degrees, from the cell to the tissue level [177].

The presence of pubescence on the surface of these herbs' leaves represents another significant mechanism of resistance (antixenosis) [175]. These tiny hairs act as a physical barrier against herbivores, such as insects and grazing animals, and reduce water loss through transpiration. They also help the plants to reflect some of the sunlight and reduce the amount of energy absorbed by the leaves, which protects them from photoinhibition or damage caused by excessive sunlight [175]. The phenylpropenes, along with terpenoids, are the primary components of essential oils that are released from the glandular trichomes of various *Lamiaceae* species [178]. In the particular case of basil, it predominantly synthesizes and stores eugenol and methyl chavicol in its glandular trichomes [179].

In addition, the three herbs have developed deep root systems, which enable them to access water and nutrients from deeper layers of soil and provide stability to the plant by anchoring it firmly in the ground. The deep roots allow the plants to survive in nutrient-poor soil and to withstand periods of drought stress by accessing water that is not available to shallower rooted plants [175]. Deep roots also provide stability to the plant by anchoring it firmly in the ground, protecting it from wind and water erosion. The root length and surface area of basil, thyme, and summer savory are positively correlated with their ability to cope with drought stress. In addition, a high root-to-shoot ratio, which is an adaptation that allows them to absorb water more efficiently and store it in their roots for future use, was observed [175].

3.10. Breeding Perspectives Regarding the Adaptability to the Main Abiotic Stressors

Genetic predisposition represents a fundamental requirement for enhancing the quality and yield of essential oils, with variety selection and plant breeding as additional factors. Induced polyploidization is one of the plant breeding techniques that can affect a plant's genome, phenotype, physiology, and metabolome, enabling us to develop novel genotypes with better morphological, physiological, and biochemical properties.

Abiotic stressors such as drought, salinity, and extreme temperatures pose major challenges for herb cultivation. To address these challenges, research needs to focus on developing new cultivars of basil, thyme, and summer savory that are more resilient to abiotic stressors. One approach has been to identify genetic markers and traits associated with stress tolerance and breed for improved adaptability to stressful conditions. Studies have identified candidate genes and quantitative trait loci (QTLs) associated with drought tolerance in basil, thyme, and summer savory and used these genetic markers to develop new cultivars with improved drought resistance.

Another promising approach has been to explore the use of plant growth-promoting rhizobacteria (PGPR) and other beneficial microorganisms to enhance the stress tolerance of these herbs. Recent studies have shown that the application of PGPR can improve the growth, yield, and quality of basil, thyme, and summer savory under stressful conditions and enhance their resistance to pests and diseases.

In addition to phenotypic screening, molecular and biochemical approaches have been used to elucidate the mechanisms of stress tolerance in basil, thyme, and summer savory, as well as the identification of the key compounds involved. Numerous investigations have highlighted the significance of aquaporins in the plant stress response. TaTIP2;2 functions as an inhibitor of drought and salinity stress, its reaction not being reliant on ABA [180]. The transcriptomic and metabolomic responses of thyme plants to heat stress were investigated, and several candidate genes and metabolites associated with thermotolerance, including heat shock proteins, proline, and flavonoids, were identified [181]. Similarly, the volatile compounds and antioxidant activity of summer savory leaves under drought stress were analyzed, and the conclusions were that some volatile terpenes, such as γ -terpinene and carvacrol, were positively correlated with drought tolerance and antioxidant capacity [43].

To identify the compounds important for stress resistance in basil, thyme, and summer savory, several approaches have been used, such as metabolomics, transcriptomics, and proteomics. Analyses of basil leaves' metabolites exposed to a water deficit found that the accumulation of phenolic acids and flavonoids was associated with drought tolerance [182]. Additionally, when the proteome of summer savory leaves under drought stress was investigated, it was found that the up-regulated proteins were related to photosynthesis, antioxidant defense, and stress response [183].

Regarding the germplasm availability, summer savory genotypes were evaluated for their tolerance to cold stress, and a promising candidate ("Mutika") was identified, exhibiting better root volume, aerial part and total fresh weights, stem height, and flower number under low-temperature conditions [184].

3.11. Breeding Perspectives Regarding the Adaptability to the Main Biotic Stressors

Pests (aphids, thrips, spider mites, and whiteflies) are major biotic stressors that can affect basil, thyme, and summer savory crops. They feed on plant sap, causing stunted growth, leaf curling, and discoloration, together with a reduction in herb yield quantity and quality. Moreover, these pests can transmit viral diseases, such as tomato spotted wilt virus (TSWV), which can cause severe damage to the crops [185]. Breeding for pest-resistant cultivars using genetic markers could be a promising solution to reduce the use of insecticides and mitigate the effects of insect pests on the crops.

Diseases, such as fungal and bacterial infections, are also significant biotic stressors that can affect basil, thyme, and savory crops. The most common fungal diseases in these herbs include powdery mildew, downy mildew, and gray mold, which can cause leaf wilting, yellowing, and necrosis. Triggering defense genes against specific pathogens is influenced by distinct environmental factors. This indicates the involvement of intricate signaling pathways that empower plants to identify and safeguard against various stressors, including pathogenic threats [186,187]. Therefore, there is a need for breeding programs that focus on developing disease-resistant varieties of these crops. However, due to the limitations of traditional breeding, such as time-consuming and limited genetic diversity, new breeding technologies, such as genome editing, could provide an efficient and more precise method to introduce disease resistance into these crops [188].

Herbivores, such as deer, rabbits, and rodents, can also cause significant damage to basil, thyme, and savory crops. These herbivores can reduce crop yield by feeding on the plants and transmit plant diseases through their saliva [183]. Developing herbivore-resistant varieties of these crops can help reduce the impact of herbivores on crop production.

Weed competition is another important biotic stress that can affect the growth and yield of basil, thyme, and savory crops. Weeds compete with the herbs for nutrients, water, and sunlight, leading to reduced yield and quality of the crops. Furthermore, weeds can also act as hosts for pests and diseases, increasing their population and spread in the crop field. However, the excessive use of herbicides can have negative impacts on the environment and human health. Therefore, breeding for herbicide-resistant cultivars could be an important strategy in future breeding perspectives.

In terms of germplasm availability, when *Ocimum* accessions were screened for resistance to downy mildew caused by *Peronospora belbahrii*, the conclusion was that some genotypes, such as "Spice", exhibited significantly lower disease severity and higher yield than others [189].

By overexpressing a gene encoding a chalcone synthase in basil plants, an increased resistance to Fusarium wilt caused by *F. oxysporum*, as well as higher levels of glyphosate-resistant basil and thyme flavonoids and phenolic acids were observed [190].

MYB and MYC proteins play a critical role in plants' ability to cope with unfavorable environmental conditions. AtMYB30 functions as an activator of the hypersensitive cell death program upon pathogenic attack [191], while AtMYB33 and AtMYB101 are associated with ABA-mediated reactions to environmental cues [192]. AtMYB96 regulates water scarcity and disease resistance by acting through the ABA signaling pathway [193], and AtMYB15 is involved in enhancing cold stress tolerance [194].

Breeding for biotic stress resistance in basil, thyme, and savory crops is essential to ensure their sustainable cultivation and production. In recent years, there have been significant advancements in molecular breeding techniques, such as marker-assisted selection (MAS) and genomic selection (GS), which can accelerate the breeding process and improve the efficiency of selecting stress-tolerant traits [195]. Moreover, the identification of stress-responsive genes and pathways in these herbs can provide valuable targets for genetic engineering and biotechnological approaches to enhance their resistance to biotic stressors [192]. The breeding of basil with resistance to *F. oxysporum* has shown promising results [196].

Molecular markers and genetic engineering techniques can be used to accelerate the breeding process and identify genes responsible for biotic stress resistance in basil, thyme, and savory. Several resistance genes from basil (Pb1A and Pb1A'), which are responsible for resistance to downy mildew, a common fungal disease, were successfully transferred by researchers [197]. Similarly, genetic engineering can be used to develop herbicide-resistant cultivars.

4. Conclusions

Selecting and breeding basil, thyme, and summer savory genotypes with enhanced tolerance to biotic and abiotic stresses is crucial for their sustainable and profitable cultivation. Phenotypic screening, molecular and biochemical approaches, and genetic engineering or exogenous application of bioactive compounds are effective strategies for identifying and enhancing stress tolerance in these herbs. Identification of key compounds and genes involved in stress resistance can also provide valuable insights for further improvement of these important crops.

The research suggests that there is a significant potential for breeding and genetic improvement, as well as the use of microbial-based strategies, to enhance the adaptability of basil, thyme, and summer savory to abiotic stressors. These efforts could have important implications for the sustainability and productivity of herb cultivation in a changing climate and could help to ensure a reliable supply of these valuable herbs for food, medicinal, and other applications.

Author Contributions: Conceptualization, D.I.A., M.C., P.M.B., C.B. and O.C.M.; methodology, D.I.A., M.C., P.M.B., C.B. and O.C.M.; validation, C.B.; resources, P.M.B. and C.B.; data curation, D.I.A.; writing—original draft preparation, D.I.A. and M.C.; writing—review and editing, D.I.A., M.C., P.M.B., C.B. and O.C.M.; visualization D.I.A. M.C., P.M.B. and C.B.; supervision, D.I.A. and C.B.; project administration, P.M.B.; funding acquisition, P.M.B. All authors have read and agreed to the published version of the manuscript.

Funding: The authors acknowledge the in-kind and cash support by the Vegetable Research and Development Station (VRDS), Bacau, Romania, Project no. 529/2018.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Availability Statements are available in section "MDPI Research Data Policies" at https://www.mdpi.com/ethics accessed on 7 February 2023. No new data were created.

Acknowledgments: This work was carried out in frame of Project no. 529/2018 National Project funded from the State budget through "Gheorghe Ionescu-Sisesti" Academy of Agricultural and Forestry Sciences and the ADER program funded by the Ministry of Agriculture and Rural Development.

Conflicts of Interest: Not the case applicable.

References

- Dragland, S.; Senoo, H.; Wake, K.; Holte, K.; Blomhoff, R. Several Culinary and Medicinal Herbs Are Important Sources of Dietary Antioxidants. J. Nutr. 2003, 133, 1286–1290. [CrossRef]
- Danesi, F.; Elementi, S.; Neri, R.; Maranesi, M.; D'antuono, L.F.; Bordoni, A. Effect of Cultivar on the Protection of Cardiomyocytes from Oxidative Stress by Essential Oils and Aqueous Extracts of Basil (*Ocimum basilicum* L.). J. Agric. Food Chem. 2008, 56, 9911–9917. [CrossRef]
- Ahmad, P.; Prasad, M.N.V. (Eds.) Abiotic Stress Responses in Plants: Metabolism, Productivity and Sustainability; Springer Science & Business Media: Berlin/Heidelberg, Germany, 2011.
- 4. Farsaraei, S.; Moghaddam, M.; Pirbalouti, A.G. Changes in growth and essential oil composition of sweet basil in response of salinity stress and superabsorbents application. *Sci. Hortic.* **2020**, 271, 109465. [CrossRef]
- Wang, W.-X.; Vinocur, B.; Altman, A. Plant responses to drought, salinity and extreme temperatures: Towards genetic engineering for stress tolerance. *Planta* 2003, 218, 1–14. [CrossRef] [PubMed]
- Ibrahim, M.F.M.; Bondok, A.M.; Al-Senosy, N.K.; Younis, R.A.A. Stimulation Some of Defense Mechanisms in Tomato Plants under Water Deficit and Tobacco mosaic virus (TMV). World J. Agric. Sci. 2015, 11, 289–302.
- Farag, R.E.; Abdelbar, O.H.; Shehata, S.A. Impact of Drought Stress on Some Growth, Biochemical and Anatomical Parameters of *Thymus vulgaris* L. *Arab. Univ. J. Agric. Sci.* 2019, 27, 37–50. [CrossRef]
- 8. Kiczorowska, B.; Klebaniuk, R.; Bakowski, M.; Al-Yasiry, M.H. Culinary Herbs-the Nutritive Value and Content of Minerals. *J. Elem.* 2015, 20, 599–608. [CrossRef]
- 9. Gebrehiwot, H.; Bachetti, R.; Dekebo, A. Chemical composition and antimicrobial activities of leaves of sweet basil (*Ocimum basilicum* L.). *Herb. Int. J. Basic Clin. Pharmacol.* **2015**, *4*, 869–875. [CrossRef]
- 10. Politeo, O.; Jukic, M.; Milos, M. Chemical composition and antioxidant capacity of free volatile aglycones from basil (*Ocimum basilicum* L.) compared with its essential oil. *Food Chem.* **2007**, *101*, 379–385. [CrossRef]
- 11. Hussain, A.I.; Anwar, F.; Sherazi, S.T.H.; Przybylski, R. Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chem.* **2008**, *108*, 986–995. [CrossRef]
- 12. Guillén, M.; Manzanos, M. Study of the composition of the different parts of a Spanish *Thymus vulgaris* L. plant. *Food Chem.* **1998**, 63, 373–383. [CrossRef]
- 13. Borugă, O.; Jianu, C.; Mişcă, C.; Goleţ, I.; Gruia, A.T.; Horhat, F.G. *Thymus vulgaris* essential oil: Chemical composition and antimicrobial activity. *J. Med. Life* **2014**, *7*, 56–60. [PubMed]

- 14. Ravindran, P.N.; Pillai, G.S.; Divakaran, M. Other Herbs and Spices: Mango Ginger to Wasabi. In *Handbook of Herbs and Spices*; Peter, K.V., Ed.; Woodhead Publishing: Cambridge, UK, 2012; Volume 2, pp. 557–582.
- 15. Fierascu, I.; Dinu-Pirvu, C.E.; Fierascu, R.C.; Velescu, B.S.; Anuta, V.; Ortan, A.; Jinga, V. Phytochemical Profile and Biological Activities of *Satureja hortensis* L.: A Review of the Last Decade. *Molecules* **2018**, *23*, 2458. [CrossRef] [PubMed]
- Skubij, N.; Dzida, K.; Jarosz, Z.; Pitura, K.; Jaroszuk-Sierocińska, M. Nutritional value of savory herb (*Satureja hortensis* L.) and plant response to variable mineral nutrition conditions in various phases of development. *Plants* 2020, *9*, 706. [CrossRef] [PubMed]
- 17. Hamidpour, R.; Hamidpour, S.; Hamidpour, M.; Shahlari, M.; Sohraby, M. Summer Savory: From the Selection of Traditional Applications to the Novel Effect in Relief, Prevention, and Treatment of a Number of Serious Illnesses Such as Diabetes, Cardiovascular Disease, Alzheimer's Disease, and Cancer. *J. Tradit. Complement. Med.* **2014**, *4*, 140–144. [CrossRef]
- 18. Tepe, B.; Cilkiz, M. A Pharmacological and Phytochemical Overview onSatureja. Pharm. Biol. 2015, 54, 375–412. [CrossRef]
- 19. Mihajilov-Krstev, T.; Radnović, D.; Kitić, D.; Zlatković, B.; Ristić, M.; Branković, S. Chemical Composition and Antimicrobial Activity of *Satureja hortensis* L. *Essential Oil. Open Life Sci.* 2009, *4*, 411–416. [CrossRef]
- Mahboubi, M.; Kazempour, N. Chemical composition and antimicrobial activity of *Satureja hortensis* and Trachyspermum copticum essential oil. *Iran. J. Microbiol.* 2011, *3*, 194–200.
- Farzaneh, M.; Kiani, H.; Sharifi, R.; Reisi, M.; Hadian, J. Chemical Composition and Antifungal Effects of Three Species of Satureja (*S. Hortensis, S. Spicigera*, and *S. Khuzistanica*) Essential Oils on the Main Pathogens of Strawberry Fruit. *Postharvest Biol. Technol.* 2015, 109, 145–151. [CrossRef]
- 22. Saeidnia, S.; Gohari, A.R.; Manayi, A.; Kourepaz-Mahmoodabadi, M. Satureja: Ethnomedicine, Phytochemical Diversity and Pharmacological Activities; Springer: Berlin/Heidelberg, Germany, 2016; pp. 31–40.
- 23. Makri, O.; Kintzios, S. *Ocimum* sp. (Basil): Botany, Cultivation, Pharmaceutical Properties, and Biotechnology. J. Herbs Spices Med. *Plants* 2008, 13, 123–150. [CrossRef]
- 24. Adham, A.N. Comparative extraction methods, phytochemical constituents, fluorescence analysis and HPLC validation of rosmarinic acid content in Mentha piperita, Mentha longifolia and Ocimum basilicum. J. Pharmacogn. Phytochem. 2015, 3, 130–139.
- 25. Shirazi, M.T.; Gholami, H.; Kavoosi, G.; Rowshan, V.; Tafsiry, A. Chemical composition, antioxidant, antimicrobial and cytotoxic activities of *T agetes minuta* and *O cimum basilicum* essential oils. *Food Sci. Nutr.* **2014**, *2*, 146–155. [CrossRef]
- 26. Koseki, P.M.; Villavicencio, A.L.C.H.; Brito, M.S.; Nahme, L.C.; Sebastião, K.I.; Rela, P.R.; de Almeida-Muradian, L.B.; Mancini-Filho, J.; Freitas, P.C. Effects of irradiation in medicinal and eatable herbs. *Radiat. Phys. Chem.* **2002**, *63*, 681–684. [CrossRef]
- 27. Hakkim, F.L.; Shankar, C.G.; Girija, S. Chemical Composition and Antioxidant Property of Holy Basil (*Ocimum sanctum* L.) Leaves, Stems, and Inflorescence and Their in Vitro Callus Cultures. J. Agric. Food Chem. 2007, 55, 9109–9117. [CrossRef]
- 28. McClatchey, W. The ethnopharmacopoeia of Rotuma. J. Ethnopharmacol. 1996, 50, 147–156. [CrossRef] [PubMed]
- 29. Stahl-Biskup, E.; Venskutonis, R.P. Thyme. In Handbook of Herbs and Spices; Woodhead Publishing: Sawston, UK, 2012; pp. 499–525.
- Thompson, J.; Charpentier, A.; Bouguet, G.; Charmasson, F.; Roset, S.; Buatois, B.; Vernet, P.; Gouyon, P.-H. Evolution of a Genetic Polymorphism with Climate Change in a Mediterranean Landscape. *Proc. Natl. Acad. Sci. USA* 2013, 110, 2893–2897. [CrossRef]
- 31. Stahl-Biskup, E.; Sáez, F. Thyme: The Genus Thymus; Taylor & Francis: London, UK, 2002; ISBN 0-415-28488-0.
- Rowland, L.S.; Smith, H.K.; Taylor, G. The potential to improve culinary herb crop quality with deficit irrigation. *Sci. Hortic.* 2018, 242, 44–50. [CrossRef]
- 33. Komaki, A.; Hoseini, F.; Shahidi, S.; Baharlouei, N. Study of the effect of extract of *Thymus Vulgaris* on anxiety in male rats. *J. Tradit. Complement. Med.* **2016**, *6*, 257–261. [CrossRef]
- Dauqan, E.M.A.; Abdullah, A. Medicinal and Functional Values of Thyme (*Thymus vulgaris* L.) Herb. J. Appl. Biol. Biotechnol. 2017, 5, 17–22. [CrossRef]
- Fachini-Queiroz, F.C.; Kummer, R.; Estevão-Silva, C.F.; Carvalho, M.D.D.B.; Cunha, J.M.; Grespan, R.; Bersani-Amado, C.A.; Cuman, R.K.N. Effects of Thymol and Carvacrol, Constituents of *Thymus vulgaris* L. Essential Oil, on the Inflammatory Response. *Evid. Based Complement. Altern. Med.* 2012, 2012, 657026. [CrossRef] [PubMed]
- Nikolić, M.; Glamočlija, J.; Ferreira, I.C.F.R.; Calhelha, R.C.; Fernandes, Â.; Marković, T.; Marković, D.; Giweli, A.; Soković, M. Chemical composition, antimicrobial, antioxidant and antitumor activity of *Thymus serpyllum* L., *Thymus algeriensis* Boiss. and Reut and *Thymus vulgaris* L. Essential Oils. *Ind. Crops Prod.* 2014, 52, 183–190. [CrossRef]
- Gontaru, L.; Plander, S.; Simándi, B. Investigation of *Satureja hortensis* L. as a possible source of natural antioxidants. *Hung. J. Ind. Chem.* 2008, 1-2, 36. [CrossRef]
- Jadczak, D. Effect of sowing date on the quantity and quality of the yield of summer savory (*Satureja hortensis* L.) grown for a bunch harvest. *Herba Pol.* 2007, 53, 22–27.
- Hadian, J.; Ebrahimi, S.N.; Salehi, P. Variability of morphological and phytochemical characteristics among Satureja hortensis L. Accessions of Iran. Ind. Crops Prod. 2010, 32, 62–69. [CrossRef]
- Hajhashemi, V.; Zolfaghari, B.; Yousefi, A. Antinociceptive and Anti-Inflammatory Activities of *Satureja hortensis* Seed Essential Oil, Hydroalcoholic and Polyphenolic Extracts in Animal Models. *Med. Princ. Pract.* 2011, 21, 178–182. [CrossRef]
- Güllüce, M.; Sökmen, M.; Daferera, D.; Ağar, G.; Özkan, H.; Kartal, N.; Polissiou, M.; Sökmen, A.; Şahin, F. In Vitro Antibacterial, Antifungal, and Antioxidant Activities of the Essential Oil and Methanol Extracts of Herbal Parts and Callus Cultures of *Satureja hortensis* L. J. Agric. Food Chem. 2003, 51, 3958–3965. [CrossRef]

- 42. Svoboda, K. Investigation of volatile oil glands of *Satureja hortensis* L. (summer savory) and phytochemical comparison of different varieties. *Int. J. Aromather.* 2003, *13*, 196–202. [CrossRef]
- Mohammadi, H.; Dashi, R.; Farzaneh, M.; Parviz, L.; Hashempour, H. Effects of beneficial root pseudomonas on morphological, physiological, and phytochemical characteristics of *Satureja hortensis* (Lamiaceae) under water stress. *Braz. J. Bot.* 2016, 40, 41–48. [CrossRef]
- Cappellari, L.D.R.; Santoro, M.V.; Nievas, F.; Giordano, W.; Banchio, E. Increase of secondary metabolite content in marigold by inoculation with plant growth-promoting rhizobacteria. *Appl. Soil Ecol.* 2013, 70, 16–22. [CrossRef]
- 45. Farzaneh, V.; Carvalho, I.S. A review of the health benefit potentials of herbal plant infusions and their mechanism of actions. *Ind. Crop. Prod.* **2015**, *65*, 247–258. [CrossRef]
- 46. Heywood, V.H. The Conservation of Genetic and Chemical Diversity in Medicinal and Aromatic Plants. *Biodiversity* **2002**, 13–22. [CrossRef]
- Saito, K.; Matsuda, F. Metabolomics for Functional Genomics, Systems Biology, and Biotechnology. Annu. Rev. Plant Biol. 2010, 61, 463–489. [CrossRef]
- 48. Flowers, T.J.; Muscolo, A. Introduction to the Special Issue: Halophytes in a changing world. AoB Plants 2015, 7, plv020. [CrossRef]
- Estaji, A.; Roosta, H.R.; Rezaei, S.A.; Hosseini, S.S.; Niknam, F. Morphological, physiological and phytochemical response of different *Satureja hortensis* L. accessions to salinity in a greenhouse experiment. *J. Appl. Res. Med. Aromat. Plants* 2018, 10, 25–33. [CrossRef]
- 50. Bakkali, F.; Averbeck, S.; Averbeck, D.; Idaomar, M. Biological effects of essential oils—A review. *Food Chem. Toxicol.* 2008, 46, 446–475. [CrossRef]
- Kokoska, L.; Kloucek, P.; Leuner, O.; Novy, P. Plant-Derived Products as Antibacterial and Antifungal Agents in Human Health Care. Curr. Med. Chem. 2019, 26, 5501–5541. [CrossRef]
- 52. Pandey, A.K.; Kumar, P.; Singh, P.; Tripathi, N.N.; Bajpai, V.K. Essential Oils: Sources of Antimicrobials and Food Preservatives. *Front. Microbiol.* **2017**, *7*, 2161. [CrossRef]
- Pavela, R.; Benelli, G. Essential Oils as Ecofriendly Biopesticides? Challenges and Constraints. *Trends Plant Sci.* 2016, 21, 1000–1007. [CrossRef]
- 54. Moghaddam, M.; Pirbalouti, A.G.; Mehdizadeh, L.; Pirmoradi, M.R. Changes in composition and essential oil yield of Ocimum ciliatum at different phenological stages. *Eur. Food Res. Technol.* **2014**, 240, 199–204. [CrossRef]
- Pirbalouti, A.G.; Malekpoor, F.; Salimi, A.; Golparvar, A. Exogenous application of chitosan on biochemical and physiological characteristics, phenolic content and antioxidant activity of two species of basil (Ocimum ciliatum and Ocimum basilicum) under reduced irrigation. *Sci. Hortic.* 2017, 217, 114–122. [CrossRef]
- 56. Riaz, U.; Kharal, M.A.; Murtaza, G.; Zaman, Q.U.; Javaid, S.; Malik, H.A.; Aziz, H.; Abbas, Z. Prospective Roles and Mechanisms of Caffeic Acid in Counter Plant Stress: A Mini Review. *Pak. J. Agric. Res.* **2018**, *32*, 8–19. [CrossRef]
- 57. Zare, M.; Ganjeali, A.; Lahouti, M. Rosmarinic and caffeic acids contents in Basil (*Ocimum basilicum* L.) are altered by different levels of phosphorus and mycorrhiza inoculation under drought stress. *Acta Physiol. Plant.* **2021**, *43*, 26. [CrossRef]
- Meot-Duros, L.; Magné, C. Antioxidant activity and phenol content of *Crithmum maritimum* L. Leaves. *Plant Physiol. Biochem.* 2009, 47, 37–41. [CrossRef]
- Scagel, C.F.; Lee, J. Phenolic Composition of Basil Plants Is Differentially Altered by Plant Nutrient Status and Inoculation with Mycorrhizal Fungi. *Hortscience* 2012, 47, 660–671. [CrossRef]
- 60. Kwee, E.M.; Niemeyer, E.D. Variations in phenolic composition and antioxidant properties among 15 basil (*Ocimum basilicum* L.) cultivars. *Food Chem.* **2011**, *128*, 1044–1050. [CrossRef]
- 61. Srivastava, S.; Conlan, X.A.; Adholeya, A.; Cahill, D.M. Elite hairy roots of *Ocimum basilicum* as a new source of rosmarinic acid and antioxidants. *Plant Cell Tissue Organ Cult. (PCTOC)* **2016**, *126*, 19–32. [CrossRef]
- Kim, Y.B.; Kim, J.K.; Uddin, M.R.; Xu, H.; Park, W.T.; Tuan, P.A.; Li, X.; Chung, E.; Lee, J.-H.; Park, S.U. Metabolomics Analysis and Biosynthesis of Rosmarinic Acid in Agastache rugosa Kuntze Treated with Methyl Jasmonate. *PLoS ONE* 2013, *8*, e64199. [CrossRef]
- 63. Hazzoumi, Z.; Moustakime, Y.; Elharchli, E.H.; Joutei, K.A. Effect of arbuscular mycorrhizal fungi (AMF) and water stress on growth, phenolic compounds, glandular hairs, and yield of essential oil in basil (*Ocimum gratissimum* L.). *Chem. Biol. Technol. Agric.* **2015**, *2*, 10. [CrossRef]
- 64. Delavari, P.M.; Baghizadeh, A.; Enteshari, S.H.; Kalantari, K.M.; Yazdanpanah, A.; Mousavi, E.A. The effects of salicylic acid on some of biochemical and morphological characteristic of *Ocimum basilicucm* under salinity stress. *Aust. J. Basic Appl. Sci.* 2010, *4*, 4832–4845.
- 65. Heidari, M. Effects of salinity stress on growth, chlorophyll content and osmotic components of two basil (*Ocimum basilicum* L.) genotypes. *Afr. J. Biotechnol.* **2011**, *11*, 379–384. [CrossRef]
- 66. Gupta, B.; Huang, B. Mechanism of Salinity Tolerance in Plants: Physiological, Biochemical, and Molecular Characterization. *Int. J. Genom.* **2014**, 2014, 701596. [CrossRef]
- 67. Tardieu, F.; Reymond, M.; Hamard, P.; Granier, C.; Muller, B. Spatial distributions of expansion rate, cell division rate and cell size in maize leaves: A synthesis of the effects of soil water status, evaporative demand and temperature. *J. Exp. Bot.* **2000**, *51*, 1505–1514. [CrossRef]

- Dos Santos, M.S.; Costa, C.A.S.; Gomes, F.P.; do Bomfim Costa, L.C.; de Oliveira, R.A.; da Costa Silva, D. Effects of water deficit on morphophysiology, productivity and chemical composition of Ocimum africanum Lour (Lamiaceae). *Afr. J. Agric. Res.* 2016, 11, 1924–1934. [CrossRef]
- 69. Damalas, C.A. Improving drought tolerance in sweet basil (*Ocimum basilicum*) with salicylic acid. *Sci. Hortic.* **2018**, 246, 360–365. [CrossRef]
- Santos, C.V. Regulation of chlorophyll biosynthesis and degradation by salt stress in sunflower leaves. *Sci. Hortic.* 2004, 103, 93–99. [CrossRef]
- 71. Yang, C.W.; Wang, P.; Li, C.Y.; Shi, D.C.; Wang, D.L. Comparison of effects of salt and alkali stresses on the growth and photosynthesis of wheat. *Photosynthetica* **2008**, *46*, 107–114. [CrossRef]
- 72. Cornic, G. Drought stress inhibits photosynthesis by decreasing stomatal aperture–not by affecting ATP synthesis. *Trends Plant Sci.* 2000, *5*, 187–188. [CrossRef]
- Maroco, J.P.; Rodrigues, M.L.; Lopes, C.; Chaves, M.M. Limitations to leaf photosynthesis in field-grown grapevine under drought—metabolic and modelling approaches. *Funct. Plant Biol.* 2002, 29, 451–459. [CrossRef] [PubMed]
- Alves, A.A.; Setter, T.L. Abscisic acid accumulation and osmotic adjustment in cassava under water deficit. *Environ. Exp. Bot.* 2004, 51, 259–271. [CrossRef]
- Golan, K.; Kot, I.; Górska-Drabik, E.; Jurado, I.G.; Kmieć, K.; Łagowska, B. Physiological Response of Basil Plants to Twospotted Spider Mite (Acari: Tetranychidae) Infestation. J. Econ. Entomol. 2019, 112, 948–956. [CrossRef]
- Gomez, K.S.; Oosterhuis, D.M.; Rajguru, S.N. and Johnson, D.R. Molecular biology and physiology. Foliar antioxidant enzyme responses in cotton after aphid herbivory. *J. Cotton Sci.* 2004, *8*, 99–104.
- Attia, H.; Arnaud, N.; Karray, N.; Lachaâl, M. Long-term effects of mild salt stress on growth, ion accumulation and superoxide dismutase expression of Arabidopsis rosette leaves. *Physiol. Plant.* 2008, 132, 293–305. [CrossRef] [PubMed]
- 78. Ben Taarit, M.; Msaada, K.; Hosni, K.; Hammami, M.; Kchouk, M.E.; Marzouk, B. Plant growth, essential oil yield and composition of sage (*Salvia officinalis* L.) fruits cultivated under salt stress conditions. *Ind. Crop. Prod.* 2009, *30*, 333–337. [CrossRef]
- Estrada, B.; Aroca, R.; Maathuis, F.J.M.; Barea, J.M.; Ruiz-Lozano, J.M. Arbuscular mycorrhizal fungi native from a Mediterranean saline area enhance maize tolerance to salinity through improved ion homeostasis. *Plant Cell Environ.* 2013, 36, 1771–1782. [CrossRef]
- 80. Trovato, M.; Mattioli, R.; Costantino, P. Multiple roles of proline in plant stress tolerance and development. *Rend. Lincei* 2008, 19, 325–346. [CrossRef]
- Seki, M.; Umezawa, T.; Urano, K.; Shinozaki, K. Regulatory metabolic networks in drought stress responses. *Curr. Opin. Plant Biol.* 2007, 10, 296–302. [CrossRef]
- Zhang, X.; Ervin, E.; Evanylo, G.; Haering, K. Impact of Biosolids on Hormone Metabolism in Drought-Stressed Tall Fescue. *Crop. Sci.* 2009, 49, 1893–1901. [CrossRef]
- 83. Ekren, S.; Sönmez, Ç.; Özçakal, E.; Kurttaş, Y.S.K.; Bayram, E.; Gürgülü, H. The effect of different irrigation water levels on yield and quality characteristics of purple basil (*Ocimum basilicum* L.). *Agric. Water Manag.* **2012**, *109*, 155–161. [CrossRef]
- Schütz, M.; Fangmeier, A. Growth and yield responses of spring wheat (*Triticum aestivum* L. cv. Minaret) to elevated CO₂ and water limitation. *Environ. Pollut.* 2001, 114, 187–194. [CrossRef]
- 85. Verslues, P.E.; Agarwal, M.; Katiyar-Agarwal, S.; Zhu, J.; Zhu, J.-K. Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status. *Plant J.* **2006**, *45*, 523–539. [CrossRef] [PubMed]
- Jones, H.G. Monitoring plant and soil water status: Established and novel methods revisited and their relevance to studies of drought tolerance. J. Exp. Bot. 2006, 58, 119–130. [CrossRef]
- Sánchez-Rodríguez, E.; Rubio-Wilhelmi, M.; Cervilla, L.M.; Blasco, B.; Rios, J.J.; Rosales, M.A.; Romero, L.; Ruiz, J.M. Genotypic differences in some physiological parameters symptomatic for oxidative stress under moderate drought in tomato plants. *Plant Sci.* 2010, 178, 30–40. [CrossRef]
- 88. Aroca, R.; Porcel, R.; Lozano, J.M.R. Regulation of root water uptake under abiotic stress conditions. *J. Exp. Bot.* **2011**, *63*, 43–57. [CrossRef]
- Liu, H.; Wang, X.; Wang, D.; Zou, Z.; Liang, Z. Effect of drought stress on growth and accumulation of active constituents in Salvia miltiorrhiza Bunge. *Ind. Crop. Prod.* 2011, 33, 84–88. [CrossRef]
- Joseph, B.; Jini, D.; Sujatha, S. Insight into the Role of Exogenous Salicylic Acid on Plants Grown under Salt Environment. Asian J. Crop. Sci. 2010, 2, 226–235. [CrossRef]
- 91. Chen, Z.; Silva, H.; Klessig, D.F. Active oxygen species in the induction of plant systemic acquired resistance by salicylic acid. *Science* **1993**, *262*, 1883–1886. [CrossRef]
- 92. Maggio, A.; Zhu, J.-K.; Hasegawa, P.M.; Bressan, R.A. Osmogenetics: Aristotle to *Arabidopsis*. *Plant Cell* **2006**, *18*, 1542–1557. [CrossRef]
- Agut, B.; Pastor, V.; Jaques, J.A.; Flors, V. Can Plant Defence Mechanisms Provide New Approaches for the Sustainable Control of the Two-Spotted Spider Mite *Tetranychus urticae*? *Int. J. Mol. Sci.* 2018, 19, 614. [CrossRef]
- 94. Marchese, A.; Orhan, I.E.; Daglia, M.; Barbieri, R.; Di Lorenzo, A.; Nabavi, S.F.; Gortzi, O.; Izadi, M.; Nabavi, S.M. Antibacterial and antifungal activities of thymol: A brief review of the literature. *Food Chem.* **2016**, *210*, 402–414. [CrossRef]

- Salehi, B.; Mishra, A.P.; Shukla, I.; Sharifi-Rad, M.; del Mar Contreras, M.; Segura-Carretero, A.; Fathi, H.; Nasrabadi, N.N.; Kobarfard, F.; Sharifi-Rad, J. Thymol, thyme, and other plant sources: Health and potential uses. *Phytother. Res.* 2018, 32, 1688–1706. [CrossRef]
- 96. Mandal, S.; DebMandal, M. Thyme (*Thymus vulgaris* L.) Oils. In *Essential Oils in Food Preservation, Flavor and Safety*; Chapter 94; Elsevier Inc.: Amsterdam, The Netherlands, 2016; pp. 825–834. [CrossRef]
- 97. Trindade, H.; Pedro, L.G.; Figueiredo, A.C.; Barroso, J.G. Chemotypes and terpene synthase genes in Thymus genus: State of the art. *Ind. Crop. Prod.* 2018, 124, 530–547. [CrossRef]
- Askary, M.; Behdani, M.A.; Parsa, S.; Mahmoodi, S.; Jamialahmadi, M. Water stress and manure application affect the quantity and quality of essential oil of *Thymus daenensis* and *Thymus vulgaris*. Ind. Crop. Prod. 2018, 111, 336–344. [CrossRef]
- Pavela, R.; Žabka, M.; Vrchotová, N.; Tříska, J. Effect of foliar nutrition on the essential oil yield of Thyme (*Thymus vulgaris* L.). *Ind. Crop. Prod.* 2018, 112, 762–765. [CrossRef]
- 100. Galovičová, L.; Borotová, P.; Valková, V.; Vukovic, N.L.; Vukic, M.; Štefániková, J.; Ďúranová, H.; Kowalczewski, P.; Čmiková, N.; Kačániová, M. *Thymus vulgaris* Essential Oil and Its Biological Activity. *Plants* 2021, 10, 1959. [CrossRef]
- Punya, H.N.; Mehta, N.; Chatli, M.K.; Wagh, R.; Panwar, H. In-vitro Evaluation of Antimicrobial and Antioxidant Efficacy of Thyme (*Thymus vulgaris* L.) Essential Oil. J. Anim. Res. 2019, 9, 443–449. [CrossRef]
- 102. Kulisic, T.; Radonić, A.; Milos, M. Antioxidant Properties of Thyme (*Thymus vulgaris* L.) and Wild Thyme (*Thymus serpyllum* L.) Essential Oils. *Ital. J. Food Saf.* 2005, 17, 315–324.
- Bistgani, Z.E.; Hashemi, M.; DaCosta, M.; Craker, L.; Maggi, F.; Morshedloo, M.R. Effect of salinity stress on the physiological characteristics, phenolic compounds and antioxidant activity of *Thymus vulgaris* L. and *Thymus daenensis* Celak. *Ind. Crop. Prod.* 2019, 135, 311–320. [CrossRef]
- 104. Al Maqtari, M. Chemical Composition and Antimicrobial Activity of Essential Oil of *Thymus Vulgaris* from Yemen. *Turk. J. Biochem.* **2011**, *36*, 342–349.
- 105. Rota, M.C.; Herrera, A.; Martínez, R.M.; Sotomayor, J.A.; Jordán, M.J. Antimicrobial activity and chemical composition of *Thymus* vulgaris, *Thymus zygis* and *Thymus hyemalis* essential oils. *Food Control* **2008**, *19*, 681–687. [CrossRef]
- 106. Imelouane, B.; Amhamdi, H.; Wathelet, J.P.; Ankit, M.; Khedid, K.; El Bachiri, A. Chemical Composition and Antimicrobial Activity of Essential Oil of Thyme (*Thymus vulgaris*) from Eastern Morocco. *Int. J. Agric. Biol.* 2009, *11*, 205–208.
- 107. Gallucci, M.N.; Oliva, M.; Casero, C.; Dambolena, J.; Luna, A.; Zygadlo, J.; Demo, M. Antimicrobial Combined Action of Terpenes against the Food-Borne Microorganisms *Escherichia Coli, Staphylococcus Aureus* and *Bacillus Cereus*. *Flavour Fragr. J.* 2009, 24, 348–354. [CrossRef]
- Gill, A.O.; Delaquis, P.; Russo, P.; Holley, R.A. Evaluation of Antilisterial Action of Cilantro Oil on Vacuum Packed Ham. *Int. J. Food Microbiol.* 2002, 73, 83–92. [CrossRef]
- Mourey, A.; Canillac, N. Anti-Listeria Monocytogenes Activity of Essential Oils Components of Conifers. *Food Control* 2002, 13, 289–292. [CrossRef]
- 110. Al-Shuneigat, J.; Al-Sarayreh, S.; Al-Saraireh, Y.; Al-Qudah, M.; Al-Tarawneh, I.; Albataineh, E. Effects of Wild *Thymus Vulgaris* Essential Oil on Clinical Isolates Biofilm-Forming Bacteria. *IOSR J. Dent. Med. Sci.* **2014**, *13*, 62–66. [CrossRef]
- Jafri, H.; Ahmad, I. Thymus Vulgaris Essential Oil and Thymol Inhibit Biofilms and Interact Synergistically with Antifungal Drugs against Drug Resistant Strains of Candida Albicans and Candida Tropicalis. J. De Mycol. Médicale 2020, 30, 100911. [CrossRef]
- Reyes-Jurado, F.; Cervantes-Rincón, T.; Bach, H.; López-Malo, A.; Palou, E. Antimicrobial Activity of Mexican Oregano (Lippia Berlandieri), Thyme (Thymus Vulgaris), and Mustard (Brassica Nigra) Essential Oils in Gaseous Phase. *Ind. Crops Prod.* 2019, 131, 90–95. [CrossRef]
- 113. Aljabeili, H.S.; Barakat, H.; Abdel-Rahman, H.A. Chemical Composition, Antibacterial and Antioxidant Activities of Thyme Essential Oil (Thymus Vulgaris). *Food Nutr. Sci.* **2018**, *9*, 433–446. [CrossRef]
- Shin, J.; Na, K.; Shin, S.; Seo, S.-M.; Youn, H.J.; Park, I.-K.; Hyun, J. Biological Activity of Thyme White Essential Oil Stabilized by Cellulose Nanocrystals. *Biomolecules* 2019, 9, 799. [CrossRef]
- 115. Chaparro, J.M.; Badri, D.V.; Vivanco, J.M. Rhizosphere Microbiome Assemblage Is Affected by Plant Development. *ISME J.* **2013**, *8*, 790–803. [CrossRef]
- 116. Abdelshafy Mohamad, O.A.; Ma, J.-B.; Liu, Y.-H.; Zhang, D.; Hua, S.; Bhute, S.; Hedlund, B.P.; Li, W.-J.; Li, L. Beneficial Endophytic Bacterial Populations Associated with Medicinal Plant *Thymus Vulgaris* Alleviate Salt Stress and Confer Resistance to Fusarium Oxysporum. *Front. Plant Sci.* 2020, 11, 44. [CrossRef] [PubMed]
- 117. Saraswathi, S.G.; Paliwal, K. Drought induced changes in growth, leaf gas exchange and biomass production in Albizia lebbeck and Cassia siamea seedlings. *J. Environ. Biol.* **2011**, *32*, 173–178. [PubMed]
- Mazars, C.; Thuleau, P.; Lamotte, O.; Bourque, S. Cross-Talk between Ros and Calcium in Regulation of Nuclear Activities. *Mol. Plant* 2010, 3, 706–718. [CrossRef] [PubMed]
- Reddy, A.R.; Chaitanya, K.V.; Vivekanandan, M. Drought-Induced Responses of Photosynthesis and Antioxidant Metabolism in Higher Plants. J. Plant Physiol. 2004, 161, 1189–1202. [CrossRef]
- Johnson, S.M.; Doherty, S.J.; Croy, R.R.D. Biphasic Superoxide Generation in Potato Tubers. A Self-Amplifying Response to Stress. *Plant Physiol.* 2003, 131, 1440–1449. [CrossRef] [PubMed]
- Slesak, I.; Libik, M.; Karpinska, B.; Karpinski, S.; Miszalski, Z. The Role of Hydrogen Peroxide in Regulation of Plant Metabolism and Cellular Signalling in Response to Environmental Stresses. *Acta Biochim. Pol.* 2007, 54, 39–50. [CrossRef]

- 122. Catala, A. Lipid Peroxidation. Available online: https://www.intechopen.com/books/2553 (accessed on 3 April 2023).
- 123. Ashapkin, V.V.; Kutueva, L.I.; Aleksandrushkina, N.I.; Vanyushin, B.F. Epigenetic Mechanisms of Plant Adaptation to Biotic and Abiotic Stresses. *Int. J. Mol. Sci.* 2020, *21*, 7457. [CrossRef]
- 124. Smirnoff, N. Ascorbic Acid: Metabolism and Functions of a Multi-Facetted Molecule. *Curr. Opin. Plant Biol.* 2000, *3*, 229–235. [CrossRef]
- 125. Müller-Moulé, P.; Golan, T.; Niyogi, K.K. Ascorbate-Deficient Mutants of Arabidopsis Grow in High Light despite Chronic Photooxidative Stress. *Plant Physiol.* **2004**, *134*, 1163–1172. [CrossRef]
- 126. Smirnoff, N.; Wheeler, G.L. Ascorbic Acid in Plants: Biosynthesis and Function. Crit. Rev. Plant Sci. 2000, 19, 267–290. [CrossRef]
- 127. Khan, T.; Mazid, M.; Mohammad, F. A Review of Ascorbic Acid Potentialities against Oxidative Stress Induced in Plants. J. Agrobiol. 2011, 28, 97–111. [CrossRef]
- Siripornadulsil, S.; Traina, S.; Verma, D.P.; Sayre, R.T. Molecular Mechanisms of Proline-Mediated Tolerance to Toxic Heavy Metals in Transgenic Microalgae. *Plant Cell* 2002, 14, 2837–2847. [CrossRef]
- 129. Agar, H.; Galatali, S.; Ozkaya, D.E.; Kaya, E. A Primary Study: Investigation of the in Vitro Salt Stress Effects on Development in Thymus Cilicicus Boiss. & Bal. *Glob. J. Bot. Sci.* **2022**, *10*, 23–27. [CrossRef]
- 130. Aziz, E.E.; Hendawy, S.F. Effect of soil type and irrigation intervals on plant growth, essential oil yield and constituents of *Thymus Vulgaris* plant. *Thymus Plant* **2008**, *4*, 443–450.
- 131. Kaur, H.; Kohli, S.K.; Khanna, K.; Bhardwaj, R. Scrutinizing the Impact of Water Deficit in Plants: Transcriptional Regulation, Signaling, Photosynthetic Efficacy, and Management. *Physiol. Plant.* **2021**, *172*, 935–962. [CrossRef]
- Haworth, M.; Killi, D.; Materassi, A.; Raschi, A.; Centritto, M. Impaired Stomatal Control Is Associated with Reduced Photosynthetic Physiology in Crop Species Grown at Elevated [CO₂]. *Front. Plant Sci.* 2016, 7, 1568. [CrossRef] [PubMed]
- 133. Frary, A. *Plant Physiology and Developmentplant*; Lincoln, T., Eduardo, Z., Ian Max, M., Angus, M., Eds.; Sinauer Associates Inc.: Sunderland, MA, USA, 2015; Volume 117, pp. 397–399. ISBN 978-1-60535-255-8. [CrossRef]
- Bielach, A.; Hrtyan, M.; Tognetti, V. Plants under Stress: Involvement of Auxin and Cytokinin. Int. J. Mol. Sci. 2017, 18, 1427. [CrossRef]
- Smirnoff, N. The Role of Active Oxygen in the Response of Plants to Water Deficit and Desiccation. New Phytol. 1993, 125, 27–58.
 [CrossRef]
- 136. Naservafaei, S.; Sohrabi, Y.; Moradi, P.; Mac Sweeney, E.; Mastinu, A. Biological Response of Lallemantia Iberica to Brassinolide Treatment under Different Watering Conditions. *Plants* **2021**, *10*, 496. [CrossRef] [PubMed]
- 137. Karimmojeni, H.; Rezaei, M.; Tseng, T.-M.; Mastinu, A. Effects of Metribuzin Herbicide on Some Morpho-Physiological Characteristics of Two Echinacea Species. *Horticulturae* 2022, *8*, 169. [CrossRef]
- 138. Rasheed, S.; Bashir, K.; Kim, J.-M.; Ando, M.; Tanaka, M.; Seki, M. The Modulation of Acetic Acid Pathway Genes in Arabidopsis Improves Survival under Drought Stress. *Sci. Rep.* **2018**, *8*, 7831. [CrossRef]
- Luo, M.; Cheng, K.; Xu, Y.; Yang, S.; Wu, K. Plant Responses to Abiotic Stress Regulated by Histone Deacetylases. *Front. Plant Sci.* 2017, *8*, 2147. [CrossRef] [PubMed]
- Ashrafi, M.; Azimi-Moqadam, M.-R.; MohseniFard, E.; Shekari, F.; Jafary, H.; Moradi, P.; Pucci, M.; Abate, G.; Mastinu, A. Physiological and Molecular Aspects of Two Thymus Species Differently Sensitive to Drought Stress. *BioTech* 2022, 11, 8. [CrossRef] [PubMed]
- 141. Moradi, P.; Ford-Lloyd, B.; Pritchard, J. Metabolic Responses of *Thymus Vulgaris* to Water Deficit Stress. *Curr. Metab.* 2018, 6, 64–74. [CrossRef]
- 142. Moori, S.; Ahmadi-Lahijani, M.J. Hormopriming Instigates Defense Mechanisms in Thyme (*Thymus Vulgaris* L.) Seeds under Cadmium Stress. *J. Appl. Res. Med. Aromat. Plants* 2020, 19, 100268. [CrossRef]
- 143. Linhart, Y.B.; Keefover-Ring, K.; Mooney, K.A.; Breland, B.; Thompson, J.D. A Chemical Polymorphism in a Multitrophic Setting: Thyme Monoterpene Composition and Food Web Structure. *Am. Nat.* **2005**, *166*, 517–529. [CrossRef]
- 144. Loxdale, H.D.; Balog, A. Aphid Specialism as an Example of Ecological-Evolutionary Divergence. *Biol. Rev.* **2017**, *93*, 642–657. [CrossRef]
- 145. Adorjan, B.; Buchbauer, G. Biological Properties of Essential Oils: An Updated Review. *Flavour Fragr. J.* 2010, 25, 407–426. [CrossRef]
- 146. Kamkar, A.; Tooryan, F.; AkhondzadehBasti, A.; Misaghi, A.; Shariatifar, N. Chemical composition of summer savory (*Satureja hortensis* L.) essential oil and comparison of antioxidant activity with aqueous and alcoholic extracts. *J. Vet. Res.* **2013**, *68*, 183–190.
- 147. Adiguzel, A.; Ozer, H.; Kilic, H.; Cetin, B. Screening of Antimicrobial Activity of Essential Oil and Methanol Extract of *Satureja hortensis* on Foodborne Bacteria and Fungi. *Czech J. Food Sci.* **2007**, *25*, 81–89. [CrossRef]
- 148. Popovici, R.A.; Vaduva, D.; Pinzaru, I.; Dehelean, C.A.; Farcas, C.G.; Coricovac, D.; Danciu, C.; Popescu, I.; Alexa, E.; Lazureanu, V.; et al. A comparative study on the biological activity of essential oil and total hydro-alcoholic extract of *Satureja hortensis* L. *Exp. Ther. Med.* 2019, *18*, 932–942. [CrossRef]
- 149. Ramos, M.; Beltrán, A.; Peltzer, M.; Valente, A.J.M.; Garrigós, M.D.C. Release and antioxidant activity of carvacrol and thymol from polypropylene active packaging films. *LWT Food Sci. Technol.* **2014**, *58*, 470–477. [CrossRef]
- 150. Chen, Q.; Gan, Z.; Zhao, J.; Wang, Y.; Zhang, S.; Li, J.; Ni, Y. In Vitro Comparison of Antioxidant Capacity of Cumin (*Cuminum cyminum* L.) Oils and Their Main Components. *LWT Food Sci. Technol.* **2014**, 55, 632–637. [CrossRef]

- Chua, L.S.; Lau, C.H.; Chew, C.Y.; Ismail, N.I.; Soontorngun, N. Phytochemical Profile of Orthosiphon Aristatus Extracts after Storage: Rosmarinic Acid and Other Caffeic Acid Derivatives. *Phytomedicine* 2018, 39, 49–55. [CrossRef] [PubMed]
- 152. Lesjak, M.; Beara, I.; Simin, N.; Pintać, D.; Majkić, T.; Bekvalac, K.; Orčić, D.; Mimica-Dukić, N. Antioxidant and Anti-Inflammatory Activities of Quercetin and Its Derivatives. J. Funct. Foods 2018, 40, 68–75. [CrossRef]
- Aksu, M.I.; Özer, H. Effects of Lyophilized Water Extract of Satureja Hortensison the Shelf Life and Quality Properties of Ground Beef. J. Food Process. Preserv. 2012, 37, 777–783. [CrossRef]
- 154. Vahidyan, H.; Sahari, M.A.; Barzegar, M.; Naghdi Badi, H. Application of Zataria multiflora Boiss. and *Satureja hortensis* L. essential oils as two natural antioxidants in Mayonnaise formulated with linseed oil. *J. Med. Plants* **2012**, *11*, 69–79.
- 155. Jafari, F.; Ghavidel, F.; Zarshenas, M.M. A Critical Overview on the Pharmacological and Clinical Aspects of Popular Satureja Species. J. Acupunct. Meridian Stud. 2016, 9, 118–127. [CrossRef]
- Hazrati, H.; Saharkhiz, M.J.; Niakousari, M.; Moein, M. Natural Herbicide Activity of Satureja hortensis L. Essential Oil Nanoemulsion on the Seed Germination and Morphophysiological Features of Two Important Weed Species. Ecotoxicol. Environ. Saf. 2017, 142, 423–430. [CrossRef] [PubMed]
- Banu, M.N.; Hoque, M.A.; Watanabe-Sugimoto, M.; Matsuoka, K.; Nakamura, Y.; Shimoishi, Y.; Murata, Y. Proline and Glycinebetaine Induce Antioxidant Defense Gene Expression and Suppress Cell Death in Cultured Tobacco Cells under Salt Stress. J. Plant Physiol. 2009, 166, 146–156. [CrossRef]
- Mehdizadeh, L.; Moghaddam, M.; Lakzian, A. Alleviating Negative Effects of Salinity Stress in Summer Savory (*Satureja hortensis* L.) by Biochar Application. *Acta Physiol. Plant.* 2019, 41, 98. [CrossRef]
- 159. Nikee, E.; Pazoki, A.; Zahedi, H. Influences of Ascorbic Acid and Gibberellin on Alleviation of Salt Stress in Summer Savory (*Satureja hortensis* L.). *Int. J. Biosci.* (*IJB*) **2014**, *5*, 245–255. [CrossRef]
- 160. Miranshahi, B.; Sayyari, M. Methyl jasmonate mitigates drought stress injuries and affects essential oil of summer savory. *J. Agric. Sci. Tech.* **2016**, *18*, 1635–1645.
- 161. Zimowska, B. Diversity of fungi occurring on savory (Satureja hortensis L.). Herba Pol. 2010, 2, 56.
- 162. Wubben, J.P.; Mulder, W.; ten Have, A.; van Kan, J.A.; Visser, J. Cloning and Partial Characterization of Endopolygalacturonase Genes from Botrytis Cinerea. *Appl. Environ. Microbiol.* **1999**, *65*, 1596–1602. [CrossRef]
- 163. Shah, P.; Gutierrez-Sanchez, G.; Orlando, R.; Bergmann, C. A Proteomic Study of Pectin-Degrading Enzymes Secreted by Botrytis Cinerea Grown in Liquid Culture. *PROTEOMICS* **2009**, *9*, 3126–3135. [CrossRef]
- 164. Zwenger, S.; Basu, C. Plant terpenoids: Applications and future potentials. Biotechnol. Mol. Biol. Rev. 2008, 3, 1.
- Heil, M.; Silva Bueno, J.C. Within-Plant Signaling by Volatiles Leads to Induction and Priming of an Indirect Plant Defense in Nature. Proc. Natl. Acad. Sci. USA 2007, 104, 5467–5472. [CrossRef]
- 166. Ton, J.; D'Alessandro, M.; Jourdie, V.; Jakab, G.; Karlen, D.; Held, M.; Mauch-Mani, B.; Turlings, T.C.J. Priming by Airborne Signals Boosts Direct and Indirect Resistance in Maize. *Plant J.* **2006**, *49*, 16–26. [CrossRef]
- 167. Cheong, J.-J.; Do Choi, Y. Methyl jasmonate as a vital substance in plants. Trends Genet. 2003, 19, 409–413. [CrossRef]
- 168. Rodríguez, A.A.; Stella, A.M.; Storni, M.M.; Zulpa, G.; Zaccaro, M.C. Effects of Cyanobacterial Extracellular Products and Gibberellic Acid on Salinity Tolerance in Oryza Satival. *Saline Syst.* **2006**, *2*, 7. [CrossRef] [PubMed]
- Maggio, A.; Barbieri, G.; Raimondi, G.; De Pascale, S. Contrasting Effects of GA3 Treatments on Tomato Plants Exposed to Increasing Salinity. J. Plant Growth Regul. 2009, 29, 63–72. [CrossRef]
- 170. Ashraf, M.; Karim, F.; Rasul, E. Interactive effects of gibberellic acid (GA3) and salt stress on growth, ion accumulation and photosynthetic capacity of two spring wheat (*Triticum aestivum* L.) cultivars differing in salt tolerance. *Plant Growth Regul.* 2002, 36, 49–59. [CrossRef]
- 171. Wen, F.-p.; Zhang, Z.-h.; Bai, T.; Xu, Q.; Pan, Y.-h. Proteomics Reveals the Effects of Gibberellic Acid (GA3) on Salt-Stressed Rice (*Oryza sativa* L.) Shoots. *Plant Sci.* **2010**, *178*, 170–175. [CrossRef]
- 172. Lu, Y.Y.; Deng, X.P.; Kwak, S.S. Over expression of CuZn superoxide dismutase (CuZn SOD) and ascorbate peroxidase (APX) in transgenic sweet potato enhances tolerance and recovery from drought stress. *Afr. J. Biotechnol.* **2015**, *9*, 8378–8391.
- 173. Desikan, R. Aba, Hydrogen Peroxide and Nitric Oxide Signalling in Stomatal Guard Cells. J. Exp. Bot. 2003, 55, 205–212. [CrossRef]
- 174. Krishnan, N.; Dickman, M.B.; Becker, D.F. Proline Modulates the Intracellular Redox Environment and Protects Mammalian Cells against Oxidative Stress. *Free. Radic. Biol. Med.* **2008**, *44*, 671–681. [CrossRef]
- 175. Lamalakshmi Devi, E.; Kumar, S.; Basanta Singh, T.; Sharma, S.K.; Beemrote, A.; Devi, C.P.; Chongtham, S.K.; Singh, C.H.; Yumlembam, R.A.; Haribhushan, A.; et al. Adaptation Strategies and Defence Mechanisms of Plants during Environmental Stress. In *Medicinal Plants and Environmental Challenges*; Ghorbanpour, M., Varma, A., Eds.; Springer: Cham, Switzerland, 2017; pp. 359–413. [CrossRef]
- 176. HongBo, S.; ZongSuo, L.; MingAn, S.; ShiMeng, S.; ZanMin, H. Investigation on Dynamic Changes of Photosynthetic Characteristics of 10 Wheat (Triticum Aestivum L.) Genotypes during Two Vegetative-Growth Stages at Water Deficits. *Colloids Surf. B Biointerfaces* 2005, 43, 221–227. [CrossRef]
- 177. Blumwald, E.; Anil, G.; Allen, G. New diections for a diverse planet. In Proceedings of the 4th international crop science congress, Brisbane, Australia, 26 September–1 October 2004. Available online: http://www.cropscience.org.au (accessed on 7 February 2023).

- 178. Croteau, R.B.; Davis, E.M.; Ringer, K.L.; Wildung, M.R. (-)-Menthol biosynthesis and molecular genetics. *Naturwissenschaften* **2005**, 92, 562–577. [CrossRef]
- 179. Gang, D.R.; Wang, J.; Dudareva, N.; Nam, K.H.; Simon, J.E.; Lewinsohn, E.; Pichersky, E. An investigation of the storage and biosynthesis of phenylpropenes in sweet basil. *Plant Physiol.* **2001**, *125*, 539–555. [CrossRef]
- 180. Xu, C.; Wang, M.; Zhou, L.; Quan, T.; Xia, G. Heterologous Expression of the Wheat Aquaporin Gene TATIP2;2 Compromises the Abiotic Stress Tolerance of Arabidopsis Thaliana. *PLoS ONE* **2013**, *8*, e79618. [CrossRef] [PubMed]
- 181. Moradi, P.; Ford-Lloyd, B.; Pritchard, J. Metabolomic Approach Reveals the Biochemical Mechanisms Underlying Drought Stress Tolerance in Thyme. *Anal. Biochem.* 2017, 527, 49–62. [CrossRef]
- Kulak, M.; Jorrín-Novo, J.V.; Romero-Rodriguez, M.C.; Yildirim, E.D.; Gul, F.; Karaman, S. Seed Priming with Salicylic Acid on Plant Growth and Essential Oil Composition in Basil (*Ocimum basilicum* L.) Plants Grown under Water Stress Conditions. *Ind. Crops Prod.* 2021, 161, 113235. [CrossRef]
- Alizadeh, A.; Moghaddam, M.; Asgharzade, A.; Sourestani, M.M. Phytochemical and Physiological Response of *Satureja hortensis* L. to Different Irrigation Regimes and Chitosan Application. *Ind. Crops Prod.* 2020, 158, 112990. [CrossRef]
- 184. Bakhshian, M.; Naderi, M.R.; Javanmard, H.R.; Bahreininejad, B. The Growth of Summer Savory (Satureja Hortensis) Affected by Fertilization and Plant Growth Regulators in Temperature Stress. *Biocatal. Agric. Biotechnol.* 2022, 43, 102371. [CrossRef]
- 185. Dikova, B. Tomato spotted wilt virus on some medicinal and essential oil-bearing plants in Bulgaria. *Bulg. J. Agric. Sci.* 2011, 17, 306–313.
- Bansal, K.C.; Singh, A.K.; Wani, S.H. Plastid Transformation for Abiotic Stress Tolerance in Plants. *Methods Mol. Biol.* 2012, 913, 351–358. [CrossRef]
- 187. Khan, H.; Wani, S.H. Molecular approaches to enhance abiotic stresses tolerance. In *Innovations in Plant Science and Biotechnology;* Wani, S.H., Malik, C.P., Hora, A., Kaur, R., Eds.; Agrobios (India): Jodhpur, India, 2014; pp. 111–152. ISBN 978-81-7754-553-1.
- 188. Gonda, I.; Faigenboim, A.; Adler, C.; Milavski, R.; Karp, M.-J.; Shachter, A.; Ronen, G.; Baruch, K.; Chaimovitsh, D.; Dudai, N. The Genome Sequence of Tetraploid Sweet Basil, Ocimum Basilicum L., Provides Tools for Advanced Genome Editing and Molecular Breeding. DNA Res. 2020, 27, dsaa027. [CrossRef] [PubMed]
- Pyne, R.M.; Koroch, A.R.; Wyenandt, C.A.; Simon, J.E. A Rapid Screening Approach to Identify Resistance to Basil Downy Mildew (*Peronospora Belbahrii*). *HortScience* 2014, 49, 1041–1045. [CrossRef]
- 190. Bi, H.H.; Song, Y.Y.; Zeng, R.S. Biochemical and molecular responses of host plants to mycorrhizal infection and their roles in plant defence. *Allelopath. J.* 2007, 20, 15–28.
- Raffaele, S.; Vailleau, F.; Leger, A.; Joubès, J.; Miersch, O.; Huard, C.; Blée, E.; Mongrand, S.; Domergue, F.; Roby, D. A MYB Transcription Factor Regulates Very-Long-Chain Fatty Acid Biosynthesis for Activation of the Hypersensitive Cell Death Response in Arabidopsis. *Plant Cell* 2008, 20, 752–767. [CrossRef]
- 192. Pérez-Clemente, R.M.; Vives, V.; Zandalinas, S.I.; López-Climent, M.F.; Muñoz, V.; Gómez-Cadenas, A. Biotechnological Approaches to Study Plant Responses to Stress. *BioMed Res. Int.* 2013, 2013, 654120. [CrossRef] [PubMed]
- 193. Seo, P.J.; Park, C.M. MYB96-mediated abscisic acid signals induce pathogen resistance response by promoting salicylic acid biosynthesis in Arabidopsis. *New Phytol.* **2010**, *186*, 471–483. [CrossRef] [PubMed]
- 194. Agarwal, M.; Hao, Y.; Kapoor, A.; Dong, C.-H.; Fujii, H.; Zheng, X.; Zhu, J.-K. A R2R3 Type MYB Transcription Factor Is Involved in the Cold Regulation of CBF Genes and in Acquired Freezing Tolerance. J. Biol. Chem. 2006, 281, 37636–37645. [CrossRef] [PubMed]
- 195. Jiang, G.-L. Molecular Markers and Marker-Assisted Breeding in Plants. Plant Breed. Lab. Fields 2013, 3, 45–83. [CrossRef]
- 196. Gonda, I.; Milavski, R.; Adler, C.; Abu-Abied, M.; Tal, O.; Faigenboim, A.; Chaimovitsh, D.; Dudai, N. Genome-Based High-Resolution Mapping of Fusarium Wilt Resistance in Sweet Basil. *Plant Sci.* **2022**, *321*, 111316. [CrossRef]
- Ben-Naim, Y.; Falach, L.; Cohen, Y. Transfer of Downy Mildew Resistance from Wild Basil (Ocimum Americanum) to Sweet Basil (O. Basilicum). Phytopathology 2018, 108, 114–123. [CrossRef]

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