Supplementary materials

Eco-friendly Synthesis of Functionalized Carbon Nanodots from Cashew Nut Skin Waste for Bioimaging

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Instrumentation Methods

Cashew nut skin waste-derived functionalized carbon nanodots (F-CNDs) were characterized by various physicochemical analytical techniques such as field emission scanning electron microscopy (FESEM) with energy-dispersive X-ray (EDX) spectroscopy, high-resolution transmittance electron microscopy (HRTEM), X-ray diffraction (XRD), Raman spectroscopy, attenuated total reflectance Fourier-transform infrared (ATR-FTIR) spectroscopy, X-ray photoelectron spectroscopy (XPS), Ultraviolet-visible (UV-vis) absorption spectroscopy, and fluorescence spectroscopy. FESEM with EDX analysis was carried out on a Hitachi S-4800 equipped with EDX at an accelerating voltage of 10/15 kV. TEM/HRTEM images were performed with a JEOL JEM transmission electron microscope with an operating accelerating voltage of 120 kV. XRD measurements were carried out using a PANalytical X'Pert3 MRD diffractometer with monochromatized Cu Kα radiation (λ = 1.54 Å) at 40 kV and 30 mA and were recorded in the range from 10 to 80° (2θ). Ra-man spectrum was recorded on XploRA Micro-Raman spectrophotometer (Horiba) with ranges between 50 and 4000 cm^{-1} at the core research support center for natural products and medical materials of Yeungnam University. ATR-FTIR spectra were recorded in transmittance mode on a Perkin Elmer Spectrum Two in the wavenumber range from 400 to 4000 cm^{-1} by the addition of 8 scans at a resolution of 8 cm^{-1}. XPS spectra were achieved using a K-Alpha (Thermo Scientific). CasaXPS software was used for the deconvolution of the high-resolution XPS spectra. UV-vis absorption spectra were recorded from 200 to 700 nm using an OPTIZEN 3220UV spectrophotometer. Excitation and emission fluorescence spectra were recorded using a Hitachi F-7000 fluorescence spectrophotometer using a 1 cm^3 path-length quartz cell. The excitation wavelength was varied to determine the maximum emission intensity. The slit width was fixed at 5 nm, and the scan speed was set to 400 nm/min. Cellular imaging was performed using a confocal laser scanning microscope (X400; LSM700, Carl Zeiss, Oberkochen, Germany).

Quantum Yield Measurement of Functionalized Carbon Nanodots

The quantum yield (QY) of the synthesized F-CNDs was calculated by using quinine sulfate in 0.1 M H_2SO_4 (QYR is 0.54) as a standard reference and was calculated using the following equation (1):

$$QY = QY_R \frac{I_S A_R (n_S)^2}{I_R A_S (n_R)^2} \quad (1)$$

where, "I" is the measured integrated fluorescent emission intensity, "n" is the refractive index of the solvent, and "A" is the absorbance (intensity). The subscript "R" and "S" refer to the known fluorescent reference and standard for the synthesized sample, respectively.

Photobleaching Measurements of Functionalized Carbon Nanodots

The photostability (photobleaching) of the synthesized F-CNDs was examined by continuous irradiation under UV light (365 nm) for 120 min with an interval of every 60 min. The fluorescence intensity of the F-CNDs aqueous solution was measured before and after UV-light irradiation.

Cell Culture, Cell Viability Assay, and Microscopy Analysis of Functionalized Carbon Nanodots

HCT-116 was cultured in Roswell Park Memorial Institute (RPMI) 1640 medium containing 25 mM HEPES buffer, 10% heat-inactivated fetal bovine serum, and 1% penicillin/streptomycin under 5% CO_2 atmosphere at 37 °C until coming to a composition, subsequently treated with different concentrations of F-CNDs (0–200 μg mL^{-1}) for 12 and 24 h. MTT (3-(4, 5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) cell viability test was performed to determine the cytotoxicity of the synthesized F-CNDs. The wells in the microtiter plate were incubated for 4 h at 37 °C in the dark in a humidified chamber with the addition of MTT solution (5 mg MTT mL^{-1} in phosphate-buffered saline (PBS)). Finally, insoluble purple-formazan crystals were observed in F-CNDs treated cells, which were subsequently dissolved in dimethyl sulfoxide (DMSO), and optical density was measured at 540 nm.

Cells were cultured on coverslips in 6-well plates for fluorescent confocal imaging. At confluence, staining was achieved by incubating the cells in the presence and absence of the F-CNDs in a humidified chamber at 37 °C for 12 and 24 h. The staining of cells was fixed with 4% p-formaldehyde, washed with PBS 3 times, and then mounted in a fluorescence mounting medium (Dako North America Inc., Carpinteria, CA, USA). Then the fluorescence of the stained cells was imaged with appropriate filter sets (blue (405 nm), green (488 nm), and red (555 nm)) as well as with bright field (BF) on confocal microscopy (X400; LSM700, Carl Zeiss, Oberkochen, Germany).

Optical Properties of Synthesized F-CNDs

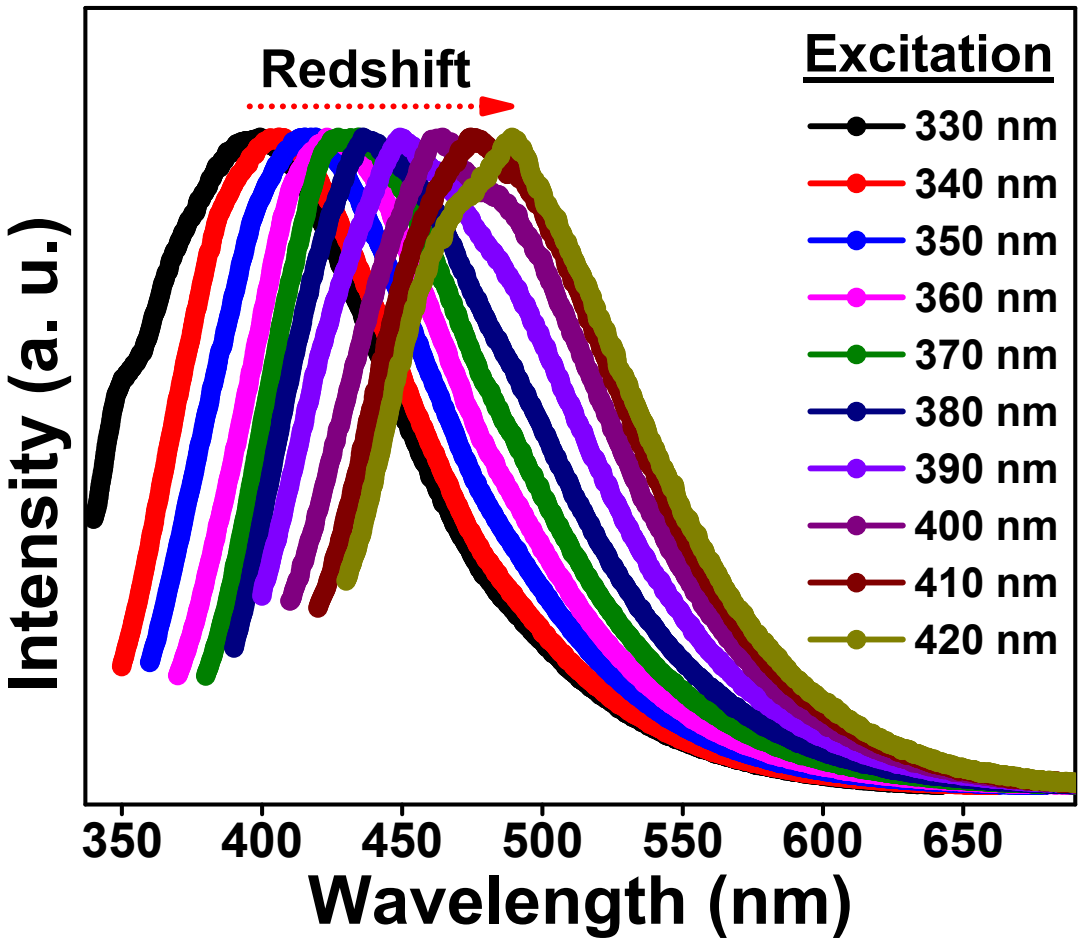

Figure S1. Fluorescence excitation-dependent emission normalized-spectra of synthesized F-CNDs.

Cell Viability Assay of Synthesized F-CNDs on Human Colon Cancer Cells

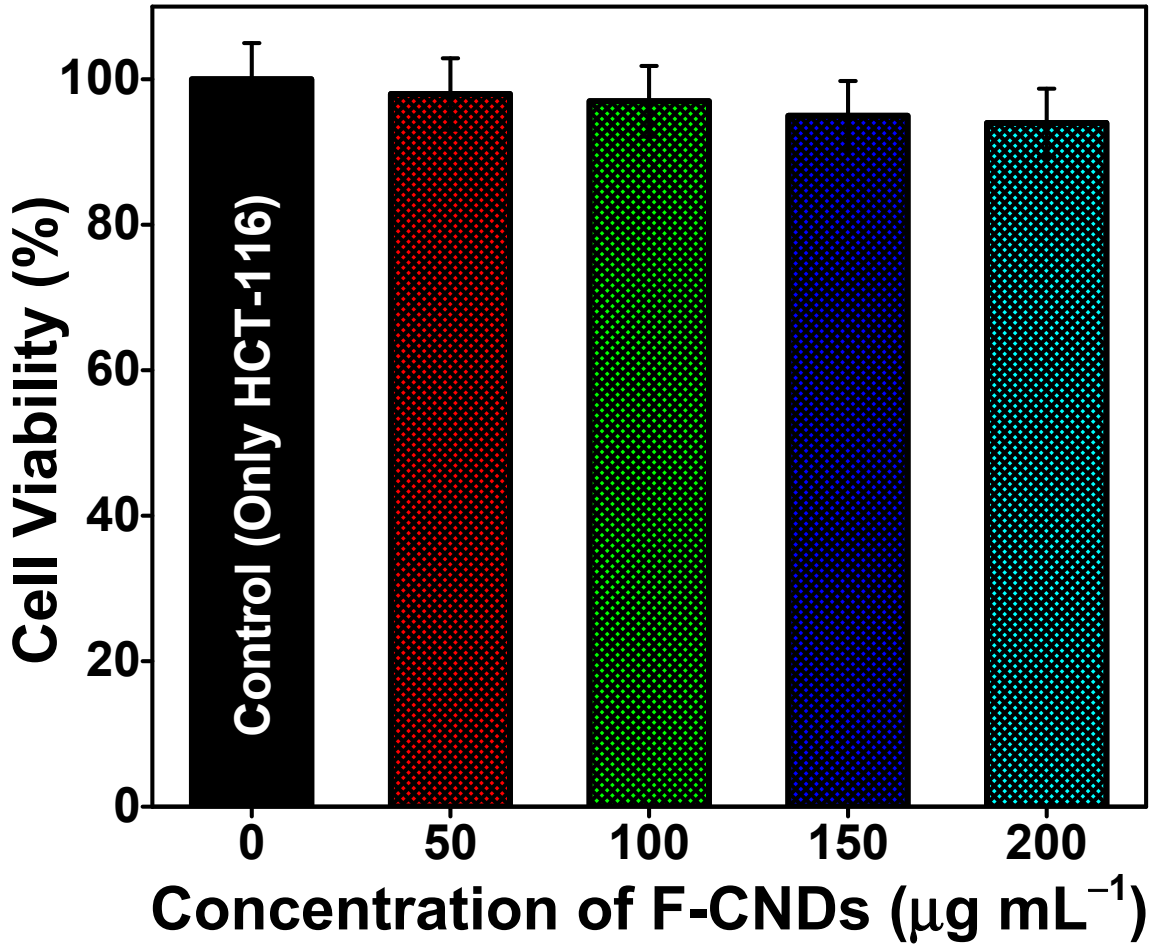

Figure S2. Cell viability values (MTT assay). The bar chart shows the comparison of viability between F-CNDs treated cells and untreated cells (control).