



Article Synthesis of Ce/MgO Catalysts for Direct Oxidation of *Hibiscus cannabinus* Stalks to Vanillin

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Abstract: One possible method of producing vanillin from biomass is through controlled oxidation of lignin. Direct oxidation of kenaf stalks was chosen without having to separate the cellulose and hemicellulose components from the lignocellulosic biomass. This makes the process greener, as well as saving time. In this paper, Ce/MgO catalysts were developed for oxidation of kenaf stalks and kenaf lignin under microwave irradiation. The catalysts were characterized for their physicochemical properties using XRD and N₂ adsorption-desorption isotherms. The synthesized MgO showed the presence of diffraction peaks assigned to cubic MgO while the 30Ce/MgO catalysts showed the presence of cubic fluorite of CeO₂. N₂ adsorption-desorption isotherms showed that all catalysts possess Type III isotherm according to IUPAC classification, indicating a nonporous structure. All catalysts were tested for direct oxidation of kenaf stalks under 300 W of microwave irradiation using H_2O_2 as the oxidizing agent at pH 11.5 and temperatures between 160 and 180 °C for 10–30 min with 5–15% catalyst loading. The highest vanillin yields of 3.70% and 2.90% for extracted lignin and direct biomass oxidation were achieved using 30Ce/MgO-48. In comparison, 7.80% and 4.45% were obtained using 2N of NaOH homogeneous catalyst for extracted lignin and direct biomass, respectively, at 170 °C for 20 min. The reusability test shows that 30Ce/MgO can be used up to three cycles without significant loss in catalytic activity. Other compounds detected were 4-vinylguaiacol, syringol and syringaldehyde.

Keywords: direct oxidation; vanillin; microwave heating; Ce/MgO catalyst

1. Introduction

Vanillin, known as 4-hydroxy-3-methoxybenzaldehyde (Figure 1), is vanilla's primary flavor constituent used as a flavoring and fragrance ingredient in the food and cosmetic industries [1]. The commercial vanillin market is served by three major sources, namely, natural vanilla from vanilla pods, vanillin produced from petrochemical sources and vanillin produced from kraft lignin [2]. The *Vanilla planifolia* A plant is the best-known natural source of vanillin, which belongs to the orchid family, mainly cultivated in Mexico, Madagascar, Reunion, Java, and Tahiti [3]. However, the production of natural vanillin from vanilla pods covers only 0.2% of the market requirement and its production cost is very high as it requires manual labor to pollinate the flower [1,4]. Furthermore, the extraction of vanillin from vanilla beans is a long and expensive process, and is inconsistent, and the demand for this universally popular flavoring cannot be satisfied by vanilla beans alone. Nowadays, vanillin produced from petroleum-based intermediates, especially guaiacol, accounts for 85% of the world supply, while the remaining 15% is produced from lignin [5].



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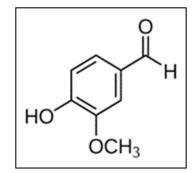


Figure 1. Chemical structure of vanillin [6].

The synthesis of vanillin from renewable sources should result in a greener and more sustainable process. Lignin could be used as a natural precursor from renewable sources to meet the growing demand for natural vanillin as far as the aroma and fragrance fields are concerned. Furthermore, vanillin produced from lignin shows 1.2 times higher aromatic intensity than petroleum-based vanillin [1]. One possible method of producing vanillin through a sustainable process is through the controlled oxidation of lignin from lignocellulosic biomass. Lignin is a three-dimensional amorphous macromolecule and is the most significant renewable material with an aromatic skeleton structure. Lignin's composition and ratio are different for different plants such as hardwood and softwood [7]. Softwoods have a lignin content of 25–35% dry weight, while the lignin content in hardwoods is 18–25% [8].

Sales and co-workers [9] reported that the oxidation of lignin to produce vanillin occurs at very alkaline pH (almost 14) and high temperatures (130 °C) with oxygen pressure higher than 3 bar. They investigated the conversion of alkaline lignin extracted from sugarcane bagasse using batch slurry and continuous fluidized-bed reactors and reported that 0.56 g of vanillin and 0.5 g of syringaldehyde were obtained from 30 g of lignin at 120 °C after 2 h [9]. Furthermore, Araujo et al. [10] reported that low-molecular weight lignin derived from softwoods is preferable to than high molecular weight lignin for obtaining a better vanillin yield. They also reported that regardless of the source of lignin, pH is the essential variable in the reaction, with lower pH values promoting vanillin degradation, thus reducing the vanillin yield.

The oxidizing agent used for the oxidation reaction also significantly influences the yield of vanillin produced, as observed by Alireza et al. [6], where 4.2% and 1.2% of vanillin yield were reported from oxidation of industrial lignin using molecular oxygen and alkaline nitrobenzene, respectively. Furthermore, Valery et al. [11] reported a vanillin yield of 18.6% when lignin extracted from powdered pine wood using standard alcohol-toluene as a solvent was oxidized using copper oxide (CuO) catalyst. On the other hand, Chen et al. [12] reported that a 4.2% vanillin yield was achieved from the direct oxidation of Japanese cedar wood particles in CuO as the catalyst and H_2O_2 solution as an oxidizing agent under microwave irradiation at 200 °C for 20 min.

Chaudhary and Dhepe [13] reported that solid acids, homogeneous acids, ionic liquid, supercritical fluids yield low molecular weight aromatic products. However, most of these methods were linked to a few problems such as high temperatures ($\geq 250 \,^{\circ}$ C) or pressures (>3 MPa) or both for operation parameters. Alkaline oxidation of lignin is considered the only approach that yields large amounts of single monomeric aldehydes that are the predominant products in the presence of base catalysts. However, the conversion of lignin to vanillin requires a very high pH and this is usually achieved by using NaOH as the solvent, resulting in problems of corrosion and pollution. A study carried out by Long et al. [14] reported a 13.2% yield phenolic monomers, including vanillin obtained during lignin depolymerization using MgO solid base catalyst, while Biswas et al. [15] stated that the properties of the high base of CaO/CeO₂ catalysts enhanced the cleavage of β -O-4 in the lignin structure, thus increasing the reaction's selectivity. Base catalysts are very interesting

for alkaline oxidation of lignin; therefore, alkaline solid catalysts based on MgO were developed. The basicity of MgO was modified using Ce to increase its basicity properties.

Kenaf (*Hibiscus cannabinus*) is a commodity crop with a high potential to produce renewable vanillin since it contains 13–15% lignin [16]. In this paper, we report the vanillin production from direct oxidation of kenaf stalks and kenaf lignin using 30Ce/MgO under microwave irradiation.

2. Result

2.1. Catalyst Characterization

Figure 2 shows the X-ray diffractograms of MgO synthesized using sol-gel and hydrothermal methods (more details are available in Figure S1). The presence of diffraction peaks at $2\theta = 29.33$, 36.90, 42.85, 62.13, 74.65 and 78.40° indicate the formation of cubic phase of MgO [10] which could be assigned to (100), (111), (200), (220), (311) and (222) planes, respectively [17,18]. Based on ICDD: 00-04500946, the diffraction peak of MgO exhibits a crystalline periclase of magnesium oxide with a cubic crystal system, as reported by Araujo et al. [10]. The result is similar to that reported by Bing et al. [19], where they observed diffraction peaks at 20 value of 36.94, 43.01, 62.21, 74.62, 78.63°, which can be indexed to the standard diffraction spectra of MgO powder. However, Araujo et al. [10] and Bing et al. [19] did not report the presence of a Mg(100) peak at a 2θ value of 29.33. On the other hand, Xiangcun et al. [20] showed that there was the formation of impurity of Mg(OH)₂ at 20 value around 30° in the XRD pattern, which could be due to high hydration properties of MgO materials. In general, the intensity of the peaks was higher when the MgO was synthesized using the hydrothermal method (MgO-18, MgO-36 and MgO-48) compared to MgO synthesized using the sol-gel method (MgO-SG). The hydrothermal method can increase crystallite size with good structure; hence the crystal quality is improved [21] compared to the sol-gel method, which results in a higher specific surface area [22].

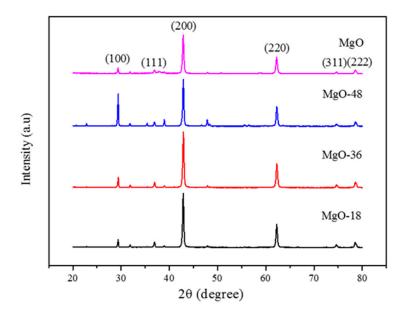


Figure 2. X-ray diffraction patterns of MgO synthesized using sol-gel and hydrothermal methods at pH 12.

From the result, MgO-36 showed the highest peak intensity at the plane (200), followed by MgO-18 and MgO-48. Increasing intensity peaks from 18 hours to 48 h synthesized duration can be related to Ahmedet al.'s study [23], which suggested that the catalyst shows the high structural ordering of mesopores in the material due to stronger intensity of peaks in the XRD spectrum. However, further extension duration to 48 h synthesis resulted in

decreased intensity peaks at the plane (200), which could be due to the structural ordering decreases in the material as the interplanar spacing increases, as reported by Ahmed [23].

When the synthesized MgO was impregnated with 30% Ce, the formation of new peaks with low intensities were observed at $2\theta = 28.51$, 33.32, 47.60, 56.35 and 78.71° (Figure 3 and Figure S2) in all 30Ce/MgO catalysts, which could be assigned to Ce (111), Ce (200), Ce (220), Ce (311) and Ce (331) planes, respectively, indicating the formation of cubic fluorite structure of CeO₂ as referenced from ICSD: 28709 [24,25]. Furthermore, the intensities of all MgO peaks is reduced while the Mg(200) and (220) peaks shifted to slightly higher 2 θ value of 43.17 and 62.94°, respectively, with lower intensities. The results are in agreement with research reported by Setoudeh et al. [26] that shows diffraction peaks of MgO would slightly shift to higher angles due to the diffusion of Ni²⁺ ions into the MgO lattice, while the diffusion of Mg²⁺ into Ni²⁺ lattice would result in the peaks to slightly lower angles. These phenomena are attributed to the presence of Ce diffused into the MgO lattice [27].

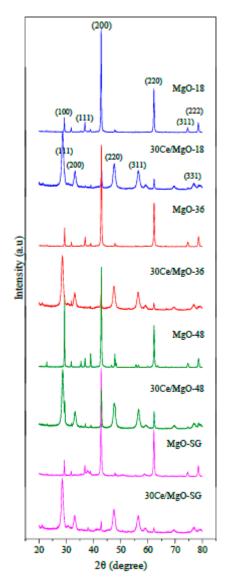


Figure 3. X-ray diffraction patterns of MgO-supported CeO₂ catalysts.

Table 1 shows the estimated crystallite size of the synthesized MgO and CeO₂, determined from the diffraction peaks for (200) and (111), respectively. The calculation of the crystallite size is based on the highest intensity peak for both MgO and CeO₂. The result showed decreased crystallite size as the hydrothermal duration process increased from 18 to 36 h. However, the crystallite size increases as the duration process are prolonged to 48 h for MgO catalysts, while MgO-36 shows a smaller crystallite size than MgO-18 and MgO-48. Soo Min et al. [28] observed that XRD peaks become sharper as the duration for hydrothermal treatment was increased as prolonged duration at hydrothermal temperature allows the crystals to grow more extensive during the hydrothermal process.

	Crystallite Size (nm)		
Catalyst –	Mg (200)	Ce (111)	
MgO-18	36.5	-	
MgO-36	29.9	-	
MgO-48	48.4	-	
MgO-SG	32.8	-	
30Ce/MgO-18	43.9	3.9	
30Ce/MgO-36	50.7	4	
30Ce/MgO-48	40.0	14.4	
30-Ce/MgO-SG	45.3	7.1	

 Table 1. Crystallite size of MgO and MgO-supported Ce catalysts.

The observation of the crystallite size trend for MgO-supported Ce catalysts shows a different pattern than for the MgO catalyst. From Table 1, it is clear that the 30Ce/MgO catalyst displays an increasing pattern, as hydrothermal duration increases from 18 h to 36 h. However, a contradiction trend in crystallite size was observed as the crystallization period was further increased to 48 for 30Ce/MgO-48 compared to 30Ce/MgO-18 and 30Ce/MgO-36 catalysts. 30Ce/MgO-48 possesses a smaller crystallite size, as the order of increasing crystallite size is 30Ce/MgO-36 > 30Ce/MgO-36 > 30Ce/MgO-18 > 30Ce/MgO-48. Crystallite size at Ce(111) plane shows a clear increasing pattern as synthesis duration increases. It could be caused by a lower state of dispersion of Ce at the site into the support surface, due to the agglomeration of impregnated metal into MgO pores/channel or surface as synthesized duration increases from 18 h to 48 h [29].

Figure 4 shows the N₂ adsorption–desorption isotherms of MgO-supported Ce catalysts, where all samples show a combination of Type II isotherms with a knee shape isotherm at a very low relative pressure (p/p_0) and Type III isotherms at higher p/p_0 according to IUPAC classification, a characteristic of non-porous materials [30,31]. Moreover, the isotherm destination turns upward at high relative pressure, which could be attributed to the voids between the particles [32]. Li et al. [33] stated that MgO exhibits a typical type-III isotherm—the N₂ adsorption–desorption isotherm, which is a typical characteristic of a porous structure with a significant H3 hysteresis loop. They also reported that observation at medium relative pressure (p/p_0) range of 0.5–1.0confirmed the presence of substantial mesopores in the MgO nanoparticles [33].

Table 2 shows the textural properties of the MgO and MgO-supported Ce catalysts where the surface area, pore volume and average pore size of MgO synthesized using the hydrothermal method increased when the crystallization period was increased from 18 to 36 h. However, when the crystallization period was further increased to 48 h, the surface area, pore volume and average pore size of the MgO decreased as the crystallite size of the MgO crystals that were formed also showed increases. The surface area and crystallite size of all catalysts showed a clear contradiction trend to each other, which agrees with the findings stated by Chowdhury et al. [33] and Li et al. [24]. Both studies reported that a bigger total surface area of the materials would result in larger interparticle porosity or voids between particles, thus reducing the crystallite size of the materials. As can be seen, MgO-36 showed the highest surface area (34.6 m²/g) with the smallest crystallite size (29.9 nm) among other MgO catalysts, while MgO-48 showed the smallest surface area (13.5 m²/g) with the biggest crystallite size (48.4 nm). The result obtained was also in agreement with the study carried out by Sui et al. [34] that showed CeO₂ support, and the Co/CeO₂ catalyst exhibited a smaller particle size correlated with a higher specific

surface area. For 30Ce/MgO catalysts, 30Ce/MgO-48 also showed the highest surface area (29.3 m²/g) compared to other 30Ce/MgO catalysts. Among the 30Ce/MgO catalysts, the surface area increases in an order of 30Ce/MgO-48 > 30Ce/MgO-SG > 30Ce/MgO-18 > 30Ce/MgO-36.

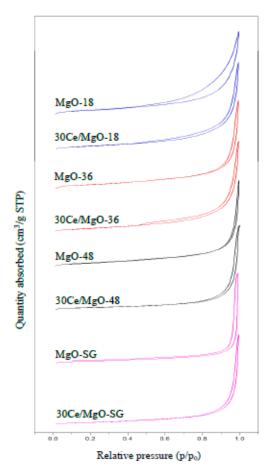


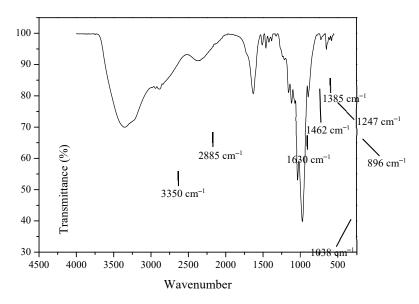
Figure 4. N₂ adsorption–desorption isotherms of the catalyst.

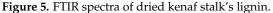
Table 2. Textural	properties of MgO a	nd 30Ce/MgO catalyst.

Catalyst	BET Surface Area (m²/g)	Pore Volume (cm ³ /g)	Average Pore Size (nm)	Crystallite Size (nm)
MgO-18	28.4	0.193	21.7	36.5
MgO-36	34.6	0.285	36.4	29.9
MgO-48	13.4	0.076	14.4	48.4
MgO-SG	27.3	0.146	20.4	32.8
30Ce/MgO-18	27.5	0.165	18.8	43.9
30Ce/MgO-36	25.9	0.158	18.2	50.7
30Ce/MgO-48	29.3	0.293	32.3	40.0
30Ce/MgO-SG	28.4	0.247	30.2	45.3

2.2. Lignin Extraction

Mohamad Aini et al. [35] reported that lignin content in kenaf is 19.2–21.2%. The lignin extracted from kenaf in this work is 23.5% compared to 20.8% reported earlier by Mohamad Aini et al. [35] for lignin extraction from kraft lignin of kenaf. On the other hand, Yang et al. [36] reported that 16.7% lignin was extracted from kenaf bast using the acid hydrolysis method. Figure 5 shows the infrared spectra of the extracted lignin, while the interpretation is tabulated in Table 3 and is compared with that reported in the literature.





According to Table 3, the broad peak at 3350 cm⁻¹ is attributed to the stretching vibration of O–H bonds [37], and the band at 2886 cm⁻¹ is attributable to the C–H stretching vibration in the methyl and methylene groups [38]. The C–H bending vibration in the methyl groups can be allocated to the band at 1462 cm⁻¹ [39]. The band 1650–1590 was due to C=C aromatic vibrations exhibiting the presence of lignin. The bands for C=O stretching vibration in the syringyl propane unit (S) and the guaiacyl propane unit (G) are at 1385 cm⁻¹ and 1247 cm⁻¹, respectively [40]. The band at 1037 cm⁻¹ can be assigned to the C=O deformation in secondary and primary alcohol or aliphatic esters [41]. An absorption band at 897 cm⁻¹ was assigned to C–O–C stretching at the β – (1, 4)–glycosidic linkage in cellulose [42].

Table 3.	Functional	group	and freq	juency of	of FTIR.

IR Band (cm ⁻¹)		n ⁻¹)	Assistment
Literature Data	[ref]	Data Obtained	- Assignment
3500-3100	[35]	3350	Stretching vibration of O-H bonds
2900-2800	[38]	2886	C–H stretching vibration in the methyl and methylene groups
1650-1590	[40]	1630	C=C aromatic vibrations
1464-1424	[39]	1462	C–H bending vibration in the methyl groups
1272-1265	[40]	1247	C=O stretching vibration in guaiacyl propane units (G)
1340–1330	[40]	1385	C=O stretching vibration in syringyl propane units (S)
1130–1035	[41]	1038	C=O deformation in secondary and primary alcohol or aliphaticesters
897-889	[42]	896	C–O–C stretching at the β-(1, 4)-glycosidic linkage in cellulose

2.3. Catalyst Testing

Figure 6 shows the HPLC chromatogram of vanillin standard, where it shows that the retention time of vanillin is 5.730 min. Based on this chromatogram, five different concentrations of standard vanillin were prepared for the calibration curve to allow for the quantitative determination of the vanillin produced from direct oxidation of kenaf stalks.

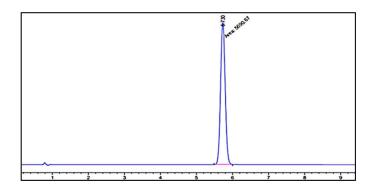


Figure 6. HPLC chromatogram of vanillin standard.

Figure 7 shows the calibration curve of vanillin standard at five different concentrations $(1.25 \times 10^{-4}, 6.25 \times 10^{-5}, 3.125 \times 10^{-5}, 1.56 \times 10^{-5} \text{ and } 7.81 \times 10^{-6})$ using HPLC. The curve was plotted to measure the peak area attained from the injected aliquot of standard vanillin solution with known concentration. The calibrated curve will be used to determine the vanillin yield quantitatively. The correlation coefficient, R² of the best-fit line displayed at 0.9995, indicates high accuracy.

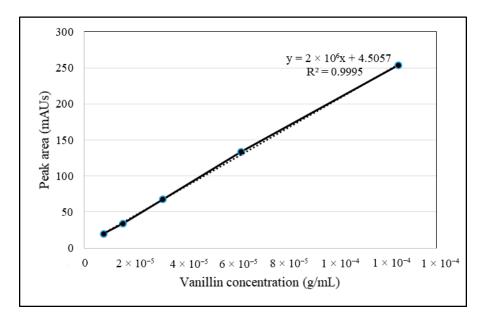


Figure 7. Calibration curve of the vanillin standard at five different concentrations.

When the catalysts were tested in the direct oxidation of kenaf stalks, the liquid produced from oxidation in the presence of 30Ce/MgO-36 shows the presence of vanillin at the retention time of 5.73 min (Figure 8). However, the chromatogram also shows the presence of three other compounds at the retention time of 1.38, 3.98 and 7.18 min. The other compounds could be 4-vinylguaiacol, syringol and syringaldehyde, respectively.

Vanillin was detected in the liquid produced from direct oxidation of kenaf stalks in all MgO-supported Ce catalysts. In contrast, no vanillin was detected when MgO was used as the catalyst, which indicates that Ce is the active component responsible for the formation of vanillin. Using the linear equation derived from the calibration curve (Figure 7), the vanillin produced from direct oxidation of kenaf stalks at 180 °C and ambient pressure can be calculated. Our earlier work on converting lignin derived from pineapple leaves shows a vanillin yield of 6% at 170 °C and pressure of 15 bars by using compressed air as an oxidizing agent [43].

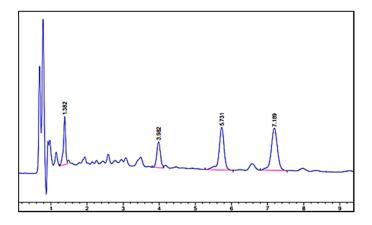


Figure 8. HPLC chromatogram of derived vanillin.

Figure 9 shows the effect of reaction time on vanillin yield. There are no significant differences in vanillin yield for all catalysts at reaction duration of 10 and 30 min. At the same time, there is an increasing trend for catalyst 30Ce/MgO-48 compared to other catalysts at a 20-minute reaction duration. The 30Ce/MgO-48 catalyst showed the highest vanillin yield of 0.684% when the reaction was performed for 20 min at 180 °C, while 30Ce/MgO-18, 30Ce/MgO-36 and 30Ce/MgO-SG gave a vanillin yield of 0.152, 0.217 and 0.202%, respectively. This is considered substantial since the vanillin is produced directly from kenaf stalks without separating the lignin from the cellulose and hemicellulose components.

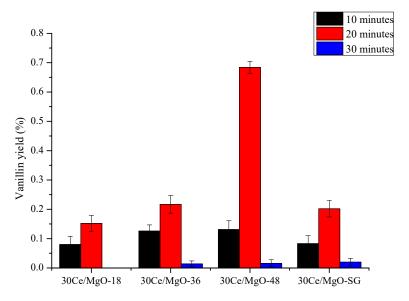


Figure 9. Reaction condition: 2 g of dried kenaf stalks, 20 mL of 0.01 N NaOH solution, 10 wt.% of catalyst, 1 mL of H_2O_2 at 180 °C.

Furthermore, the reaction was performed at pH 11.5, which is lower than pH 12–14 as reported by other researchers [1,10]. Direct production of vanillin has also been achieved by Chen et al. [12]. Japanese cedar wood was used as a biomass source that gives 5% vanillin yield. The CuO catalyst combined with H_2O_2 solution as an oxidant showed the best yield for 20 min under 180 °C. From this result, 20 min was chosen as the optimum reaction time. Das et al. [44] stated that further extension duration might lead to low product yield due to repolymerization of depolymerized lignin or over-oxidized products that form during the time while the reaction occurs. The research properties were continued to determine the effect of reaction temperature on the vanillin yield.

There was no vanillin produced using MgO catalyst; however, by using ceria loading, the presence of vanillin was detected. Nolan et al. [45] reported that ceria could be an

active catalyst due to its high oxygen storage capacity, which can absorb/release oxygen under oxidizing/reducing conditions. It is related to the comparatively facile formation of O vacancies in CeO₂ [46]. Several studies have investigated and confirmed the presence of peroxide species on CeO₂ surfaces, but these have been solely related to adsorbed molecular oxygen [47]. Oxidation reactions continuously occur because oxidized surfaces of ceria are being provided by peroxide, which acts as an oxidant in the reaction. The low mass transfer and low concentration rate of O₂ during the reaction are also being provided by H₂O₂ [44].

Figure 10 shows the effect of reaction temperature on vanillin yield using all the MgO-supported Ce catalysts. Vanillin yields increase with increasing reaction temperature from 160 to 170 °C using all catalysts. However, the vanillin yield decreased when the temperature was further increased to 180 °C regardless of the catalyst used, which could be attributed to the degradation of vanillin at high temperatures, as reported by Chen et al. [12]. Based on Yuting et al. [37], prolonging the reaction time did not improve the vanillin yield, which could be caused by low selectivity due to tandem reaction. According to Tarabanko et al. [48], increasing the temperature of aspen wood oxidation to more than 160 °C can give a higher yield of vanillin and syringaldehyde, while another study showed a high aromatic aldehyde yield in aspen wood oxidation when the temperature was raised from 110 °C to 160 °C [48,49].

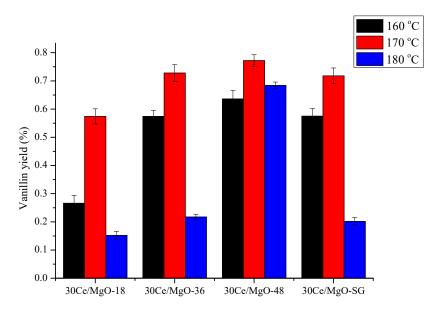


Figure 10. Reaction condition: 2 g of dried kenaf stalks, 20 mL of 0.01 N NaOH solution, 10 wt.% of catalyst, 1 mL of H₂O₂ at 20 min.

Figure 10 shows that 170 °C reaction temperature with a 20-minute reaction time presents the high vanillin yield, which was 0.772% in the presence of 30-Ce/MgO-48 catalyst and followed by 0.728, 0.718, 0.716% for 30Ce/MgO-36, 30Ce/MgO-SG, 30Ce/MgO-18 catalysts, respectively.

Based on the vanillin yield obtained in Figures 9 and 10, 30Ce/MgO-48 shows the higher vanillin yield for reaction time and temperature. Hence, 30Ce/MgO-48 was chosen as the best catalyst for further optimization on catalyst loading with a fixed amount of biomass. Figure 11 shows the vanillin yield obtained on the different amounts (5, 10 and 15 wt.%) of 30Ce/MgO-48 catalyst loading synthesized with 2 g of kenaf dried stalks for 20 min at 170 °C under microwave heating. The result shows the increased number of yields with the increased amount of catalyst loading with vanillin yield of 0.141% (5 wt.%), 0.772% (10 wt.%) and 1.37% (15 wt.%). These were due to the higher total contact frequency of catalysts with lignin, as reported by Chen et al. [12]. In addition, higher catalyst dosage indicates a much more catalytic active center resulting in higher catalytic performance [50].

This result is also in agreement with the research carried out by Prado et al. [51], which stated that increasing catalyst loading from 1% to 20% preceded to greater vanillin yield from 0.015 to 0.22%. It is also speculated that increases in catalyst loading resulted in more active sites available to break down lignin into aromatic aldehyde products.

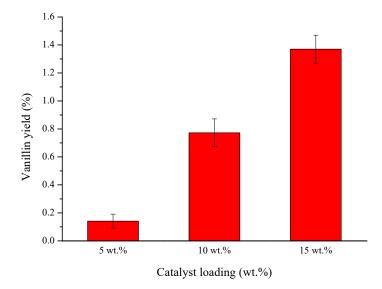


Figure 11. Reaction condition: 2 g of dried kenaf stalks, 20 mL of 0.01 N NaOH solution, 1 mL of H_2O_2 at 20 min for 170 °C.

Figure 12 shows vanillin production from extracted lignin and biomass synthesized using 15 wt.% of heterogeneous and homogeneous catalyst for 20 min at 170 °C under microwave heating. The optimum operating conditions for a time, temperature and catalyst loading were chosen to synthesize the extracted lignin and direct biomass for vanillin production in the presence of heterogeneous (30Ce/MgO-48) and homogeneous (NaOH) catalysts. This study was performed to compare their catalytic performance and obtained yield on vanillin production. The vanillin yield obtained in the presence of 30CeMgO-48 as heterogeneous catalysts were 3.70% and 2.90% for oxidations of extracted lignin and the direct oxidation of biomass, respectively. On the other hand, 7.80% and 4.45% were obtained in the presence of 2N NaOH as a homogeneous catalyst for oxidations of extracted lignin and direct biomass, respectively.

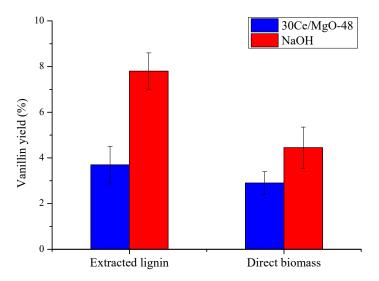
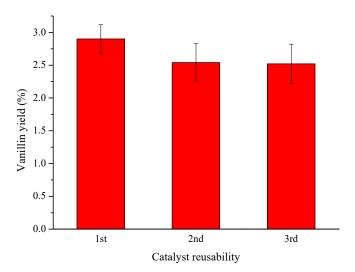


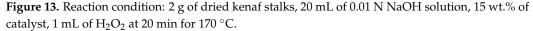
Figure 12. Reaction condition: 2 g of dried kenaf stalks and 1.06 g of extracted lignin, 20 mL of 0.01 N NaOH solution, 15 wt.% of catalyst, 1 mL of H_2O_2 at 20 min for 170 °C.

Extracted lignin obtained a higher vanillin yield than direct biomass because of the isolation of lignin as an active compound for conversion to vanillin. The vanillin production is caused by the breaking down of aryl ether linkages in β -O-4 bonds and C_{α} - C_{β} cleavage between lignin structures through the oxidation process. Compared to biomass, the oxidation process directly to biomass without separating lignin with cellulose and hemicellulose was the cause of lower vanillin yield. According to Lu et al. [52], more than 50% of aryl ether bonds (β -O-4) are contained in the lignin structure. Furthermore, the (β -O-4) bonds are a stable linkage (52–75 kcal mol⁻¹) and require high energy for the cleavage process to occur [53]. In this regard, the use of catalysts is vital for the efficient and selective cleavage of (β -O-4) bonds because of the unique characteristics of catalysts that render them able to control the activation routes of cleavage [54]. Cleavage (β -O-4) bonds in the lignin structure from direct biomass are challenging because the lignin structure is not separated from cellulose and hemicellulose components.

As reported by previous studies, the use of supported or unsupported metal catalysts such as Pd [52], Ce [55], V [56], Ni [52], Cu [57] and Ru [58] as heterogeneous catalysts was successful in producing vanillin with high selectivity and conversion percentage. However, there are limitations during the reaction process where the reaction cannot proceed for a new turn when the lignin fragment is saturated on the catalyst's surface. On the other hand, the homogeneous catalyst is counted as flexible a catalyst that allows them to freely contact the C_{Ar} -O bonds without high steric limitation, since the reaction occurs solely in a liquid phase. This condition results in a reduction in mass transfer limitations [59], which is one of the reasons for the high production yield of vanillin in the presence of homogeneous catalysts compared to heterogeneous catalysts. Moreover, NaOH was used as a homogeneous catalyst during the oxidation reaction. A study reported by Lopes et al. [54] stated that using Ionic Liquids (ILs) as a green framework, ground-breaking homogeneous catalyst at the same time. They also stated that the cleavage of lignin ether bonds was promoted by acidic ILs approaches.

The reusability of the 30Ce/MgO-48 catalyst was determined by the repeated use of the catalyst for the direct oxidation of Kenaf stalks to vanillin production three times with the new reactants and optimized reaction conditions. After the first use, the recovered catalyst was washed with ethanol to remove the organic deposits such as unreacted biomass and derived biomass product, then dried at 100 °C for 12 h. Figure 13 shows that the 30Ce/MgO-48 catalyst is chemically stable and can be repeated for a third time with a slightly significant loss in activity compared to vanillin's yield in the first use of catalyst. There are 0.5% yield differences between the first and second cycle with 2.90% and 2.54%, while there are no significant in yield obtained for the third cycle with 2.52% vanillin. The slight decrease in vanillin production for the second cycle may be attributed to the saturated surface of catalyst by side product of biomass, including a major lignin-like oligomers product from vanillin oxidative polymerization [60,61].





3. Materials and Methods

3.1. Catalyst Preparation

MgO was synthesized using sol-gel and hydrothermal methods. First, 1 M magnesium nitrate solution was prepared using magnesium nitrate hexahydrate (Mg(NO₃)₂·6H₂O) salts in the sol-gel method. Then, 1 M sodium hydroxide (NaOH) was added dropwise to the magnesium nitrate solution under vigorous stirring to adjust the pH to pH 12. The mixture was stirred until a white solution was formed. Then, the white solution was dried at 120 °C for 12 h followed by calcination at 500 °C for 5 h. The sample is denoted as MgO-SG. For the synthesis of MgO under the hydrothermal method, the same procedure was repeated until the formation of a white solution. The white solution was then transferred to a Teflon-lined stainless steel hydrothermal vessel and left for crystallization at 120 °C for 18–48 h, followed by drying at 120 °C for 5 h and calcination at 500 °C for 5 h. The catalysts were denoted as MgO-18, MgO-36 and MgO-48 for 18, 36 and 48 h crystallization time. The calcined MgO was impregnated with 30 wt.% of Ce using Ce(NO₃)₃.6H₂O via incipient wetness impregnation method. Typically, the required amount of metal salt was dissolved in a sufficient amount of distilled water before MgO was added to the solution and stirred for 3 h at room temperature. The slurry was then dried at 120 °C for 12 h and calcined at 500 °C for 5 h.

3.2. Catalyst Characterization

The crystallinity of the MgO-supported Ce catalysts was determined using Bruker X-ray diffraction (XRD) model X'Pert³ Powder and Empyrean (PANanalytical, Billerica, MA, USA). The catalysts were recorded on an X-ray Diffractometer system between 20 value of 20–80° with 227 s/step exposure time and 0.105°/step step size. Surface area and pore size of catalysts were analyzed using Brunauer-Emmett-Teller (BET) (Micromeritics ASAP 2020, Norcross, GA, USA). The catalysts were degassed at 200 °C for 24 h, prior to N₂ adsorption measurement at -77 °C. The crystallite size of synthesized MgO particles was determined according to Scherrer's equation: D = k λ/β cos θ . Crystallite size were analyzed using HighScore Plus software (version 3.0), Malvern, UK with X-ray wavelength of CuK α radiation at $\lambda = 1.54$ A°, where θ is Bragg angle. β is the full width at half maximum in radians was taken corresponding to the 2 θ value at plane (100) and (200). The unknown shape factor, k, was assumed as 0.89, the reflecting peak at 2 θ was chosen for the entire sample.

3.3. Lignin Extraction and Characterization

Dried kenaf stalks were broken into 10 cm lengths before being grinded into small pieces using a grinder. The extractive compounds such as protein, waxes, resin, fatty acids and lipid were removed from the dried kenaf stalks using a mixture of ethanol and water with (2:1) ratio via Soxhlet extraction for 8 h. Extracted sample was dried in the oven for 24 h at 60 °C. The dried residue was then soaked in an NaOH solution for 5 min with a sample to the solvent ratio (6: 100 g/mL). The mixture was stirred and placed into a hydrothermal pressure vessel prior for the cooking process at 121 °C for 2 h to break the bonds between lignin and hemicellulose. Heated mixture left to cool at room temperature before the separation of solid residue through filtration. The dark brown sample fluid (black liquor) was kept overnight at 4 °C (refrigerator). Black liquor was acidified by H₃PO₄ (20%) at pH 2. Lignin precipitates into semi-solid form at around pH 14. The precipitated lignin was then vacuum filtrated using a 0.45 µm pore filter and washed using distilled water and 20% H₃PO₄. The isolated was dried in an oven at 55 °C for 48 h [43]. The sample was characterized using FTIR (Perkin Elmer Frontier 01, Cheshire, UK) between 500 and 4000 cm⁻¹ for 50 scans with a resolution of 4 cm⁻¹.

3.4. Catalytic Oxidation of Raw Kenaf

The performance of the catalyst was tested for the direct oxidation of kenaf stalks and extracted lignin to vanillin under microwave irradiation. Two grams of dried kenaf stalks and 1.06 g of dried lignin were soaked in 20 mL of 0.01 N NaOH solution in a screw-cap test Then, tube for 24 h before it was transferred to a Teflon tube. 5 wt.%–15 wt.% of catalyst and 1 mL of H_2O_2 were added to the mixture and the pH was adjusted to pH 11.5 and stirred at 600 rpm for 10 s at room temperature using a magnetic stirrer before the mixture was heated in the 300W of microwave heating (Milestone Srl, Milan, Italy, MicroSYNTH MA143) at 160–200 °C for 10–30 min. Then, the mixture was cooled to room temperature and filtered through filter paper to remove the insoluble materials. The residue was washed with 20 mL of 0.01 N NaOH solution twice. The same procedure was performed by changing the heterogeneous catalyst to a homogeneous sodium hydroxide catalyst to study vanillin production for direct oxidation from Kenaf stalks and extracted lignin. According to the obtained result, 15 wt.% was chosen as optimized catalyst loading for further study using 2N of sodium hydroxide catalyst. The filtrate was transferred to a glass test tube with a cap. At the same time, the catalyst residue was washed several times with ethanol to further investigate catalyst reusability under optimized reaction conditions, followed by a drying process under an oven at 100 °C for 2 h. Concentrated hydrochloric acid (37%) was then added to the filtrate at a 1:2 ratio. The mixture was then agitated using a vortex agitator at 5000 rpm for 15 s, followed by centrifugation for 15 min at 1000 rpm. Next, ethyl acetate was added to the supernatant at a 1:1 ratio. The low molecular weight compounds were extracted into the organic phase, including vanillin. The mixture was agitated for 60 s at 5000 rpm followed by centrifugation at 1000 rpm for 5 min, after which the mixture separated into two phases. The upper phase was transferred to a vial. Excess solvent was removed using a rotary evaporator at 50 °C and 400 mbar [12].

The brownish solution containing vanillin was analyzed using Agilent High Performance Liquid Chromatography (HPLC) with a UV-VIS detector at a wavelength of 280 nm. The HPLC (Agilent 1200 series, Santa Clara, CA, USA) was equipped with Hypersil C18 column, Santa Clara, CA, USA (particle size of 5 μ m, 150 × 4.6 mm inside diameter). The column temperature was set at 35 °C and injection volume of 20 μ L with a mixture of acetonitrile: water (1:8 v/v) containing 1% acetic acid used as the mobile phase at a flow rate of 2 mL min⁻¹. Vanillin (Sigma-Aldrich, purity 99.7%, Darmstadt, Germany) was used as standard [61].

4. Conclusions

Vanillin was successfully produced from direct oxidation of kenaf stalks and lignin extract from kenaf stalks under microwave heating using H_2O_2 as an oxidizing agent in the presence of MgO-supported Ce catalysts at pH 11.5. The vanillin yield of 3.70% was achieved from extracted lignin using 30Ce/MgO-48 catalyst at 170 °C for 20 min with catalyst loading of 15%, while direct oxidation from kenaf stalks 2.90%. On the other hand, the highest vanillin was obtained when NaOH was used as a homogeneous catalyst with 7.80% and 4.45% for oxidations of extracted lignin and direct biomass, respectively. The heterogeneous 30Ce/MgO-48 catalyst is chemically stable and can be repeated use for 3 times.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10.3390/catal11121449/s1, Figure S1. X-ray diffraction patterns of MgO. (a) MgO-18; (b) MgO-36; (c) MgO-48; (d) MgO-SG, Figure S2. X-ray diffraction patterns of 30Ce/MgO. (a) 30Ce/MgO-18; (b) 30Ce/MgO-36; (c) 30Ce/MgO-48; (d) 30Ce/MgO-SG, Figure S3. Gas-chromatogram of degradation wood product.

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