

Supporting Information: Integrating Soft Hydrogel with Nanostructures Reinforces Stem Cell Adhesion and Differentiation

Bohan Yin ¹, Hongrong Yang ² and Mo Yang ^{1,*}

¹ Department of Biomedical Engineering, The Hong Kong Polytechnic University, Hong Kong 999077, China; bohanyin93@gmail.com

² Department of Bioengineering, College of Engineering, Northeastern University, Boston, MA 02115, USA; donnayeung0722@gmail.com

* Correspondence: mo.yang@polyu.edu.hk; Tel.: +852-2766-4946

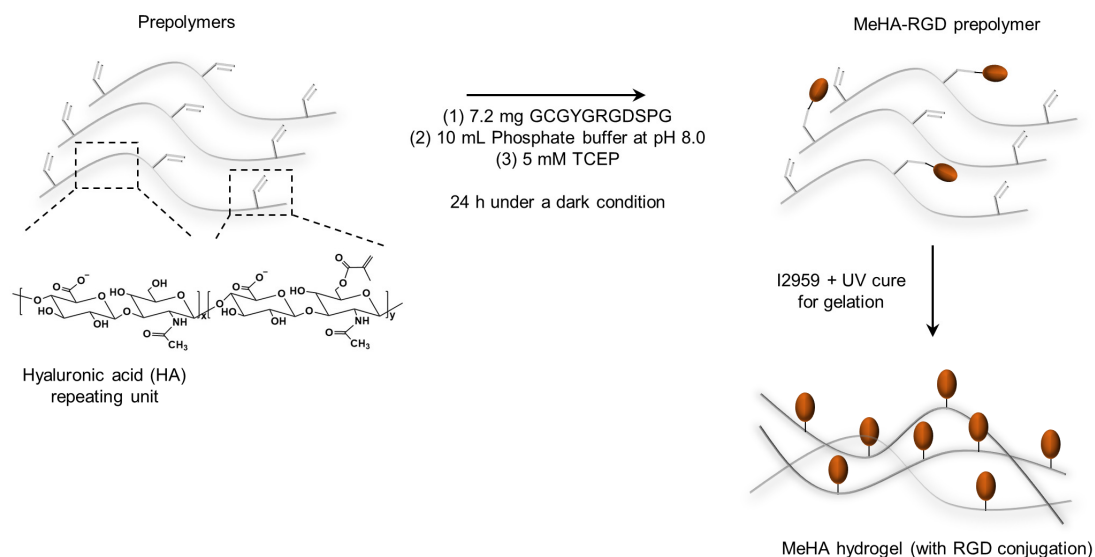


Figure S1. Synthetic route of RGD-coupled methacrylated hyaluronic acid (MeHA) hydrogel.

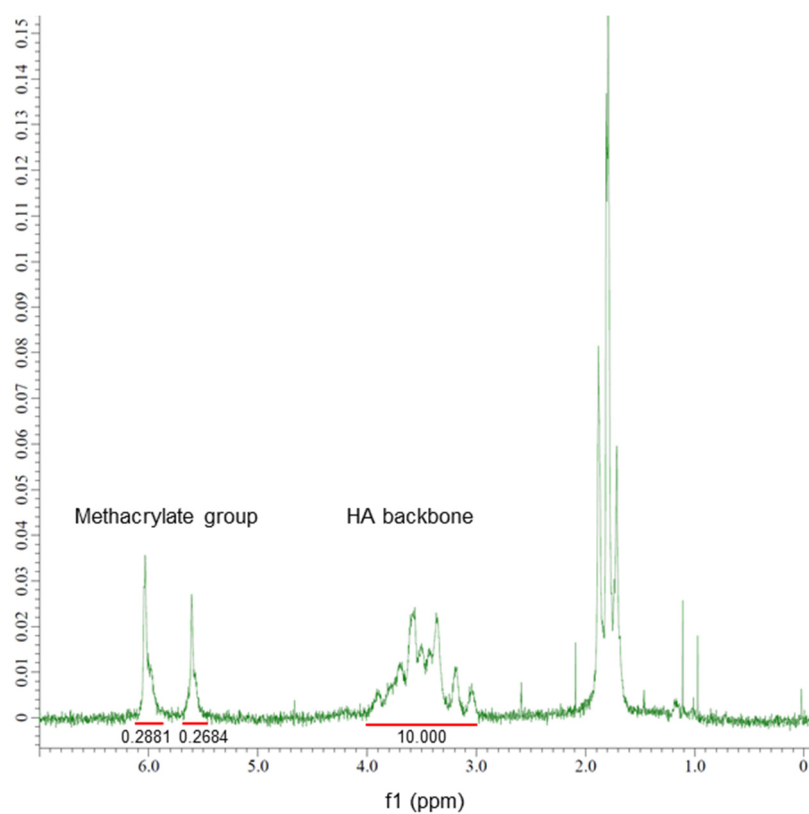


Figure S2. ^1H NMR spectra of the methacrylated hyaluronic acid (MeHA). The integration of the spectrum shows that methacrylation degree is ~30%, confirming the successful methacrylation in this study.

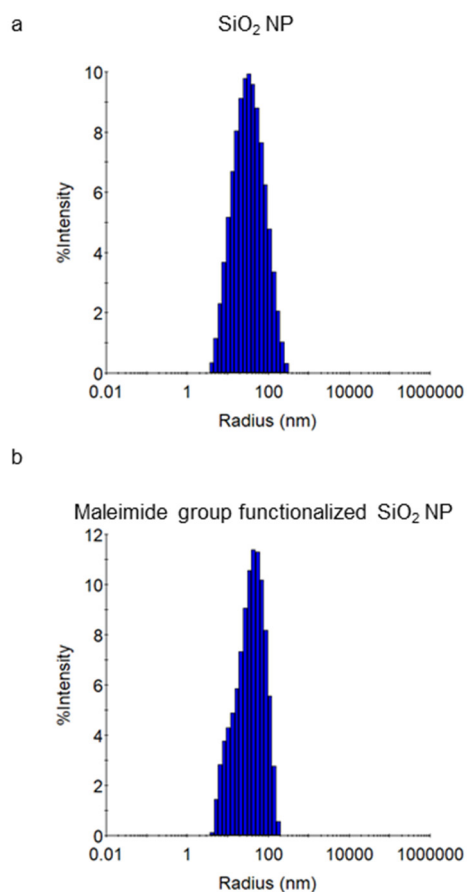


Figure S3. Dynamic light scattering (DLS) characterization for the size of (a) SiO NPs, (b) maleimide group functionalized SiO₂ NPs. The average hydrodynamic measured diameters of (a) and (b) were 31.4 ± 2.5 nm and 39.5 ± 5.4 nm, respectively.

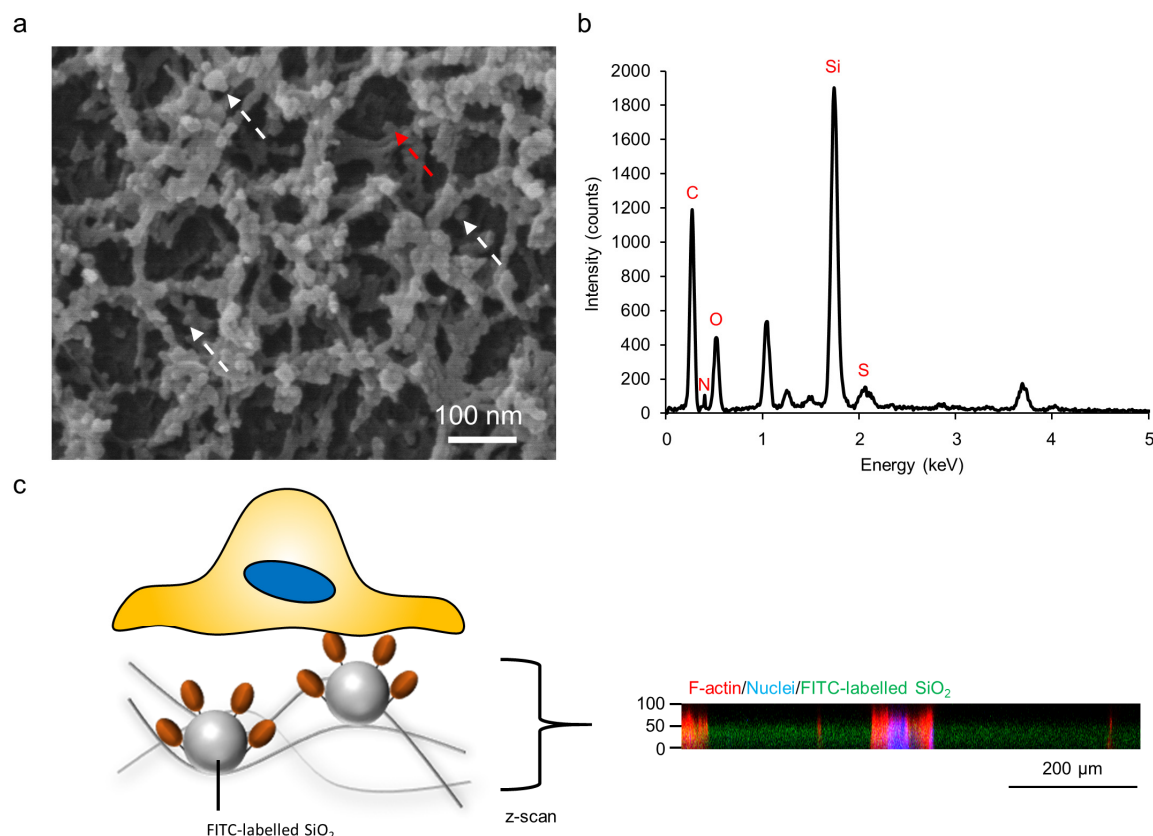


Figure S4. Exploration of immobilized SiO₂ NPs in MeHA hydrogel network. (a) Magnified SEM image of MeHA-SiO₂ hydrogel. White arrows indicate the immobilized SiO₂ NPs distributed throughout different layers of MeHA hydrogel. (b) EDS spectrum from red arrow in (a), indicating high content of SiO₂ inside the MeHA network. (c) Cells cultured on MeHA-SiO₂ hydrogel where SiO₂ NPs were labelled with FITC. The right panel shows the z-axis stacking images of the FITC-labelled SiO₂ (green) conjugated on MeHA hydrogel, with the presence of cells cultured on the hydrogel (6 h of cell culture). The cells were also stained with F-actin (red) and nuclei (blue).

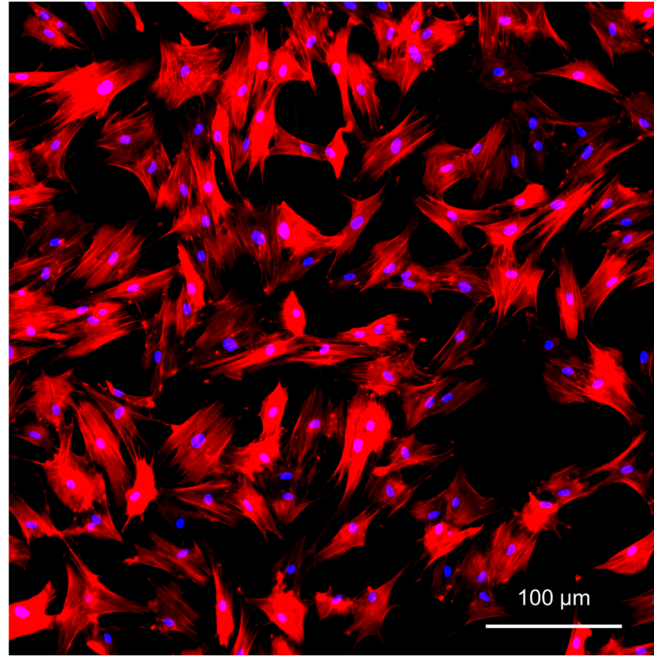


Figure S5. Cells cultured on a glass substrate were considered as a positive control group as the cells spread well on it. The spreading area was $5841 \pm 518 \mu\text{m}^2$.

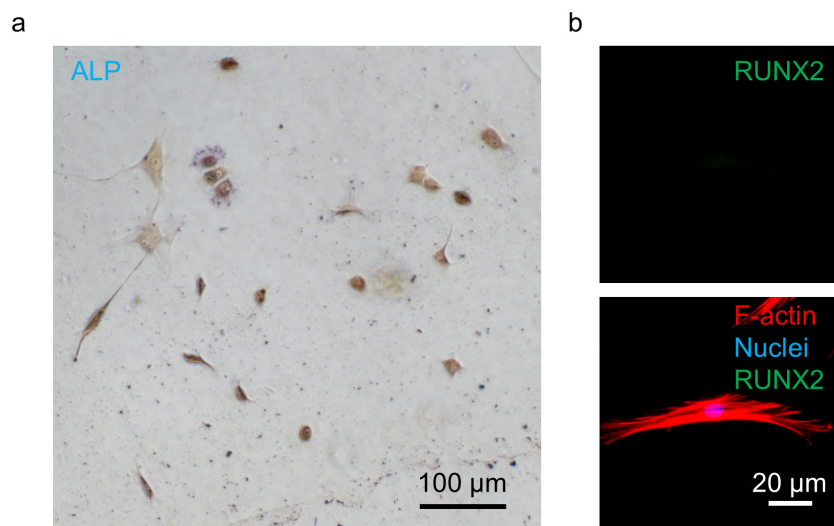


Figure S6. Examination of the expression level of ALP and RUNX2, the early-stage markers of osteogenic differentiation of stem cells, in hMSCs without the osteogenic medium by immunostaining on MeHA hydrogel (with RGD conjugation). (a) ALP staining result. There were only 8.41 ± 2.13 % of positively stained cells for expressing ALP. (b) Immunostaining result RUNX2 (green) merging with F-actin and nuclei staining. The nuclear localization ratio of RUNX2 was 0.37 ± 0.82 in the cells. These results suggest that undifferentiated hMSCs show minimal basal expression of these two markers.