



Article Feed and Disease at Olive Flounder (Paralichthys olivaceus) Farms in Korea

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Abstract: The objective of this study was to conduct field experiments comparing formulated (extruded pellets (E.P)) and raw-fish (moist pellets (M.P)) feeds at two flounder aquaculture farms for six months to obtain the basic data necessary for improving aquafarmers' awareness of feed-quality issues and firmly establish and expand the use of formulated feed. According to the results, the M.P group was higher in weight gain, feed efficiency ratio, protein efficiency ratio, and specific growth rate and lower in mortality rate than the E.P group. All cases of mortality were caused by six kinds of disease, four of which (*Vibrio* sp., *Edwardsiella* sp., *Streptococcus* sp., and *Scuticociliates*) were common to the two groups, whereas *Lymphocystis* arose only in the M.P group and abdominal inflation only in the E.P group at both farms. As for mortality in the present experiment, the numbers were 1047 at Da-Hae farm and 1167 at Global farm, with more fish dying in the E.P tanks than in the M.P tanks. By multiplying the number of deaths by selling price, the economic losses were \$9650 and \$10,756, respectively. Therefore, it should be considered an urgent priority to develop flounder-exclusive formulated feed for improved digestion and absorption rate and also to establish a water-quality-improvement management plan.

Keywords: olive flounder in Korea; moist pellets; extruded pellets; Lymphocystis; abdominal inflation

1. Introduction

Marine aquaculture has developed rapidly around the world, and is now regarded as an essential component of the global marine food supply. Global aquaculture production increased approximately three-fold, from 26.7 million metric ton in 1996 to 73.8 million metric ton in 2014, and in that year, fish accounted for 44% of global production from capture fisheries and aquaculture [1,2]. In the Republic of Korea, which ranked 7th among the world's top 25 aquaculture producers in 2014, flounder, the most produced fish, accounted for 46.2% (37,240 metric ton) of the total fish farming yield (80,530 metric ton) in 2018 [3].

A variety of interdisciplinary studies entailing selective breeding for improvement of growth [4,5], feed [6–8], and vaccine development [9–13] have been carried out to develop flounder as a national strategic breed. The government has been promoting the use of formulated feeds since 2004 in order to develop a sustainable aquaculture industry. One result of this development is the Standard Guide Book, which was published in 2006 for olive flounder (*Paralichthys olivaceus*) culturing and again in 2007 for rockfish (*Sebastes schlegeli*); it includes information on aquaculture technology, techniques, and processes such as fish farming, parenting management, seedling production, nurturing management, feeding

systems, disease control, and economic analysis of regional and scale forms. Despite such efforts, however, the use of formulated feed for fish farming in 2018 was only 23% (155,897 metric ton) of total fish feed (670,631 metric ton), while the rate at flounder farms accounted for only 7.5% (20,656 metric ton) of total flounder feed (274,599 metric ton) consumed at flounder farms [14].

In developed countries, the aquaculture industry and economy has been revitalized by, in part, the development and distribution of low-cost high-quality/efficiency formulated feeds through long-term field experiments. In Japan, for example, changes in awareness of the use of formulated feed by aquaculture producers due to increasing raw-fish feed have encouraged active formulated feed research and development by feed companies, which has led to a reduction in feed prices. In Norway, overall aquaculture costs have been reduced by reducing fishmeal content in feed from 75 to 20–25% in salmon farming. The utilization rate of formulated feed is more than 90% for red sea bream and salmon at Japanese and Norwegian farms, respectively, and more than 65% for yellow tail/amberjack at Japanese farms. [15]. However, in Korea, despite the various problems incurred by the use of raw fish feed (i.e., moist pellets (M.P)), which include, for example, overfishing of small fish such as anchovies, sardines, and herring, and deterioration of water quality and environmental pollution, M.P is still preferred at flounder farms due mainly to its low cost and high growth rate effect for flounder compared with formulated feed (i.e., extruded pellets (E.P)) [16].

Therefore, the objective of this study was to conduct field experiments comparing formulated (E.P) and raw-fish (M.P) feeds at two flounder aquaculture farms to obtain the basic data necessary for improving aquafarmers' awareness of feed-quality issues and firmly establish and expand the use of formulated feed.

2. Results and Discussion

2.1. Evaluations of Extruded Pellet (E.P) and Moist Pellet (M.P) Systems for Fish-Growth Performance

The growth performance and body compositions of the flounder fed with either of the two diets (E.P or M.P) are provided in Table 1 and Figure 1. The monthly weight gain rate relative to the stock weight (78.0 g) at Da-Hae was 56% (August), 221% (September), 437% (October), 625% (November), and 765% (December) in the control group (M.P) and in the experimental group (E.P) it was 56% (August), 155% (September), 300% (October), 473% (November), and 606% (December). Meanwhile, the rate of monthly weight gain rate relative to the stock weight (113.8 g) at Global was 47% (August), 198% (September), 291% (October), 434% (November), and 472% (December) in the control group (M.P) and in the experimental group (E.P) it was 44% (August), 170% (September), 271% (October), 367% (November), and 401% (December). Overall, there was a significant difference in the values of weight gain (WG), feed efficiency ratio (FE), protein efficiency ratio (PER), and specific growth rate (SGR) between the two diets at both farms.

Protein in feed for flounder is important to meet amino acid and energy requirements, since this carnivorous fish has high protein availability but low carbohydrate and fat availability [17–19]. The results of relevant previous studies [20–27] indicated that the range of protein requirements varied according to fish size (4–307 g); however, the difference in protein content between the two feeds (> 59.9% in the present study) was not considered to have affected fish growth, given the fact that the range of protein content in feed was 45–60% as well as the Fish Weight theory's contention that the correlation between fish growth and protein content in diet is not always proportional (40%, flounder) [18]. In studies on lipid supply in feed [28–32], the proper requirements for lipids also varied (9–17%) according to fish size (18–597 g); however, as the lipid content in the feed increased, fish growth and the lipid contents in the fish whole body tended to be higher. Therefore, in the present study, the higher growth (Table 1 and Figure 1A,B) and lipid contents in the whole body (Table 1) of the fish in the control group (M.P) relative to the experimental group (E.P) was considered to have been due to their feed's high lipid content.

Cholesterol, a major component of the cell membranes of animal cells, is a lipid found mainly in animal tissues. It is reported that it plays an important role in the immune system as a precursor of hormone and vitamin D [33,34]. Previous studies have shown that an increase in blood cholesterol by addition of fat sources in feed improved the disease resistance and immunity of fish against the pathogen *Aeromonas hydrophila* [35] and improved immune cell activation [36]. Indeed, the lipid content in feed and blood cholesterol are correlated, and, accordingly, the higher value of blood cholesterol in the fish in the control group (M.P) compared to those in the experimental group (E.P) was considered to have been due to the difference in the lipid contents between the two feeds (Table 2 and Figure 1C,D).

		Da-	Hae	Global			
		E.P ²	M.P ³	E.P ²	M.P ⁴		
Growth performance & Feed efficiency	Initial mean weight (g fish ⁻¹)	78.0 ± 0.9	78.0 ± 2.4	113.8 ± 3.0	113.8 ± 6.1		
	Initial number (fish tank ⁻¹)	6000	6000	5000	5000		
	Final mean weight (g fish ⁻¹)	551.0 ± 10.9 b	674.9 ± 8.3^{a}	570.5 ± 4.4 ^b	651.4 ± 19.9 ^a		
	WG (%) ⁵	606.4 ^b	765.3 ^a	401.3 ^b	472.4 ^a		
	FE (%) ⁶	99.4 ^b	113.8 ^a	90.1 ^b	100.5 ^a		
	PER ⁷	1.66 ^b	1.99 ^a	1.51 ^b	1.58 ^a		
	SGR (%/day) ⁸	1.24 ^b	1.37 ^a	1.09 ^b	1.18 ^a		
	Mortality rate (%) ⁹	35.6 ^b	30.4 ^a	25.2 ^b	18.2 ^a		
Whole-body proximate	Moisture ¹⁰	70.3	66.4	72.8	70.3		
	Crude protein ¹⁰	20.9	21.9	19.6	19.5		
composition	Crude lipid ¹⁰	3.2 ^b	8.7 ^a	4.3 ^b	6.2 ^a		

Table 1. Growth performance, feed efficiency, and whole-body proximate composition at the end of the 5-month experiment at two farms raising olive flounder fed with experimental diets (extruded pellets (E.P) and moist pellets (M.P))¹.

¹ Values are the mean of triplicate groups and presented as mean \pm SD. Values in the same row having different superscript letters (^{a,b}) are significantly different (p < 0.05). ² Commercial feed containing 59.9% protein and 11.2% lipid, Republic of Korea. ³ Raw fish feed (composed of mackerel and herring) containing 61.2% protein and 22.0% lipid. ⁴ Raw fish feed (composed of mixed fish such as cutlass, jack mackerel, croaker, mackerel, and longsnout pricklebacks) containing 63.8% protein and 22.8% lipid. ⁵ Weight gain (WG, %) = (final weight – initial weight) × 100/initial weight. ⁶ Feed efficiency ratio (FE, %) = (wet weight gain/dry feed intake) × 100. ⁷ Protein efficiency ratio (PER) = wet weight gain/protein intake. ⁸ Specific growth rate (SGR, %) = (loge final weight – loge initial weight) × 100/days. ⁹ Mortality rate (%) = dead fish × 100/initial total fish. ¹⁰ Wet weight basis.



Figure 1. Cont.



Figure 1. Variation of weight gain (WG) and total cholesterol (TCHO): (**A**,**C**) Da-Hae and (**B**,**D**) Global. Values are mean of triplicate groups and presented as mean \pm SD. Values having different superscript letters (^{a,b}) are significantly different (*p* < 0.05).

Table 2. Formulation and proximate chemical compositions of experimental diets: extruded pellets (E.P) and moist pellets (M.P) ^{1, 2}.

	БР 3	M.P					
	L.I	Da-Hae ⁴	Global ⁵				
Ingredients (%)							
Animal protein ⁶	> 52						
Plant protein ⁷	< 33						
Grains ⁸	< 10						
Additives ⁹	> 5						
Raw fish		> 80	> 80				
Additives ¹⁰		> 20	> 20				
Proximate analysis (%)							
Dry matter	90.7 ^a	30.1 ^b	29.2 ^b				
Crude protein	59.9	61.2	63.8				
Crude lipid	11.2 ^b	22.0 ^a	22.8 ^a				
Ash	12.8	13.8	12.6				

¹ Values are the mean of triplicate groups and presented as mean \pm SD. Values in the same row having different superscript letters (^{a,b}) are significantly different (p < 0.05). ² Dry matter basis. ³ Commercial feed containing 59.9% protein and 11.2% lipid, Republic of Korea. ⁴ Raw fish feed (composed of mackerel and herring) containing 61.2% protein and 22.0% lipid. ⁵ Raw fish feed (composed of mixed fish such as cutlass, jack mackerel, croaker, mackerel, and longsnout pricklebacks) containing 63.8% protein and 22.8% lipid. ⁶ Fishmeal, shrimp-, squid-, and meat-powder. ⁷ Soybean, gluten, and yeast. ⁸ Wheat flour. ⁹ Vitamin, mineral, calcium phosphate lecithin, and choline chloride. ¹⁰ Fishmeal, wheat flour, vitamin, mineral, etc. (farm-specific).

2.2. Mortalities at Farms

Pathogens (pathogenic microorganisms) causing mortality were monitored by the aquatic-animal disease center in charge of each farm. All of the fish deaths at both farms were caused by disease (Table 3). Similar mortal diseases at flounder farms, among the approximately 100 diseases found in Korean aquatic life since the 1980s [37,38], have been reported, most commonly as single instances or combinations of four bacterial diseases (*Vibrio* sp., *Edwardsiella* sp., *Streptococcus* sp., *Flexibacter* sp.) and two parasitic diseases (*Scuticociliates*, Emaciation disease). *Vibrio* sp., *Edwardsiella* sp., *Streptococcus* sp., *streptococcus* sp., and *Scuticociliates* arose in both the control and experimental groups at both farms, which were considered to have originated in the influx of natural, untreated seawater that is used at both farms. In particular, mortality caused by *Lymphocystis* disease (Figure 2A) occurred only in the control group, and mortality caused by abdominal inflation (Figure 2B) occurred only in the control group (M.P), possibly via inflow from the raw fish feed or via an infected carrier fish, is in contradiction to previous reports that *Lymphocystis* disease transmission occurs through the skin and gills of fish by

direct contact or by waterborne exposure [39–41]. The present finding on abdominal inflation, that it arose only in the experimental group (E.P), could possibly have been related to the digestibility of the E.P diet, irrespective of earlier reports of abdominal inflation symptoms having been caused by *Streptococcus* sp. or *Lactococcus* sp. [42,43]. Further study, however, will be needed for accurate identification of the causes of disease in both cases (*Lymphocystis* disease and abdominal inflation).

Microorganisms are particularly sensitive to ambient temperature; that is, when the temperature is lower than is proper, catalytic activity is halted, deforming cells and inhibiting microorganism growth, whereas at higher-than-normal temperatures, enzymes, transport proteins, and other proteins are denatured, leading to cell-membrane decomposition [44,45]. Similar to the results of aquaculture disease monitoring conducted by the Korean government [14,42], the incidence rate of disease peaked at high water temperature and then decreased as the water temperature decreased (Figure 2C,D), this phenomenon is considered to have been due to the above-noted effect of temperature on microorganisms. Nevertheless, in January (water temperature < 10 °C), mortality rarely occurred (<10 fish/month) in the control group (M.P), whereas in the experimental group (E.P), it occurred continuously (>100 fish/month) due to the same reason (abdominal inflation symptoms) at both farms (Figure 2C,D), which is, as mentioned earlier, considered to be owed to the correlation between feed and digestibility.

Table 3. Disease status of olive flounder fed with experimental diets (E.P and M.P): "V" (positive) and "-" (negative).

		Aug		Sep		Oct		Nov		Dec		Jan	
		E.P	M.P										
Da-Hae	Scuticociliates	-	-	V	V	V	V	-	-	-	-	-	-
	Vibrio sp.	V	V	V	V	V	V	-	-	-	-	-	-
	Abdominal inflation	V	-	V	-	V	-	V	-	V	-	V	-
	Lymphocystis	-	-	-	V	-	V	-	V	-	V	-	-
	Edwardsiella sp.	V	V	V	V	V	V	-	-	-	-	-	-
	Streptococcus sp.	-	-	V	V	V	V	V	V	-	-	-	-
	Flexibacter sp.	-	-	-	-	V	V	-	-	-	-	-	-
Global	Scuticociliates	-	-	-	-	-	-	-	-	-	-	-	-
	Vibrio sp.	V	V	V	V	-	-	-	-	-	-	-	-
	Abdominal inflation	V	-	V	-	V	-	V	-	V	-	V	-
	Lymphocystis	-	-	-	V	-	V	-	V	-	V	-	-
	Edwardsiella sp.	V	V	V	V	-	-	-	-	-	-	-	-
	Streptococcus sp.	-	-	V	V	V	V	V	V	-	-	-	-
	Flexibacter sp.	-	-	-	-	-	-	-	-	-	-	-	-



Figure 2. Cont.

Figure 2. Morphological disease diagnoses of (**A**) *Lymphocystis* disease (M.P) and (**B**) abdominal inflation (E.P). Mortality rate and water temperature change in (**C**) Da-Hae and (**D**) Global. (**E**) Annual water temperature in Wando (*Korea Hydrographic and Oceanographic Agency*). Values are the mean of triplicate groups and are presented as mean \pm SD. Values having different superscript letters (^{a,b}) are significantly different (p < 0.05).

2.3. Histopathological Examination

The results for hepatic fibrosis, steatosis (fatty change), inflammation, and necrosis or apoptosis of hepatocytes, as compared between flounder weight and feed (600-800 g, M.P and E.P) are shown in Figure 3. In the experimental group (E.P), microfragmented fat cells and balloon-shaped deformed adipocytes were observed weakly around the central vein, but showed a similar pattern to that of normal liver tissue (Figure 3A,B). On the other hand, in the control group (M.P), the nucleus was biased to one side, the cell body was enlarged, and transparent cytoplasmic vacuoles were clearly identified as adipocytes were lysed during Hematoxylin & Eosin staining (Figure 3C,D). Neither portal inflammation, mononuclear cell collection, nor apoptosis was observed. Steatosis (fatty change or hepatocytic vacuolization), caused by abnormal accumulation of lipids in cells due to defects in the synthesis and removal of fat, was observed. Owing to the increase of adipocytes, normal hepatic parenchymal cells on the periphery of the hepatic central vein tended to be very few, whereas there appeared to be many round vacuoles in the form of lipids accumulated in the cytoplasm of hepatocytes, and tissues were contracted as dead hepatocytes had been removed by phagocytosis. In summation, the morphological differences of the flounder hepatocytes in the present study were considered to be due to the effects of the lipid content in feed, since the correlation between fat and hepatocytes is already known [46-48].

Figure 3. Optical-microscopy analysis of hepatocytes in liver tissue from olive flounder. Images representative of the sections of the steatosis group: (**A**,**B**) mixture of fatty change and cytoplasmic glycogen; (**C**,**D**) mild fatty change group. Hematoxylin & Eosin staining, scale bars indicate 50 μ m (×400).

2.4. Flounder Aquaculture in Korea

Most flounder in Korea are produced on Jeju Island (22,171 metric ton, 59.5%) and Wando (11,887 metric ton, 30.2%). The proper water temperature for flounder growth is 14–23 °C [49–51]; the average annual seawater temperature on Jeju Island is 20 °C (9–27.4 °C), and at Wando it is 15.6 °C (7–25 °C, Figure 2E; water temperature varies in different areas of Wando) [3]. Jeju Island, where there is underground seawater (annual average water temperature 18 °C), can produce flounder all year round due to maintenance of water temperature by mixing with natural seawater depending on the season; Wando, though, is allowed approximately six months (May–July, September–November) of farming a year. Therefore, flounder farming to sale size (1–2 kg) takes approximately 15 months in Wando. In the meantime, Wando must pass through one winter (< 9 °C, feeding 2–3 times a week to maintain growth/life, most farms do not carry out normal operation) and two summers (>24 °C), during which time most cases of mortality occur. As for mortality in the present experiment, the numbers were 1047 at Da-Hae farm and 1167 at Global farm, with more fish dying in the E.P tanks than in the M.P tanks. By multiplying the number of deaths by selling prices, the economic losses were \$9650 and \$10,756, respectively.

In most flounder aquafarms using moist pellets (M.P) in Korea, it is common to use extruded pellets (E.P) for the fry period, and then to convert to moist pellets as fish enter the growing period. Past surveys of aquafarmers, including those at the present two farms using M.P pellets, showed, on the one hand, a distrust of the quality of E.P, especially with regard to its low digestibility after the fry period, and on the other hand, there was consensus on the need for feed conversion to E.P from M.P in the sense that the use of E.P would consequentially protect the marine environment in terms of prevention of the catching of small fish, fishery resources protection, and reduction of pollutants in effluents. Flounder producers in Korea have, as mentioned in the introduction, preferred to use

M.P owing to its association with faster growth and lower mortality relative to E.P, despite their full awareness of the benefits of E.P, including nutritional value, easy feed storage/handing and feeding control, reduction of feeding time and workload, stable supply, and others. Therefore, development of flounder-exclusive formulated feed suitable for each growth stage and season as well as dissemination of more advanced feed manuals for improved digestion and absorption rate are considered to be urgent priorities. In the case of disease outbreak, there was a strong perception that it was not due to feed but rather to environmental impacts, especially water quality, which was supported by the fact that high mortality was observed in farms using untreated seawater, while low mortality was found in farms using treated water [14] (Figure 4). Therefore, a water-quality-improvement management plan should also be established urgently.

Figure 4. Electrolysis of natural seawater: (**A**) before and (**B**) after at Bi-bong Co., Ltd. (33°18′28.9″ N 126°49′35.5″ E, Jeju island).

3. Materials and Methods

The analyses were performed in triplicate (at the minimum). All of the experimental procedures involving fish were performed in accordance with the guidelines approved by the Institutional Animal Care and Use Committee, Kunsan National University.

3.1. Fish and Rearing Conditions

The experiment was carried out at two local flounder farms in Wando, Republic of Korea, for six months (August, 2019 to January, 2020). The flounder were cultured in the traditional way at each farm. The same commercial diet exclusively for flounder containing 59.9% protein and 11.2% lipid (Republic of Korea) and raw fish feed containing 61.2% (Da-Hae) or 63.8% (Global) protein and 22.0% (Da-Hae) or 22.8% (Global) lipid, were supplied for six months at the two farms, but the sources of raw fish feed between the two farms (Da-Hae and Global) were different (Table 2). The dissolved oxygen (DO) level within the tanks was maintained at an average of 8 by utilizing liquid oxygen, and 100% natural seawater (without any water treatment) was used during the experiment. The water temperature in all of the tanks was maintained at the natural seawater temperature for the given month (from 24.9 °C in August to 10.1 °C in January).

3.1.1. Da-Hae Flounder Farm (34°17'31.9" N 126°45'21.7" E, Wando, Jeonnam Province)

Growing flounder (*Paralichthys olivaceus*) averaging 78 g (mean \pm SD) were randomly distributed in triplicate for two treatments (6000 fish per 100 metric ton at 100 m² tank/total 18,000 fish, Figure 5A). Commercial feed (E.P) was used for the experimental group, while raw fish feed (M.P, composed

mainly of mackerel and herring) containing 61.2% protein and 22.0% lipid was supplied to the control group (Table 2). The fish were fed 2 times daily (9:00, 17:00) to apparent satiation at a rate of 0.82–2.14% of wet body weight per day for the 6-month duration of the experiment.

Figure 5. Experiment farms: (A) Da-Hae and (B) Global.

3.1.2. Global Flounder Farm (34°21′54.5″ N 126°53′21.5″ E, Wando, Jeonnam Province)

Growing flounder (*Paralichthys olivaceus*) fish averaging 123.8 g (mean \pm SD) were randomly distributed in triplicate to two treatments (5000 fish per 100 metric ton at 100 m² tank/total 15,000 fish, Figure 5B). Commercial feed (E.P) was used for the experimental group, while raw fish feed (M.P, composed mainly of mixed fish such as cutlass, jack mackerel, croaker, mackerel, and longsnout pricklebacks) containing 63.8% protein and 22.8% lipid was supplied for the control group (Table 2). The fish were fed 2 times daily (9:00, 17:00) to apparent satiation at a rate of 0.49–2.34% of wet body weight per day for the 6-month duration of the experiment.

3.2. Growth Performance and Analysis Methods

A total of 500 fish were randomly selected monthly from each tank and weighed to determine the weight gain (WG), feed efficiency (FE), specific growth rate (SGR), and survival rate. At the end of the feeding trial, a proximate composition analysis of three fish randomly selected monthly from each tank and diets (E.P (monthly); MP (every 10 days)) at two farms was performed using the standard AOAC (Association of Official Analytical Chemists) methods [52]. Preparatorily, samples were freeze-dried for 48 h. The moisture contents were determined using a dry oven at 105 °C, and the ash contents by means of combustion at 550 °C. The crude protein was analyzed by the Kjedahl method, and the crude lipid was analyzed by soxhlet extraction using the soxhlet system 1046 (Tecator AB, Hoganas, Sweden) [53]. The activities of total cholesterol (TCHO-P) in serum (blood) were measured using a chemical analyzer (FujiDRI-CHEM 3500i, Fuji Photo Film, Ltd., Tokyo, Japan). The samples were allowed to clot at room temperature for 30 min. Then, the serum was separated by 5000× g centrifugation (10 min).

3.3. Histopathological Examination of Liver

The fish were dissected to observe the overall external morphology (e.g., lesion appearance) and the health of the internal organs, and histological observation was performed by general staining after general tissue processing in order to identify the morphology of the cells and their changes. After anesthetization of the fish by 20 ppm of 2-phenoxyethanol (Sigma, St. Louis, Missouri, MO, USA) followed by dissection and tissue ablation, the liver was immersed in 10% neutral buffered formalin solution (Sigma, St. Louis, Missouri, MO, USA) for 24 h. In addition, the paraffin was infiltrated, according to a conventional method, to produce a paraffin block. Each block was cut to a thickness of 5 μ m with a microtom (Leica, Wetzlar, Germany), and then the tissue sections were attached to slide glass coated with 3-aminopropyl triethoxysilane (Sigma, St. Louis, Missouri, MO, USA). The attached tissue sections were deparaffinized and hydrolyzed using xylene and ethanol in staged concentrations, followed by Hematoxylin & Eosin staining. After staining, the tissue sections were dehydrated

and cleared and then sealed using Synthetic Mountant (Shandon, Runcorn, UK). The stained tissue sections were observed under optical microscopy and photographed using a digital camera (Olympus, Tokyo, Japan).

3.4. Statistical Analysis

All of the data were analyzed by one-way ANOVA (SPSS Inc., Chicago, IL, USA) to test for the effects of the treatments. When a significant treatment effect was observed, a Tukey HSD test was applied for comparison of the means. The treatment effects were considered at the 5% level of significance (p < 0.05).

4. Conclusions

The objective of this study was to conduct field experiments comparing formulated (E.P) and raw-fish (M.P) feeds at two flounder aquaculture farms to obtain the basic data necessary for improving aquafarmers' awareness of feed-quality issues and firmly establish and expand the use of formulated feed. In the results, there were significant differences in growth performance and mortality rate/number between the two groups. The M.P group showed higher growth and lower mortality relative to the E.P group. All cases of mortality were caused by six kinds of disease, four of which (*Vibrio* sp., *Edwardsiella* sp., *Streptococcus* sp., and *Scuticociliates*) were found to be common to the two groups; *Lymphocystis*, meanwhile, occurred only with M.P group and abdominal inflation only with E.P at both farms. With total deaths of 1047 (Da-Hae) and 1,167 (Global), more fish died in the E.P tanks than M.P tanks. By multiplying the number of deaths by selling price, the economic losses were \$9,650 and \$10,756, respectively. Therefore, it should be considered an urgent priority to develop flounder-exclusive formulated feed (E.P) for improved digestion and absorption rate and also to establish a water-quality-improvement management plan.

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