

Supplementary Materials

Phenazine-Based Compound as a Universal Water-Soluble Anolyte Material for the Redox Flow Batteries

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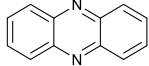
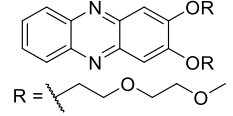
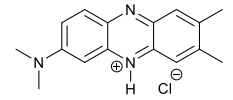
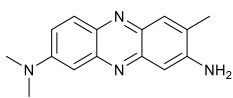
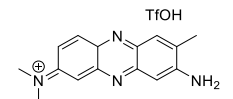
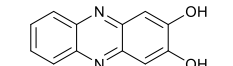
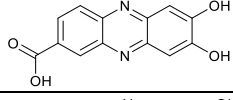
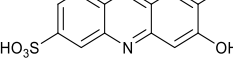
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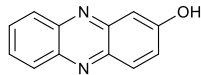
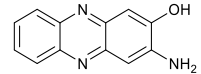
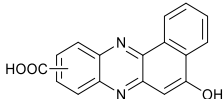
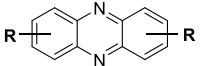
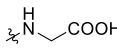
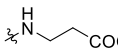
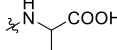
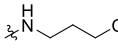
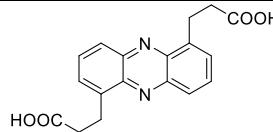
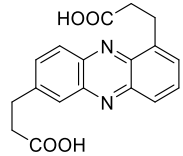
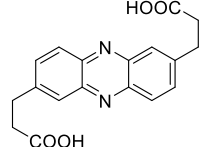
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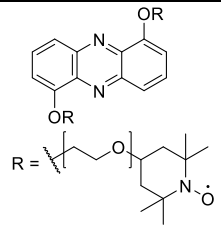
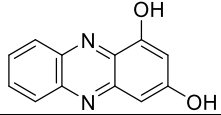
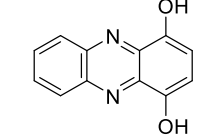
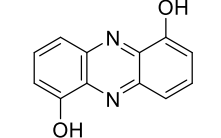
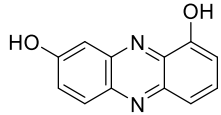
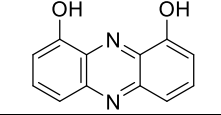
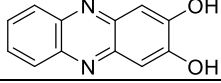
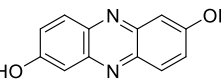
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1. Literature review

Table S1. Summary of the investigated phenazine-based compounds, their solubilities, and literature references (SE = supporting electrolyte).

| Compound | Code | Tested in | Solubility | Redox potential vs. Ag/AgCl (for water-based systems) | Best redox flow battery test | Reference |
|---|------------|-----------|---|---|---|-----------|
|  | - | MeCN | 6.8×10 ⁻⁵ M in pure water | -1.5 V vs. Ag/Ag ⁺ | - | [S1,S2] |
|  | M1 | MeCN | 2.2 M in MeCN 1.0 M in 5.0 M TBABF ₄ in MeCN | -1.72 V vs. Ag/Ag ⁺ | Anolyte – M1 Catholyte – triarylamine derivative Concentration - 0.01 M (50 cycles) SE – 0.1 M TBABF ₄ | [S3] |
|  | BR-5 | water | 0.1 M in 1.0 M H ₂ SO ₄ 0.054 M in 4.0 M methanesulfonic acid | 0.0 V | Anolyte - d-BR-5 Catholyte - Ce(CH ₃ SO ₃) ₃ Concentration - 0.05 M (200 cycles) SE - 4.0 M methanesulfonic acid | [S4,S5] |
|  | d-BR-5 | | 0.34 M in 1.0 M H ₂ SO ₄ 0.206 M in 4.0 M methanesulfonic acid | -0.013 V | | |
|  | BR-5- TfOH | | 0.11 M in 1.0 M H ₂ SO ₄ 0.175 M in 4.0 M methanesulfonic acid | -0.012 V | | |
|  | DHP | | 0.1 M in 1.0 M NaOH | -1.02 V | | |
|  | DHPC | | 0.95 M in 1.0 M NaOH | -1.06 V | | |
|  | DHPS | | 1.45 M in 1.0 M NaOH | -1.08 V | Anolyte – DHPS Catholyte – K ₄ Fe(CN) ₆ Concentration - 0.1 M (200 cycles) Concentration – 1.4 M (500 cycles) SE - 1.0 M NaOH | [S1] |

| | | | | | | |
|---|--|-------|---------------------------------------|--|--|------|
|  | HP | water | 1.70 M in 1.0 M KOH | -0.88 V | Anolyte – BHPC Catholyte – K ₄ Fe(CN) ₆ Concentration - 0.5 M (1305 cycles) SE - 1.0 M KOH | [S6] |
|  | AHP | | 0.43 M in 1.0 M KOH | -0.99 V | | |
|  | BHPC | | 1.55 M in 1.0 M KOH | -0.99 V | | |
|  <p>R selected from:</p>     | 2,7-DGAP 2,7-DAAP 2,7-DPAP 2,7-DBAP 1,8-DGAP 1,6-DGAP 1,6-DAAP 1,6-DPAP 1,6-DBAP | water | From 1.047 M to 0.401 M in pure water | 2,7-DGAP - 0.73 V 2,7-DAAP - 0.75 V 2,7-DPAP -0.73 V 2,7-DBAP -0.74 V 1,8-DGAP -0.78 V 1,6-DGAP -0.73 V 1,6-DAAP -0.72 V 1,6-DPAP -0.77 V 1,6-DBAP -0.74 V | Anolyte – 1,6-DPAP Catholyte – K ₄ Fe(CN) ₆ Concentration - 0.1 M (192 cycles) SE - 1.0 M KCl pH=12 Concentration – 0.5 M (70 cycles) SE - 1.0 M KCl pH=8 | [S7] |
|  | 1,6-PFP | water | 1.01 M in 1.0 M KOH | -0.77 V | Anolyte – 1,8-PFP Catholyte – K ₄ Fe(CN) ₆ Concentration - 0.1 M (340 cycles) SE - 1.0 M KOH | [S8] |
| | | | 1.02 M in pure water | | | |
|  | 1,8-PFP | | 1.46 M in 1.0 M KOH | -0.78 V | | |
| | | | 1.09 M in pure water | | | |
|  | 2,7-PFP | | 0.84 M in 1.0 M KOH | -0.82 V | | |
| | | | 0.79 M in pure water | | | |

| | | | | | | |
|---|---------|-----------------|--------------------------------|---------|--|-------|
|  | - | water + diglyme | - | -0.62 V | Anolyte – compound itself Catholyte – compound itself Concentration - 0.01 M (1800 cycles) SE – 0.5 M NaCl | [S9] |
|  | 1,3-DHP | water | 0.3 M 1M NaOH 1.6 M 2M KOH | -0.81 V | Anolyte – 1,4-DHP Catholyte – K ₄ Fe(CN) ₆ Concentration - 0.42 M (1000 cycles) SE - 2.0 M KOH Anolyte – 1,6-DHP Catholyte – K ₄ Fe(CN) ₆ Concentration - 0.36 M (1000 cycles) SE - 2.0 M KOH | [S10] |
|  | 1,4-DHP | | 0.3 M 1M NaOH 0.5 M 2M KOH | -0.72 V | | |
|  | 1,6-DHP | | 0.07 M 1M NaOH 0.6 M 2M KOH | -0.91 V | | |
|  | 1,8-DHP | | 1.8 M 1M NaOH 1.9 M 2M KOH | -1.02 V | | |
|  | 1,9-DHP | | 0.1 M 1M NaOH 0.3 M 2M KOH | -0.81 V | | |
|  | 2,3-DHP | | 1.6 M 1M NaOH 2.0 M 2M KOH | -1.06 V | | |
|  | 2,7-DHP | | 0.1 M 1M NaOH 0.2 M 2M KOH | -0.96 V | | |

2. Experimental methods

2.1. NMR characterization

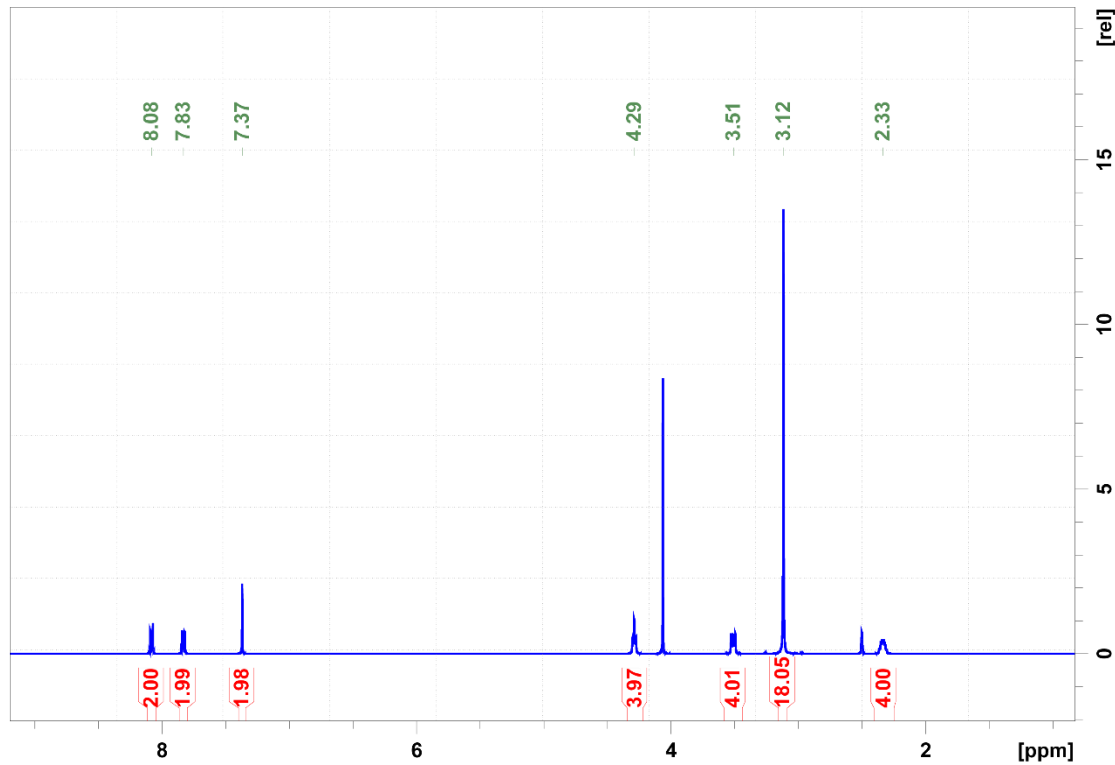


Figure S1. Full ^1H NMR spectrum of **M1** compound.

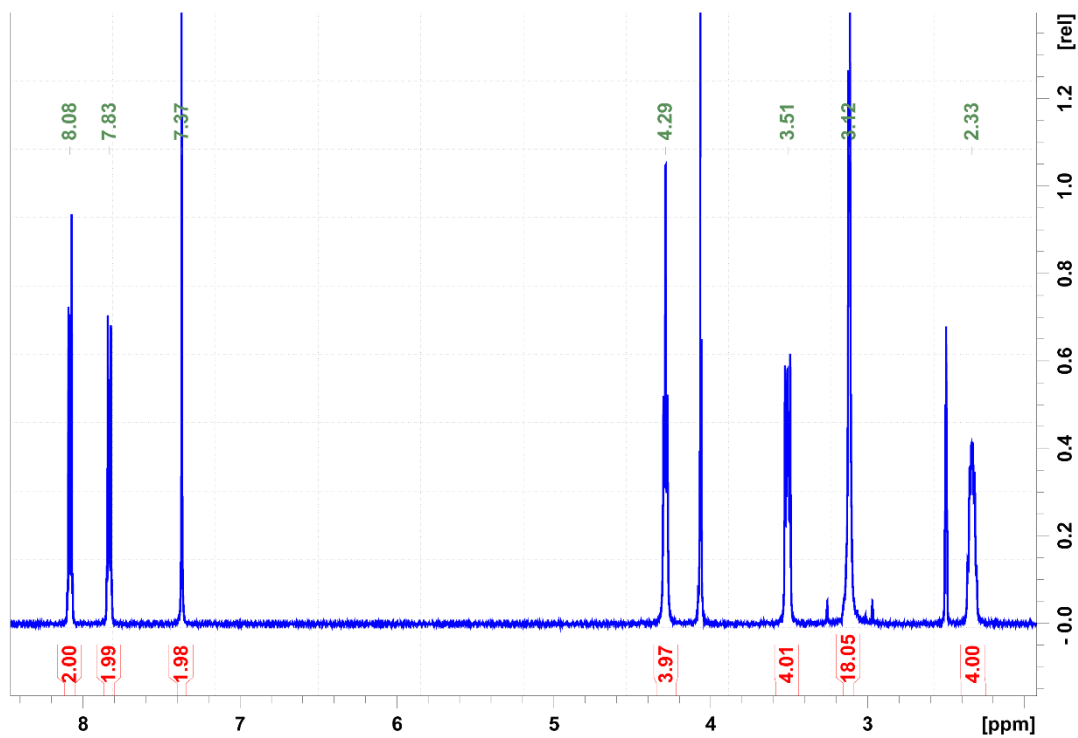


Figure S2. Zoomed ^1H NMR spectrum of **M1** compound.

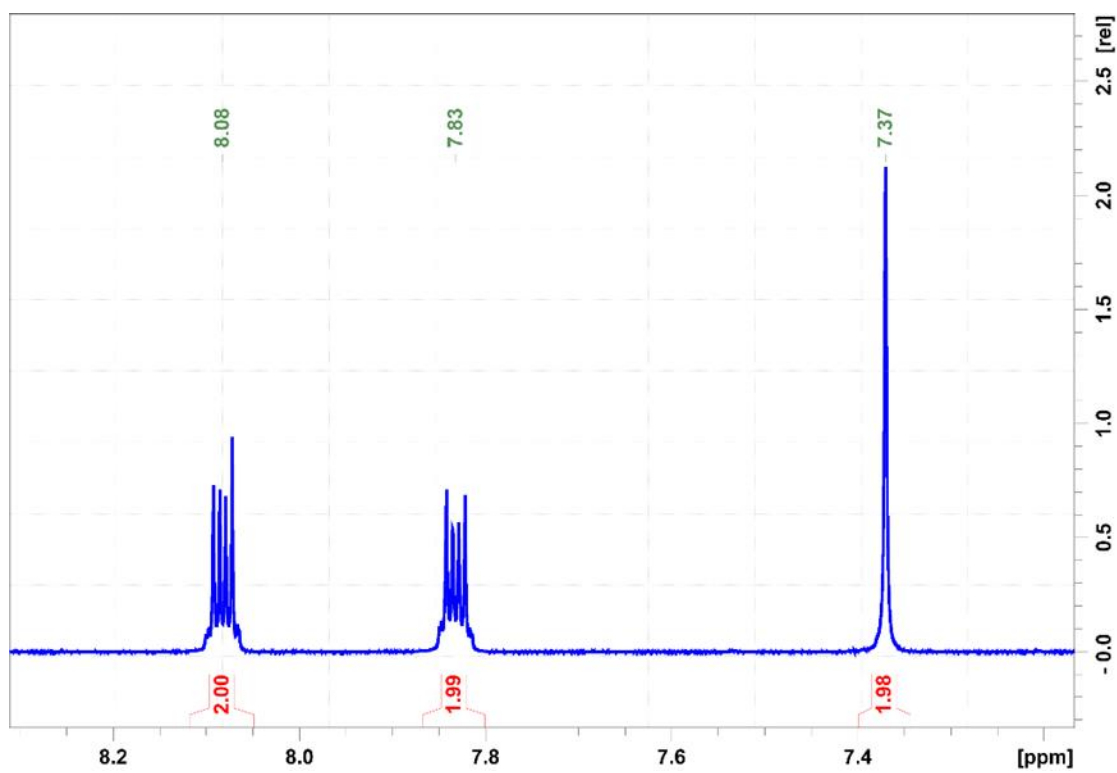


Figure S3. Aromatic region of ^1H NMR spectrum of M1 compound.

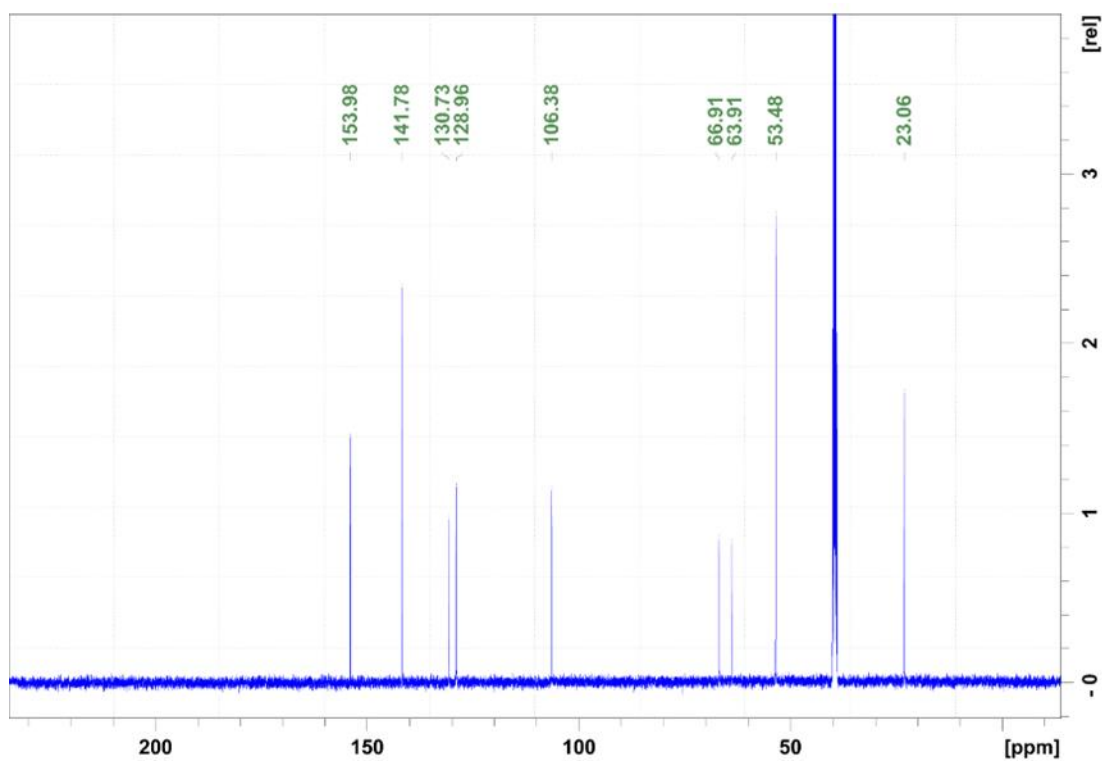


Figure S4. ^{13}C NMR spectrum of M1 compound.

2.2. Melting point determination

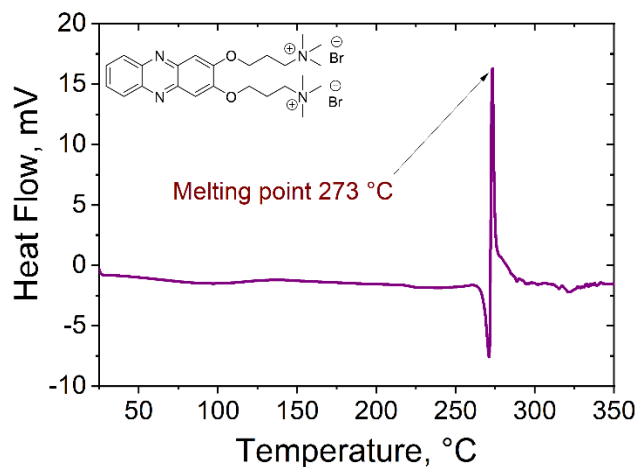


Figure S5. Differential scanning calorimetry graph for **M1** compound.

2.3. H-cell tests

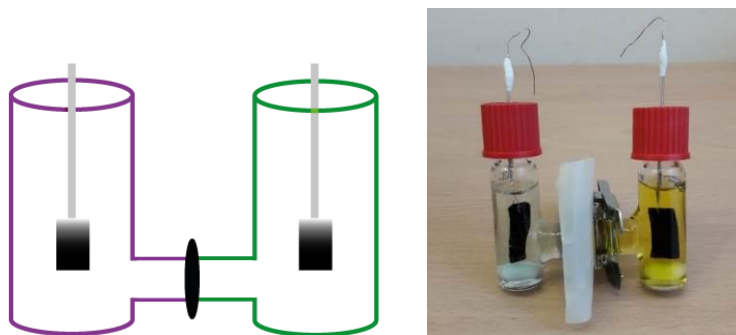


Figure S6. Schematic representation and photo of the h-cell used.

2.4. Redox flow battery tests



Figure S7. Schematic representation of the redox flow battery and the photos of a laboratory setup. The redox flow cell consisted of metal cell frames, Teflon cell frames, graphite/Teflon (20/80) current collectors, Teflon flow frames with graphite felt electrodes (6 mm thick graphite felt, geometric active area of 4.00 cm²), and Neosepta AHA anion-exchange membrane.

3. CVs of the background electrolytes

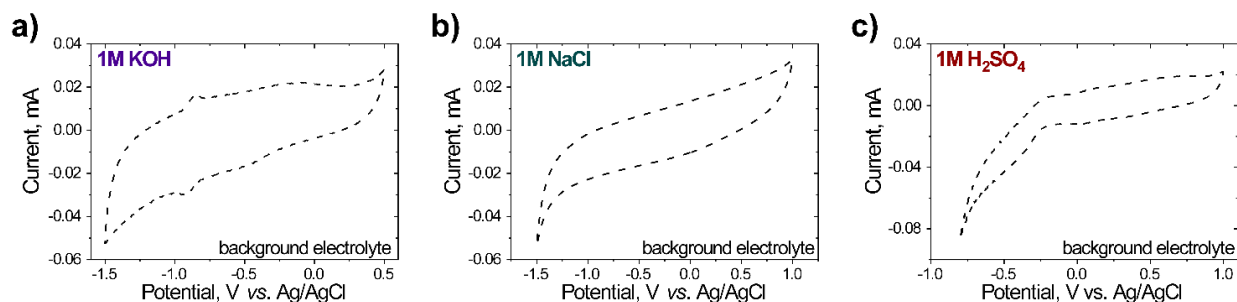


Figure S8. Cyclic voltammogram of the background electrolytes: (a) 1.0 M KOH, (b) 1.0 M NaCl and (c) 1.0 M H₂SO₄.

4. CV curves of M1 in different supporting electrolytes (long potential range)

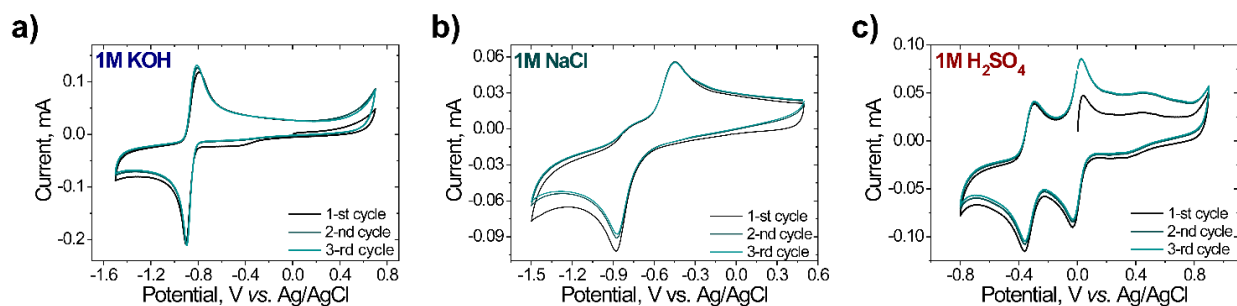


Figure S9. Cycling voltammograms were obtained for 30.0 mM aqueous solution of M1 with (a) 1.0 M KOH, (b) 1.0 M NaCl and (c) 1.0 M H₂SO₄ as supporting electrolytes at a scan rate of 100 mV s⁻¹.

5. Pourbaix diagrams

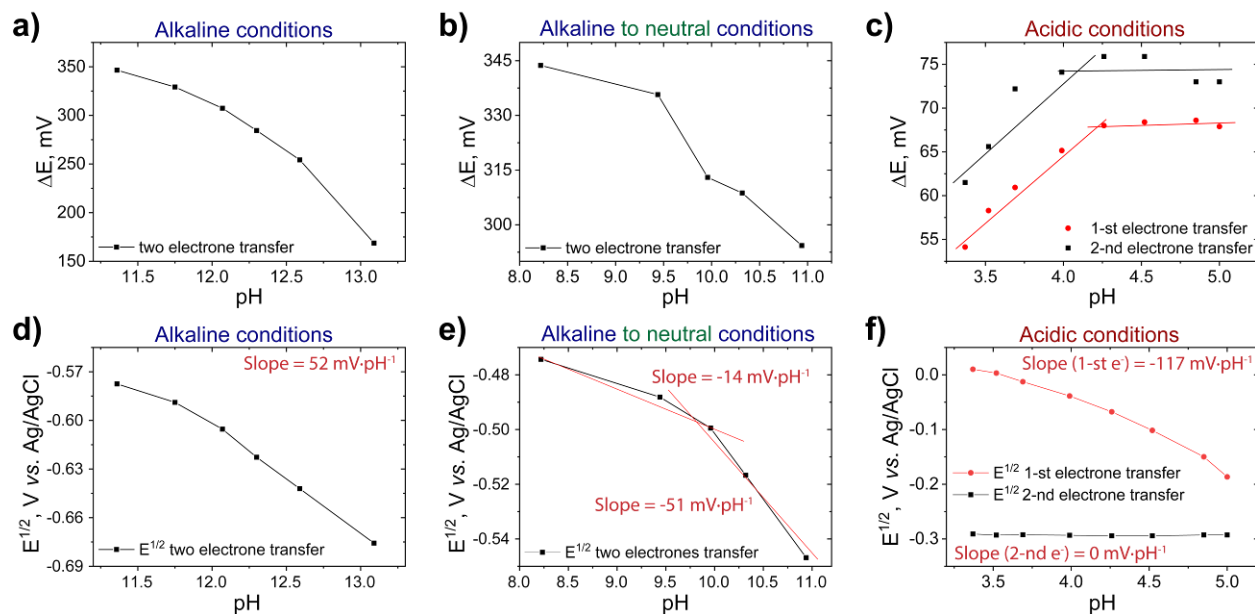


Figure S10. Dependence of peak splitting (ΔE) of the pH for **M1** in alkaline (**f**), alkaline to neutral (**b**) and acidic (**c**) conditions. Dependence of half-wave potential ($E^{1/2}$) of the pH for the alkaline (**d**), alkaline to neutral (**e**) and acidic (**f**) conditions.

6. Pourbaix diagrams: reference electrodes calibration

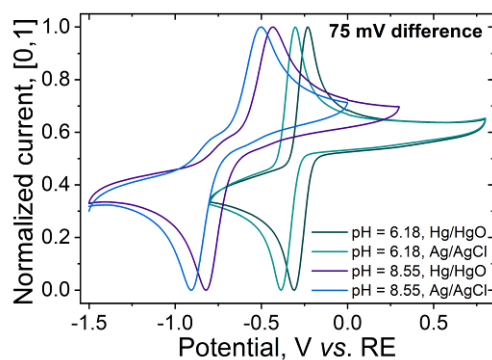
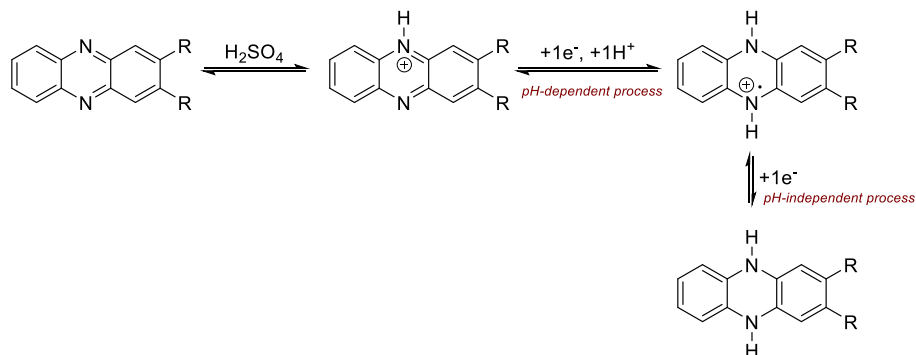


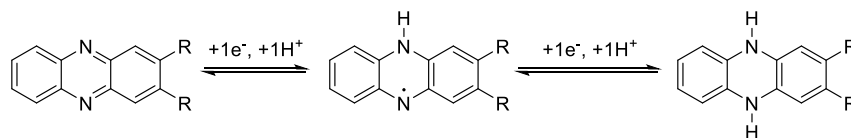
Figure S11. Calibration of Hg/HgO electrode vs. Ag/AgCl electrode. CV curves are measured for the **M1** compound at 100 mV s⁻¹ scan rate at different pH values.

7. Pourbaix diagrams: proposed electrochemical mechanism and DFT calculations

Acidic conditions



Neutral conditions



Basic conditions

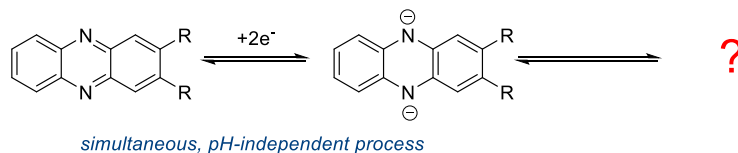


Figure S6. Proposed mechanism and thermodynamically favored protonation pathway for **M1** at acidic, neutral, and alkaline conditions.

DFT calculations - Neutral conditions

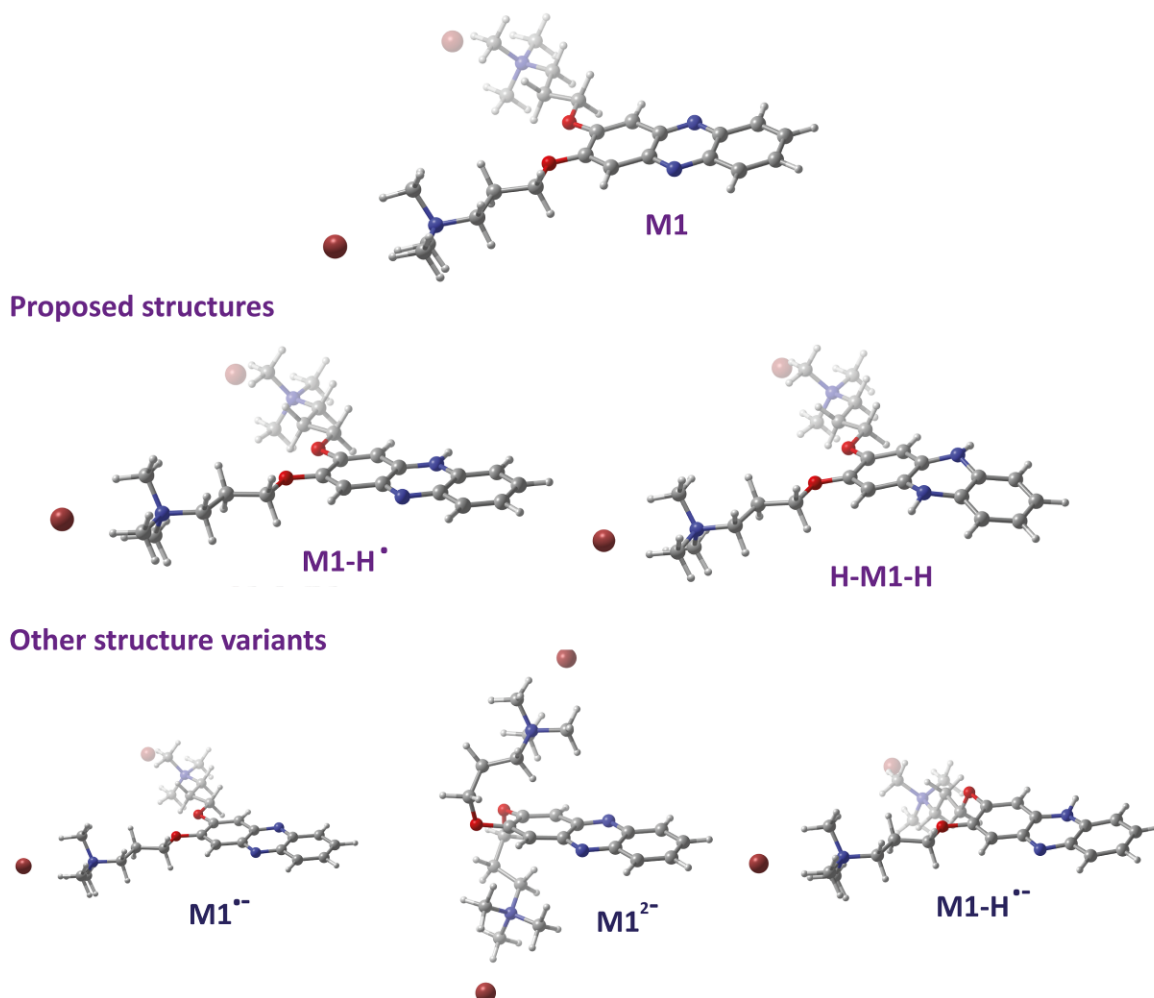


Figure S13. Optimised structures for **M1** molecule, reduced **M1-H[•]** molecule (+ 1e⁻ + 1H⁺), reduced **H-M1-H** molecule (+ 2e⁻ + 2H⁺) and other theoretically possible structure variants.

Table S2. Calculated and measured redox potentials for the **M1** reduction in neutral conditions.

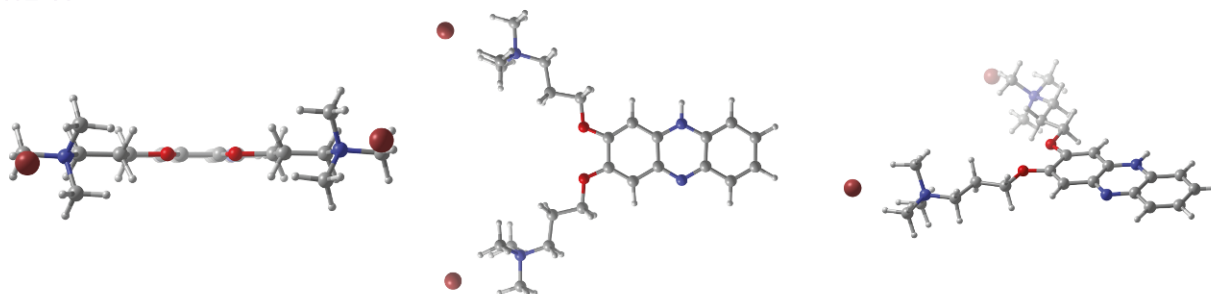
| Redox process | $\text{M1} + \text{e}^- + \text{H}^+ \rightarrow \text{M1-H}^\bullet$ | $\text{M1-H}^\bullet + \text{e}^- + \text{H}^+ \rightarrow \text{H-M1-H}$ |
|---|---|---|
| Calculated redox potential (V) vs. NHE | -0.446 V | -0.065 V |
| Measured redox potential (V) vs. NHE 1M NaCl Supporting electrolyte | -0.460 V | |

Table S3. Calculated redox potentials for other possible redox reaction for the **M1** reduction in neutral conditions.

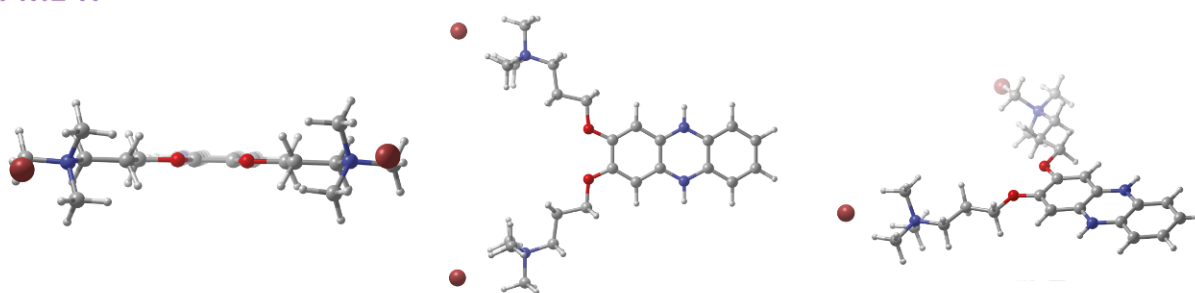
| Redox process | $\text{M1} + \text{e}^- \rightarrow \text{M1}^{\bullet-}$ | $\text{M1} + 2\text{e}^- \rightarrow \text{M1}^{2\bullet-}$ | $\text{M1} + 2\text{e}^- + \text{H}^+ \rightarrow \text{M1-H}^\bullet$ |
|--------------------------------------|---|---|--|
| Computed Reduction Potential vs. NHE | -2.615 V | -3.942 V | -1.422 V |

DFT calculations - Acidic conditions

M1-H⁺



H-M1-H⁺



H-M1-H

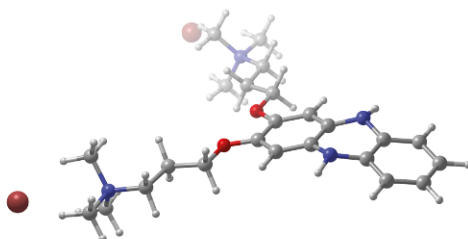


Figure S14. Optimised structures for **M1-H⁺** molecule, reduced **M1-H[•]** molecule (+ 1e⁻ + 1H⁺), and reduced **H-M1-H** molecule (+ 2e⁻ + 2H⁺).

Table S4. Calculated and measured redox potentials for the **M1** reduction in acidic conditions.

| Redox process | M1-H ⁺ + e ⁻ + H ⁺ → H-M1-H [•] | H-M1-H [•] + e ⁻ → H-M1-H |
|--|---|---|
| Computed Reduction Potential vs. NHE | 0.298 V | -0.073 V |
| Measured redox potential vs. NHE 1M H ₂ SO ₄ | 0.26 V | -0.05V |

8. Determination of the diffusion coefficient for M1 in 0.5 M KOH from the RDE measurements

The diffusion coefficient was calculated according to Levich equation for rotation disk electrode measurements. The kinematic viscosity for 0.5 M KOH solution is $1.03 \times 10^{-2} \text{ cm}^2 \text{ s}^{-1}$ [S11].

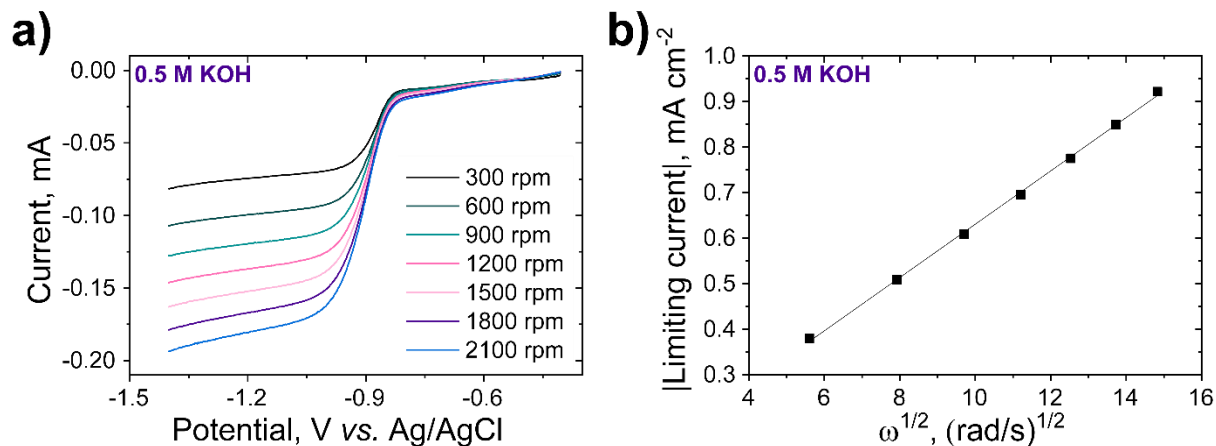


Figure S15. Linear sweep voltammetry test with a glassy carbon rotating disk electrode for **M1** in 0.5 M KOH supporting electrolyte at a scan rate of 10 mV s^{-1} (a), and linear plot of the current as a function of the square root of the rotation rate (ω) (b). Concentration of the redox-active compound was 15.0 mM.

Table S5. Data set used for limiting current - square root of the rotation rate plot and evaluation of diffusion coefficient.

| ω , rpm | ω , rad s^{-1} | Limiting current, mA (at -1.2 V) |
|----------------|--------------------------------|----------------------------------|
| 300 | 31.41 | -0.0745 |
| 600 | 62.83 | -0.0998 |
| 900 | 94.25 | -0.1195 |
| 1200 | 125.66 | -0.1364 |
| 1500 | 157.08 | -0.1521 |
| 1800 | 188.49 | -0.1668 |
| 2100 | 219.91 | -0.1809 |

The slopes were found to be:

| Electron | Slope | Error |
|----------------------|-----------------------|-----------------------|
| 2 electron reduction | 5.85×10^{-2} | 7.68×10^{-4} |

Diffusion coefficient is:

| | 2 e ⁻ reduction |
|-----------|---|
| 0.5 M KOH | $1.94 \times 10^{-6} \pm 2.93 \times 10^{-9}$ |

9. Determination of the diffusion coefficient for M1 in 0.5 M NaCl from the RDE measurements

The diffusion coefficient was calculated according to Levich equation for rotation disk electrode measurements. The kinematic viscosity for 0.5 M NaCl solution is $0.010 \text{ cm}^2 \text{ s}^{-1}$ [S6].

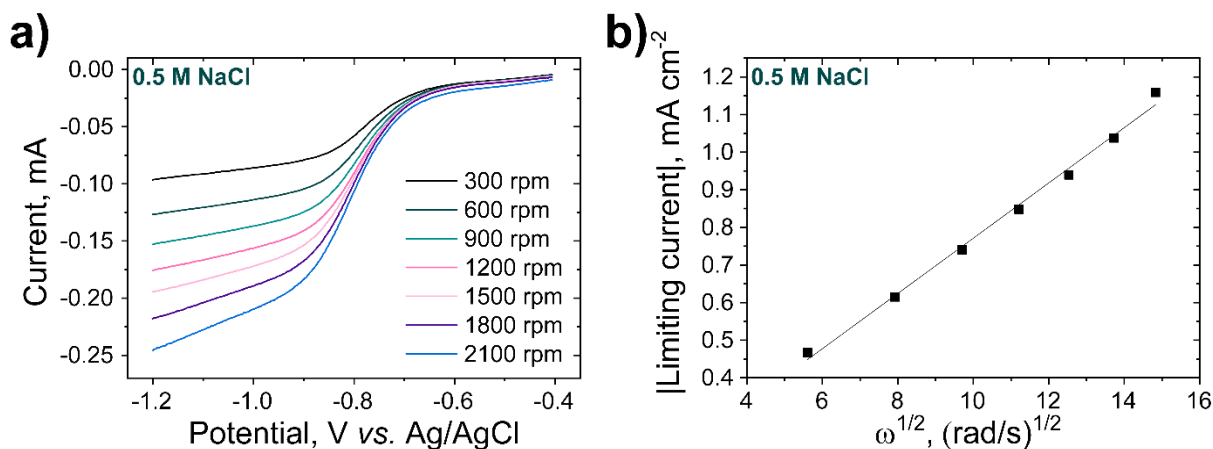


Figure S16. Linear sweep voltammetry test with a glassy carbon rotating disk electrode for **M1** in 0.5 M NaCl supporting electrolyte at a scan rate of 10 mV s^{-1} (a), and linear plot of the current as a function of the square root of the rotation rate (ω) (b). Concentration of the redox-active compound was 15.0 mM.

Table S6. Data set used for limiting current - square root of the rotation rate plot and evaluation of diffusion coefficient.

| ω , rpm | ω , rad s^{-1} | Limiting current, mA (at -1.1 V) |
|----------------|--------------------------------|----------------------------------|
| 300 | 31.41 | -0.0916 |
| 600 | 62.83 | -0.1207 |
| 900 | 94.25 | -0.1453 |
| 1200 | 125.66 | -0.1665 |
| 1500 | 157.08 | -0.1844 |
| 1800 | 188.49 | -0.2037 |
| 2100 | 219.91 | -0.2275 |

The slopes were found to be:

| Electron | Slope | Error |
|----------------------|-----------------------|-----------------------|
| 2 electron reduction | 7.45×10^{-2} | 2.52×10^{-3} |

Diffusion coefficient is:

| | 2 e ⁻ reduction |
|------------|---|
| 0.5 M NaCl | $9.64 \times 10^{-7} \pm 6.13 \times 10^{-9}$ |

10. Determination of the diffusion coefficient for M1 in 0.5 M H₂SO₄ from the RDE measurements

The diffusion coefficient was calculated according to Levich equation for rotation disk electrode measurements. The kinematic viscosity for 0.5 M H₂SO₄ solution is 0.0098 cm² s⁻¹ [S12].

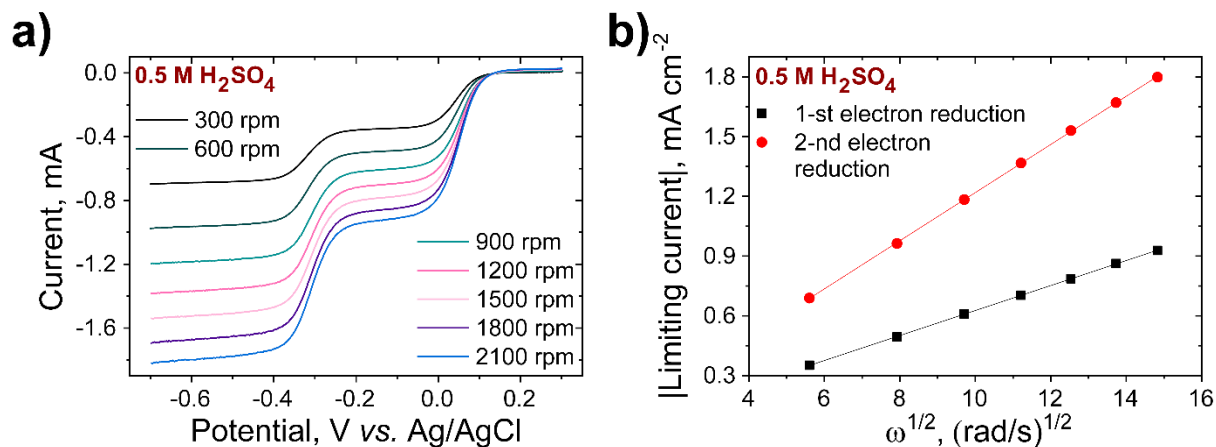


Figure S17. Linear sweep voltammetry test with a glassy carbon rotating disk electrode for **M1** in 0.5 M H₂SO₄ supporting electrolyte at a scan rate of 10 mV s⁻¹ (a), and linear plots of the currents as a function of the square root of the rotation rate (ω) (b). Concentration of the redox-active compound was 15.0 mM.

Table S7. Data set used for limiting current - square root of the rotation rate plot, and evaluation of diffusion coefficient.

| ω , rpm | ω , rad s ⁻¹ | Limiting current for the 1-st electron reduction, mA (at -0.15 V) | Limiting current for the 2-nd electron reduction, mA (at -0.60 V) |
|----------------|--------------------------------|---|---|
| 300 | 31.41 | -0.351 | -0.689 |
| 600 | 62.83 | -0.493 | -0.963 |
| 900 | 94.25 | -0.609 | -1.180 |
| 1200 | 125.66 | -0.703 | -1.370 |
| 1500 | 157.08 | -0.786 | -1.530 |
| 1800 | 188.49 | -0.862 | -1.670 |
| 2100 | 219.91 | -0.927 | -1.800 |

The slopes were found to be:

| Electron | Slope | Error |
|-------------------------|-----------------------|-----------------------|
| 1-st electron reduction | 6.29×10^{-2} | 2.45×10^{-4} |
| 2-nd electron reduction | 1.20×10^{-1} | 4.66×10^{-5} |

Diffusion coefficients are:

| | 1-st e ⁻ reduction | 2-nd e ⁻ reduction |
|--------------------------------------|--|---|
| 0.5 M H ₂ SO ₄ | $2.09 \times 10^{-6} \pm 5.16 \times 10^{-10}$ | $5.66 \times 10^{-6} \pm 1.35 \times 10^{-9}$ |

11. Determination of the standard rate constant for the M1 in 1.0 M KOH by RDE studies and Koutecký-Levich analysis

The standard electrochemical rate constant was measured by use of the Koutecký-Levich method.

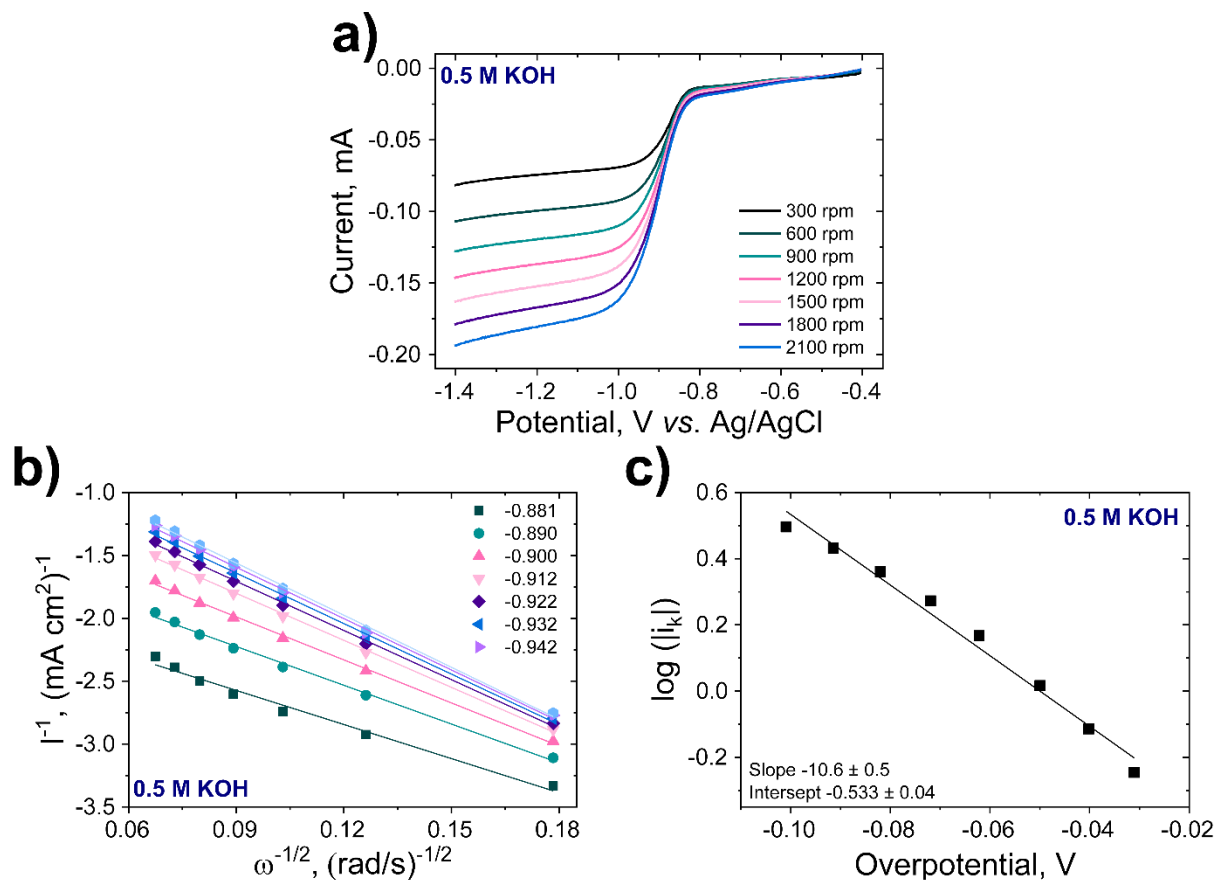


Figure S18. (a) Rotating disk electrode measurement of **M1** (15 mM), 0.5 M NaCl in distilled water, scan rate 10 mV s⁻¹, rotating rates from 300 to 2100 rpm; (b) Koutecký-Levich plot for different overpotentials yielding the mass-transfer-independent current i_k ; (c) Tafel plot yielding $k_0 = 5.16 \times 10^{-4}$ cm s⁻¹ and $\alpha = 0.90$.

12. Determination of the standard rate constant for the M1 in 0.5 M NaCl by RDE studies and Koutecký-Levich analysis

The standard electrochemical rate constant was measured by use of the Koutecký-Levich method.

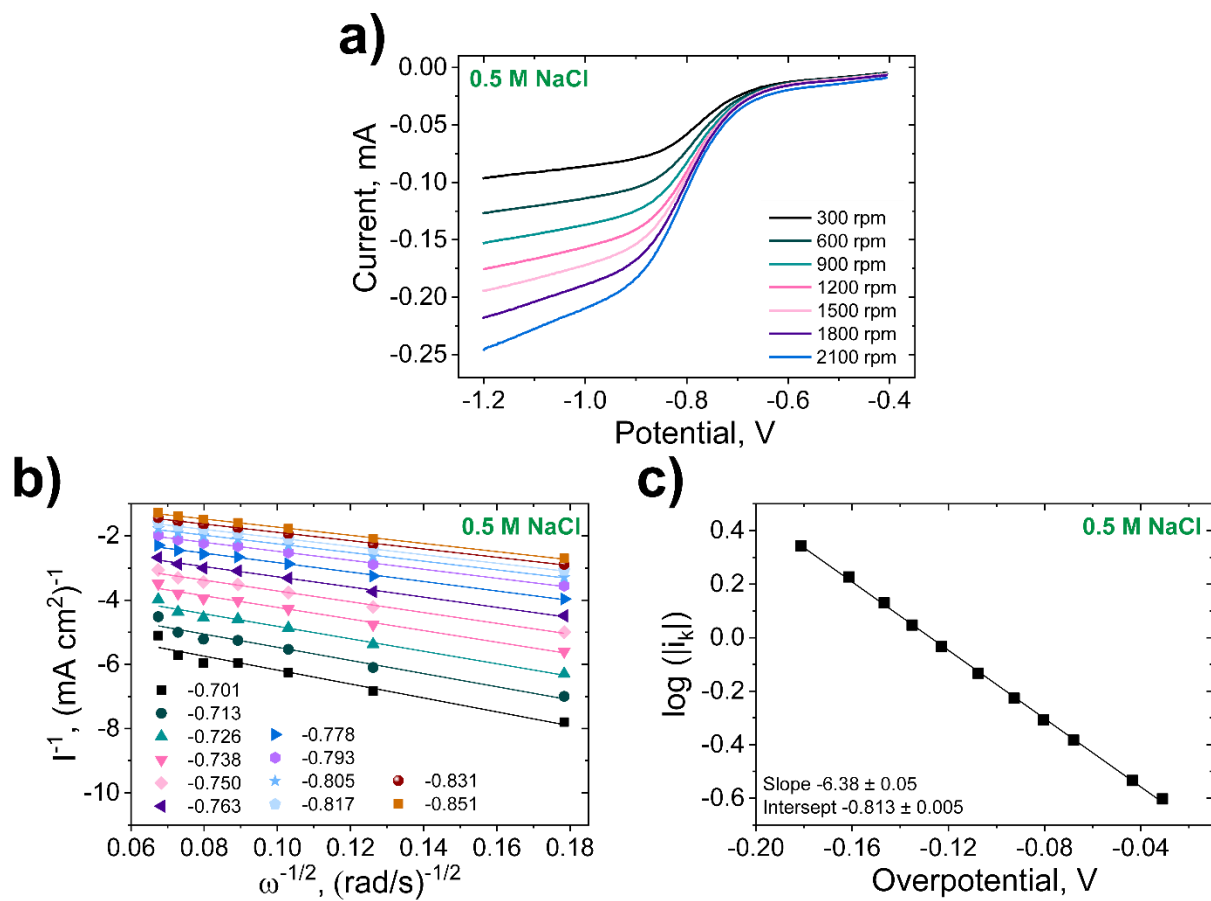


Figure S19. (a) Rotating disk electrode measurement of **M1** (15 mM), 0.5 M NaCl in distilled water, scan rate 10 mV s^{-1} , rotating rates from 300 to 2100 rpm; (b) Koutecký-Levich plot for different overpotentials yielding the mass-transfer-independent current i_k ; (c) Tafel plot yielding $k_0 = 2.59 \times 10^{-4} \text{ cm s}^{-1}$ and $\alpha = 0.94$.

13. Membrane resistance measurements

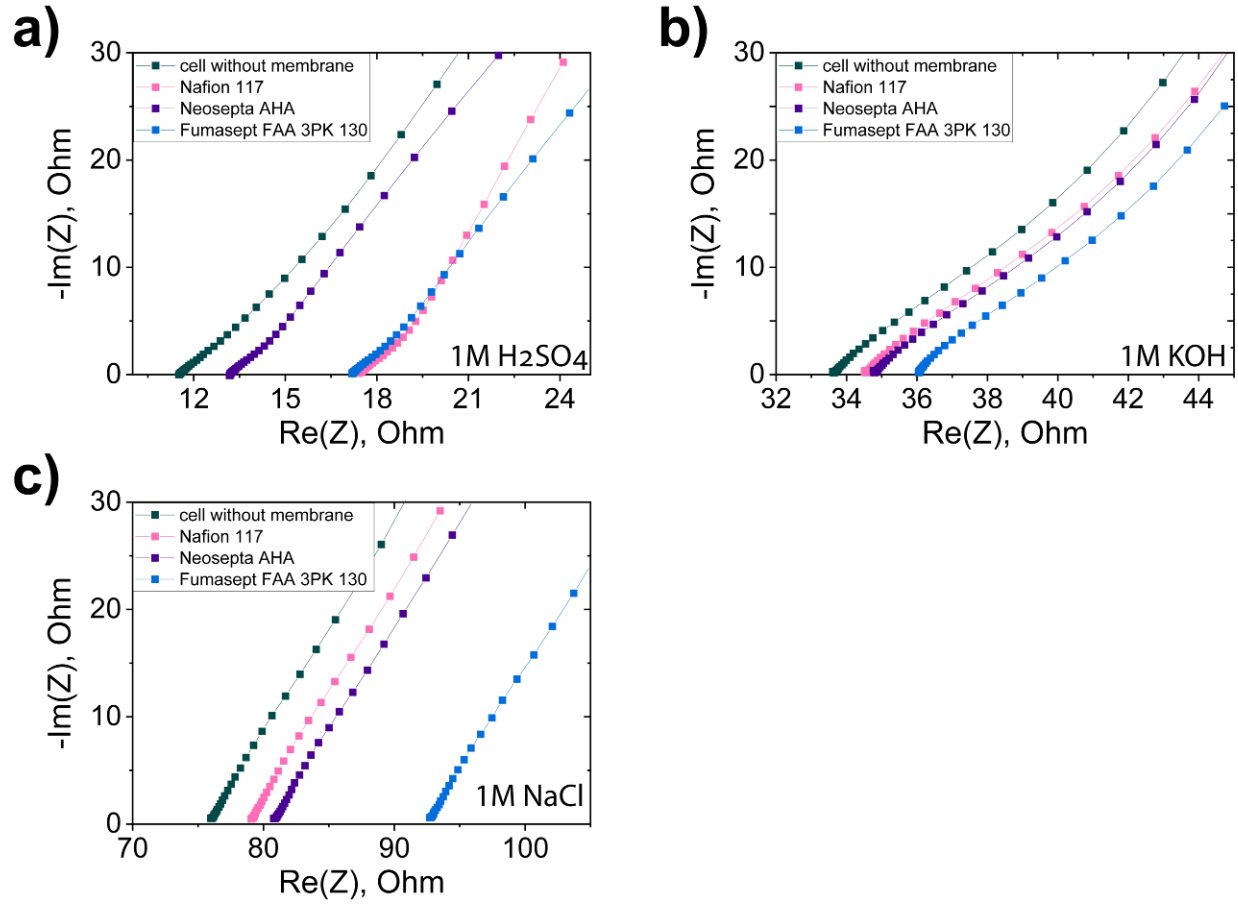


Figure S20. The Nyquist Plots of the non-flow cell with different membranes under with different supporting electrolytes **(a)** 1.0 M H_2SO_4 , **(b)** 1.0 M KOH, **(c)** 1.0 M NaCl.

14. CV of M1 and TEMPOL in neutral conditions

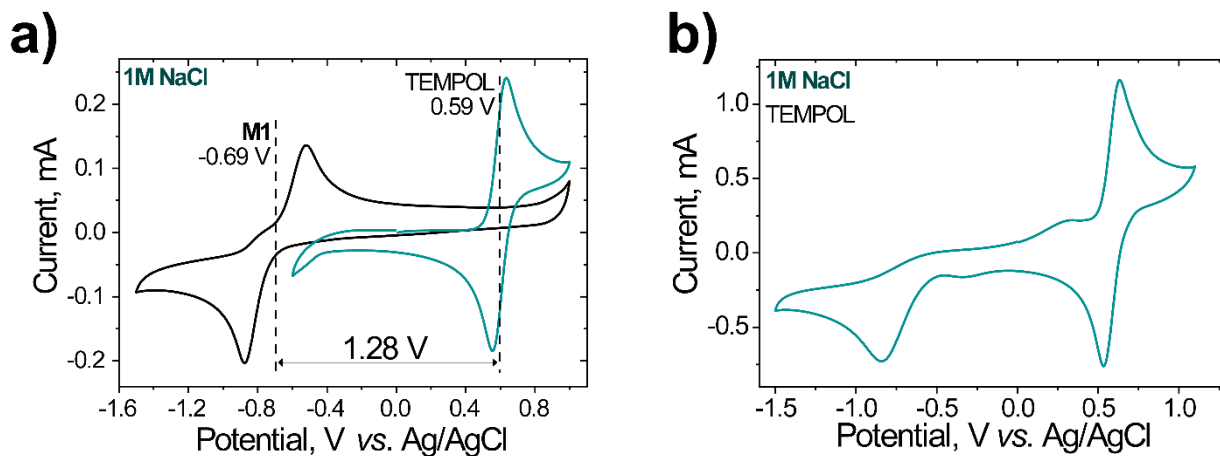


Figure S21. Cyclic voltammograms of pure **M1** (long potential range) and TEMPOL; **(b)** Cyclic voltammograms of pure TEMPOL (long potential range).

15. Investigation of the h-cell with Nafion 117 membrane in neutral conditions

h-cell, Nafion 117 membrane, unmixed electrolytes

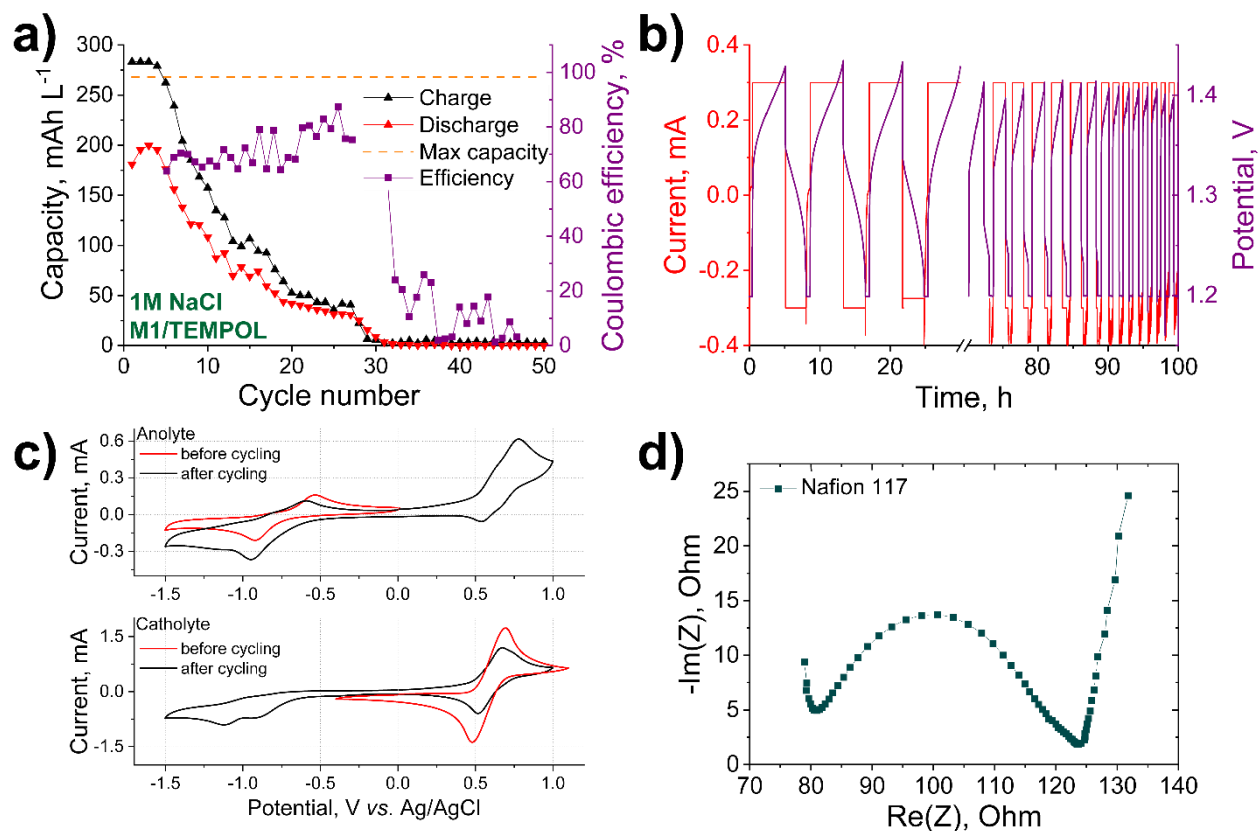


Figure S22. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon charge-discharge cycling of the h-cell based on **M1** anolyte and TEMPOL catholyte in 1M NaCl water solution with Nafion 117 membrane (unmixed electrolytes were used); (b) Cell voltage and cell current versus time profile for first 30 hours and 70-100 hours of cycling; (c) CVs of the anolyte and catholyte before and after cycling; (d) The Nyquist Plot of the redox flow battery after cycling.

Electrolyte's composition: mixed electrolytes, **M1** – 5.0 mM, **TEMPOL** -50.0 mM, NaCl – 1.0 M in deionised water. Battery charging and discharging time was limited to 18 000 seconds, that is 108% of theoretical battery capacity.

16. Investigation of the h-cell with Fumasept FAA-3PK-130 membrane in neutral conditions

h-cell, Fumasept FAA-3PK-130 membrane, unmixed electrolytes

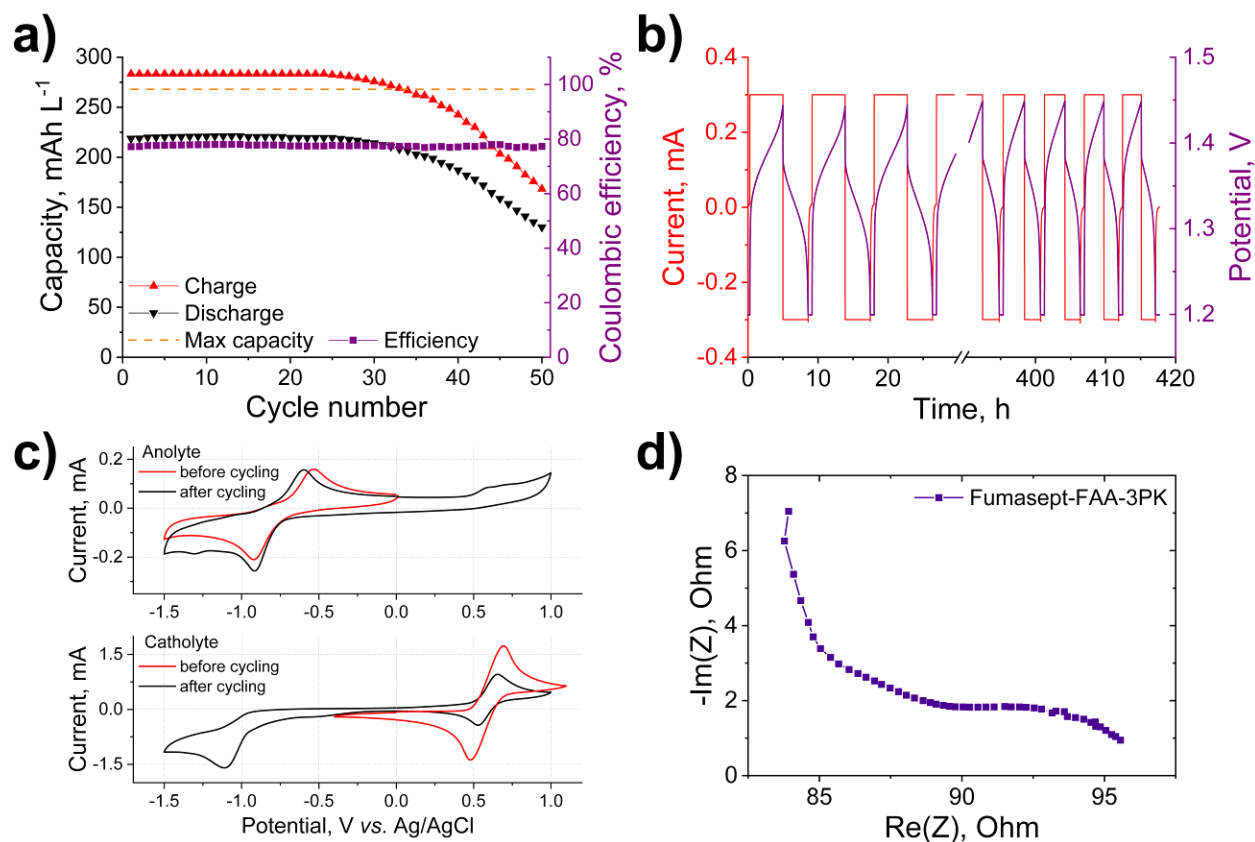


Figure S23. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon charge-discharge cycling for the h-cell based on **M1** anolyte and TEMPOL catholyte in 1M NaCl water solution with Fumasept FAA-3PK-130 membrane (unmixed electrolytes were used); (b) Cell voltage and cell current versus time profile for first 30 hours and 390-420 hours of cycling; (c) CVs of the anolyte and catholyte before and after cycling; (d) The Nyquist Plot of the redox flow battery after cycling.

Electrolyte's composition: mixed electrolytes, **M1** – 5.0 mM, **TEMPOL** -50.0 mM, NaCl – 1.0 M in deionised water. Battery charging and discharging time was limited to 18 000 seconds, that is 108% of theoretical battery capacity.

17. Investigation of the h-cell with Neosepta AHA membrane in neutral conditions

h-cell, Neosepta AHA membrane, unmixed electrolytes

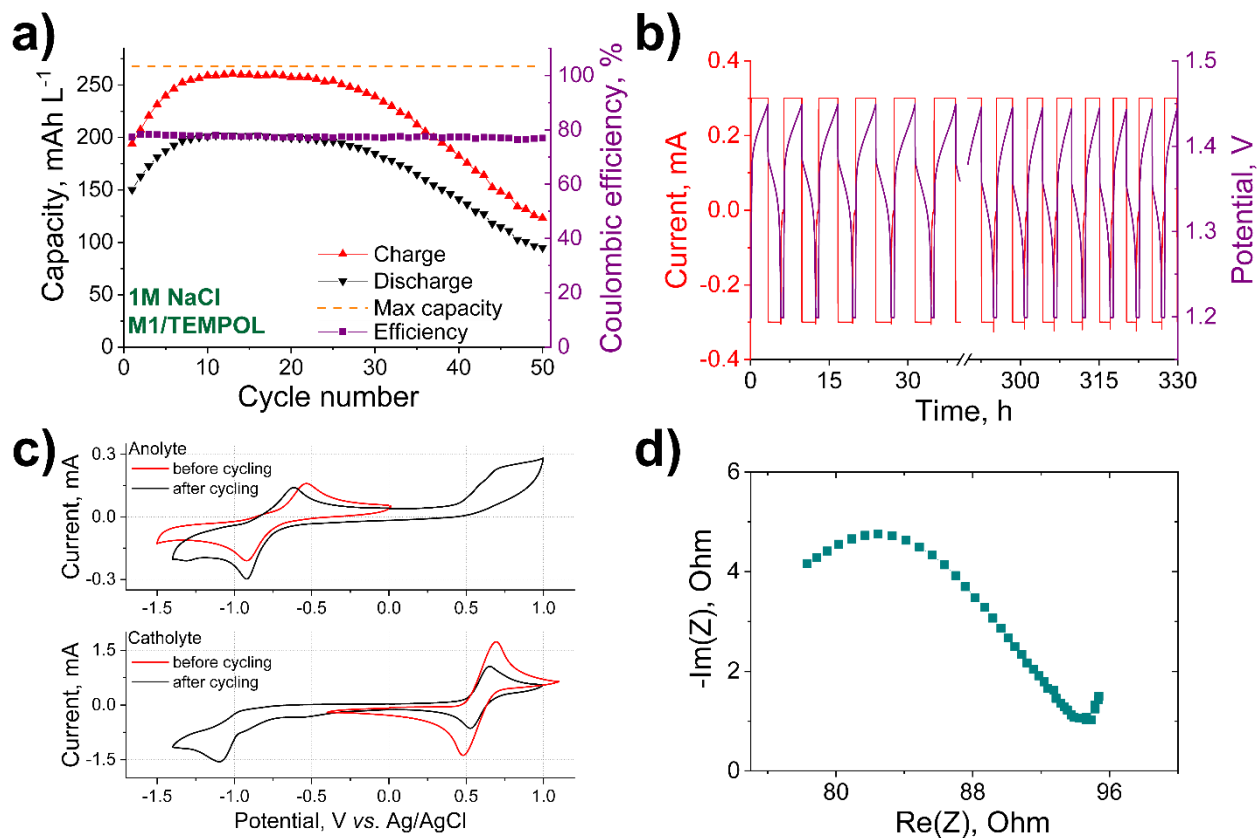


Figure S24. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon charge-discharge cycling for the h-cell based on **M1** anolyte and TEMPOL catholyte in 1M NaCl water solution with Neosepta AHA membrane (unmixed electrolytes were used); (b) Cell voltage and cell current versus time profile for first 40 hours and 190-330 hours of cycling; (c) CVs of the anolyte and catholyte after cycling; (d) The Nyquist Plot of the redox flow battery after cycling.

Electrolyte's composition: mixed electrolytes, **M1** – 5.0 mM, **TEMPOL** -10.0 mM, NaCl – 1.0 M in deionised water. Battery charging and discharging time was limited to 18 000 seconds, that is 108% of theoretical battery capacity.

18. Investigation of the RFB based on M1 anolyte in neutral conditions

RFB, Neosepta AHA membrane, unmixed electrolytes

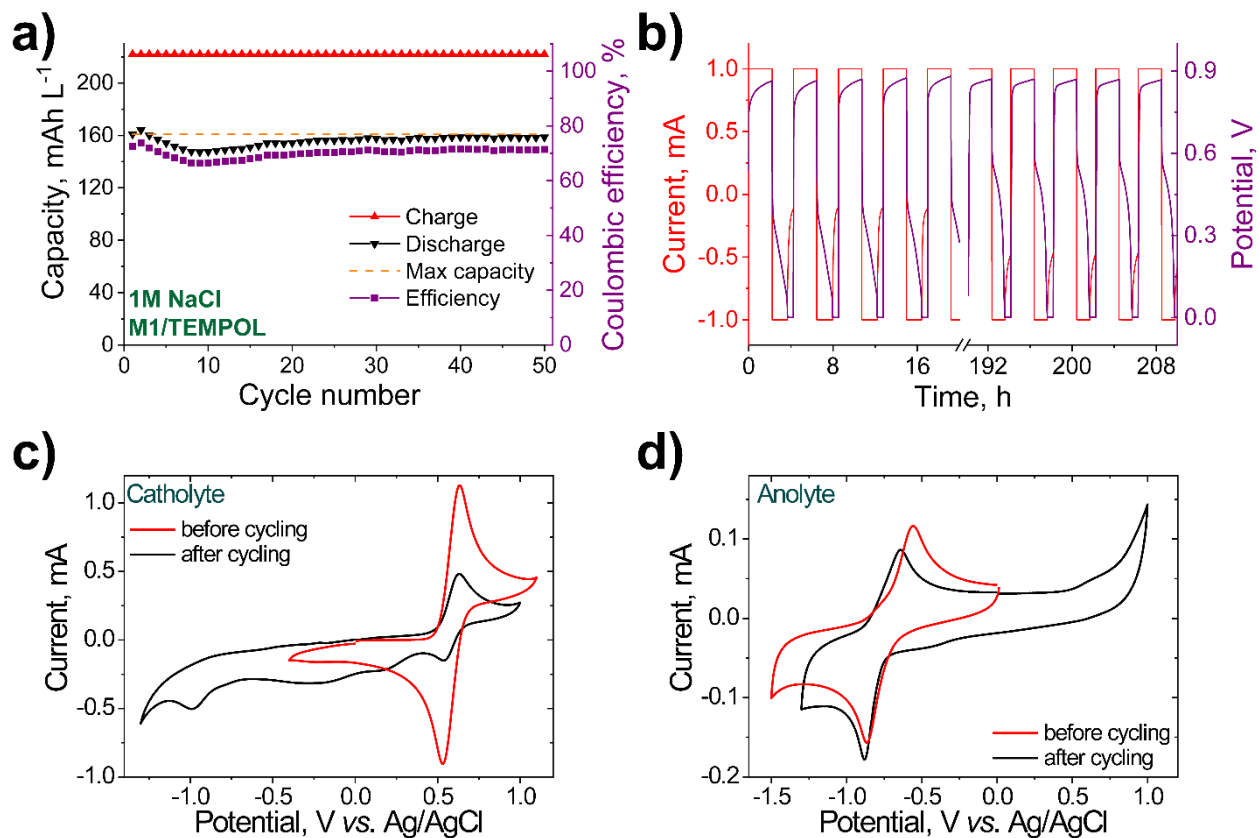


Figure S25. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon charge-discharge cycling for the redox flow battery based on **M1** anolyte and TEMPOL catholyte in 1M NaCl water solution with Neosepta AHA membrane and unmixed electrolytes; (b) Cell voltage and cell current versus time profile for first 20 hours and 190-210 hours of cycling; CVs of the catholyte (c) and anolyte (d) before and after cycling.

Electrolyte's composition: mixed electrolytes, **M1** – 3.0 mM, **TEMPOL** -30.0 mM, NaCl – 1.0 M in deionised water. Battery charging and discharging time was limited to 8000 seconds, that is 138% of theoretical battery capacity.

19. Investigation of the long cycling stability of the RFB based on M1 anolyte in neutral conditions

RFB, Neosepta AHA membrane, unmixed electrolytes

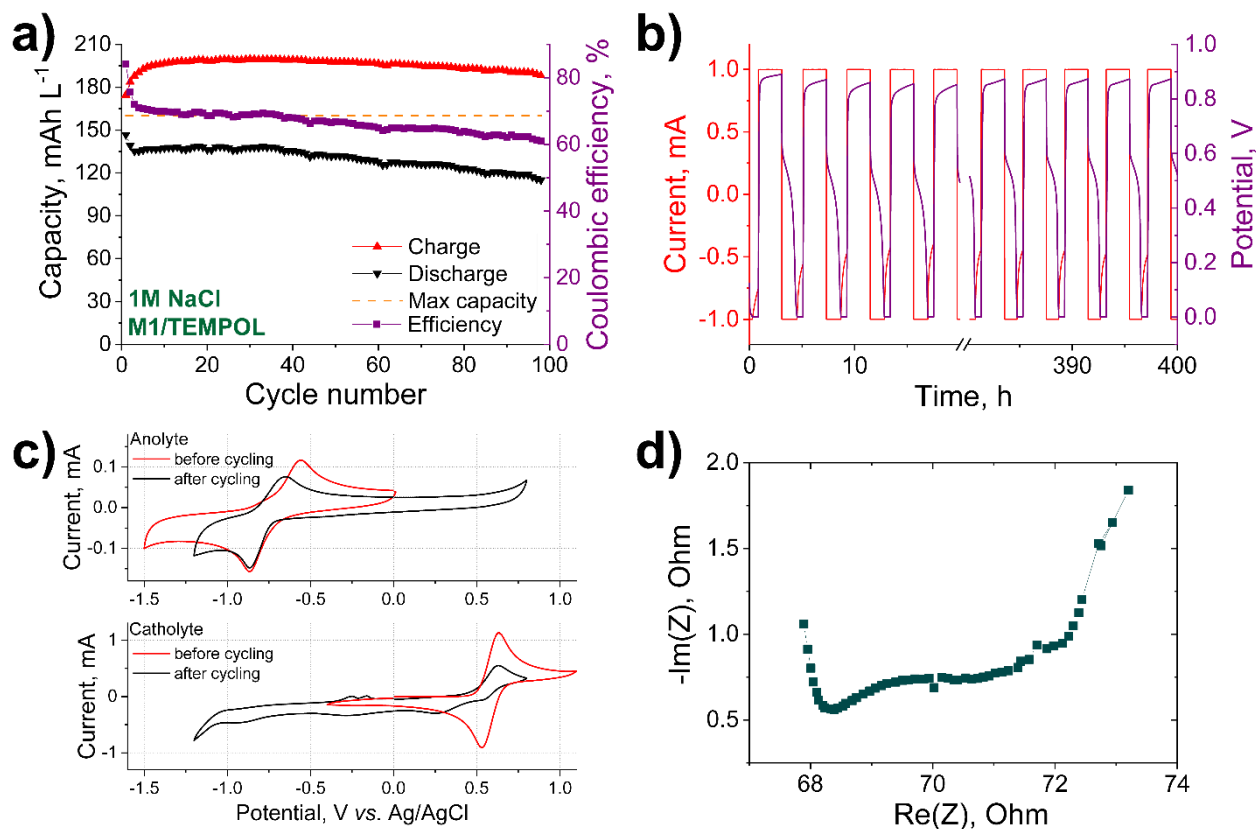


Figure S26. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon charge-discharge cycling for the redox flow battery based on **M1** anolyte and TEMPOL catholyte in 1M NaCl water solution with Neosepta AHA membrane and unmixed electrolytes; (b) Cell voltage and cell current versus time profile for first 20 hours and 190-210 hours of cycling; (c) CVs of the anolyte and catholyte before and after cycling; (d) The Nyquist Plot of the redox flow battery after cycling.

Electrolyte's composition: mixed electrolytes, **M1** – 3.0 mM, **TEMPOL** -30.0 mM, NaCl – 1.0 M in deionised water. Battery charging and discharging time was limited to 8000 seconds, that is 138% of theoretical battery capacity.

20. CV of M1 and $\text{K}_4\text{Fe}(\text{CN})_6$ in base conditions

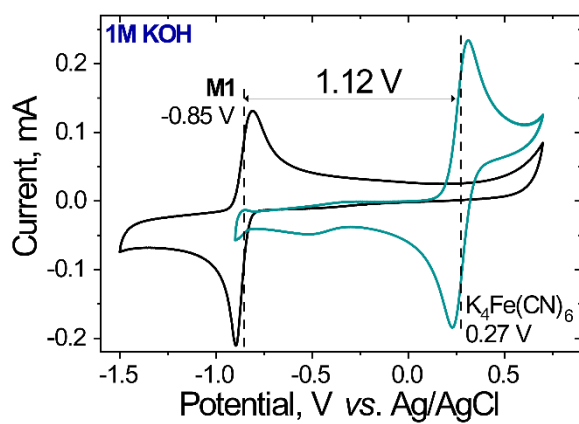


Figure S27. Cyclic voltammograms of pure **M1** and $\text{K}_4\text{Fe}(\text{CN})_6$.

21. Investigation of the h-cell based on M1 anolyte in basic conditions upon potentiostatic cycling

h-cell, Neosepta AHA membrane, unmixed electrolytes

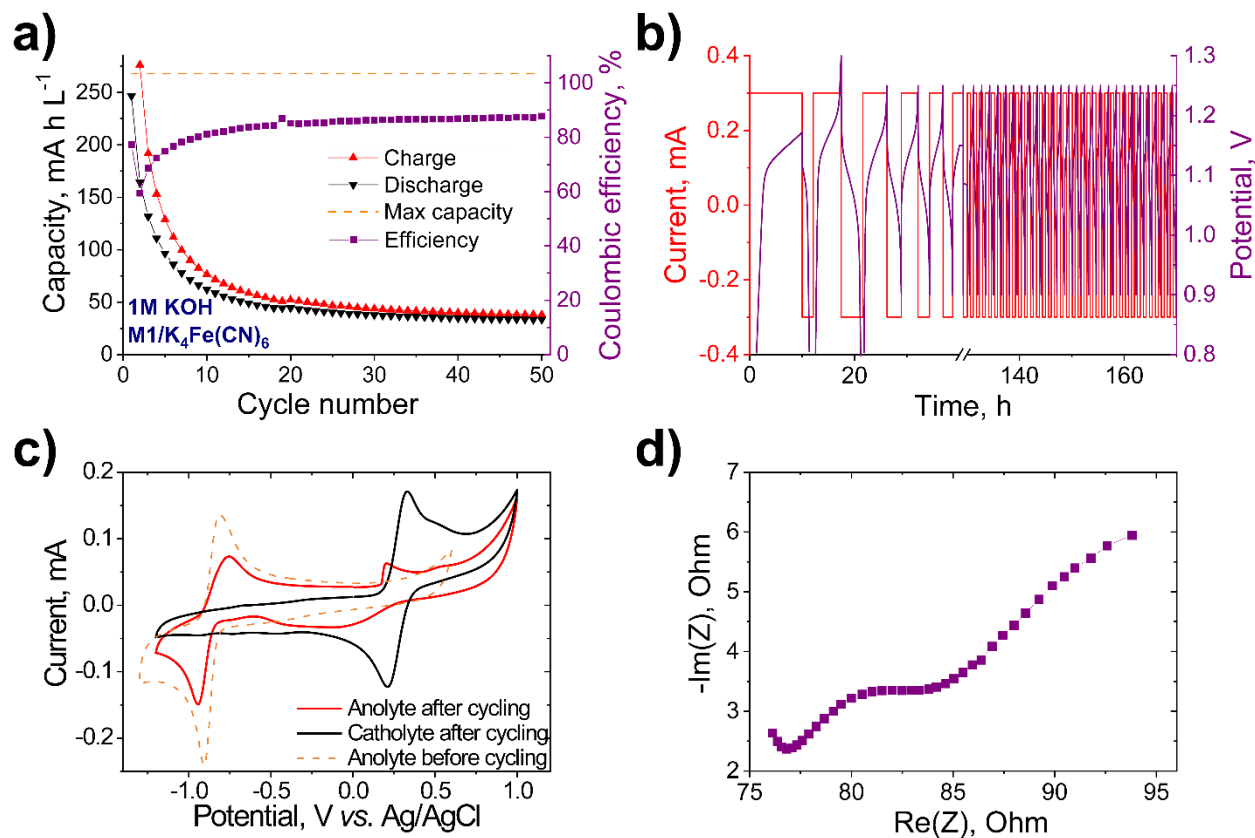


Figure S28. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon potentiostatic charge-discharge cycling for the h-cell based on **M1** anolyte and K₄Fe(CN)₆ catholyte in 1M KOH water solution with Neosepta AHA membrane (unmixed electrolytes were used); (b) Cell voltage and cell current versus time profile for first 30 hours and 140-170 hours of cycling; (c) CVs of the catholyte after cycling and anolyte before and after cycling; (d) The Nyquist Plot of the h-cell after cycling.

Electrolyte's composition: anolyte **M1** – 5.0 mM, KOH – 1.0 M in deionised water; catholyte K₄Fe(CN)₆– 15 mM, K₃Fe(CN)₆– 7.5 mM, KOH – 1.0 M in deionised water.

22. Investigation of the h-cell based on M1 anolyte in basic conditions upon mixed potentiostatic + galvanostatic cycling

h-cell, Neosepta AHA membrane, unmixed electrolytes

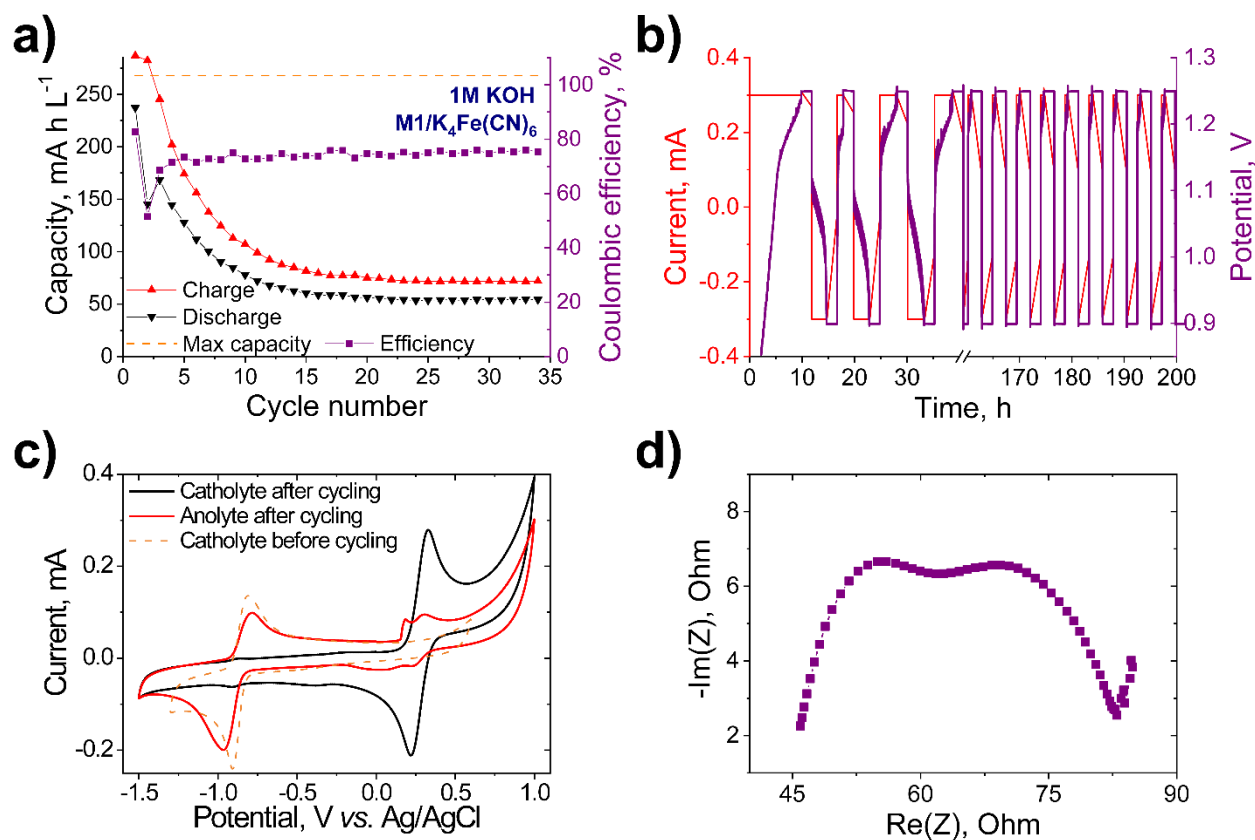


Figure S29. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon mixed potentiostatic + galvanostatic charge-discharge cycling for the h-cell based on **M1** anolyte and K₄Fe(CN)₆ catholyte in 1M KOH water solution with Neosepta AHA membrane and unmixed electrolytes; (b) Cell voltage and cell current versus time profile for first 40 hours and 160-200 hours of cycling; (c) CVs of the catholyte after cycling and anolyte before and after cycling; (d) The Nyquist Plot of the h-cell after cycling.

Electrolyte's composition: anolyte **M1** – 5.0 mM, KOH – 1.0 M in deionised water; catholyte K₄Fe(CN)₆ – 15 mM, K₃Fe(CN)₆ – 7.5 mM, KOH – 1.0 M in deionised water.

23. Investigation of the h-cell based on M1 anolyte in acidic conditions upon potentiostatic cycling

h-cell, Neosepta AHA membrane, unmixed electrolytes

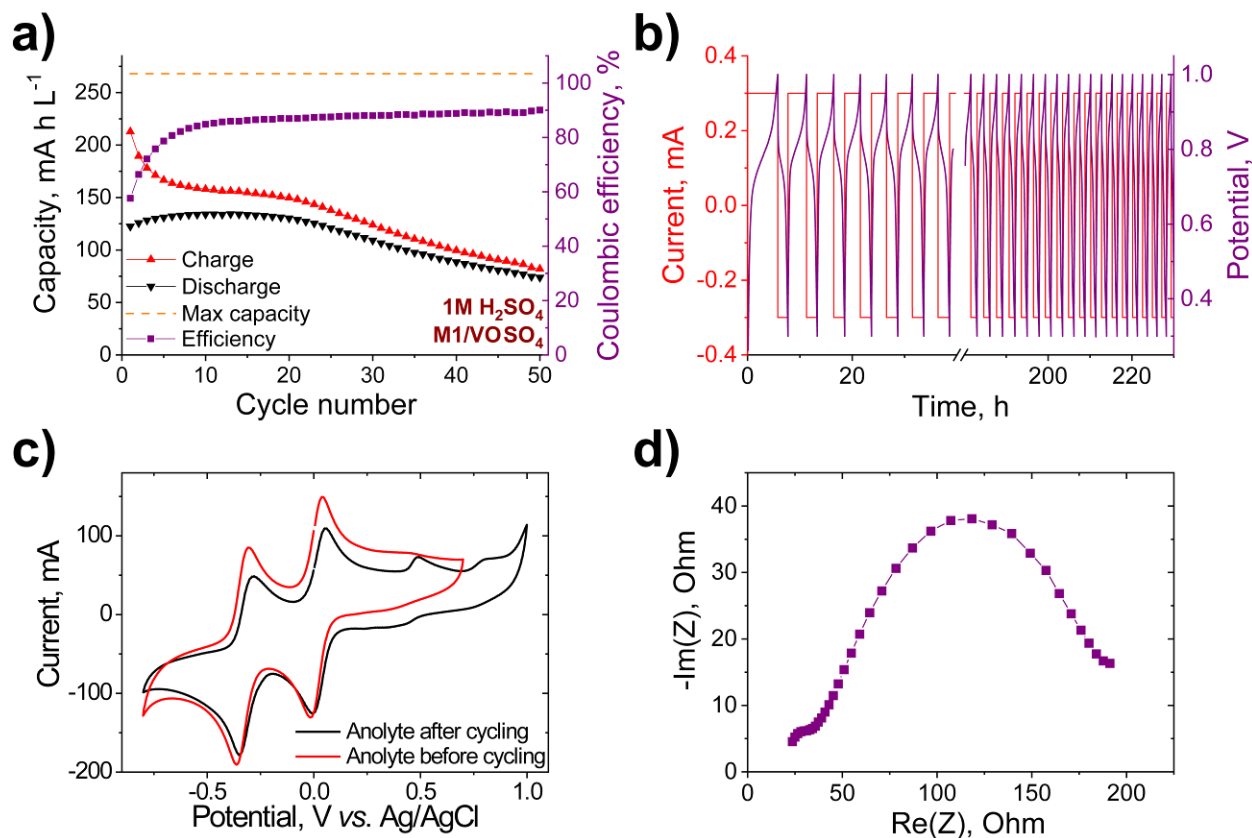


Figure S30. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon potentiostatic charge-discharge cycling for the h-cell based on **M1** anolyte and VOSO₄ catholyte in 1M H₂SO₄ water solution with Neosepta AHA membrane and unmixed electrolytes; (b) Cell voltage and cell current versus time profile for first 30 hours and 200-230 hours of cycling; (c); CVs of the anolyte before and after cycling; (d) The Nyquist Plot of the h-cell after cycling.

Electrolyte's composition: anolyte **M1** – 5.0 mM, H₂SO₄ – 1.0 M in deionised water; catholyte VOSO₄ – 15 mM, H₂SO₄ – 1.0 M in deionised water.

24. Investigation of the h-cell based on M1 anolyte in acidic conditions upon mixed potentiostatic + galvanostatic cycling

h-cell, Neosepta AHA membrane, unmixed electrolytes

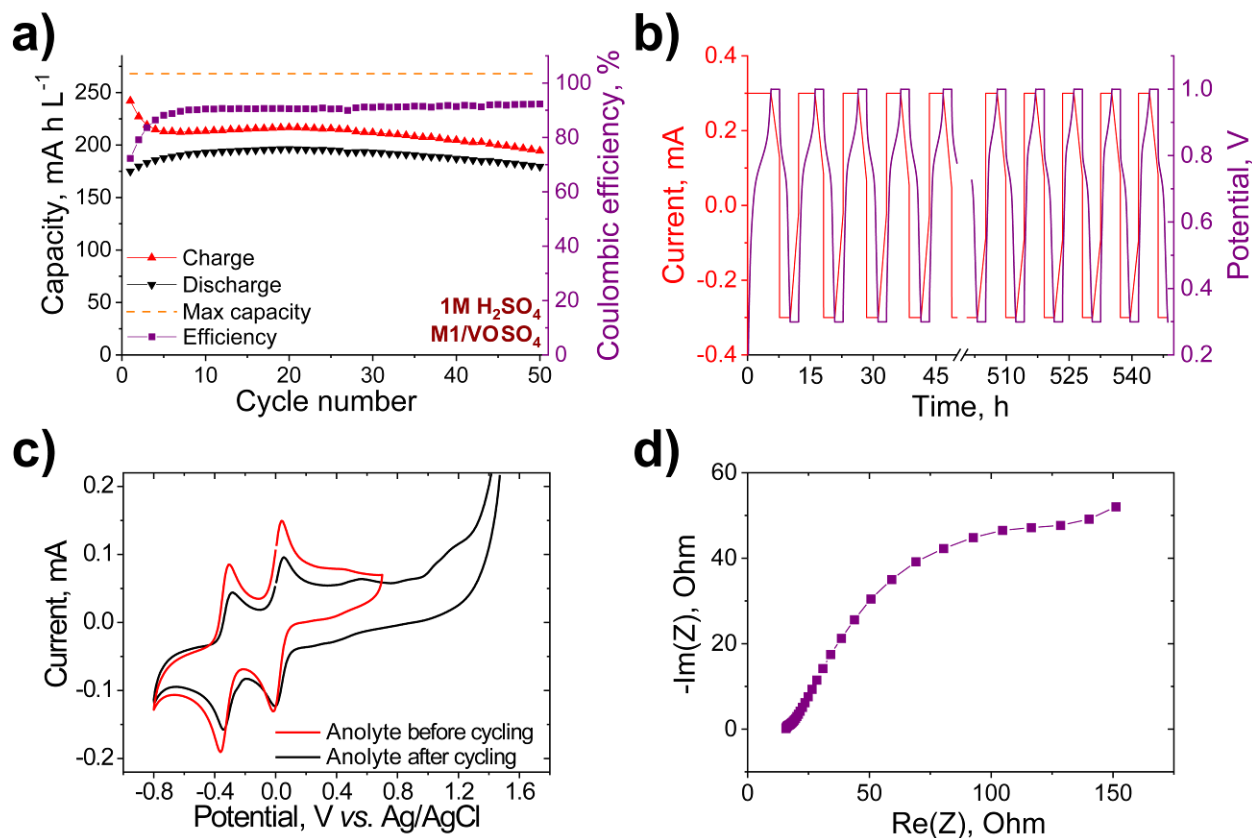


Figure S31. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon mixed potentiostatic + galvanostatic charge-discharge cycling for the h-cell based on **M1** anolyte and VOSO₄ catholyte in 1M H₂SO₄ water solution with Neosepta AHA membrane and unmixed electrolytes; (b) Cell voltage and cell current versus time profile for first 40 hours and 510-550 hours of cycling; (c) CVs of the anolyte before and after cycling; (d) The Nyquist Plot of the the h-cell after cycling.

Electrolyte's composition: anolyte **M1** – 5.0 mM, H₂SO₄ – 1.0 M in deionised water; catholyte VOSO₄ – 15 mM, H₂SO₄ – 1.0 M in deionised water.

25. Investigation of the h-cell based on M1 anolyte in acidic conditions under different charge-discharge currents

h-cell, Neosepta AHA membrane, unmixed electrolytes

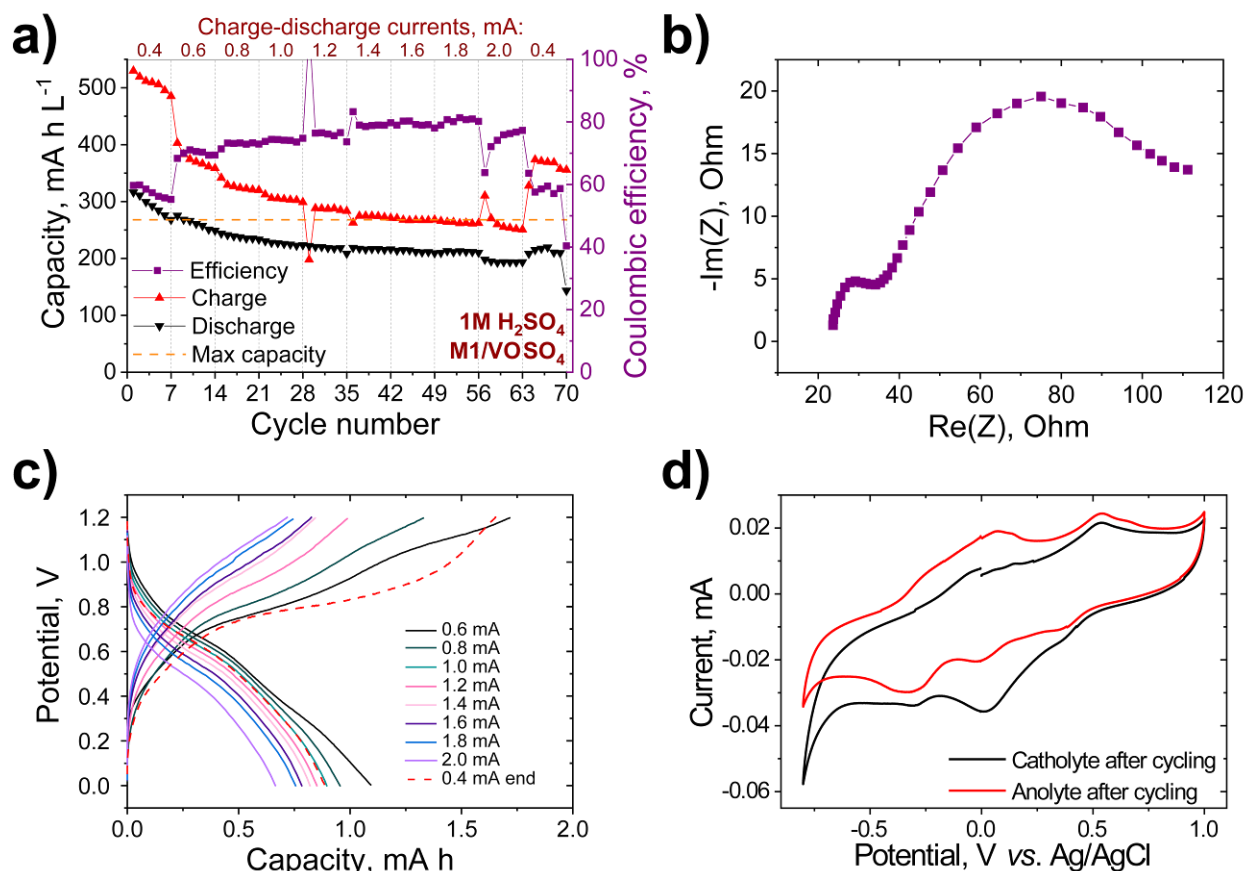


Figure S32. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon charge-discharge cycling at the different currents for the h-cells based on **M1** anolyte, VOSO₄ catholyte in 1M H₂SO₄ water solution with Neosepta AHA membrane; (b) The Nyquist Plot of the the h-cell after cycling; (c) Evolution of the charge and discharge capacities upon charge-discharge cycling at the different currents for the h-cells based on **M1** anolyte, VOSO₄ catholyte in 1M H₂SO₄ water solution with Neosepta AHA membrane; (d) CVs of the anolyte and catholyte after cycling.

Electrolyte's composition: anolyte **M1** – 5.0 mM, H₂SO₄ – 1.0 M in deionised water; catholyte VOSO₄ – 15 mM, H₂SO₄ – 1.0 M in deionised water.

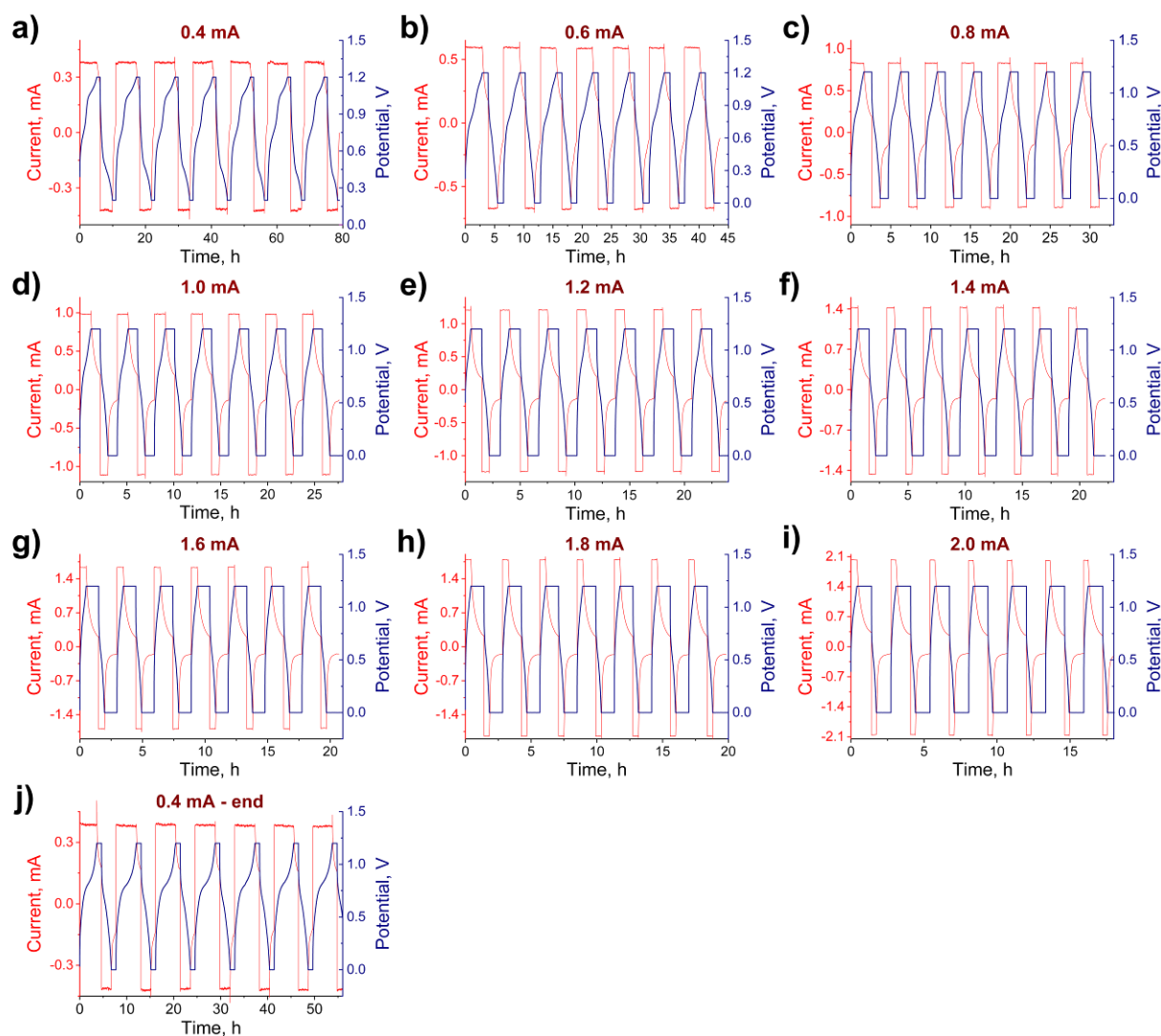


Figure S33. Potential-time dependence curves for the h-cell cycling based on **M1** anolyte, VOSO_4 catholyte in 1M H_2SO_4 water solution with Neosepta AHA membrane under different charge-discharge currents: **(a)** 0.4 mA, **(b)** 0.6 mA, **(c)** 0.8 mA, **(d)** 1.0 mA, **(e)** 1.2 mA, **(f)** 1.4 mA, **(g)** 1.6 mA, **(h)** 1.8 mA, **(i)** 2.0 mA, **(j)** 0.4 mA return to the starting value.

26. Investigation of the cycling stability of the RFB with Neosepta AHA membrane in acidic conditions

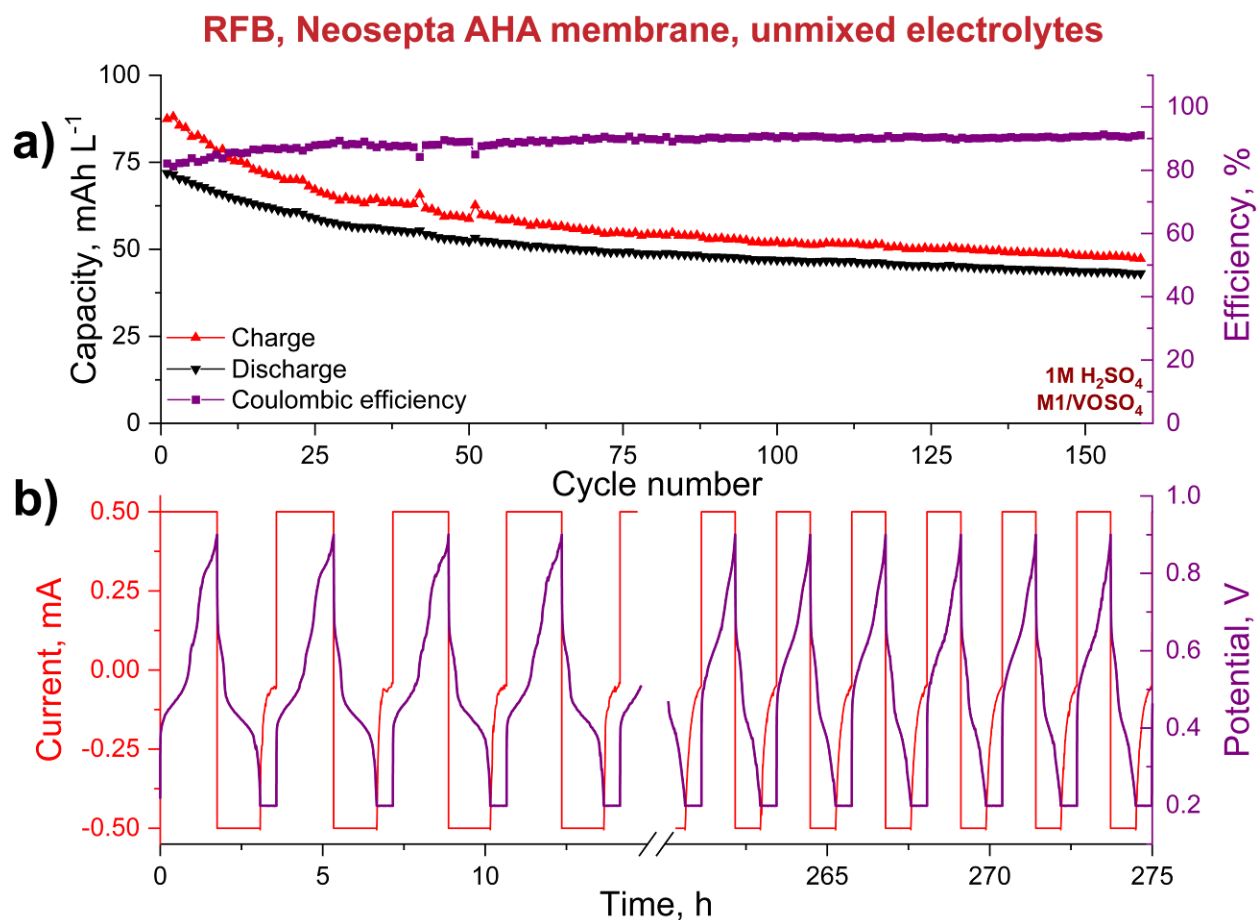


Figure S34. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon mixed potentiostatic + galvanostatic charge-discharge cycling for the h-cell based on **M1** anolyte and VOSO_4 catholyte in 1M H_2SO_4 water solution with Neosepta AHA membrane and unmixed electrolytes; **(b)** Cell voltage and cell current versus time profile for first 20 hours and 255-275 hours of cycling.

Electrolyte's composition: anolyte **M1** – 3.0 mM, H_2SO_4 – 1.0 M in deionised water; catholyte VOSO_4 – 10 mM, H_2SO_4 – 1.0 M in deionised water.

27. Investigation of the long cycling stability of the RFB with Neosepta AHA membrane in acidic conditions

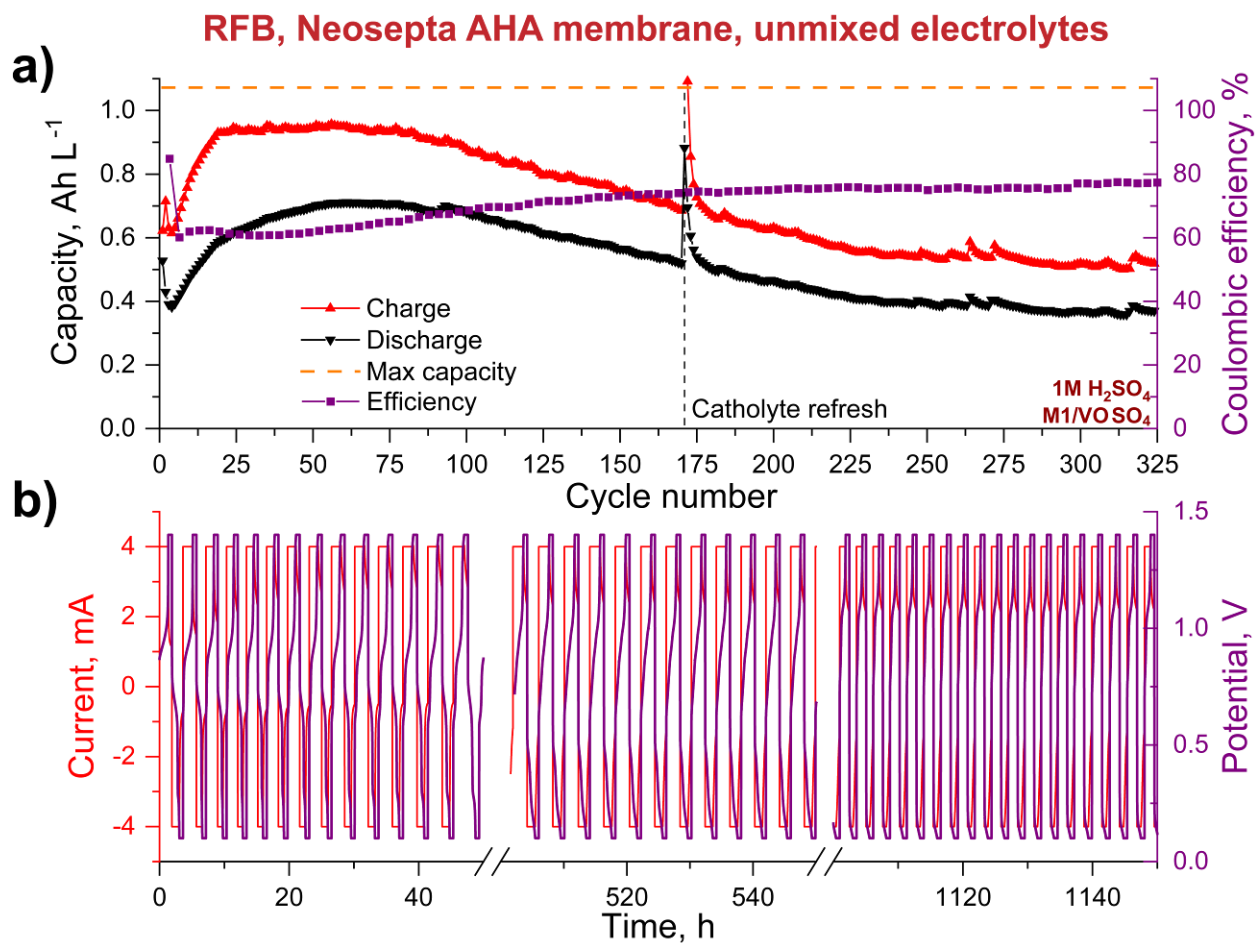


Figure S35. (a) Evolution of the Coulombic efficiency, charge and discharge capacities upon mixed potentiostatic + galvanostatic charge-discharge cycling for the RFB based on **M1** anolyte and VOSO₄ catholyte in 1M H₂SO₄ water solution with Neosepta AHA membrane and unmixed electrolytes; **(b)** Cell voltage and cell current versus time profile for first 50 hours, 500-550 hours and 1100-1150 hours of cycling.

Electrolyte's composition: anolyte **M1** – 20.0 mM, H₂SO₄ – 1.0 M in deionised water; catholyte VOSO₄ – 60 mM, H₂SO₄ – 1.0 M in deionised water.

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