

Article

The Second-Generation Biomethane from Mandarin Orange Peel under Cocultivation with Methanogens and the Armed *Clostridium cellulovorans*

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Abstract: This study demonstrates that the consortium, which consists of the microbial flora of methane production (MFMP) and *Clostridium cellulovorans* grown with cellulose, can perform the direct conversion of cellulosic biomass to methane. The MFMP was taken from a commercial methane fermentation tank and was extremely complicated. Therefore, *C. cellulovorans* grown with cellobiose could not perform high degradation ability on cellulosic biomass due to competition by various microorganisms in MFMP. Focusing on the fact that *C. cellulovorans* was cultivated with cellulose, which is armed with cellulosome, so that it is now armed *C. cellulovorans*; the direct conversion was carried out by the consortium which consisted of MFMP and the armed *C. cellulovorans*. As a result, the consortium of *C. cellulovorans* grown with cellobiose and MFMP (CCeM) could not degrade the purified cellulose and mandarin orange peel. However, MFMP and the armed *C. cellulovorans* reduced 78.4% of the total sugar of the purified cellulose such as MN301, and produced 6.89 mL of methane simultaneously. Furthermore, the consortium consisted of MFMP and the armed *C. cellulovorans* degraded mandarin orange peel without any pretreatments and produced methane that was accounting for 66.2% of the total produced gas.

Keywords: methanogenesis; cellulosic biomass degradation; consortia; armed *C. cellulovorans*

1. Introduction

Although the first-generation biofuels, which are made from corn and sugarcane, have become widespread, there is concern about competition with food supply. Without competing for food, second-generation biofuels are produced from non-edible biomass such as agricultural wastes and cellulosic substrates [1,2].

A plant cell wall is composed of cellulose, hemicellulose, lignin, pectin, etc. Cellulose is a fiber of D-glucose monomers and has strong crystalline [3]. Moreover, cellulose, hemicellulose, and lignin compose the rigid and complex structures [4]. Hemicellulose is a heteropolymer such as xylan, glucuronoxylan, arabinoxylan, glucomannan, and xyloglucan. In addition, lignin, phenol compounds, reinforces the structure of cellulose and hemicellulose, making it more difficult to degrade. Thus, since rigid and complex structures are constructed in cellulosic biomass, it is very difficult to degrade them enzymatically.

Orange juice is one of the major fruit juices and almost the same amount of orange waste as orange juice comes out as a byproduct in orange juice factories. Therefore, it has been considered that such orange wastes are available for non-edible biomass all over the world. However, d-limonene, which is included

in citrus, has an extremely toxic effect on fermenting microorganisms [5,6]. Depending on the type of citrus, the orange peel contains approximately 3% limonene. Therefore, it was necessary to separate d-limonene before the cultivation or to protect microorganisms from d-limonene by encapsulation or immobilization [7,8]. Recently, it was reported that *Clostridium cellulovorans* can degrade orange wastes in the culture including d-limonene and *Clostridium beijerinckii* can carry out isopropanol–butanol–ethanol (IBE) fermentation, which is a bacterial fermentation process producing isopropanol instead of acetone on acetone–ethanol–butanol (ABE) fermentation [9–11]. Thus, the breakthrough was obtained utilizing orange wastes for second-generation biofuels without any pre-treatment.

Some *Clostridia*, such as *C. cellulovorans* and *Clostridium thermocellum*, are known to have the ability to degrade cellulosic biomass efficiently using cellulosomes and secreted non-cellulosomal enzymes [12]. Among those species, we have been studying *C. cellulovorans*, which is a mesophilic and anaerobic cellulolytic bacterium [13]. *C. cellulovorans* degrades not only cellulose but also hemicelluloses consisting of xylose, fructose, galactose, and mannose [14–16]. Whole-genome sequencing of *C. cellulovorans* and the exoproteome profiles revealed 57 cellulosomal protein-encoding genes and 168 secreted-carbohydrase-encoding genes [17,18]. Furthermore, the high degradation ability on plant cell walls has so far been reported [19]. *C. cellulovorans* grown in the culture with cellulose has large protuberances on its surface [20] and the protuberances contain cellulosomes [21]; on the other hand, *C. cellulovorans* grown in the culture with cellobiose does not have the protuberances (Figure 1a,b). *C. cellulovorans* acquires a high ability to degrade the plant cell wall with cellulosome, in other words, *C. cellulovorans* is armed with cellulosome to attack the plant cell wall. Therefore, it can be said that it is the armed *C. cellulovorans* that grow with cellulose. Although *C. cellulovorans* has high degradation ability on plant cell walls in the *C. cellulovorans* monoculture, there are challenges in that *C. cellulovorans* cannot perform high degradation ability in the co-culture or consortia of other microorganisms. Most of the reports used *C. cellulovorans* grown with cellobiose, and these challenges were most likely derived using *C. cellulovorans* grown with cellobiose. Therefore, this study focuses on utilizing the armed *C. cellulovorans* especially for the consortium of other microorganisms.



Figure 1. (a) Cell surface of *C. cellulovorans* grown with cellobiose and (b) cell surface of *C. cellulovorans* grown with cellulose. White bar indicates 2 μm . Modified from [21].

Many studies on methane (CH_4) fermentation have been reported in a wide range of study fields [22]. Methane fermentation using cellulosic biomass is also second-generation biomethane. Since methane production is carried out by the complex microbial flora including methanogens, the second-generation biomethane process needs the consortium to be constructed with microbial flora of methane production (MFMP) and microorganisms which can degrade cellulosic biomass, such as *C. cellulovorans*. The consortium of *C. cellulovorans* with MFMP (CCeM) can degrade sugar beet pulp, which is the residue in a sugar refinery factory, and ferments biogas included methane simultaneously [23]. However, the relict sugars in sugar beet pulp were possible to help *C. cellulovorans* to survive and coexist with MFMP, and the same CCeM could not carry out degrading the purified cellulose and producing methane.

In the present study, we investigated the degradation ability on cellulosic biomass of CCeM that was consistent with the armed *C. cellulovorans* (ACCeM).

2. Materials and Methods

2.1. Materials

Mandarin oranges were purchased at a grocery store in Japan in 2017. Flavedo and albedo, hereafter called removed peel, were cut into strips with scissors just after removing from a mandarin orange. The strip size was approximately 10 mm in length and 2 mm in width (Figure 2). The peel was used as fresh, not dried, and ground. Furthermore, the peel was not treated by any chemicals. The dried weight of the removed peel was measured, it contained 71.6% water. The substrate concentration of removed peel, the purified celluloses, such as Avicel (Sigma, St. Louis, MO, USA) and MN301 (MACHEREY-NAGEL, Düren, Deutschland), and cellobiose (Sigma, St. Louis, MO, USA) was 0.5% (*w/v*) of the dry weight. Avicel is crystalline cellulose powder that is industrially refined from natural cellulose, and the particle size of Avicel is less than 50 μm . MN301 is also industrially refined cellulose powder, and 80% of the particle size is less than 160 μm .

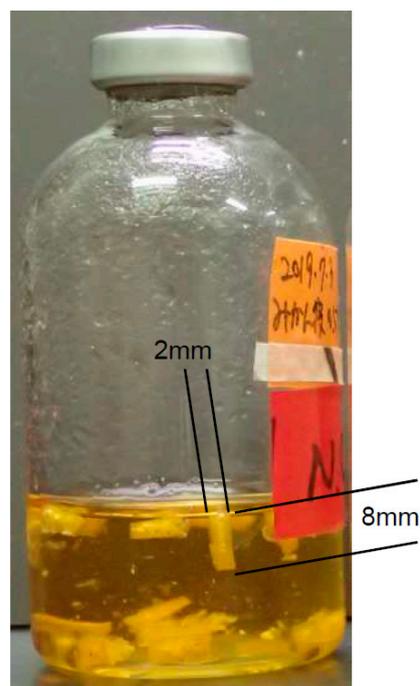
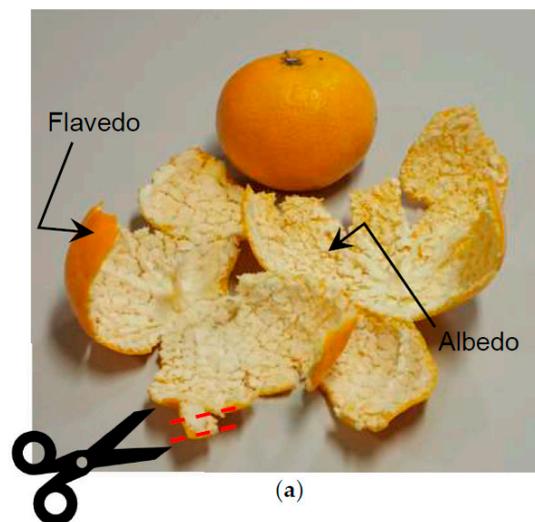


Figure 2. (a) Removed peel before cutting. (b) Strips of removed peel in the medium.

2.2. Microorganism and Culture Condition

The medium was partially modified by *Clostridium cellulovorans* medium [11]. One liter medium contained 4 g of yeast extract, 1 mg of Resazurin salt, 1 g of L-cysteine-HCl, 5 g of NaHCO₃, 0.45 g of K₂HPO₄, 0.45 g of KH₂PO₄, 0.3675 g of NH₄Cl, 0.9 g of NaCl, 0.1575 g of MgCl₂·6H₂O, 0.12 g of CaCl₂·2H₂O, 0.85 mg of MnCl₂·4H₂O, 0.942 mg of CoCl₂·6H₂O, 5.2 mg of Na₂EDTA, 1.5 mg of FeCl₂·4H₂O, 0.07 mg of ZnCl₂, 0.1 mg H₃BO₃, 0.017 mg of CuCl₂·2H₂O, 0.024 mg of NiCl₂·6H₂O, 0.036 mg of Na₂MoO₄·2H₂O, 6.6 mg of FeSO₄·7H₂O, and 0.1 g of p-aminobenzoic acid and was adjusted to pH 7. *C. cellulovorans* 743B (ATCC 35296) was used and anaerobically cultivated in 0.5% (*w/v*) cellobiose, Avicel and MN301 at 37 °C stationary for 19 h, for 4 days and 2 days respectively. The MFMP was obtained from methane fermentation digested liquid in January 2017 at Gifu in Japan. The MFMP was anaerobically cultivated in *Clostridium cellulovorans* medium with 0.5% (*w/v*) glucose (Wako) and 0.25% (*w/v*) cellobiose, or 0.5% (*w/v*) ryegrass leaves at 37 °C for 19 h stationary. The ryegrass leaves were obtained in May 2017 at Aichi in Japan, and were dried and grained to a powder.

2.3. Data Deposition

The sequences of MFMP that are reported in this paper have been deposited in the DNA Data Bank of Japan (DDBJ) (accession 104 no. DRR160954).

2.4. Measurement of Total Sugar Concentration

The total sugar concentration was measured from the precipitation after centrifugation which was 20,000 rpm at 4 °C for 5 min. The precipitation was washed by phosphate buffered salts and 5N NaOH. The total sugar concentration of the washed precipitation was measured by the phenol-sulfuric acid method as D-glucose equivalents.

2.5. Gas Concentration

The produced gas volume of the head space in the culture vial was collected and measured by a syringe (Terumo, Tokyo, Japan). The concentrations of CH₄, H₂, and CO₂ were measured by a gas chromatograph GC-8A (Shimadzu, Kyoto, Japan) with a Thermal Conductivity Detector (TCD) and a column SINCARBON ST (full length 6 m, inner diameter 3 mm; Shinwa, Kyoto, Japan). The column temperature was 200 °C. Argon was a carrier gas and set at a flow rate of 50 mL/min. The injection volume of each sample was 5 mL. The concentration of CH₄ was also measured by a gas chromatograph GC-2010plus (Shimadzu, Kyoto, Japan) with a capillary column Rt-Q-BOND (30 m, inner diameter. 0.32 mm; RESTEK, Centre, PA, USA). The oven temperature was 250 °C and the column temperature was 150 °C. Nitrogen was the carrier gas and set at a flow rate of 1.21 mL/min. The injection volume of each sample was 0.5 mL.

2.6. Organic Acid Concentration

The concentration of organic acids was measured by high-performance liquid chromatography (HPLC), CBM-20A, LC-20AD, CTO-20AC, SPD-20A, and DGU-20A3 (Shimadzu, Kyoto, Japan) with a UV detector and a column KC-811 (300 mm × 2 mm, inner diameter. 8 mm; Showa Denko, Tokyo, Japan). The column temperature was 60 °C. The Bromothymol blue (BTB) post-column method was used. The eluent was 2 mM perchloric acid, and the flow rate was 1.0 mL/min. The reagent was 0.2 mM BTB and 15 mM disodium hydrogen phosphate, and the flow rate was 1.2 mL/min at the wavelength of 445 nm. The injection volume of each sample was 20 µL.

2.7. Cell Growth

Cell growth was measured by Lumitester PD-20, LuciPac Pen and adenosine triphosphate (ATP) eliminating enzyme (Kikkoman Biochemifa, Tokyo, Japan). It is known that integrated intracellular ATP concentration correlates with cell growth [24]. Cell growth was estimated by measuring ATP

concentration of 0.1 mL of cell culture according to the manufacturer’s instruction and was expressed by the relative light unit (RLU) value.

2.8. Statistics

The data were analyzed for statistical significance using Welch’s *t*-test. The difference was assessed with a two-sided test with an α level of 0.05.

3. Results

3.1. Degrading Cellulose and Removed Peel under Cocultivation with Methanogens and Non-Armed *C. cellulovorans*

C. cellulovorans was pre-cultivated in media containing 0.5% (*w/v*) of cellobiose, which meant *C. cellulovorans* was not armed with cellulosome. Anaerobic batch cultivations of *C. cellulovorans*, CCeM, and MFMP were carried out in a 50-mL medium containing 0.5% (*w/v*) of Avicel and removed peel at 37 °C without shaking.

According to measured cell growth of precultures, inoculation amounts for monocultures of *C. cellulovorans* and MFMP were decided so that initial RLU values of each monoculture came close to 2000. RLU values of the preculture of *C. cellulovorans* and MFMP were 45,310 and 50,494, respectively. The inoculation volume was decided as 2 mL for 50 mL monoculture; it was 26 times the dilution so that the initial RLU value of monoculture of *C. cellulovorans* and MFMP were 1743 and 1942, respectively. CCeM was inoculated 2 mL each from both precultures so that concentrations of cell growth against substrate became the same as the monoculture.

RLU values increased for 1 day of cultivation and then cell growth was observed in all cultures (Figure 3a,b). Interestingly, RLU profiles of CCeM and MFMP were quite similar, even CCeM included *C. cellulovorans* and MFMP both. The total sugar concentrations were significantly reduced in *C. cellulovorans* monocultures with Avicel and removed peel after 11 days of cultivation, it was demonstrated that *C. cellulovorans* could degrade Avicel and removed peel. However, the total sugar concentrations in CCeM with Avicel and removed peel were almost the same as that of MFMP; *C. cellulovorans* could not degrade Avicel and removed peel in cultures included MFMP (Figure 3c,d). It was thought that *C. cellulovorans* did not grow in CCeM because cell growths of CCeM and MFMP were almost the same. Methane in the head space was not detected in CCeM with removed peel and that was slightly detected in CCeM and MFMP with Avicel (Figure 3e,f).

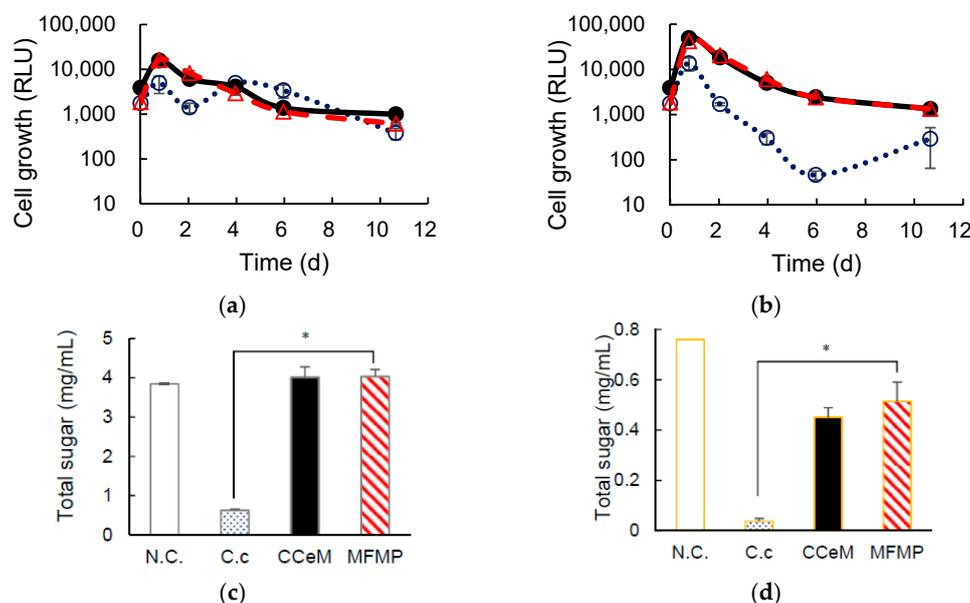


Figure 3. Cont.

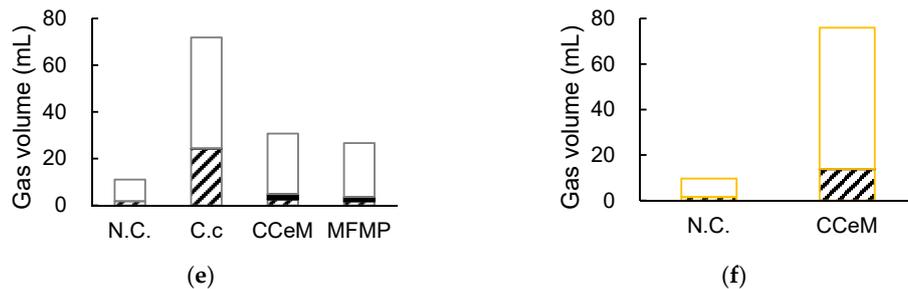


Figure 3. Cultivation of *C. cellulovorans*, cellobiose and MFMP (CCeM) and microbial flora of methane production (MFMP) with Avicel. (a) Cell growth, where *C. cellulovorans* (○_open circle), CCeM (●_closed circle) and MFMP (△_open triangle) are included; (c) total sugar concentration; (e) gas production of H₂ (hatched bar), CH₄ (closed bar), and CO₂ (open bar) are included. Cultivation of *C. cellulovorans*, CCeM, and MFMP with removed peel. (b) Cell growth, where *C. cellulovorans* (○_open circle), CCeM (●_closed circle), and MFMP (△_open triangle) are included; (d) total sugar concentration; (f) gas production of H₂ (hatched bar), CH₄ (closed bar), and CO₂ (open bar) are included. Values with error bars are mean ± SE of three independent samples. SE means a standard error. An asterisk indicates a significant difference ($p < 0.05$).

3.2. Cellulose Degradation and Methane Production under Cocultivation with Methanogens and the Armed *C. cellulovorans*

C. cellulovorans was pre-cultivated in media containing 0.5% (w/v) of MN301 for 2 days, which meant *C. cellulovorans* was armed with cellulosome. Anaerobic batch cultivations of the armed *C. cellulovorans*, ACCeM, and MFMP were carried out in a 40-mL medium containing 0.5% (w/v) of MN301 at 37 °C without shaking. RLU values of the preculture of the armed *C. cellulovorans* and MFMP were 6786 and 38,538, respectively. RLU value of the armed *C. cellulovorans* culture did not increase over 10,000 like the non-armed *C. cellulovorans*. Therefore, inoculation amounts for monocultures of the armed *C. cellulovorans* and MFMP were decided so that initial RLU values of each monoculture became close to 1000. Inoculation volumes of the armed *C. cellulovorans* and MFMP were decided as 7 mL and 1.2 mL, respectively. The armed *C. cellulovorans* was 6.69 times dilution and MFMP was 40.17 times dilution so that initial RLU values of monoculture of the armed *C. cellulovorans* and MFMP were 1014 and 959, respectively. ACCeM was inoculated in 7 mL of the armed *C. cellulovorans* preculture and 1.2 mL of MFMP preculture so that concentrations of cell growth against substrate became the same as the monoculture. Cell growths in the armed *C. cellulovorans* monoculture, ACCeM culture, and MFMP monoculture were measured for 7 days of cultivation. RLU values in ACCeM culture and MFMP monoculture rapidly increased more than 100,000 for 10 h of cultivation. The RLU value in the armed *C. cellulovorans* monoculture increased for 2 days of cultivation and then decreased slowly. Interestingly, the RLU value in ACCeM culture was higher than that in MFMP monoculture after 2 days of cultivation. It suggested that the armed *C. cellulovorans* grew for a couple of days of cultivation and coexisted with MFMP (Figure 4a). The total sugar concentration in the armed *C. cellulovorans* monoculture decreased to 0.38 mg/mL from 5 mg/mL for 7 days of cultivations; it indicated that 92.4% of MN301 was degraded by the armed *C. cellulovorans* for 1 week (Figure 4b). The total sugar concentration in ACCeM culture did not decrease for 3 days of cultivation, however it rapidly decreased from 4 days of cultivation. It also suggested that the armed *C. cellulovorans* survived for 3 days using brought in cellulosome while adapting to coexisting MFMP. Total sugar concentration in ACCeM culture decreased to 1.08 mg/mL for 7 days of cultivation, therefore it was demonstrated that ACCeM could degrade 78.4% of MN301 for 1 week. The total sugar concentration in MFMP monoculture did not decrease for 27 days of cultivation, it indicated that MFMP did not have a capability to degrade MN301. The total sugar concentrations in the armed *C. cellulovorans* monoculture and ACCeM culture was significantly low than that in MFMP for 7 days of cultivation (Figure 4c). Total gas volume in the ACCeM culture rapidly increased for 10 h of cultivation, and the total gas volume kept for 1–7 days of cultivation. Interestingly, the methane proportion in the total gas increased for 1–7 days of cultivation, even though total gas volume did not increase. Furthermore, methane volume continuously increased for 21 days of

cultivation. Finally, methane volume increased 6.89 mL for 1–21 days of cultivation, therefore it revealed that methane production rate from MN301 was 0.014 mL/h (Figure 4d). Organic acid concentrations in the culture supernatant of ACCeM were measured. Lactic acid concentration was slightly detected for 1 day of cultivation and was not detected after 2 days of cultivation (Figure 4e). Formic acid concentration was temporally increased at 1 day of cultivation, and then it was not detected after 2 days of cultivation. Acetic acid concentration increased for 6 days of cultivation that was the same term when the total sugar concentration reached 78.4%. Acetic acid concentration turned to decrease from 8 days of cultivation. On the other hand, propionic acid concentration increased but did not turn to decrease. It was suggested that MN301 was degraded by *C. cellulovorans* and acetic acid and methane were produced, and methane was continuously produced utilizing acetic acid after cellulose was degraded. Moreover, it suggested that acetic acid was converted to methane in preference to propionic acid.

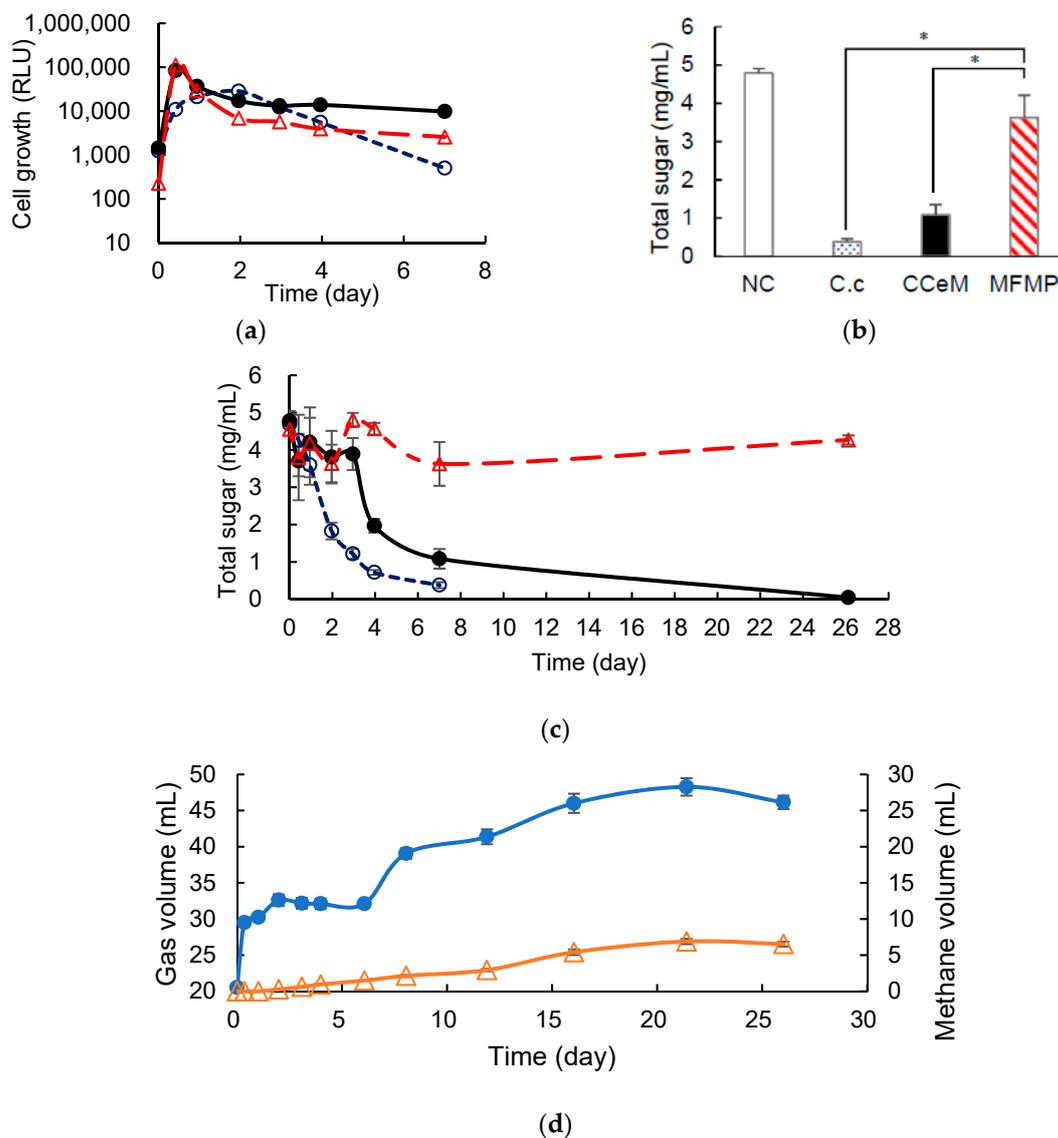


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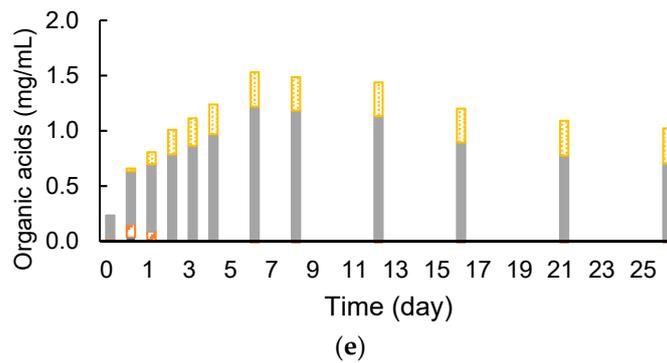


Figure 4. Cultivation of *C. cellulovorans*, CCeM, and MFMP with Avicel. (a) Cell growth, where *C. cellulovorans* (○_open circle), CCeM (●_closed circle), and MFMP (△_open triangle) are included; (b) total sugar concentration; (c) time course of total sugar, where *C. cellulovorans* (○_open circle), CCeM (●_closed circle), and MFMP (△_open triangle) are included; (d) time course of gas production, where total gas volume (●_closed circle) and methane volume (△_open triangle) are included; (e) time course of organic acids concentration of CCeM, where lactic acid (closed bars), formic acid (open bars), acetic acid (closed bars), and propionic acid (dotted bars) are included. Values with error bars are mean \pm SE of three independent samples. SE means a standard error. An asterisk indicates a significant difference ($p < 0.05$).

3.3. Removed Peel Degradation and Methane Production under Cocultivation with Methanogens and the Armed *C. cellulovorans*

It was confirmed that the armed *C. cellulovorans* could degrade cellulose and MFMP also worked to produce methane simultaneously. Anaerobic batch cultivations of ACCeM was carried out in a 20-mL medium containing 0.5% (w/v) of removed peel, which was an actual agricultural waste, at 37 °C without shaking. The armed *C. cellulovorans* was pre-cultivated in media containing 0.5% (w/v) of MN301 for 2 days. According to measured cell growth of precultures, inoculation amounts for monocultures of the armed *C. cellulovorans* and MFMP were decided so that initial RLU values of each monoculture closely became 1000. RLU values of the preculture of the armed *C. cellulovorans* and MFMP were 3448 and 38,538, respectively. Inoculation volumes of the armed *C. cellulovorans* and MFMP were decided at 8.5 mL and 0.8 mL, respectively. The armed *C. cellulovorans* was 3.44 times dilution and MFMP was 36.6 times dilution so that the initial RLU values of ACCeM were 1002 for the armed *C. cellulovorans* and 1052 for MFMP. The degradation of removed peel was observed for 5 days of cultivation (Figure 5a). Gas production started immediately after inoculation and continued for 4 days of cultivation. However, methane was not detected for these early 4 days of cultivation (Figure 5b). Methane began to be detected after 12 days of cultivation. Interestingly, the increment of methane was 4.7 mL for 12–26 days of cultivation against that the total gas was 7.1 mL for the same term, therefore methane occupied 66.2% of the increased gas. This methane concentration is good performance as the fuel for a biogas power generation. Furthermore, methane productivity was 0.014 mL/h for 12–26 days of cultivation, which was equal to that of MN301 substrate. Acetic acid concentration in the culture supernatant increased the same amount as the MN301 substrate. However, the acetic acid concentration was 2.5 times higher than that in the culture supernatant with MN301 (Figure 5c). It suggested that there was room to convert much acetic acid to methane to improve methane production.

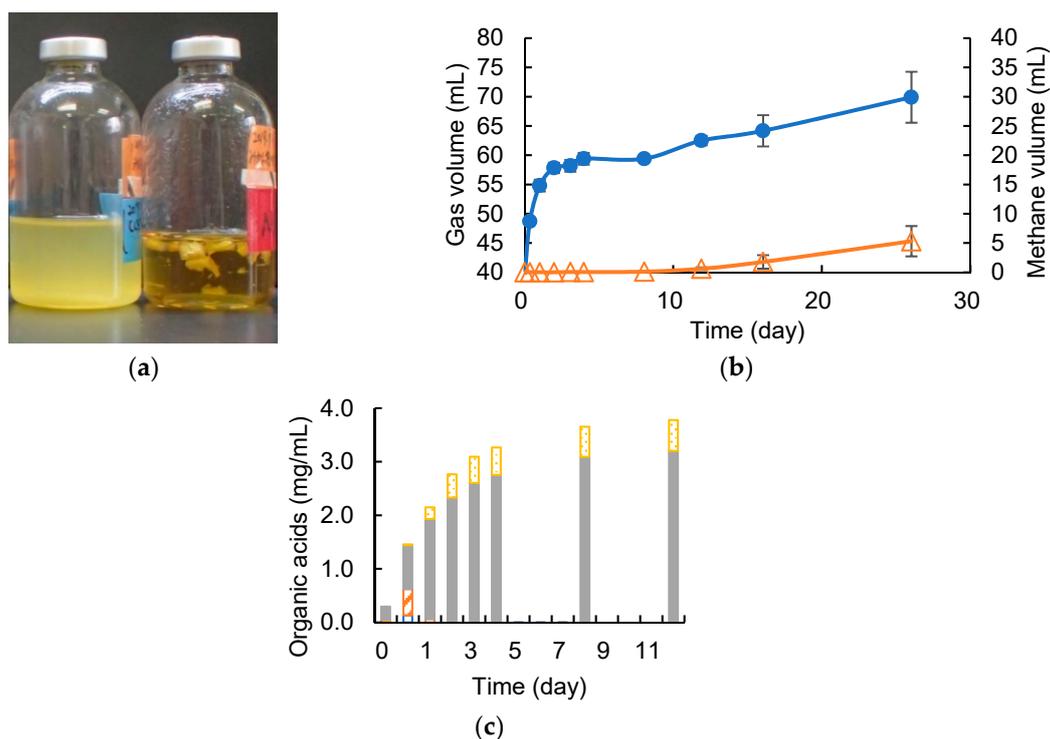


Figure 5. (a) CCEM culture (left), negative control (right); (b) time course of gas production, where total gas volume (●_closed circle) and methane volume (Δ_open triangle) are included; (c) time course of organic acids concentration of CCEM, where lactic acid (closed bars), formic acid (open bars), acetic acid (closed bars), and propionic acid (dotted bars) are included. Values with error bars are mean \pm SE of three independent samples. SE means a standard error. An asterisk indicates a significant difference ($p < 0.05$).

4. Discussion

Since reducing carbon dioxide to conserve the global environment is one of the purposes to replace fossil fuels with biofuels, it is desirable to adopt the biofuel process that has a low environmental load. A sulfuric acid degradation of cellulosic biomass brings about corrosion of a reactor and needs a waste liquid treatment, and the steam explosion process requires a lot of energy to create high pressure and temperature [25,26], therefore these are not low environmental load methods. The enzymatic saccharification is environmental-friendly, because it is a mesophilic process and does not use acids. However, the degradation cost is high because the purified enzymes are expensive and used in large quantities. Instead of using expensive enzymes, the cost will be reduced to utilize microorganisms that can produce enzymes. Furthermore, since *C. cellulovorans* has many genomes related to cellulosome and non-cellulosomal [27], there is potential to improve the efficiency of the enzymatic saccharification. In addition, when the sulfuric acid process or the steam explosion process is used to saccharide cellulosic biomass, various microorganisms attached to the biomass and brought into the saccharification process are not a problem, because they are treated by high temperature and heat or acid. However, there is the key issue that the microorganisms which produce enzymes must survive and perform its saccharification ability coexisting other various microorganisms, because a lot of microorganisms are brought into the culture apparatus together with cellulosic biomass. In this respect, the present study provided a method that enables the co-culture of various microbiota and *C. cellulovorans*, which has been difficult until now. Although there are few reports only co-culture *C. cellulovorans* and one other microbe [28], many experiences can be performed from cellulose to methane using a consortium and the armed *C. cellulovorans*. Notably, this armed *C. cellulovorans* is non-GMO (non- Genetically Modified Organisms) and is not modified with genome editing, such as CRISPR/Cas9 [29], therefore anyone can easily cultivate the armed *C. cellulovorans* from a cellulosic substrate medium and use it anywhere. It is useful and powerful

that the armed *C. cellulovorans*, which anyone can use, degrades orange wastes including d-limonene and the production of methane, which accounts for 66% of the total gas volume, was produced from the consortia.

However, for stable biogas production, it is essential that the carbon, nitrogen, phosphorus, sulfur, and a carbon-to-nitrogen ratio (C/N ratio) in the culture medium have the desired values. In order for sustainable biogas production from cellulosic biomass, it is necessary to investigate in detail how these components are indispensable for microbial growth change. On the other hand, since the sulfur content contained in the cellulosic biomass is small, simplification of a desulphurization equipment in the biogas plant can be expected.

Moreover, an orange contains beta-cryptoxanthin (β -cry), especially in the orange peel [30]. β -cry is a natural carotenoid pigment, and carotenoids are known to contribute to the defense system of the human body against reactive oxygen species. Inverse associations of serum β -cry with the risk for cancer, diabetes, and liver dysfunction have been reported [31–34]. The enzymatic degradation of orange peel can extract carotenoids contained in the culture. This means orange peel, which was disposed of so far, can be converted to useful raw material to have functional foods. In addition, the enzymatic saccharification does not decompose the compound by high temperature and pressure and by acid, therefore the enzymatic saccharification is the most advanced process to extract the useful compounds.

5. Conclusions

This study revealed that *C. cellulovorans* by growing with cellulose instead of cellobiose can coexist with complex microbiota such as methanogenic microbiota. Therefore, it provided a useful method for researching the co-culture of *C. cellulovorans* with various other microorganisms. Furthermore, *C. cellulovorans* and methanogenic microflora coexisted with each other while keeping the cellulose degradation ability and the methane production ability. As a result, the biogas was produced containing 66.2% of methane using mandarin orange peel as a carbon source, and when the biogas contains 60% or more of methane, the biogas can be used directly as fuel for a gas engine.

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