

Table S7. Fungal taxa isolated from Solid sediment (SS), Watery sediment (WS) and microplastics (MPs) and recovered from other marine environments.

Taxa	SS	WS	MPs	Marine environments
<i>Acremonium antarcticum</i> (Spegazzini) D. Hawksworth	x			FR
<i>Acremonium pilosum</i> Giraldo, Guarro, Cano & Gene	x			FR
<i>Acremonium sclerotigenum</i> (Moreau & R. Moreau ex Valenta) W. Gams	x			[1,2]
<i>Acremonium tubakii</i> W. Gams	x			[3,4]
<i>Acrostalagmus luteoalbus</i> (Link) Zare, W. Gams & Schroers	x	x		[5,6]
<i>Albifimbria verrucaria</i> (Alb. & Schwein.) L. Lombard & Crous		x		FR
<i>Alternaria alternata</i> (Fr.) Keissl.	x	x		[5,7]
<i>Alternaria</i> sp.		x		-
<i>Amesia nigricolor</i> (L.M. Ames) X. Wei Wang & Samson	x			[8]
<i>Annulohypoxylon multiforme</i> (Fr.) Y.M. Ju, J.D. Rogers & H.M. Hsieh	x			FR
<i>Apiospora arundinis</i> (Corda) Pintos & P. Alvarado		x		[9,10]
<i>Apiospora marii</i> (Larrondo & Calvo) Pintos & P. Alvarado		x		[11]
<i>Apiospora stipae</i> Crous & R.K. Schumach.	x			FR
<i>Apiotrichum dulcitum</i> (Berkhout) Yurkov & Boekhout		x		[12]
<i>Apiotrichum laibachii</i> (Windisch) Yurkov & Boekhout		x		FR
<i>Armillaria mellea</i> (Vahl) P. Kumm.	x			FR
<i>Arthopyrenia salicis</i> A. Massal.	x			FR
<i>Arthrinium arundinis</i> (Corda) Dyko & B. Sutton	x			[9,13,14]
<i>Arthrinium kogelbergense</i> Crous		x		FR
<i>Arthrinium marii</i> Larrondo & Calvo	x			[11]
<i>Arthrinium phaeospermum</i> (Corda) M.B. Ellis	x	x		[14]
<i>Ascochyta</i> sp.		x		-
Ascomycota sp.1	x			-
Ascomycota sp.2	x			-
Ascomycota sp.3	x			-
Ascomycota sp.4	x			-
Ascomycota sp.5	x			-
<i>Aspergillus aureolatus</i> Munt.-Cvetk. & Bata	x			[15,16]
<i>Aspergillus domesticus</i> Sklenar, Houbraken, Zalar & Hubka			x	FR
<i>Aspergillus europaeus</i> Hubka, A. Nováková, Samson, Houbraken, Frisvad, M. Kolařík		x		[17,18]
<i>Aspergillus flavipes</i> (Bainier & Sartory) Thom & Church	x			[19]
<i>Aspergillus fumigatus</i> Fresen.	x			[4,5,11]
<i>Aspergillus germanicus</i> Frisvad, Varga & Samson	x			FR
<i>Aspergillus heyangensis</i> Z.T. Qi, Z.M. Sun & Yu X. Wang	x	x		[11]
<i>Aspergillus insuetus</i> (Bainier) Thom & Church	x	x		[5,20]
<i>Aspergillus jensenii</i> Jurjević, S.W. Peterson & B.W. Horn	x		x	[21]
<i>Aspergillus niger</i> Tiegh.	x			[5,22,23]
<i>Aspergillus pseudodeflectus</i> Samson & Mouch.	x			[5,24,25]
<i>Aspergillus pseudoglaucus</i> Blochwitz	x			[9,26]
<i>Aspergillus</i> sp.	x			-
<i>Aspergillus spelaeus</i> A. Nováková, Hubka, M. Kolařík & S.W. Peterson		x		[27]
<i>Aspergillus tabacinus</i> Nakaz., Y. Takeda, Simo & A. Watan.	x			[26]
<i>Aspergillus templicola</i> Visagie, Hirooka & Samson	x			FR
<i>Aspergillus terreus</i> Thom	x			[5]

<i>Aspergillus tubingensis</i> Mosseray		x		[5]
<i>Aspergillus ustus</i> (Bainier) Thom & Church	x			[5]
<i>Aspergillus versicolor</i> (Vuill.) Tirab.	x			[5]
<i>Aureobasidium pullulans</i> (de Bary) G. Arnaud	x	x	x	[5,13]
<i>Auxarthron conjugatum</i> (Kuehn) G.F. Orr & Kuehn	x			FR
<i>Auxarthron thaxteri</i> (Kuehn) G.F. Orr & Kuehn	x			FR
<i>Beauveria bassiana</i> (Bals.-Criv.) Vuill.	x			[5]
<i>Beauveria brongniartii</i> (Sacc.) Petch	x			[5]
<i>Beauveria felina</i> (DC.) J.W. Carmich.	x			[5,28]
<i>Beauveria varroae</i> S.A. Rehner & R.A. Humber	x			FR
<i>Biatriospora</i> sp. (MUT 4883)	x			-
Bionectriaceae sp	x			-
<i>Bjerkandera adusta</i> (Willd.) P. Karst.	x		x	[29]
<i>Brunneomyces brunnescens</i> (W. Gams) Giraldo, Gene & Guarro	x			FR
<i>Cadophora luteo-olivacea</i> (J.F.H. Beyma) T.C. Harr. & McNew	x			[30]
<i>Candida membranifaciens</i> (Lodder & Kreger) Wick. & K.A. Burton	x			[5,31,32]
<i>Candida norvegica</i> (Reiersöl) S.A. Mey. & Yarrow	x	x		[5,32]
<i>Candida pseudolambica</i> M.T. Sm. & Poot	x			[5,33]
<i>Candida</i> sp.	x			-
<i>Cephalotrichum asperulum</i> (J.E. Wright & S. Marchand) Sand.-Den., Guarro & Gené		x		FR
<i>Cephalotrichum columnare</i> (H.J. Swart) S.P. Abbott	x	x		FR
<i>Cephalotrichum gorgonifer</i> (Bainier) Sand.-Den., Gené & Guarro	x			[5]
<i>Cephalotrichum inflatum</i> Y.L. Jiang & T.Y. Zhang		x		-
<i>Cephalotrichum stemonitis</i> (Pers.) Nees	x			[34]
<i>Chaetomium cochliodes</i> Palliser		x		FR
<i>Chaetomium globosum</i> Kunze	x			[5,35]
<i>Chrysosporium lobatum</i> Scharapov	x			[9,36]
<i>Cistella</i> sp.	x			-
<i>Cladosporium aggregatocicatricatum</i> Bensch, Crous & U. Braun	x	x		[29]
<i>Cladosporium allicinum</i> (Fr.) Bensch, U. Braun & Crous	x	x		[13,37]
<i>Cladosporium asperulatum</i> Bensch, Crous & U. Braun	x			FR
<i>Cladosporium cladosporioides</i> (Fresen.) G.A. de Vries	x	x	x	[5,14,34]
<i>Cladosporium halotolerans</i> Zalar, de Hoog & Gunde-Cim.	x	x	x	[11,14,38]
<i>Cladosporium inversicolor</i> Bensch, Crous & U. Braun		x		FR
<i>Cladosporium limoniforme</i> Bensch, Crous & U. Braun	x	x		[39]
<i>Cladosporium perangustum</i> Bensch, Crous & U. Braun	x	x		[40]
<i>Cladosporium pseudocladosporioides</i> Bensch, Crous & U. Braun	x	x	x	[14,37]
<i>Cladosporium psychrotolerans</i> Zalar, de Hoog & Gunde-Cim.	x	x		[41]
<i>Cladosporium ramotenellum</i> K. Schub., Zalar, Crous & U. Braun		x	x	[42]
<i>Cladosporium</i> sp.2	x			-
<i>Cladosporium</i> sp1	x			-
<i>Cladosporium sphaerospermum</i> Penz		x		[5,43]
<i>Cladosporium tenellum</i> K. Schub., Zalar, Crous & U. Braun		x		FR
<i>Cladosporium velox</i> Zalar, de Hoog & Gunde-Cim.		x		FR
<i>Cladosporium westerdijkiae</i> Bensch & Samson		x		FR
<i>Cladosporium xylophilum</i> Bensch, Shabunin, Crous & U. Braun		x		[11]
<i>Clonostachys rosea</i> (Link) Schroers, Samuels, Seifert & W. Gams	x	x		[44]
<i>Clonostachys rosmaniae</i> Schroers	x			FR
<i>Clonostachys</i> sp.	x			-

Coniochaeta sp.	x	-	
Cyberlindnera saturnus (Klöcker) Minter	x	[5]	
Cystobasidium slooffiae (E.K. Novák & Vörös-Felkai) Yurkov, Kachalkin, H.M. Daniel, M. Groenew., Libkind, V. de Garcia, Zalar, Gouliam., Boekhout & Begerow	x	[45]	
Debaryomyces hansenii (Zopf) Lodder & Kreger	x	x	[5]
Debaryomyces marasmius Di Menna	x	FR	
Diaporthe columnaris (D.F. Farr & Castl.) Udayanga & Castl.	x	FR	
Dichotomopilus funicola (Cooke) X. Wei Wang & Samson	x	[5]	
Discula destructiva Redlin	x	FR	
Doratomyces microsporus (Sacc.) F.J. Morton & G. Sm.	x	FR	
Emericella pluriseminata Stchigel & Guarro	x	FR	
Emericellopsis maritima Beliakova	x	[5,38]	
Emericellopsis minima Stolk	x	[5]	
Epicoccum nigrum Link	x	[14,38]	
Eupenicillium crustaceum F. Ludw.	x	FR	
Exophiala equina (Pollacci) de Hoog, Vicente, Najafzadeh, Harrak, Badali & Seyedmousavi	x	FR	
Exophiala pisciphila McGinnis & Ajello	x	[5]	
Exophiala xenobiotica de Hoog, J.S. Zeng, Harrak & Deanna A. Sutton	x	[5]	
Filobasidium magnum (Lodder & Kreger-van Rij) Xin Zhan Liu, F.Y. Bai, M. Groenew. & Boekhout	x	[46]	
Furcasterigmium furcatum (Gams) Giraldo López & Crous	x	[5]	
Fusarium brachygibbosum Padwick	x	[14]	
Fusarium cerealis (Cooke) Sacc.	x	FR	
Fusarium incarnatum/equiseti sp. complex	x	[5]	
Fusarium cortaderiae O'Donnell, T. Aoki, Kistler & Geiser	x	FR	
Fusarium culmorum (Wm.G. Sm.) Sacc.	x	x	[5]
Fusarium equiseti (Corda) Sacc.	x	[5]	
Fusarium flocciferum Corda	x	FR	
Fusarium incarnatum (Roberge ex Desm.) Sacc.	x	[5]	
Fusarium oxysporum Schltdl.	x	x	[47,48]
Geotrichum candidum Link	x	[49]	
Gibellulopsis nigrescens (Pethybr.) Zare, W. Gams & Summerb.	x	x	[9]
Gliomastix murorum (Corda) S. Hughes	x	[5]	
Halosphaeriaceae sp.	x	-	
Hongkongomyces thailandica Phukhams. & K.D. Hyde	x	FR	
Hypocreales sp.	x	-	
Kiflimonium curvulum (W. Gams) Summerb., J.A. Scott, Guarro & Crous	x	FR	
Kondoa aerea Á. Fonseca, J.P. Samp. & Fell	x	[50–52]	
Lasionectria marigotensis Lechat & J. Fourn.	x	FR	
Lecanicillium fusisporum (W. Gams) Zare & W. Gams	x	[5]	
Lentitheciaceae sp.	x	-	
Malbranchea circinata Sigler & J.W. Carmich.	x	FR	
Marquandomyces marquandii (Masse) Samson, Houbraken & Luangsa-ard	x	[53]	
Massarina rubi (Fuckel) Sacc.	x	[54]	
Massarina sp.1	x	-	
Massarina sp.2	x	-	
Massarina sp.3	x	-	
Metarhizium brunneum Petch	x	FR	

<i>Microascus paisii</i> (Pollacci) Sand.-Den., Gené & Guarro	x			FR
<i>Microascus</i> sp.	x			-
<i>Microdiplodia</i> sp.	x			-
<i>Microdochium seminicola</i> M. Hern.-Restr., Seifert, Clear & B. Dorn	x			FR
<i>Mortierella alpina</i> Peyronel		x		FR
<i>Mucor circinelloides</i> Tiegh.		x		[55]
<i>Mucor heterogamus</i> Vuill.		x		FR
<i>Mucor hiemalis</i> Wehmer		x		FR
<i>Mucor plumbeus</i> Bonord.		x		FR
<i>Myrmecridium phragmitis</i> Crous	x			FR
<i>Myrmecridium schulzeri</i> (Sacc.) Arzanlou, W. Gams & Crous	x			[5]
<i>Myrothecium</i> sp.	x			-
<i>Nectria inventa</i> Pethybr.	x			[56]
<i>Nectria pseudotrichia</i> Berk. & M.A. Curtis		x		-
<i>Neocosmospora solani</i> (Mart.) L. Lombard & Crous	x			[57]
<i>Neopestalotiopsis clavispora</i> (G.F. Atk.) Maharachch., K.D. Hyde & Crous		x		FR
<i>Niesslia tenuis</i> (W. Gams) W. Gams	x			FR
<i>Nothophoma quercina</i> (Syd. & P. Syd.) Qian Chen & L. Cai		x		FR
<i>Papiliotrema laurentii</i> (Kuff.) Xin Zhan Liu, F.Y. Bai, M. Groenew. & Boekhout		x		FR
<i>Paraconiothyrium cyclothyrioides</i> Verkley	x			[5]
<i>Parasarocladium debruyinii</i> L. Lombard	x			FR
<i>Parasarocladium radiatum</i> (Sukapure & Thirum.) Summerb., J.A. Scott, Guarro & Crous	x			FR
<i>Parasarocladium wereldwijsianum</i> Hern.-Restr.	x			FR
<i>Parathielavia kuwaitensis</i> (Moustafa) X. Wei Wang & Houbraken	x			[58]
<i>Parathyridaria flabelliae</i> E. Bovio, A. Poli, Prigione & Varese	x			[59]
<i>Parengyodontium album</i> (Limber) C.C. Tsang, J.F.W. Chan, W.M. Pong, J.H.K. Chen, A.H.Y. Ngan, M. Cheung, C.K.C. Lai, D.N.C. Tsang, S.K.P. Lau & P.C.Y. Woo		x	x	[5,60,61]
<i>Penicillium jacksonii</i> K.G. Rivera, Houbraken & Seifert		x		[5]
<i>Penicillium antarcticum</i> A.D. Hocking & C.F. McRae	x	x		[13,21,37,62]
<i>Penicillium atramentosum</i> Thom	x	x		[47]
<i>Penicillium bialowiezense</i> K.W. Zaleski			x	[63]
<i>Penicillium brevicompactum</i> Dierckx	x	x	x	[4,5,13]
<i>Penicillium canescens</i> Sopp		x		[5,14]
<i>Penicillium canis</i> S.W. Peterson	x			FR
<i>Penicillium chrysogenum</i> Thom	x	x		[5,14]
<i>Penicillium cinereoatrum</i> Chalab.	x			[64]
<i>Penicillium citreonigrum</i> Dierckx	x			[65]
<i>Penicillium coprophilum</i> (Berk. & M.A. Curtis) Seifert & Samson	x			[65]
<i>Penicillium coralligerum</i> Nicot & Pionnat		x		[66]
<i>Penicillium crustosum</i> Thom	x	x		[67]
<i>Penicillium cyclopium</i> Westling		x		[68]
<i>Penicillium daleae</i> K.W. Zaleski		x		FR
<i>Penicillium echinulatum</i> Raper & Thom ex Fassat.	x	x		[69]
<i>Penicillium egyptiacum</i> J.F.H. Beyma		x		FR
<i>Penicillium expansum</i> Link		x		[5]
<i>Penicillium fellutanum</i> Biourge	x			[70]
<i>Penicillium freii</i> Frisvad & Samson		x		FR

<i>Penicillium glabrum</i> (Wehmer) Westling	x	x	[5,64]
<i>Penicillium glandicola</i> (Oudem.) Seifert & Samson		x	FR
<i>Penicillium griseofulvum</i> Dierckx		x x	[71]
<i>Penicillium canescens</i> Sopp		x	[5,14]
<i>Penicillium herquei</i> Bainier & Sartory		x	[72]
<i>Penicillium hoeksii</i> Houbraken		x	FR
<i>Penicillium hordei</i> Stolk		x	FR
<i>Penicillium jacksonii</i> K.G. Rivera, Houbraken & Seifert		x	FR
<i>Penicillium janczewskii</i> K.W. Zaleski	x		[5]
<i>Penicillium javanicum</i> J.F.H. Beyma	x		[73]
<i>Penicillium manginii</i> Duché & R. Heim		x	FR
<i>Penicillium melanoconidium</i> (Frisvad) Frisvad & Samson		x	FR
<i>Penicillium menonorum</i> S.W. Peterson	x		FR
<i>Penicillium murcianum</i> C. Ramírez & A.T. Martínez	x		[64]
<i>Penicillium nalgiovense</i> Laxa		x	FR
<i>Penicillium olsonii</i> Bainier & Sartory		x	FR
<i>Penicillium osmophilum</i> Stolk & Veenb.-Rijks		x	FR
<i>Penicillium oxalicum</i> Currie & Thom	x		[5]
<i>Penicillium pancosmum</i> Houbraken, Frisvad & Samson		x	FR
<i>Penicillium paneum</i> Frisvad	x		[5]
<i>Penicillium parvulum</i> S.W. Peterson & B.W. Horn	x		FR
<i>Penicillium restrictum</i> J.C. Gilman & E.V. Abbott	x		[5]
<i>Penicillium roseopurpureum</i> Dierckx	x		[9]
<i>Penicillium sacculum</i> E. Dale	x		[5]
<i>Penicillium scabrosum</i> Frisvad, Samson & Stolk		x	FR
<i>Penicillium simplicissimum</i> (Oudem.) Thom	x	x	[5]
<i>Penicillium sizovae</i> Baghd.	x	x	[74]
<i>Penicillium solitum</i> Westling	x	x	[5]
<i>Penicillium</i> sp.1	x		-
<i>Penicillium</i> sp.2	x		-
<i>Penicillium spathulatum</i> Frisvad & Samson		x	FR
<i>Penicillium steckii</i> K.W. Zaleski		x	[64]
<i>Penicillium yezoense</i> Hanzawa		x	FR
<i>Penicillium ubiquetum</i> Houbraken, Frisvad & Samson		x	[75]
<i>Penicillium vancouverense</i> Houbraken, Frisvad & Samson		x	FR
<i>Penicillium venetum</i> (Frisvad) Frisvad		x	FR
<i>Penicillium viridicatum</i> Westling		x	FR
<i>Penicillium waksmanii</i> K.W. Zaleski	x		[5]
<i>Penicillium yarmokense</i> Baghd.	x	x	FR
<i>Periconia byssoides</i> Pers.	x		[5]
<i>Phaeosphaeria</i> sp.	x		-
<i>Phaeosphaeriaceae</i> sp.	x		-
<i>Pholiota gummosa</i> (Lasch) Singer	x		FR
<i>Phoma leveillei</i> Boerema & G.J. Bollen	x		[76]
<i>Pichia methanolica</i> Makig.	x		FR
<i>Pichia occidentalis</i> (Kurtzman, M.J. Smiley & C.J. Johnson) Kurtzman, Robnett & Basehoar-Powers	x		[5]
<i>Plectosphaerella cucumerina</i> (Lindf.) W. Gams	x		[77]
<i>Pleosporales</i> sp.1	x		-
<i>Pleosporales</i> sp.2	x		-

<i>Preussia</i> sp.	x	-
<i>Priceomyces carsonii</i> (Phaff & E.P. Knapp) M. Suzuki	x	[5]
<i>Pseudeurotium bakeri</i> C. Booth	x	[21]
<i>Pseudeurotium ovale</i> Stolk	x	FR
<i>Pseudogymnoascus pannorum</i> (Link) Minnis & D.L. Lindner	x	[5]
<i>Purpureocillium lilacinum</i> (Thom) Luangsa-ard, Houbraken, Hywel-Jones & Samson	x	[5]
<i>Pyrenochaetopsis tabarestanensis</i> M. Papizadeh, Soudi, Amini, Wijayaw. & K.D. Hyde	x	FR
<i>Rhizopus oryzae</i> Went & Prins. Geerl.	x	[78,79]
<i>Rhodotorula diobovata</i> (S.Y. Newell & I.L. Hunter) Q.M. Wang, F.Y. Bai, M. Groenew. & Boekhout	x	[80–83]
<i>Rhodotorula mucilaginosa</i> (A. Jörg.) F.C. Harrison	x	[5,82]
<i>Rhodotorula sphaerocarpa</i> (S.Y. Newell & Fell) Q.M. Wang, F.Y. Bai, M. Groenew. & Boekhout	x	[82]
<i>Roussoella</i> sp.	x	-
<i>Roussoellaceae</i> sp.	x	-
<i>Sakaguchia dacryoidea</i> (Fell, I.L. Hunter & Tallman) Y. Yamada, K. Maeda & Mikata	x	[84–87]
<i>Sarocladium kiliense</i> (Grütz) Summerbell	x	[88]
<i>Scedosporium dehoogii</i> Gilgado, Cano, Gené & Guarro	x	[89]
<i>Scytalidium lignicola</i> Pesante	x	[90]
<i>Sesquicillium microsporum</i> (Jaap) Veenb.-Rijks & W. Gams	x	[91]
<i>Solicoccozyma aerea</i> (Saito) Yurkov	x	FR
<i>Sordariomycetes</i> sp.	x	-
<i>Sporothrix inflata</i> de Hoog	x	[11]
<i>Sporothrix nigrograna</i> (Masuya) Z.W. de Beer, T.A. Duong & M.J. Wingf.	x	FR
<i>Stachybotryna excentrica</i> Gené, M. Caldusch, Abdullah & Guarro	x	FR
<i>Stachybotrys chartarum</i> (Ehrenb.) S. Hughes	x	[5]
<i>Stachybotrys chlorohalonata</i> B. Andersen & Thrane	x	[5]
<i>Stachylidium bicolor</i> Link	x	[5]
<i>Stagonosporopsis cucurbitacearum</i> (Fr.) Aveskamp, Gruyter & Verkley	x	[38]
<i>Stilbella fimetaria</i> (Pers.) Lindau	x	FR
<i>Stilbella</i> sp.	x	-
<i>Suhomyces ambrosiae</i> (Kurtzman) M. Blackw. & Kurtzman	x	FR
<i>Suhomyces</i> sp.	x	-
<i>Symmetrospora coprosmae</i> (Hamamoto & Nakase) Q.M. Wang, F.Y. Bai, M. Groenew. & Boekhout	x	FR
<i>Talaromyces albobiverticillius</i> (H.M. Hsieh, Y.M. Ju & S.Y. Hsieh) Samson, Yilmaz, Frisvad & Seifert	x	[92]
<i>Talaromyces allahbadensis</i> (B.S. Mehrotra & D. Kumar) Samson, N. Yilmaz & Frisvad	x	FR
<i>Talaromyces argentinensis</i> Jurjević & S.W. Peterson	x	FR
<i>Talaromyces borbonicus</i> Houbraken	x	FR
<i>Talaromyces catalonicus</i> Guevara-Suarez, Gené & Guarro	x	FR
<i>Talaromyces flavus</i> (Klöcker) Stolk & Samson	x	[5]
<i>Talaromyces minioluteus</i> (Dierckx) Samson, Yilmaz, Frisvad & Seifert	x	[5]
<i>Talaromyces muroii</i> Yaguchi, Someya & Udagawa	x	FR
<i>Talaromyces verruculosus</i> (Peyronel) Samson, Yilmaz, Frisvad & Seifert	x	[5]
<i>Talaromyces versatilis</i> P.F. Cannon, Bridge & Buddie	x	[93]
<i>Talaromyces wortmannii</i> (Klöcker) C.R. Benj.	x	FR

-
- [3] E.B.G. Jones, S. Suetrong, J. Sakayaroj, A.H. Bahkali, M.A. Abdel-Wahab, T. Boekhout, K.L. Pang, Classification of marine Ascomycota, Basidiomycota, Blastocladiomycota and Chytridiomycota, *Fungal Diversity*. 73 (2015). <https://doi.org/10.1007/s13225-015-0339-4>.
- [4] L. Panno, M. Bruno, S. Voyron, A. Anastasi, G. Gnani, L. Miserere, G.C. Varese, Diversity, ecological role and potential biotechnological applications of marine fungi associated to the seagrass *Posidonia oceanica*, *New Biotechnology*. 30 (2013). <https://doi.org/10.1016/j.nbt.2013.01.010>.
- [5] E.B.G. Jones, S. Suetrong, J. Sakayaroj, A.H. Bahkali, M.A. Abdel-Wahab, T. Boekhout, K.L. Pang, Classification of marine Ascomycota, Basidiomycota, Blastocladiomycota and Chytridiomycota, *Fungal Diversity*. 73 (2015). <https://doi.org/10.1007/s13225-015-0339-4>.
- [6] J. Cao, X.M. Li, L.H. Meng, B. Konuklugil, X. Li, H.L. Li, B.G. Wang, Isolation and characterization of three pairs of indoleketopiperazine enantiomers containing infrequent N-methoxy substitution from the marine algal-derived endophytic fungus *Acrostalagmus luteoalbus* TK-43, *Bioorganic Chemistry*. 90 (2019). <https://doi.org/10.1016/j.bioorg.2019.103030>.
- [7] S.T. Fang, F.P. Miao, X.H. Liu, Y.P. Song, N.Y. Ji, Two new tricycloalternarene acids from the marine-derived fungus *Alternaria alternata* ICD5-11, *Phytochemistry Letters*. 23 (2018). <https://doi.org/10.1016/j.phytol.2017.12.009>.
- [8] J.M. Wong Chin, D. Puchooa, T. Bahorun, R. Jeewon, Antimicrobial properties of marine fungi from sponges and brown algae of Mauritius, *Mycology*. 12 (2021). <https://doi.org/10.1080/21501203.2021.1895347>.
- [9] A. Poli, E. Bovio, L. Ranieri, G.C. Varese, V. Prigione, Fungal Diversity in the Neptune Forest: Comparison of the Mycobiota of *Posidonia oceanica*, *Flabellia petiolata*, and *Padina pavonica*, *Frontiers in Microbiology*. 11 (2020). <https://doi.org/10.3389/fmicb.2020.00933>.
- [10] Marine *Athrinium* spp. Isolated from *Sargassum* sp. (Brown Algae) in Jeju Island and Unrecorded Species in Korea, *The Korean Journal of Mycology*. 44 (2016). <https://doi.org/10.4489/kjm.2016.44.4.259>.
- [11] L. Garzoli, A. Poli, V. Prigione, G. Gnani, G.C. Varese, Peacock's tail with a fungal cocktail: first assessment of the mycobiota associated with the brown alga *Padina pavonica*, *Fungal Ecology*. 35 (2018) 87–97. <https://doi.org/10.1016/j.funeco.2018.05.005>.
- [12] L.M.Y. Mitchison-Field, J.M. Vargas-Muñiz, B.M. Stormo, E.J.D. Vogt, S. van Dierdonck, J.F. Pelletier, C. Ehrlich, D.J. Lew, C.M. Field, A.S. Gladfelter, Unconventional Cell Division Cycles from Marine-Derived Yeasts, *Current Biology*. 29 (2019). <https://doi.org/10.1016/j.cub.2019.08.050>.
- [13] G. Gnani, L. Garzoli, A. Poli, V. Prigione, G. Burgaud, G.C. Varese, The culturable mycobiota of *Flabellia petiolata*: First survey of marine fungi associated to a Mediterranean green alga, *PLoS ONE*. 12 (2017). <https://doi.org/10.1371/journal.pone.0175941>.
- [14] E. Bovio, G. Gnani, V. Prigione, F. Spina, R. Denaro, M. Yakimov, R. Calogero, F. Crisafi, G.C. Varese, The culturable mycobiota of a Mediterranean marine site after an oil spill: Isolation, identification and potential application in bioremediation, *Science of the Total Environment*. 576 (2017) 310–318. <https://doi.org/10.1016/j.scitotenv.2016.10.064>.
- [15] J. Peng, X. Zhang, W. Wang, T. Zhu, Q. Gu, D. Li, Austalides S-U, new meroterpenoids from the sponge-derived fungus *Aspergillus aureolatus* HDN14-107, *Marine Drugs*. 14 (2016). <https://doi.org/10.3390/md14070131>.
- [16] W. Sun, W. Wu, X. Liu, D.A. Zaleta-Pinet, B.R. Clark, Bioactive compounds isolated from marine-derived microbes in China: 2009-2018, *Marine Drugs*. 17 (2019). <https://doi.org/10.3390/md17060339>.
- [17] X. Du, D. Liu, J. Huang, C. Zhang, P. Proksch, W. Lin, Polyketide derivatives from the sponge associated fungus *Aspergillus europaeus* with antioxidant and NO inhibitory activities, *Fitoterapia*. 130 (2018). <https://doi.org/10.1016/j.fitote.2018.08.030>.
- [18] J. Xu, M. Yi, L. Ding, S. He, A review of anti-inflammatory compounds from marine fungi, 2000-2018, *Marine Drugs*. 17 (2019). <https://doi.org/10.3390/md17110636>.
- [19] E. Bovio, L. Garzoli, A. Poli, V. Prigione, D. Firsova, G.P. McCormack, G.C. Varese, The culturable mycobiota associated with three Atlantic sponges, including two new species: *Thelebolus balaustiformis* and *T. spongiae*, *Fungal Systematics and Evolution*. 1 (2018). <https://doi.org/10.3114/fuse.2018.01.07>.
- [20] E. Cohen, L. Koch, K.M. Thu, Y. Rahamim, Y. Aluma, M. Ilan, O. Yarden, S. Carmeli, Novel terpenoids of the fungus *Aspergillus insuetus* isolated from the Mediterranean sponge *Psammocinia* sp. collected along the coast of Israel, in: *Bioorganic and Medicinal Chemistry*, 2011. <https://doi.org/10.1016/j.bmc.2011.05.045>.
- [21] E. Bovio, L. Garzoli, A. Poli, V. Prigione, D. Firsova, G.P. McCormack, G.C. Varese, The culturable mycobiota associated with three Atlantic sponges, including two new species: *Thelebolus balaustiformis* and *T. spongiae*, *Fungal Systematics and Evolution*. 1 (2018). <https://doi.org/10.3114/fuse.2018.01.07>.
- [22] C. Raghukumar, S. Raghukumar, Barotolerance of fungi isolated from deep-sea sediments of the Indian Ocean, *Aquatic Microbial Ecology*. 15 (1998). <https://doi.org/10.3354/ame015153>.
- [23] Y. Khambhaty, K. Mody, S. Basha, B. Jha, Kinetics, equilibrium and thermodynamic studies on biosorption of hexavalent chromium by dead fungal biomass of marine *Aspergillus niger*, *Chemical Engineering Journal*. 145 (2009). <https://doi.org/10.1016/j.cej.2008.05.002>.
- [24] Y.M. Lee, M.J. Kim, H. Li, P. Zhang, B. Bao, K.J. Lee, J.H. Jung, Marine-Derived *Aspergillus* Species as a Source of Bioactive Secondary Metabolites, *Marine Biotechnology*. 15 (2013). <https://doi.org/10.1007/s10126-013-9506-3>.
- [25] S. Dobretsov, H. Al-Shibli, S.S.N. Maharachchikumbura, A.M. Al-Sadi, The presence of marine filamentous fungi on a copper-based antifouling paint, *Applied Sciences (Switzerland)*. 11 (2021). <https://doi.org/10.3390/app11188277>.
- [26] S. Lee, M.S. Park, Y.W. Lim, Diversity of marine-derived *Aspergillus* from tidal mudflats and Sea Sand in Korea, *Mycobiology*. 44 (2016). <https://doi.org/10.5941/MYCO.2016.44.4.237>.

-
- [27] P. Marchese, L. Garzoli, G. Gnani, E. O'Connell, A. Bouraoui, M. Mehiri, J.M. Murphy, G.C. Varese, Diversity and bioactivity of fungi associated with the marine sea cucumber *Holothuria poli*: disclosing the strains potential for biomedical applications, *Journal of Applied Microbiology*. 129 (2020). <https://doi.org/10.1111/jam.14659>.
- [28] F.Y. Du, X.M. Li, Z.C. Sun, L.H. Meng, B.G. Wang, Secondary Metabolites with Agricultural Antagonistic Potentials from *Beauveria felina*, a Marine-Derived Entomopathogenic Fungus, *Journal of Agricultural and Food Chemistry*. 68 (2020). <https://doi.org/10.1021/acs.jafc.0c05696>.
- [29] J. Álvarez-Barragán, C. Cravo-Laureau, L.Y. Wick, R. Duran, Fungi in PAH-contaminated marine sediments: Cultivable diversity and tolerance capacity towards PAH, *Marine Pollution Bulletin*. 164 (2021). <https://doi.org/10.1016/j.marpolbul.2021.112082>.
- [30] P. Marchese, L. Garzoli, R. Young, L. Allcock, F. Barry, M. Tuohy, M. Murphy, Fungi populate deep-sea coral gardens as well as marine sediments in the Irish Atlantic Ocean, *Environmental Microbiology*. 23 (2021) 4168–4184. <https://doi.org/10.1111/1462-2920.15560>.
- [31] D. Greetham, A.S. Zaky, C. Du, Exploring the tolerance of marine yeast to inhibitory compounds for improving bioethanol production, *Sustainable Energy and Fuels*. 3 (2019). <https://doi.org/10.1039/c9se00029a>.
- [32] L. Wang, Z. Chi, L. Yue, Z. Chi, D. Zhang, Occurrence and diversity of *Candida* genus in marine environments, *Journal of Ocean University of China*. 7 (2008). <https://doi.org/10.1007/s11802-008-0416-3>.
- [33] X. Zou, Y. Wei, K. Dai, F. Xu, H. Wang, X. Shao, Yeasts from intertidal zone marine sediment demonstrate antagonistic activities against *Botrytis cinerea* in vitro and in strawberry fruit, *Biological Control*. 158 (2021). <https://doi.org/10.1016/j.biocontrol.2021.104612>.
- [34] L. Panno, M. Bruno, S. Voyron, A. Anastasi, G. Gnani, L. Miserere, G.C. Varese, Diversity, ecological role and potential biotechnological applications of marine fungi associated to the seagrass *Posidonia oceanica*, *New Biotechnology*. 30 (2013). <https://doi.org/10.1016/j.nbt.2013.01.010>.
- [35] C.M. Cui, X.M. Li, C.S. Li, P. Proksch, B.G. Wang, Cytoglobosins A-G, cytochalasins from a marine-derived endophytic Fungus, *Chaetomium globosum* QEN-14, *Journal of Natural Products*. 73 (2010). <https://doi.org/10.1021/np900569t>.
- [36] G. le Goff, P. Lopes, G. Arcile, P. Vlachou, E. van Elslande, P. Retailleau, J.F. Gallard, M. Weis, Y. Benayahu, N. Fokialakis, J. Ouazzani, Impact of the Cultivation Technique on the Production of Secondary Metabolites by *Chrysosporium lobatum* TM-237-S5, Isolated from the Sponge *Acanthella cavernosa*, *Marine Drugs*. 17 (2019). <https://doi.org/10.3390/md17120678>.
- [37] P. Marchese, L. Garzoli, R. Young, L. Allcock, F. Barry, M. Tuohy, M. Murphy, Fungi populate deep-sea coral gardens as well as marine sediments in the Irish Atlantic Ocean, *Environmental Microbiology*. 23 (2021) 4168–4184. <https://doi.org/10.1111/1462-2920.15560>.
- [38] G. Cecchi, L. Cutroneo, S. di Piazza, M. Capello, M. Zotti, Culturable fungi from dredged and marine sediments from six ports studied in the framework of the SEDITERRA Project, *Journal of Soils and Sediments*. 21 (2021). <https://doi.org/10.1007/s11368-021-02884-4>.
- [39] A. Maamar, M.E. Lucchesi, S. Debaets, N.N. van Long, M. Quemener, E. Coton, M. Bouderbala, G. Burgaud, A. Matallah-Boutiba, Highlighting the crude oil bioremediation potential of marine fungi isolated from the Port of Oran (Algeria), *Diversity (Basel)*. 12 (2020). <https://doi.org/10.3390/D12050196>.
- [40] Z. Fan, Z.H. Sun, H.X. Liu, Y.C. Chen, H.H. Li, W.M. Zhang, Perangustols A and B, a pair of new azaphilone epimers from a marine sediment-derived fungus *Cladosporium perangustum* FS62, *Journal of Asian Natural Products Research*. 18 (2016). <https://doi.org/10.1080/10286020.2016.1181623>.
- [41] P. Zalar, G.S. de Hoog, H.J. Schroers, P.W. Crous, J.Z. Groenewald, N. Gunde-Cimerman, Phylogeny and ecology of the ubiquitous saprobe *Cladosporium sphaerospermum*, with descriptions of seven new species from hypersaline environments, *Studies in Mycology*. 58 (2007). <https://doi.org/10.3114/sim.2007.58.06>.
- [42] A. Patyshakuliyeva, D.L. Falkoski, A. Wiebenga, K. Timmermans, R.P. de Vries, Macroalgae derived fungi have high abilities to degrade algal polymers, *Microorganisms*. 8 (2020). <https://doi.org/10.3390/microorganisms8010052>.
- [43] G.H. Yu, G.W. Wu, T.J. Zhu, Q.Q. Gu, D.H. Li, Cladosins F and G, two new hybrid polyketides from the deep-sea-derived *Cladosporium sphaerospermum* 2005-01-E3, *Journal of Asian Natural Products Research*. 17 (2015). <https://doi.org/10.1080/10286020.2014.940330>.
- [44] L. Garzoli, G. Gnani, F. Tamma, S. Tosi, G.C. Varese, A.M. Picco, Sink or swim: Updated knowledge on marine fungi associated with wood substrates in the Mediterranean Sea and hints about their potential to remediate hydrocarbons, *Progress in Oceanography*. 137 (2015). <https://doi.org/10.1016/j.pocean.2015.05.028>.
- [45] V.N. Gonçalves, G.A. Vitoreli, G.C.A. de Menezes, C.R.B. Mendes, E.R. Secchi, C.A. Rosa, L.H. Rosa, Taxonomy, phylogeny and ecology of cultivable fungi present in seawater gradients across the Northern Antarctica Peninsula, *Extremophiles*. 21 (2017). <https://doi.org/10.1007/s00792-017-0959-6>.
- [46] B. Valderrama, J.J. Ruiz, M.S. Gutiérrez, K. Alveal, M. Caruffo, M. Oliva, H. Flores, A. Silva, M. Toro, A. Reyes-Jara, P. Navarrete, Cultivable yeast microbiota from the marine fish species *genypterus chilensis* and *seriolella violacea*, *Journal of Fungi*. 7 (2021). <https://doi.org/10.3390/jof7070515>.
- [47] G. Greco, G. Cecchi, S. di Piazza, L. Cutroneo, M. Capello, M. Zotti, Fungal characterisation of a contaminated marine environment: the case of the Port of Genoa (North-Western Italy), *Webbia*. 73 (2018). <https://doi.org/10.1080/00837792.2017.1417964>.
- [48] V. Nenkep, K. Yun, B.W. Son, Oxysporizoline, an antibacterial polycyclic quinazoline alkaloid from the marine-mudflat-derived fungus *Fusarium oxysporum*, *Journal of Antibiotics*. 69 (2016). <https://doi.org/10.1038/ja.2015.137>.
- [49] Samuel, Prabakaran, Antibacterial Activity of Marine derived Fungi Collected from South East Coast of Tamilnadu, India, 2011. <http://scholarsresearchlibrary.com/archive.html>.

-
- [50] R. Fotedar, A. Kolecka, T. Boekhout, J.W. Fell, A. Al-Malki, A. Zeyara, M. al Marri, Fungal diversity of the hypersaline Inland Sea in Qatar, *Botanica Marina*. 61 (2018) 595–609. <https://doi.org/10.1515/bot-2018-0048>.
- [51] M.M. Francis, V. Webb, G.C. Zuccarello, Marine yeast biodiversity on seaweeds in New Zealand waters, *New Zealand Journal of Botany*. 54 (2016) 30–47. <https://doi.org/10.1080/0028825X.2015.1103274>.
- [52] Minegishi H, Usami R, Abe F, Phylogenetic analysis of pectin degrading yeasts from deep-sea environments, 2006.
- [53] G.M. Cabrera, M. Butler, M.A. Rodriguez, A. Godeas, R. Haddad, M.N. Eberlin, A sorbicillinoid urea from an intertidal *Paecilomyces marquandii*, *Journal of Natural Products*. 69 (2006). <https://doi.org/10.1021/np060315d>.
- [54] A. Poli, E. Bovio, L. Ranieri, G.C. Varese, V. Prigione, Fungal Diversity in the Neptune Forest: Comparison of the Mycobiota of *Posidonia oceanica*, *Flabellia petiolata*, and *Padina pavonica*, *Frontiers in Microbiology*. 11 (2020). <https://doi.org/10.3389/fmicb.2020.00933>.
- [55] L. Garzoli, A. Poli, V. Prigione, G. Gnani, G.C. Varese, Peacock's tail with a fungal cocktail: first assessment of the mycobiota associated with the brown alga *Padina pavonica*, *Fungal Ecology*. 35 (2018) 87–97. <https://doi.org/10.1016/j.funeco.2018.05.005>.
- [56] M.R.Z. Passarini, C. Santos, N. Lima, R.G.S. Berlinck, L.D. Sette, Filamentous fungi from the Atlantic marine sponge *Drummacidon reticulatum*, *Archives of Microbiology*. 195 (2013). <https://doi.org/10.1007/s00203-012-0854-6>.
- [57] G. Greco, G. Cecchi, S. di Piazza, L. Cutroneo, M. Capello, M. Zotti, Fungal characterisation of a contaminated marine environment: the case of the Port of Genoa (North-Western Italy), *Webbia*. 73 (2018). <https://doi.org/10.1080/00837792.2017.1417964>.
- [58] X.W. Wang, F.Y. Bai, K. Bensch, M. Meijer, B.D. Sun, Y.F. Han, P.W. Crous, R.A. Samson, F.Y. Yang, J. Houbraken, Phylogenetic re-evaluation of *Thielavia* with the introduction of a new family *Podosporaceae*, *Studies in Mycology*. 93 (2019). <https://doi.org/10.1016/j.simyco.2019.08.002>.
- [59] A. Poli, E. Bovio, L. Ranieri, G.C. Varese, V. Prigione, News from the sea: A new genus and seven new species in the pleosporalean families roussoellaceae and thyridariaceae, *Diversity (Basel)*. 12 (2020). <https://doi.org/10.3390/D12040144>.
- [60] W. Wang, S. Li, Z. Chen, Z. Li, Y. Liao, J. Chen, Secondary Metabolites Produced by the Deep-Sea-Derived Fungus *Engyodontium album*, *Chemistry of Natural Compounds*. 53 (2017). <https://doi.org/10.1007/s10600-017-1957-8>.
- [61] L.A. Giddings, D.J. Newman, Extremophilic Fungi from Marine Environments: Underexplored Sources of Antitumor, Anti-Infective and Other Biologically Active Agents, *Marine Drugs*. 20 (2022). <https://doi.org/10.3390/md20010062>.
- [62] L. Garzoli, A. Poli, V. Prigione, G. Gnani, G.C. Varese, Peacock's tail with a fungal cocktail: first assessment of the mycobiota associated with the brown alga *Padina pavonica*, *Fungal Ecology*. 35 (2018) 87–97. <https://doi.org/10.1016/j.funeco.2018.05.005>.
- [63] Q. Zhang, B. Yang, F. Li, M. Liu, S. Lin, J. Wang, Y. Xue, H. Zhu, W. Sun, Z. Hu, Y. Zhang, Mycophenolic acid derivatives with immunosuppressive activity from the coral-derived fungus *penicillium bialowiezense*, *Marine Drugs*. 16 (2018). <https://doi.org/10.3390/md16070230>.
- [64] E. Bovio, E. Stecci, A. Poli, G. Gnani, V. Prigione, T. Lacour, M. Mehiri, G.C. Varese, The culturable mycobiota associated with the Mediterranean sponges *Aplysina cavernicola*, *Crambe crambe* and *Phorbas tenacior*, *FEMS Microbiology Letters*. 366 (2020). <https://doi.org/10.1093/femsle/fnaa014>.
- [65] D. Reeb, P.B. Best, A. Botha, K.J. Cloete, M. Thornton, M. Mouton, Fungi associated with the skin of a southern right whale (*Eubalaena australis*) from South Africa, *Mycology*. 1 (2010). <https://doi.org/10.1080/21501203.2010.492531>.
- [66] K. Takahashi, K. Sakai, Y. Nagano, S. Orui Sakaguchi, A.O. Lima, V.H. Pellizari, M. Iwatsuki, K. Takishita, K. Nonaka, K. Fujikura, S. Omura, Cladomarine, a new anti-saproplegniasis compound isolated from the deep-sea fungus, *Penicillium coralligerum* YK-247, *Journal of Antibiotics*. 70 (2017). <https://doi.org/10.1038/ja.2017.58>.
- [67] J. Álvarez-Barragán, C. Cravo-Laureau, L.Y. Wick, R. Duran, Fungi in PAH-contaminated marine sediments: Cultivable diversity and tolerance capacity towards PAH, *Marine Pollution Bulletin*. 164 (2021). <https://doi.org/10.1016/j.marpolbul.2021.112082>.
- [68] Y.H. Li, X.M. Li, X. Li, S.Q. Yang, X.S. Shi, H.L. Li, B.G. Wang, Antibacterial Alkaloids and Polyketide Derivatives from the Deep Sea-Derived Fungus *Penicillium cyclopium* SD-413, *Mar Drugs*. 18 (2020). <https://doi.org/10.3390/md18110553>.
- [69] A.I. Khusnullina, E.N. Bilanenko, A. v. Kurakov, Microscopic Fungi of White Sea Sediments, *Contemporary Problems of Ecology*. 11 (2018). <https://doi.org/10.1134/S1995425518050062>.
- [70] R. Nicoletti, A. Trincone, Bioactive compounds produced by strains of *Penicillium* and *Talaromyces* of marine origin, *Marine Drugs*. 14 (2016). <https://doi.org/10.3390/md14020037>.
- [71] L.N. Zhou, T.J. Zhu, S.X. Cai, Q.Q. Gu, D.H. Li, Three new indole-containing diketopiperazine alkaloids from a deep-ocean sediment derived fungus *Penicillium griseofulvum*, *Helvetica Chimica Acta*. 93 (2010). <https://doi.org/10.1002/hlca.200900443>.
- [72] S.Q. Yang, A. Mándi, X.M. Li, H. Liu, X. Li, S. Balázs Király, T. Kurtán, B.G. Wang, Separation and configurational assignment of stereoisomeric phenalenones from the marine mangrove-derived fungus *Penicillium herquei* MA-370, *Bioorganic Chemistry*. 106 (2021). <https://doi.org/10.1016/j.bioorg.2020.104477>.
- [73] H. al Nasrawi, Biodegradation of Crude Oil by Fungi Isolated from Gulf of Mexico, *Journal of Bioremediation & Biodegradation*. 03 (2012). <https://doi.org/10.4172/2155-6199.1000147>.

-
- [74] M. da Luz Calado, J. Silva, C. Alves, P. Susano, D. Santos, J. Alves, A. Martins, H. Gaspar, R. Pedrosa, M.J. Campos, Marine endophytic fungi associated with *Halopteris scoparia* (Linnaeus) Sauvageau as producers of bioactive secondary metabolites with potential dermocosmetic application, *PLoS ONE*. 16 (2021). <https://doi.org/10.1371/journal.pone.0250954>.
- [75] T.P.T. Hoang, C. Roullier, G. Genta-Jouve, M.C. Boumard, T. Robiou du Pont, H. Nazih, Y.F. Pouchus, O. Grovel, C25 steroids from the marine mussel-derived fungus *Penicillium ubiqueum* MMS330, *Phytochemistry Letters*. 34 (2019). <https://doi.org/10.1016/j.phytol.2019.09.002>.
- [76] Z. Paz, M. Komon-Zelazowska, I.S. Druzhinina, M.M. Aveskamp, A. Shnaiderman, Y. Aluma, S. Carmeli, M. Ilan, O. Yarden, Diversity and potential antifungal properties of fungi associated with a Mediterranean sponge, *Fungal Diversity*. 42 (2010). <https://doi.org/10.1007/s13225-010-0020-x>.
- [77] G. Carr, W. Tay, H. Bottriell, S.K. Andersen, A.G. Mauk, R.J. Andersen, Plectosphaeric acids A, B, and C, Lndoleamine 2,3-dioxygenase inhibitors produced in culture by a marine isolate of the fungus *plectosphaerella cucumerina*, *Organic Letters*. 11 (2009). <https://doi.org/10.1021/ol900972j>.
- [78] A.K. Vala, Intra-and extracellular biosynthesis of gold nanoparticles by a marine-derived fungus *Rhizopus oryzae*, *Synthesis and Reactivity in Inorganic, Metal-Organic and Nano-Metal Chemistry*. 44 (2014). <https://doi.org/10.1080/15533174.2013.799492>.
- [79] A.D. Pilevneli, S.S. Ebada, B. Kaşkatepe, B. Konuklugil, Penicic acids H-J, three new mycophenolic acid derivatives from the marine-derived fungus: *Rhizopus oryzae*, *RSC Advances*. 11 (2021). <https://doi.org/10.1039/d1ra07196c>.
- [80] C. Kaewkrajay, S. Putchakarn, S. Limtong, Cultivable yeasts associated with marine sponges in the Gulf of Thailand, South China Sea, Antonie van Leeuwenhoek, *International Journal of General and Molecular Microbiology*. 114 (2021). <https://doi.org/10.1007/s10482-021-01518-6>.
- [81] A. Poli, A. Vizzini, V. Prigione, G.C. Varese, Basidiomycota isolated from the Mediterranean Sea – Phylogeny and putative ecological roles, *Fungal Ecology*. 36 (2018) 51–62. <https://doi.org/10.1016/j.funeco.2018.09.002>.
- [82] C.F. Chang, C.F. Lee, K.Y. Lin, S.M. Liu, Diversity of yeasts associated with the sea surface microlayer and underlying water along the northern coast of Taiwan, *Research in Microbiology*. 167 (2016). <https://doi.org/10.1016/j.resmic.2015.08.005>.
- [83] Y. Gao, X. Du, W. Xu, R. Fan, X. Zhang, S. Yang, X. Chen, J. Lv, Z. Luo, Fungal Diversity in Deep Sea Sediments from East Yap Trench and Their Denitrification Potential, *Geomicrobiology Journal*. (2020). <https://doi.org/10.1080/01490451.2020.1789778>.
- [84] M. Gadanho, J.M. Almeida, J. Paulo, S. Sampaio, S. Sampaio, Assessment of yeast diversity in a marine environment in the south of Portugal by microsatellite-primed PCR, n.d. www.crem.fct.unl.pt.
- [85] E. Breyer, M. Böhm, M. Reitbauer, C. Amano, M. Heitger, F. Baltar, Autofluorescence is a common trait in different oceanic fungi, *Journal of Fungi*. 7 (2021). <https://doi.org/10.3390/jof7090709>.
- [86] M.M. Francis, V. Webb, G.C. Zuccarello, Marine yeast biodiversity on seaweeds in New Zealand waters, *New Zealand Journal of Botany*. 54 (2016) 30–47. <https://doi.org/10.1080/0028825X.2015.1103274>.
- [87] J.W. Fell, Sakaguchia Y: Yamada, Maeda & Mikata (1994), in: *The Yeasts*, Elsevier, 2011: pp. 1541–1544. <https://doi.org/10.1016/B978-0-444-52149-1.00128-2>.
- [88] W. Guo, S. Wang, N. Li, F. Li, T. Zhu, Q. Gu, P. Guo, D. Li, Saroclides A and B, Cyclic Dipeptides from the Mangrove-Derived Fungus *Sarocladium kilense* HDN11-112, *Journal of Natural Products*. 81 (2018). <https://doi.org/10.1021/acs.jnatprod.7b00644>.
- [89] K.C. Hu, M.Y. Xu, H.J. Li, J. Yuan, G. Tang, J. Xu, D.P. Yang, W.J. Lan, Discovery of aromadendrane analogues from the marine-derived fungus: *Scedosporium dehoogii* F41-4 by NMR-guided isolation, *RSC Advances*. 6 (2016). <https://doi.org/10.1039/c6ra21142a>.
- [90] I. Risnari, R. Batubara, A. Nuryawan, F.A. Lubis, O. Firmadi, M.A. Maulana, Deterioration of wood exposed to mangrove ecosystem, in: *IOP Conference Series: Earth and Environmental Science*, 2021. <https://doi.org/10.1088/1755-1315/782/3/032031>.
- [91] A.L. Grunwald, C. Cartmell, R.G. Kerr, Auyituaqamides A-D, Cyclic Dipeptides from *Sesquicillium microsporium* RKAG 186 Isolated from Frobisher Bay Sediment, *Journal of Natural Products*. 84 (2021). <https://doi.org/10.1021/acs.jnatprod.0c00966>.
- [92] M. Venkatachalam, L. Gérard, C. Milhau, F. Vinale, L. Dufossé, M. Fouillaud, Salinity and temperature influence growth and pigment production in the marine-derived fungal strain *Talaromyces albobiverticillius* 30548, *Microorganisms*. 7 (2019). <https://doi.org/10.3390/microorganisms7010010>.
- [93] J. xing Li, X. xin Lei, Y. hong Tan, Y. hong Liu, B. Yang, Y. qiu Li, Two new bioactive polyphenols from the soft coral-derived fungus *Talaromyces* sp. SCSIO 041201, *Natural Product Research*. 35 (2021). <https://doi.org/10.1080/14786419.2020.1836632>.
- [94] K. Kim, Y.M. Heo, S. Jang, H. Lee, S.L. Kwon, M.S. Park, Y.W. Lim, J.J. Kim, Diversity of *Trichoderma* spp. In marine environments and their biological potential for sustainable industrial applications, *Sustainability (Switzerland)*. 12 (2020). <https://doi.org/10.3390/su12104327>.
- [95] Y. Zhao, D. Liu, P. Proksch, D. Zhou, W. Lin, Truncateols O-V, further isoprenylated cyclohexanols from the sponge-associated fungus *Truncatella angustata* with antiviral activities, *Phytochemistry*. 155 (2018) 61–68. <https://doi.org/10.1016/j.phytochem.2018.07.017>.
- [96] D. Xu, M. Luo, F. Liu, D. Wang, X. Pang, T. Zhao, L. Xu, X. Wu, M. Xia, X. Yang, Cytochalasan and Tyrosine-Derived Alkaloids from the Marine Sediment-Derived Fungus *Westerdykella dispersa* and Their Bioactivities, *Scientific Reports*. 7 (2017). <https://doi.org/10.1038/s41598-017-12327-1>.