

Figure S1. Generation and identification of *Cmhyd1* mutants in *Cordyceps militaris*. (A) Schematic diagram showing the generation of $\Delta Cmhyd1$ via homologous recombination and southern blot analysis. Bases are numbered starting with the translational initiation codon for *Cmhyd1* (CCM_03537). Hydrophobin_2 was the functional domains in CmHYD1. CCM_03536 and CCM_03538 are the up- and downstream-genes of *Cmhyd1*, respectively. *Hyg* is the hygromycin resistance gene. Probe is for the southern blot hybridization. The primers 1 (Cmhyd1-verti-F/ Cmhyd1-verti-R), 2 (Hyg-F/Hyg-R), and 3 (Cmhyd1-up-F/ Cmhyd1-down-R) were for identification. (B) Confirmation of $\Delta Cmhyd1$ by PCR. PCR1, PCR2, and PCR3 were performed with the primers 1, 2 and 3 respectively. PCR4 and PCR5 were performed with the primers 3F/2R and 2F/3R respectively. WT, the wild-type strain. DNA fragments with lengths of 421, 800, 4443, 2287 and 2956 bp were obtained using primer sets 1, 2, 3, 3F/2R and 2F/3R from the $\Delta Cmhyd1$ strain, respectively. (C) The transcript levels of *Cmhyd1* in the WT and *Cmhyd1* mutants by RT-PCR. Total RNA extraction and cDNA synthesis were performed as described in the experimental procedures. Transcript levels of *Cmhyd1* and *rpb1* were detected with the primers qCmhyd1-F/qCmhyd1-R and qrp1-F/qrp1-R, respectively. The expression levels of *Cmhyd1* relative to *rpb1* were presented by optical density ratio using ImageJ. (D) Southern blot assay validating the *Cmhyd1* gene deletion and complementation strains. Lane 1: WT, Lanes 2: $\Delta Cmhyd1$ strains, Lane 3 and 4: $\Delta Cmhyd1$ complementation strain. Genomic DNAs were digested with *HindIII*/HindIII for probing *Cmhyd1*.

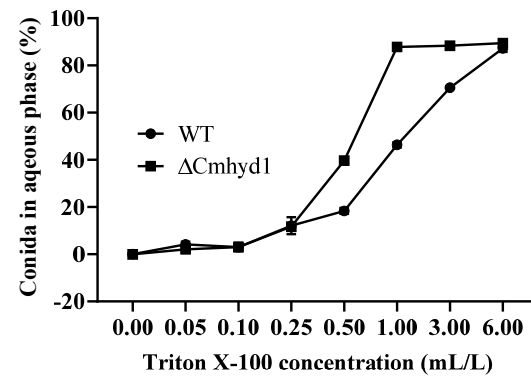


Figure S2. Hydrophobicity of the conidia determined using a MATH assay. Conidia of WT (●) and $\Delta Cmhyd1$ (■) strains were suspended in PBS/hexadecane mixture and shaken vigorously. The amount of conidia in the aqueous phase in the presence of various amounts of Triton X-100 was shown (each point was the mean of three replicates). The more hydrophobic conidia required higher concentrations of detergent to release them from the oil-water interface.

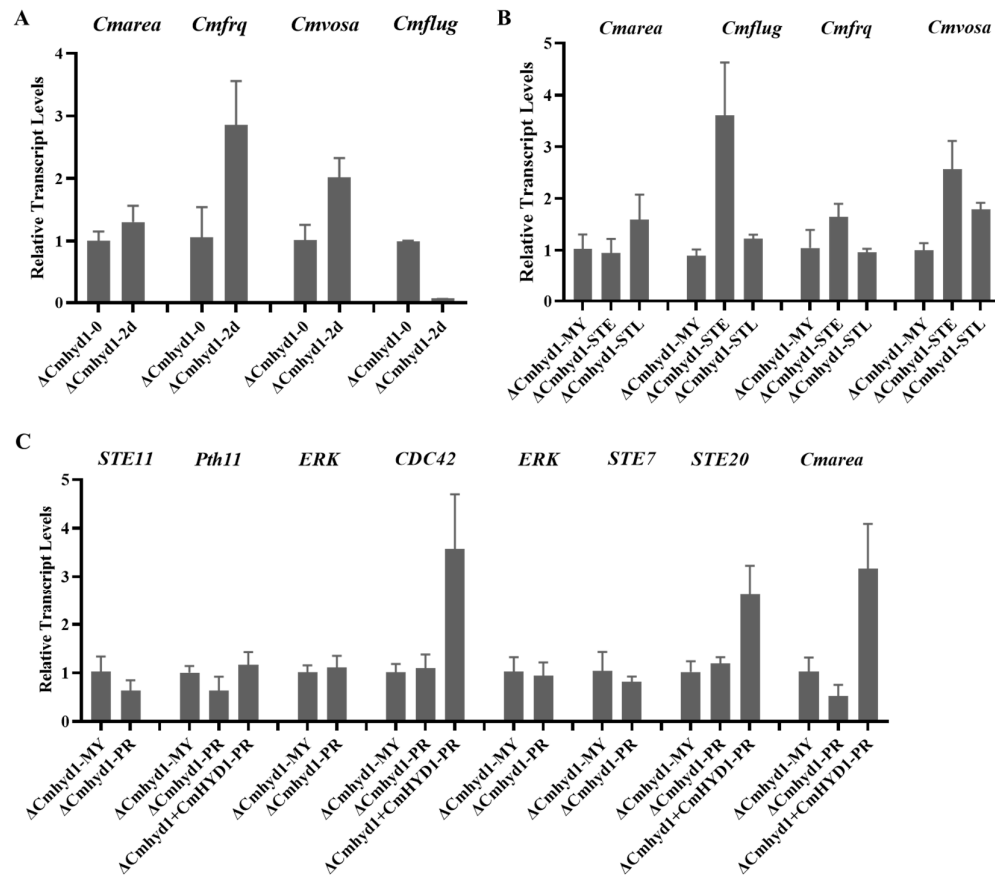


Figure S3. Relative transcript levels in the $\Delta Cmhyd1$ strain. (A) The relative transcript levels of *Cmhyd1* and conidiation-related genes after being exposed to light for 2 d. All results were based on the standard levels of darkness stage of the $\Delta Cmhyd1$ strain ($\Delta Cmhyd1$ -0). (B) The relative transcript levels of *Cmhyd1* and infection-related genes at the different cuticle-bypassing infection stages. All results were based on the standard levels of the MY stage of $\Delta Cmhyd1$ strain ($\Delta Cmhyd1$ -MY). (C) The relative transcript levels of *Cmarea* and fruiting body development related genes at the primordium stage. All results were based on the standard levels of the MY stage of $\Delta Cmhyd1$ strain ($\Delta Cmhyd1$ -MY). Error bars indicate the standard deviation (SD) of three biological replicates with two technical replicates.

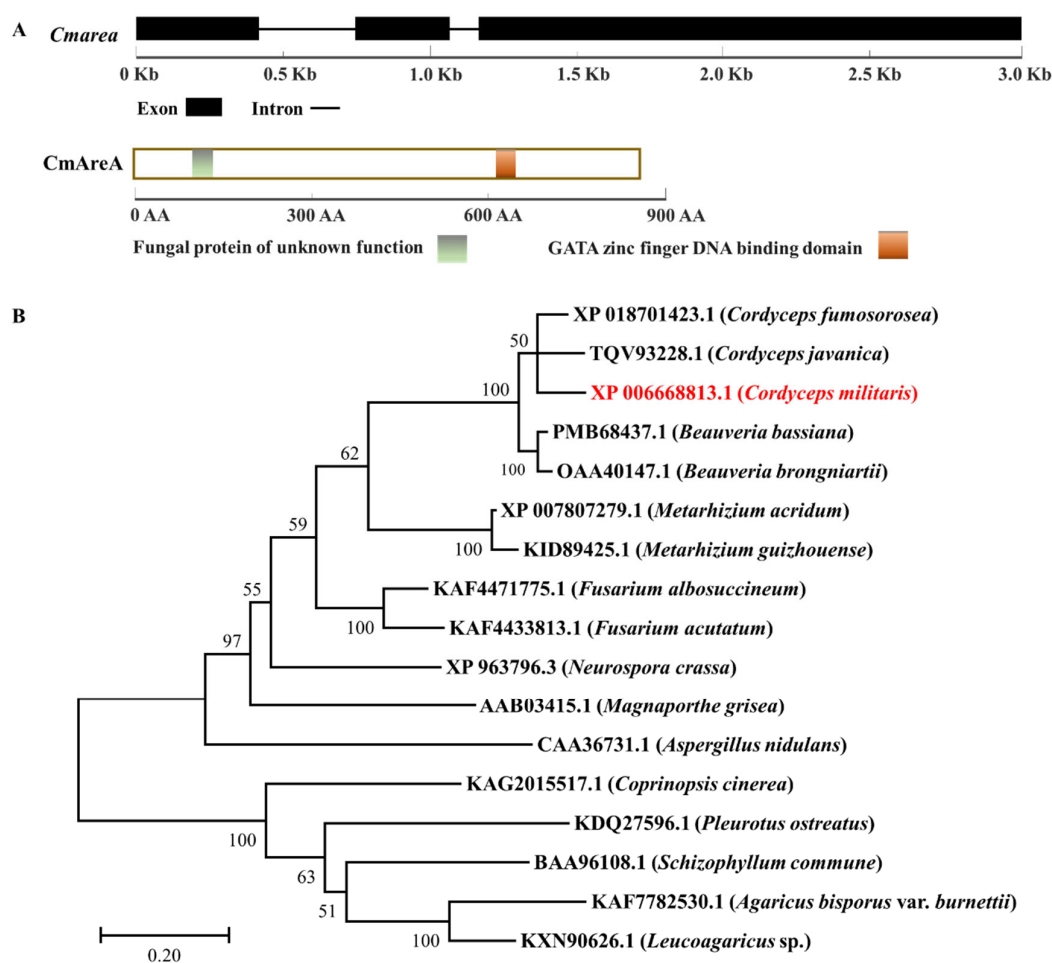


Figure S4. Characterization of the GATA zinc finger (ZnF) transcript factor gene *Cmarea* in *Cordyceps militaris*. (A) The gene structure and protein domains of *CmAreaA*. (B) Phylogenetic relationships of *CmAreaA* (red colored) and its homologs. The amino acid sequences were aligned with MEGAX, and a neighbor-joining tree was generated with 1000 bootstrap replicates using the program MEGAX.

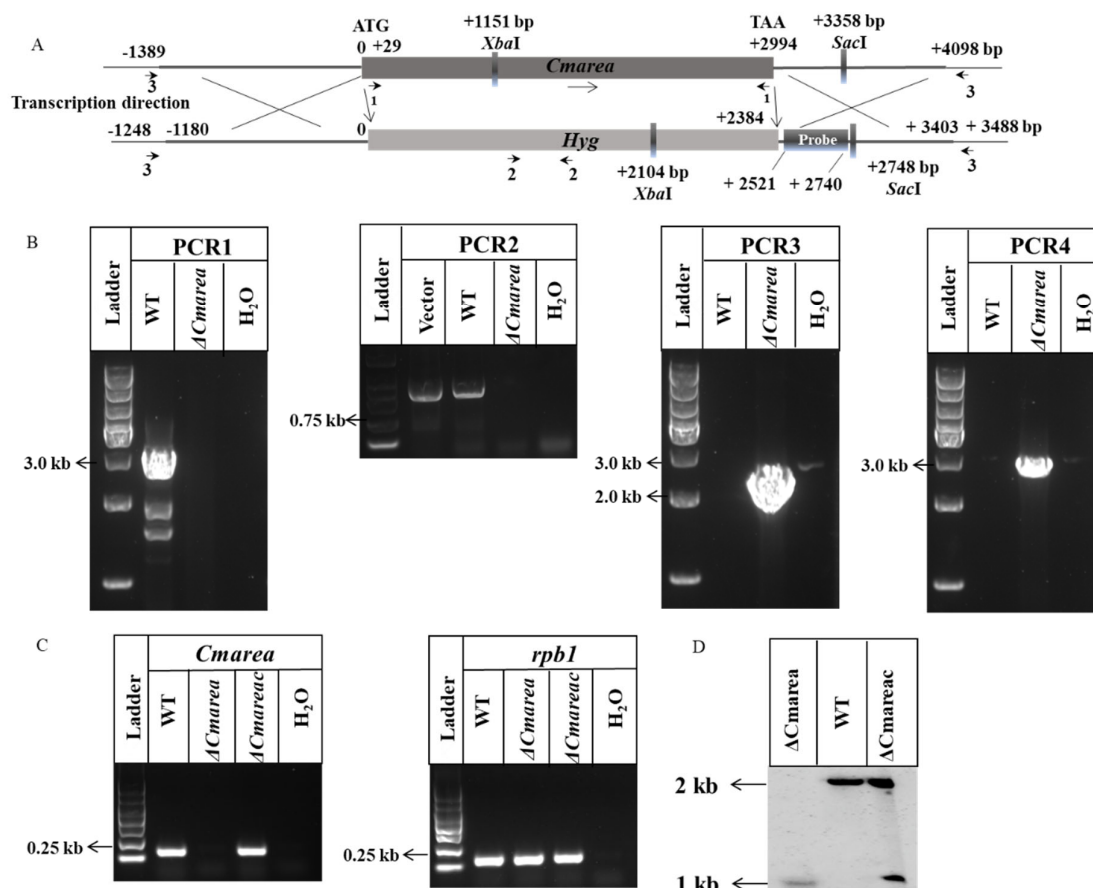


Figure S5. Generation and identification of *Cmarea* mutants in *Cordyceps militaris*. **(A)** Schematic diagram showing the generation of $\Delta Cmarea$ via homologous recombination and southern blot analysis. **(B)** Confirmation of $\Delta Cmarea$ by PCR. PCR1, PCR2, and PCR3 were performed with the primers 1, 2, 3F/2R and 2F/3R respectively. WT, the wild-type strain. DNA fragments with lengths of 2664, 800, 2417 and 3119 bp were obtained using primer sets 1, 2, 3F/2R and 2F/3R from the $\Delta Cmhyd1$ strain respectively. **(C)** The transcript levels of *Cmarea* in the WT, $\Delta Cmarea$ and $\Delta Cmarea$ ac strains using RT-PCR. Total RNA extraction and cDNA synthesis were performed as described in the experimental procedures. Transcript levels of *Cmarea* and *rpb1* were detected with the primers qCmarea-F/qCmarea-R and qrpbl-F/qrpbl-R, respectively. **(D)** Southern blot assay validating the *Cmarea* gene deletion and complementary strains. Lane 1: $\Delta Cmarea$ strain for 1254 bp, Lane 2: WT strain for 2207 bp, Lane 3: $\Delta Cmarea$ complementary strain for 1254 and 2207 bp. Genomic DNAs were digested with Xba I /Sac I for probing *Cmarea* in southern blot hybridization.

Table S1. Primers used in this study.

Primer Name	Primer Sequence (5' to 3')
rpb1-F	CTGTTCCCCCTCCTCCTGTG
rpb1-R	ATGTTGCGGCGATCCTTCTC
qPCR- <i>Cmhyd1</i> -F	CCTCTACTCCAACCCCATCTG
qPCR- <i>Cmhyd1</i> -R	AGGACTGCCTGGTCAATCAC
<i>Cmhyd1</i> -verti-F1	GCAAGTCCTTCTCATTGCTAC
<i>Cmhyd1</i> -verti-R1	GCCTGGTCAATCTGTTTCAGATG
<i>Hyg</i> -verti-F2	CTGTTCGAGAAGTTTCTGATCG
<i>Hyg</i> -verti-R2	CTGATAGAGTTGGTCAAGACC
<i>Cmhyd1</i> -verti-F3	GCTTTCTGCATAGTTTTCTCCATT

<i>Cmhyd1</i> -verti-R3	ACCAAGTGTTACATTAATTGAAAGC
Sothern Blot- <i>Cmhyd1</i> -F	TGGGCTGTTCTTGTCTCTGC
Sothern Blot- <i>Cmhyd1</i> -R	GGCGAAATGTAAACGGAGC
qPCR- <i>Cmare</i> a-F	TGCCATCCATCTCGTCCAC
qPCR- <i>Cmare</i> a-R	TGAGCCACTCCCATTCTG
<i>Cmare</i> a-verti-F1	AACCAGCAACGCATGGAAAACCTC
<i>Cmare</i> a-verti-R1	TTACAGACTCATCGTGAGCCACTC
<i>Cmare</i> a-verti-F3	TTCCAACAGCCAGACAAACAAGAC
<i>Cmare</i> a-verti-R3	ATAGCCTTGTTGCTACGTTTACGAG
Sothern Blot- <i>Cmare</i> a-F	CTACTCTTGCGTGTTTCTTTTATAC
Sothern Blot- <i>Cmare</i> a-R	GTAGTCCAAGGAGATTAAGCGC
<i>Cmare</i> a-verti-F	AACCAGCAACGCATGGAAAACCTC
<i>Cmare</i> a-verti-R	TTACAGACTCATCGTGAGCCACTC
Biotin probe 1	TTCGCCTTCTCACCATCCCAAGATACTCCCATCAGCCAAGATGCAA
Biotin probe 2	GAGGACAACGATCCTCTCCTTGATACTCACAGGCCTGACTTCAAA C
Unlable	CAGTGAGACGGGACCGGCATCGATAGAACGCTGCAGCCACTGCC AA