

Supplementary informations

1. Effect of *P. aeruginosa* siderophore pyochelin and pyoverdine on *A. fumigatus*.

a. Strains used in this study

Souches	Génotype	Reference
Parental strain: CEA17 Δ akuB ^{KU80}	CEA17 Δ akuB ^{KU80}	[1]
Parental strain: ATCC 46645	Souche sauvage	American Type Culture Collection
Δ hapX	ATCC 46645 Δ hapX::hph	[2]
Δ sidC	ATCC 46645 Δ sidC::hph	[3]
Δ sidD	ATCC 46645 Δ sidD::hph	[3]
Δ sidF	ATCC 46645 Δ sidF::hph	[3]
Parental strain: AF14	Souche sauvage	[4]
Δ zrfA Δ zrfB Δ zrfC	AF14	[5]

b. Medium composition

The medium used to test the effect of pyochelin on *A. fumigatus* growth was minimal medium (MM) containing 10g/L glucose, 0.92g/L ammonium tartrate dibasic, 1ml/L trace-element-solution and 20ml/L 50x salt solution, pH 6.5. Trace-element-solution contained Na₂B₄O₇ x10 H₂O 40 mg/L, CuSO₄ x5 H₂O 400 mg/L, MnSO₄ x4 H₂O 800 mg/L, Na₂MoO₄ x10 H₂O 800 mg/L, ZnSO₄ x7 H₂O 8g/L, 5 μ M FeCl₃ (2 M stock solution in 1 M HCl) and 0.2 mM HCl. Salt solution contained KCl 26 g/L, MgSO₄ x7 H₂O 26 g/L and KH₂P₄O₇ 76 g/L). To get an excess of iron in MM, 250 μ M FeCl₃ were added.

c. Minimal Inhibitory Concentration (MIC) determination

The susceptibility of *A. fumigatus* to pyochelin or pyoverdine was undertaken by measuring the MIC in 96-well flat bottom plates. Briefly, the assay mixture was prepared by adding 1 volume of conidial suspension (1x10⁵ conidia/mL in 0.05% Tween20-water) to 12 volumes of assay medium (MM containing 0.1% Tween 20, and 1% methanol. Twofold dilutions of pyochelin or pyoverdine were prepared with this assay mixture. The plates were incubated at 37°C for 20 h. Biofilm biomass was assessed using crystal violet protocol previously described in [6]. Briefly, after incubation, the culture medium was removed and the biofilms were washed with water. 130 ml of 0.01% (w/v) crystal violet solution (Sigma Aldrich) was added to each well for 20 min at room temperature. The solution was then removed and the biofilms were washed with water until the supernatant was clear. The plates were air-dried. The biofilms were destained by the addition of 130 ml of 30% acetic acid in for 20 min under agitation. The acetic acid was transferred to clean 96 well plates and the absorbance at 560 nm was measured (Thermo Labsystems Multiskan EX).

d. Penetration of pyochelin in *A. fumigatus* cell

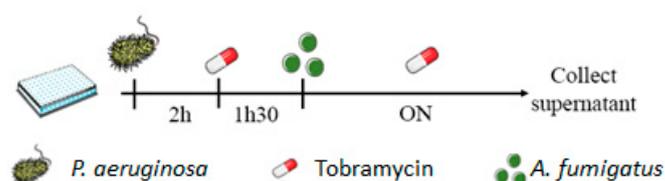
To test the penetration of pyochelin-4-nitrobenzo[1,2,5]oxadiazole (PCH-NBD) in *A. fumigatus*, swollen conidia were obtained after 4 h 30 min incubation into MM at 37°C under shaking and then incubated with 250 µM PCH-NBD (pyochelin MIC) in MM for 1 h 30 min at 37°C. the fluorescence was observed by Epifluorescence microscopy using the filter excitation 472 ± 30 nm, emission 520 ± 30 nm and dichroic 502-730 nm.

e. Reactive oxidants species (ROS) and reactive nitrogen species (RNS) assays

To test the induction of ROS and RNS in *A. fumigatus* cells by pyochelin, we used the $O_2^{\cdot-}$ fluorescent probe using 2',7'-dichlorodihydrofluorescein diacetate (H2DCFDA) and dihydrorhodamine 123 (DHR123) probes which become fluorescent in presence of ROS and RNS respectively following the method described previously for phenazines inducing ROS and RNS in *A. fumigatus* [6]. Briefly, H2DCFDA and DHR123, ROS and RNS fluorescent probes respectively, were added to swollen conidia followed by the addition of 250 µM pyochelin for 1 h in the darkness at 37°C. Fluorescence of the respective reduced products DCF and Fluorescein 123 was visualized using a fluorescence microscope Leica DMLB with Leica filter I3, a filter excitation BP 450-490 nm, FT 510 nm and emission LP 515 nm. For control, swollen conidia were incubated with H2DCFDA and DHR123 in absence of pyochelin.

2. Ex vivo human macrophage model of co-infection with *P. aeruginosa* and *A. fumigatus*

In multi-well plates, human GM-CSF or M-CSF differentiated monocyte-derived macrophages [7] were growing in RPMI 1640 supplemented with 10% fetal bovine serum (Sigma) until confluence and then infected with *P. aeruginosa* (2.5×10^5 CFU/mL or MOI 0.05) for 2 h. Bacteria were eliminated by renewing the medium and adding tobramycin, an aminoglycoside antibiotic used in clinic to treat *P. aeruginosa* infection. After 90 min, supernatants were removed and macrophages infected with *A. fumigatus* (2.5×10^6 conidia/mL or MOI 10). Cells were incubated overnight and supernatant collected and used for cytokines quantification [7].



References

1. Ferreira, M.E. da S.; Kress, M.R.V.Z.; Savoldi, M.; Goldman, M.H.S.; Härtl, A.; Heinekamp, T.; Brakhage, A.A.; Goldman, G.H. The akuBKU80 Mutant Deficient for Nonhomologous End Joining Is a Powerful Tool for Analyzing Pathogenicity in *Aspergillus fumigatus*. *Eukaryot. Cell* **2006**, *5*, 207–211.
2. Schrettl, M.; Beckmann, N.; Varga, J.; Heinekamp, T.; Jacobsen, I.D.; Jöchl, C.; Moussa, T.A.; Wang, S.; Gsaller, F.; Blatzer, M.; et al. HapX-mediated adaption to iron starvation is crucial for virulence of *Aspergillus fumigatus*. *PLoS Pathog.* **2010**, *6*, e1001124.
3. Schrettl, M.; Bignell, E.; Kragl, C.; Sabiha, Y.; Loss, O.; Eisendle, M.; Wallner, A.; Arst, H.N.J.; Haynes, K.; Haas, H. Distinct roles for intra- and extracellular siderophores during *Aspergillus fumigatus* infection. *PLoS Pathog.* **2007**, *3*, 1195–1207.
4. Vicentefranqueira, R.; Moreno, M.A.; Leal, F.; Calera, J.A. The *zrfA* and *zrfB* genes of *Aspergillus fumigatus* encode the zinc transporter proteins of a zinc uptake system induced in an acid, zinc-depleted environment. *Eukaryot. Cell* **2005**, *4*, 837–848.
5. Amich, J.; Vicentefranqueira, R.; Mellado, E.; Ruiz-Carmuega, A.; Leal, F.; Calera, J.A. The *ZrfC* alkaline zinc transporter is required for *Aspergillus fumigatus* virulence and its growth in the presence of the Zn/Mn-chelating protein calprotectin. *Cell. Microbiol.* **2014**, *16*, 548–564.
6. Briard, B.; Bomme, P.; Lechner, B.E.; Mislin, G.L.A.; Lair, V.; Prévost, M.-C.; Latgé, J.-P.; Haas, H.; Beauvais, A. *Pseudomonas aeruginosa* manipulates redox and iron homeostasis of its microbiota partner *Aspergillus fumigatus* via phenazines. *Sci. Rep.* **2015**, *5*, 8220.
7. Camilli, G.; Eren, E.; Williams, D.L.; Amanianda, V.; Meunier, E.; Quintin, J. Impaired phagocytosis directs human monocyte activation in response to fungal derived β -glucan particles. *Eur. J. Immunol.* **2018**, *48*, 757–770.