

Supplementary information for:

Downscaling industrial-scale syngas fermentation to simulate frequent and irregular dissolved gas concentration shocks

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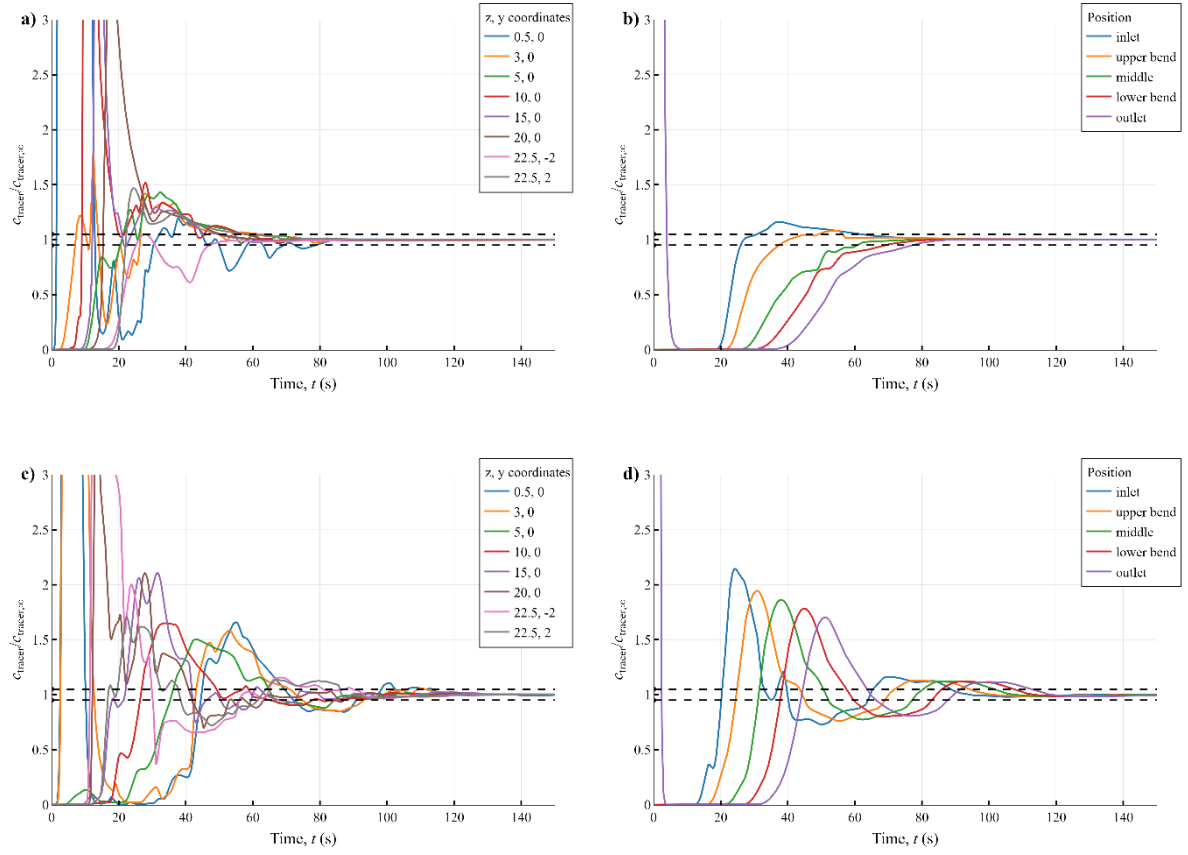


Figure S1. Mixing study in the EL-GLR. A tracer with the same properties as the fluid phase was injected at the outlet of the downcomer and its normalized concentration recorded at several positions in the riser (a, c – coordinates in m) and downcomer (b, d). The 95% mixing time t_m is obtained when all curves are within the dashed lines representing a 5% deviation around the final tracer concentration. t_m was recorded for a case without reaction (only the hydrodynamic model was enabled) (a, b) and a case including mass transfer and reaction at 25 g L⁻¹ biomass without CO₂ production (c, d). See (Puiman et al., 2022) for more details about the hydrodynamic model and the reactor geometry.

t_m without reaction is around 80 s, while with reaction it is around 115 s. This difference is explained by the lower gas hold-up in the case with reaction since the syngas is consumed. The circulation time t_c is estimated from the mixing time as $t_m \sim 3\text{-}5 t_c$ (van't Riet and van der Lans, 2011).

Table S1. Characteristics of the scale-down simulator (Garcia-Ochoa and Gomez, 2009; Zanghi et al., 2017).

Reactor type and dimensions	Applikon 3L Glass bioreactor
Volume reactor (L)	3
Height (m)	0.234
Diameter (m)	0.13
Impeller diameter, d_i (m)	0.06
Power number, N_{Po}	1.5
Liquid volume, V_L (L)	2
Mass transfer parameters	
α	0.783
β	0.459
Fixed operating conditions	
Temperature (°C)	37
Pressure (kPa)	101
pH	5 (for solventogenesis)
Ethanol concentration (g L ⁻¹)	50 (if possible because of inhibition)
Acetate concentration (g L ⁻¹)	12
Dilution rate (h ⁻¹)	0.021
Gas flow rate (vvm)	0.05

Statistical analysis of lifelines

It was determined how many lifelines were required to obtain statistically independent probability density distributions of the residence time in peaks and valleys. This was done by calculating the Kullback-Leibler divergence D_{KL} (Equation S1) or the relative entropy for the variables of interest (e.g. the residence time distribution in peaks and valleys for CO and H₂). D_{KL} is a common tool in data sciences to measure dissimilarity between two probability distributions (Bishop, 2006). By minimizing D_{KL} of a distribution with respect to a given (“true”) distribution, it can be determined how close a distribution is to the true distribution (Akaike, 1998; Bishop, 2006). As a “true” distribution (with $N_p, N_{t_c} \rightarrow \infty$) was not available, it was assumed that the distribution obtained with all peaks and valleys, $P(N_p N_{t_c})_\infty$, is sufficiently representative of the “true” distribution.

$$D_{KL}\left(P(N_p N_{t_c})_\infty \parallel P(N_p N_{t_c})\right) = \sum_{i=1}^k P_i(N_p N_{t_c})_\infty \ln \left[\frac{P_i(N_p N_{t_c})_\infty}{P_i(N_p N_{t_c})} \right] \quad (S1)$$

From the analysis became clear that with $N_p N_{t_c} \sim O(10^5)$ the D_{KL} approaches zero (Figure S2) for the probability distributions describing the duration of a peak in CO concentration. One could argue that even around $N_p N_{t_c} \sim O(10^4)$ the information gain by extending the set of lifelines is small. These results give an indication of the required number of particles and runtime for lifeline analysis of environmental fluctuations, which is considerably lower than the numbers typically applied for such analysis (e.g. $N_p N_{t_c} > O(10^6)$ (Haringa et al., 2016; Kuschel and Takors, 2020; Siebler et al., 2019)). As an exception, McClure et al. (2016) only used $N_p N_{t_c} \sim O(10^4)$ while obtaining statistically satisfactory results. It must be noted that our analysis is only applicable when particles act as passive tracers. When the simulated biomass particles affect the Eulerian flow or concentration field (e.g. two-way coupling with metabolic models) (Haringa et al., 2018; Lapin et al., 2004), a higher N_p is required to achieve a sufficiently homogeneous spatial biomass distribution (Haringa et al., 2017). In syngas fermentation modelling, this could be applicable if one wants to study the effect of local microbial CO₂ production or consumption, using a detailed kinetic metabolic model.

By calculating D_{KL} , we saved a significant amount of simulation and data processing time: the analyses with 10 and 25 g L⁻¹ (650 s and 40,000 particles) only required 20 GB of data instead of 200 GB with the 5 g L⁻¹ case (1000 s and 160,000 particles). Next to checking whether the Lagrangian averages matches the Eulerian average (Siebler et al., 2019), which is – in case of sufficient mixing – usually after 3 mixing times per particle, we suggest calculating D_{KL} as well to check variability of the results.

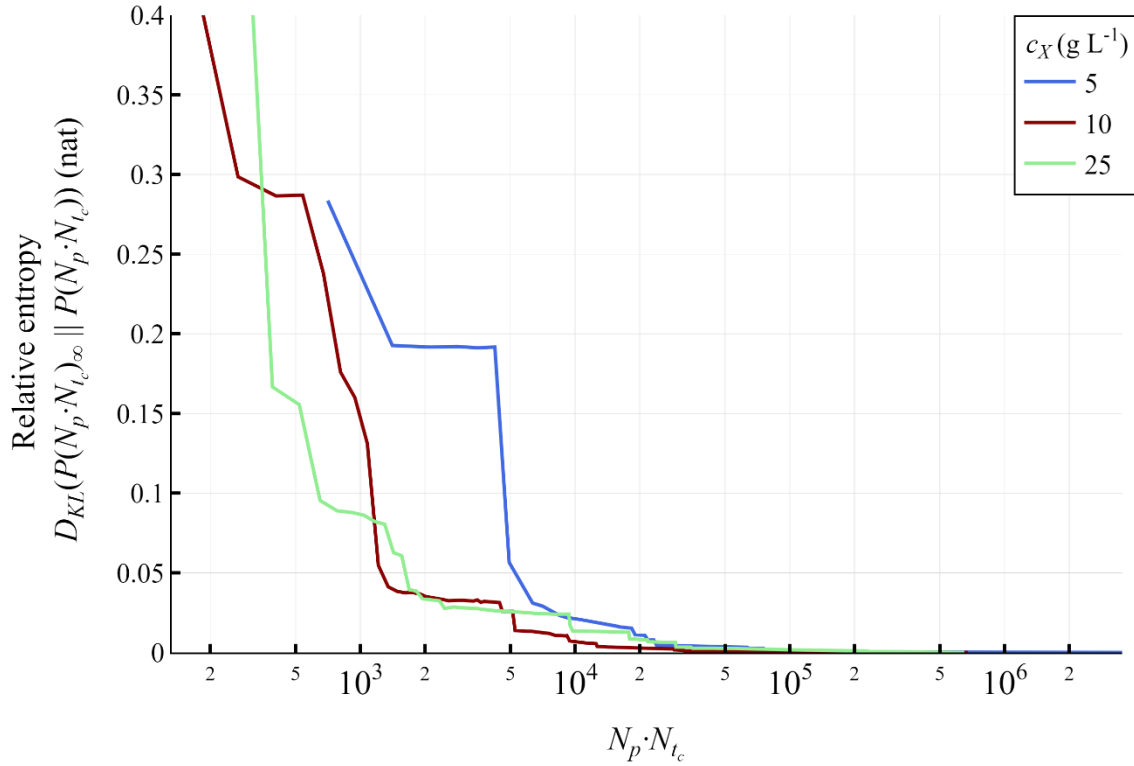


Figure S2. Kullback-Leibler divergence D_{KL} determined using an increasing $N_p N_{t_c}$ of the residence time distribution in a CO peak. Blue: D_{KL} obtained from distributions derived from lifelines with 5 g L⁻¹ of biomass with $(N_p N_{t_c})_\infty \sim 3 \cdot 10^6$. Green: 10 g L⁻¹ and $(N_p N_{t_c})_\infty \sim 6 \cdot 10^5$, Red: 25 g L⁻¹ and $(N_p N_{t_c})_\infty \sim 6 \cdot 10^5$.

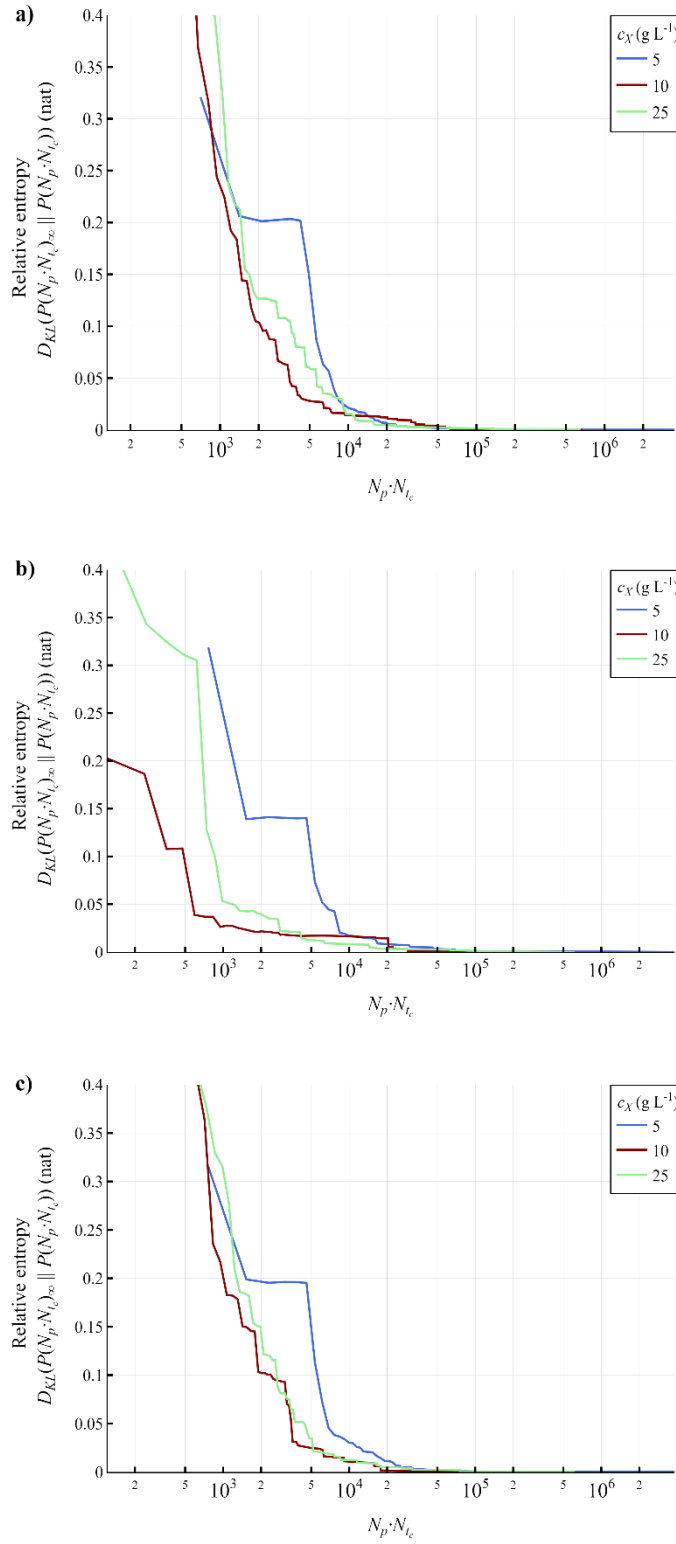


Figure S3. Kullback-Leibler divergence D_{KL} determined using an increasing $N_p N_{t_c}$ of the residence time distribution a) in a CO valley, b) H₂ peak, and c) H₂ valley. Blue: D_{KL} obtained from distributions derived from lifelines with 5 g L⁻¹ of biomass with $(N_p N_{t_c})_{\infty} \sim 3 \cdot 10^6$. Green: 10 g L⁻¹ and $(N_p N_{t_c})_{\infty} \sim 6 \cdot 10^5$, Red: 25 g L⁻¹ and $(N_p N_{t_c})_{\infty} \sim 6 \cdot 10^5$.

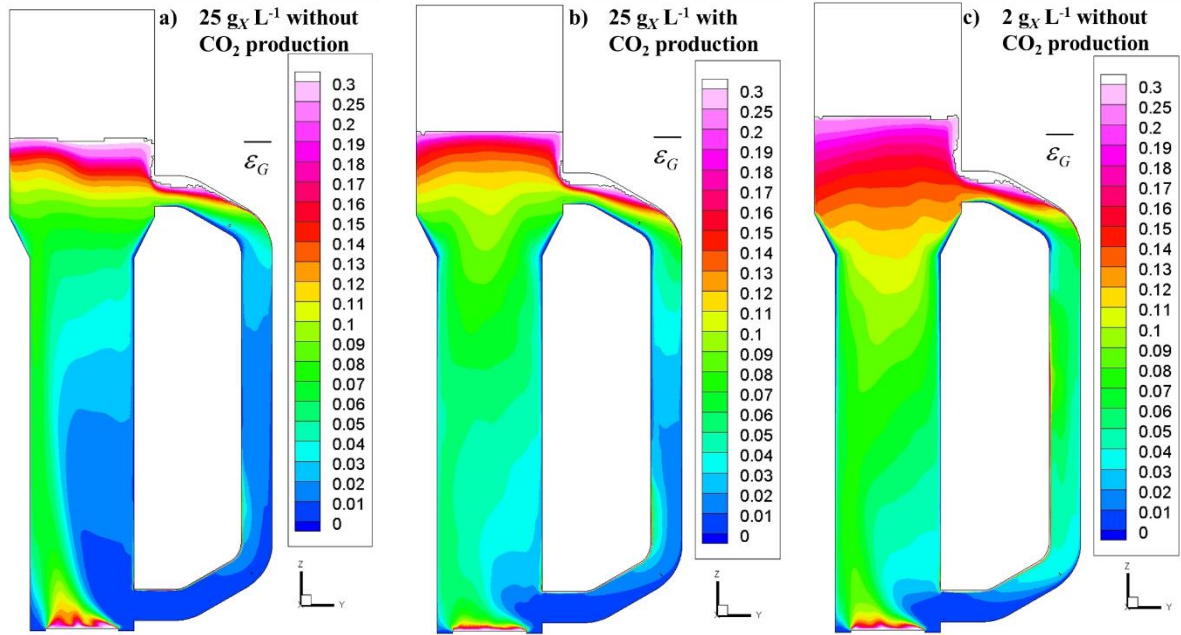


Figure S4. Surface plots of the 200 s time-averaged gas hold-up in the zy-plane ($x = 0$) of the EL-GLR, obtained a) with $25 \text{ g}_X \text{ L}^{-1}$ biomass without including CO_2 production, b) $25 \text{ g}_X \text{ L}^{-1}$ biomass while including CO_2 production and c) $2 \text{ g}_X \text{ L}^{-1}$ biomass without CO_2 production.

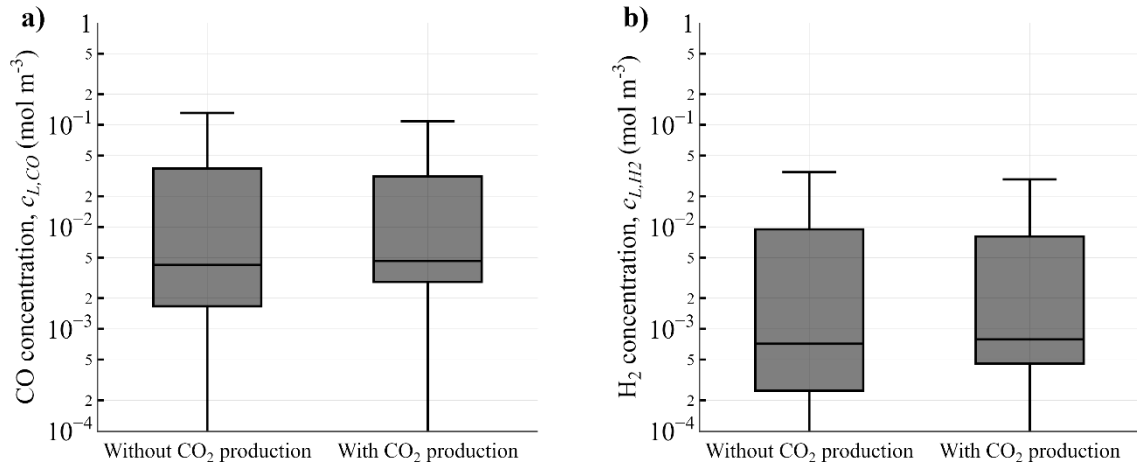


Figure S5. Variation of a) dissolved CO and b) dissolved H_2 concentrations within the EL-GLR reactor volume in the cases without and with CO_2 production (with $25 \text{ g}_X \text{ L}^{-1}$ biomass). In the boxplot each quartile represents 25% of the dispersion volume with a specific concentration.

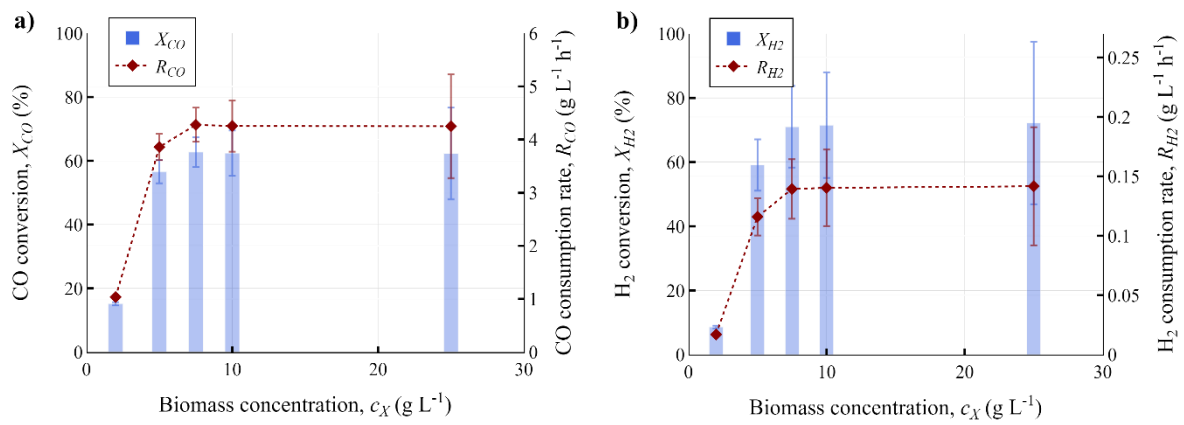


Figure S6. For the varying biomass concentrations the Eulerian results for a) CO and b) H₂ conversion (blue bars) and consumption rates (red markers) are given.

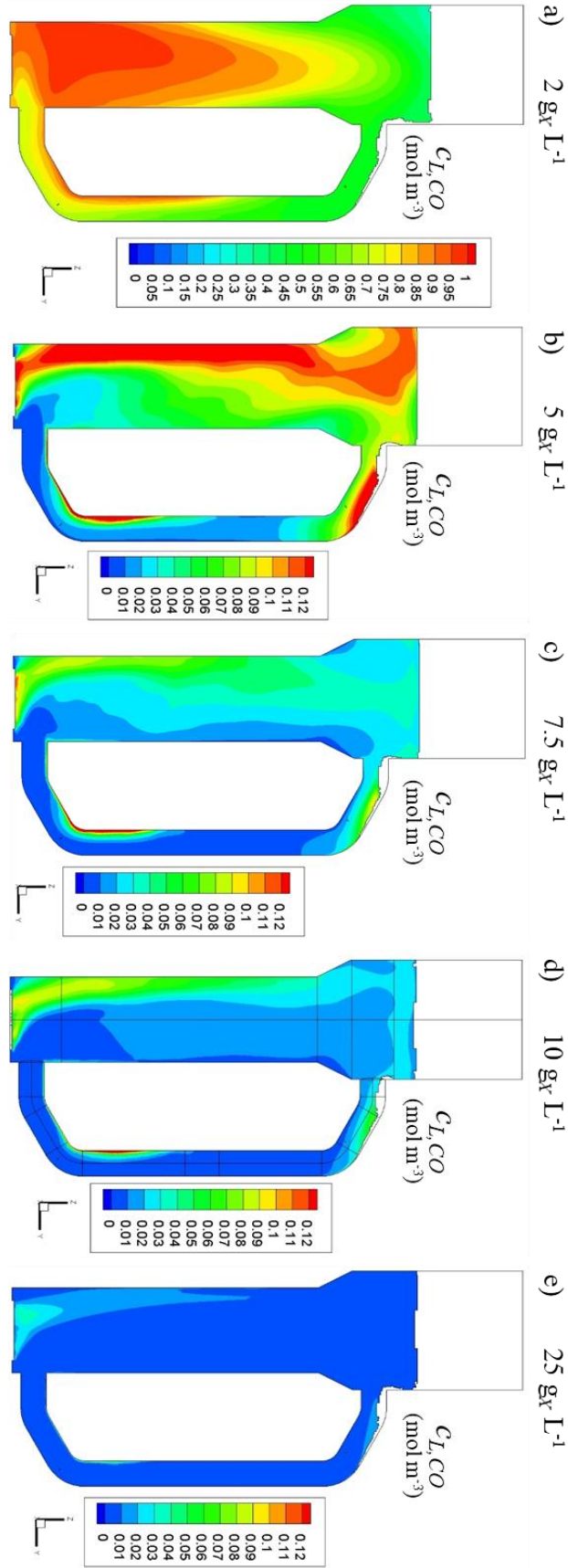


Figure S7. Surface plots of the dissolved CO concentration in the zy -plane ($x=0$) of the EL-GLR, for several concentrations of biomass (2, 5, 7.5, 10, 25 g L^{-1}) (a-e).

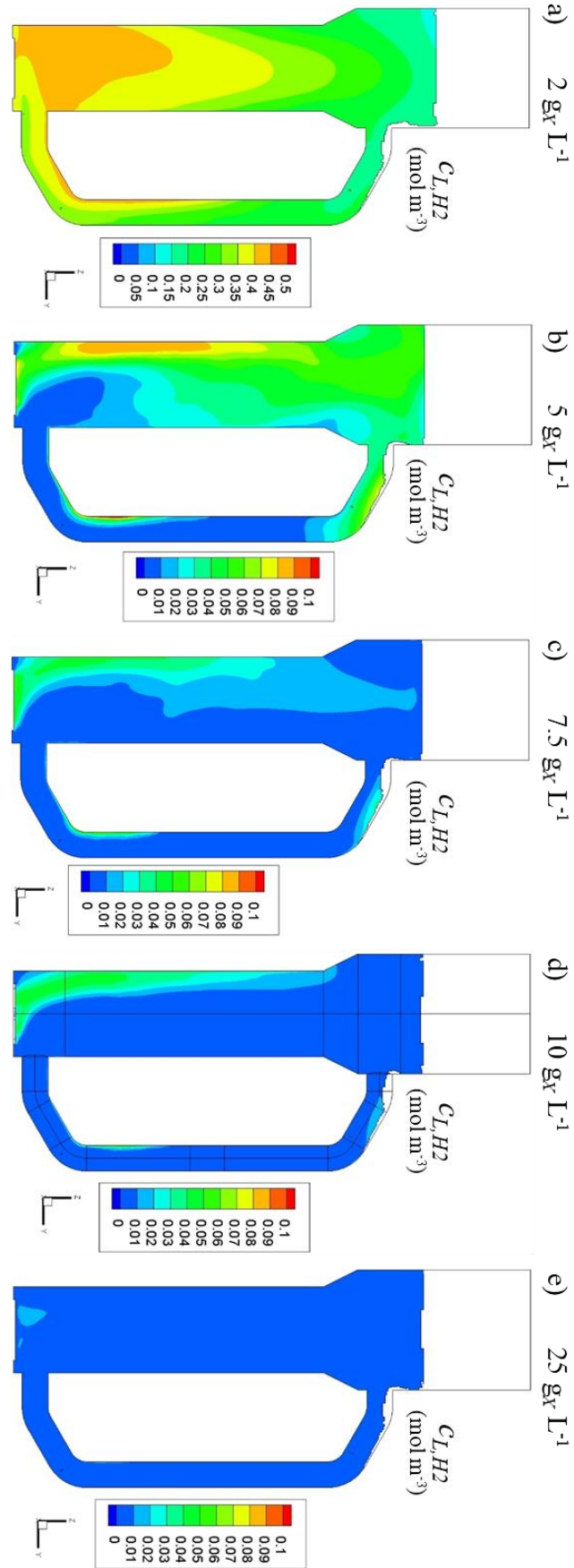


Figure S8. Surface plots of the dissolved H_2 concentration in the zy -plane ($x=0$) of the EL-GLR, for several concentrations of biomass (2, 5, 7.5, 10, 25 $g\ L^{-1}$) (a-e).

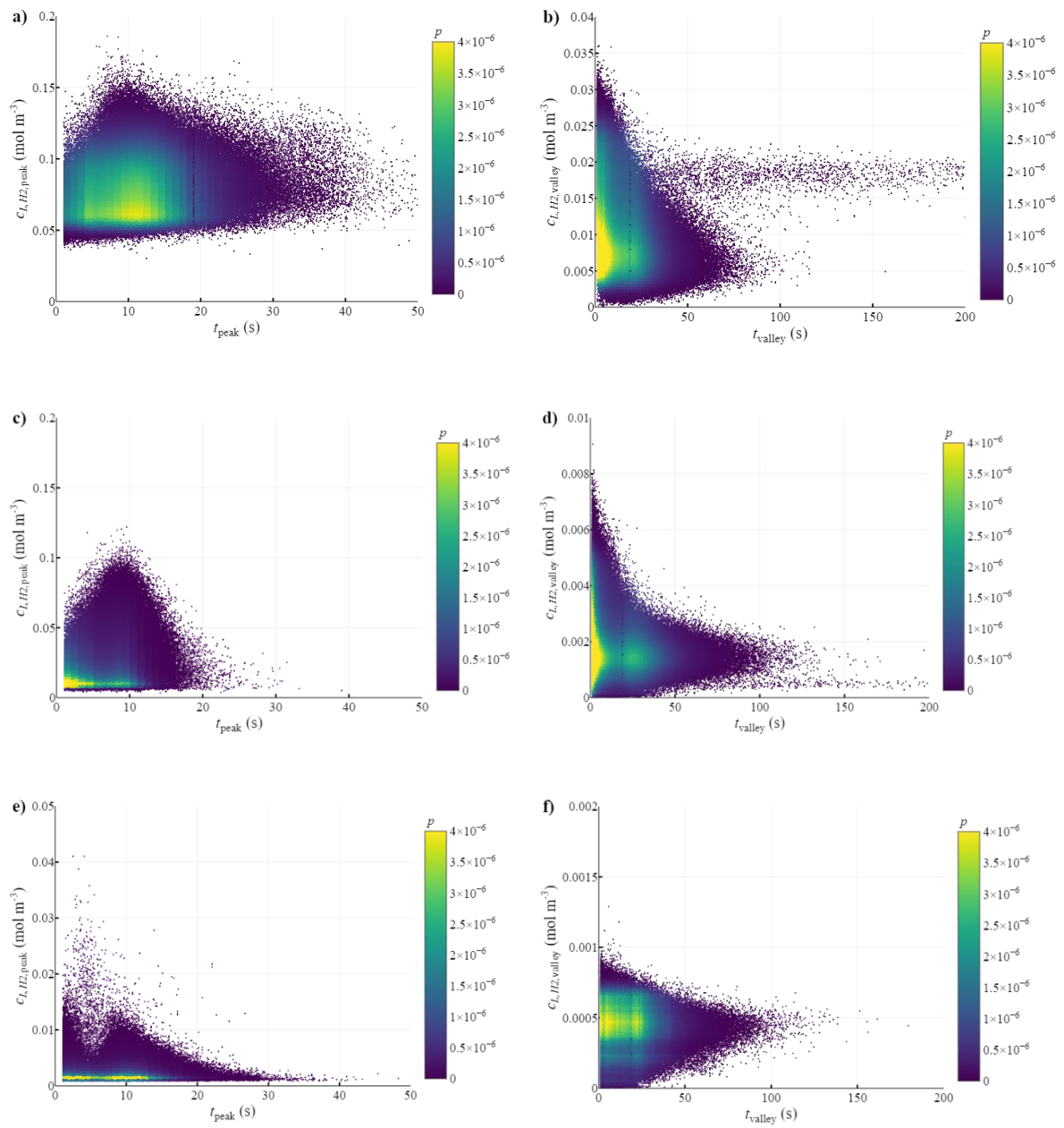


Figure S9. The probability of a microbe to experience a specific H_2 concentration peak or valley. Each dot represents a peak or valley with such a concentration and time, and is coloured by the probability of occurrence. Each row represents data obtained with a specific concentration: (a, b) 5, (c, d) 10 and (e, f) 25 g L⁻¹, with the peaks in the left column (a, c, e) and the valleys at the right (b, d, f).

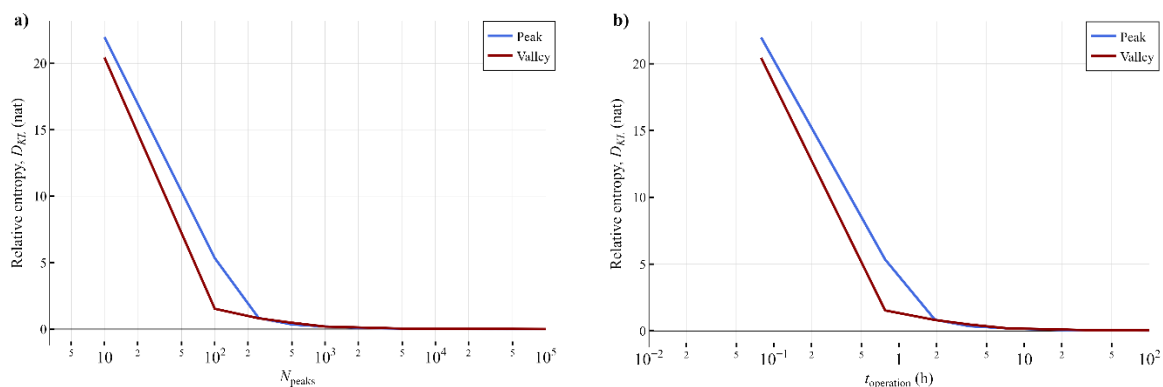


Figure S10. Minimization of the Kullback-Leibler divergence for a) the number of peaks used as input for the scale-down simulator, and b) the operational time of the scale-down simulator. D_{KL} was calculated by comparing the probability distribution of the residence time in the peak (blue line) or valley (red line) from the scale-down simulator with their respective CFD-derived counterpart. The probability distributions from the scale-down simulator were derived without assuming mass transfer and reaction.

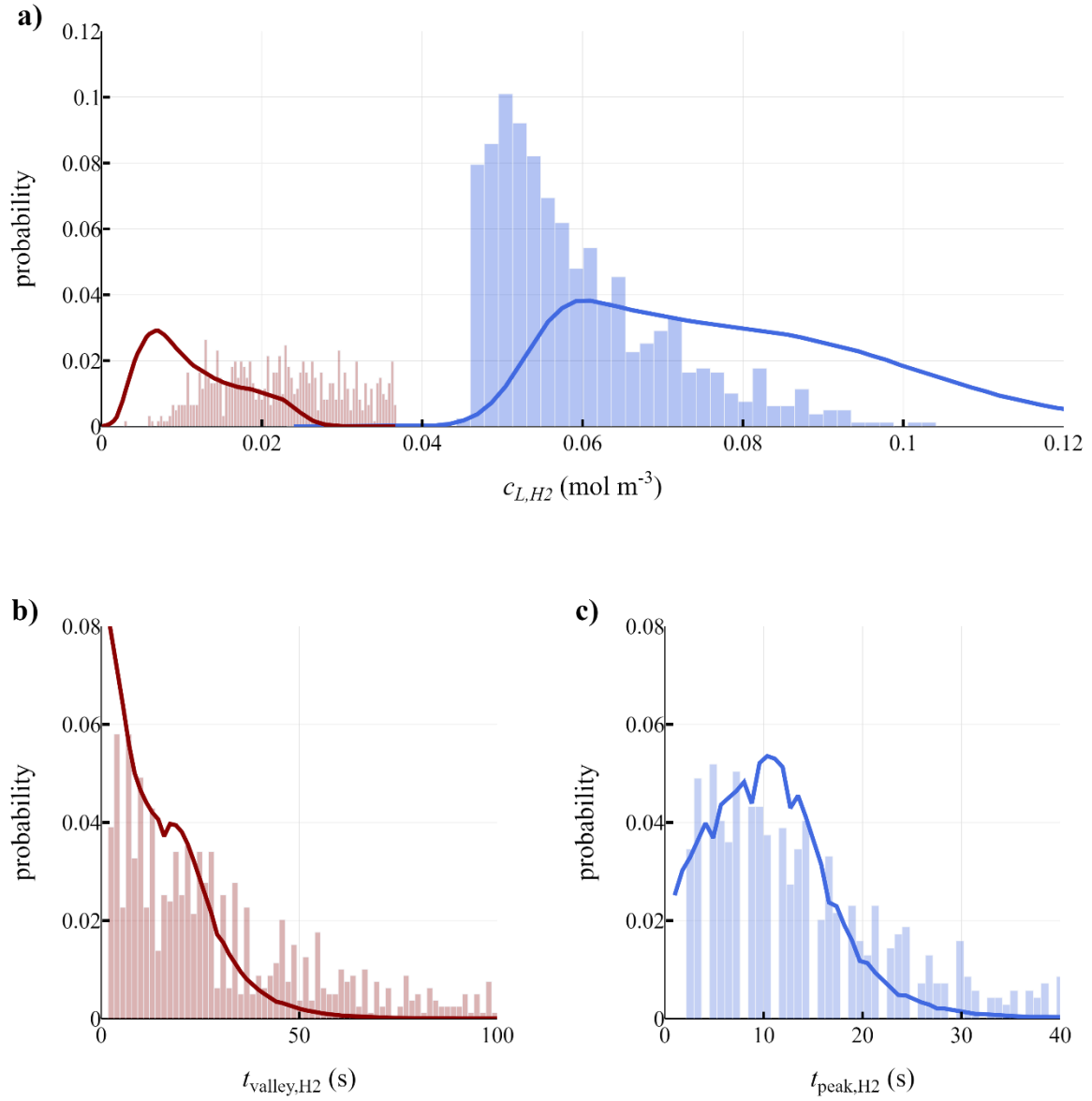


Figure S11. Comparison of the probability density functions obtained by the scale-down simulator (bars) with the CFD results (lines). Probability density functions for a) the concentration of dissolved H_2 during the peaks (blue) and the valleys (red), as well as the residence time in a b) valley or c) peak, obtained with 5 g L^{-1} biomass and simulating 2000 peaks.

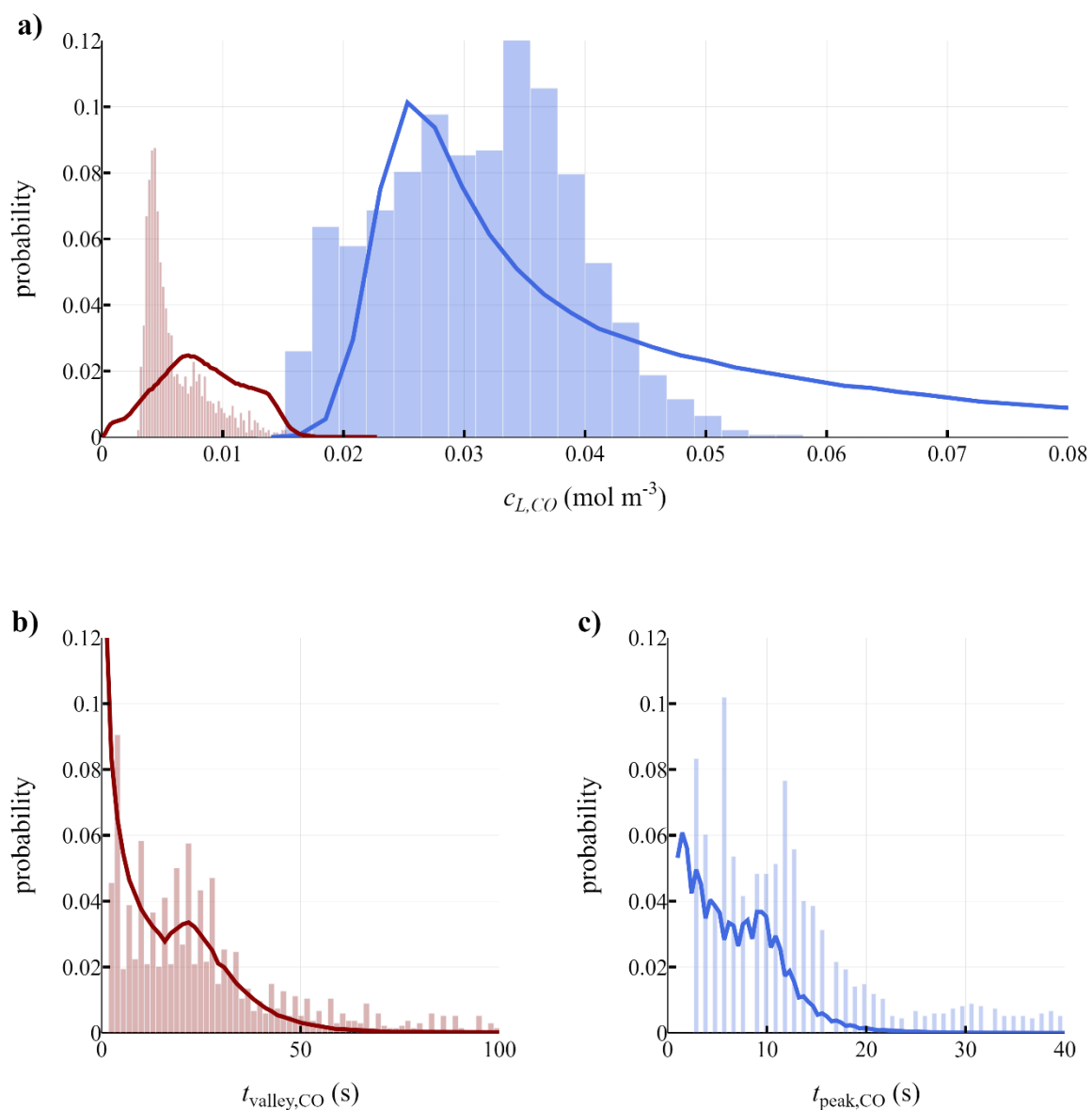


Figure S12. Comparison of the probability density functions obtained by the scale-down simulator (bars) with the CFD results (lines). Probability density functions for a) the concentration of dissolved CO during the peaks (blue) and the valleys (red), as well as the residence time in a b) valley or c) peak, obtained with 10 g L⁻¹ biomass and simulating 2000 peaks.

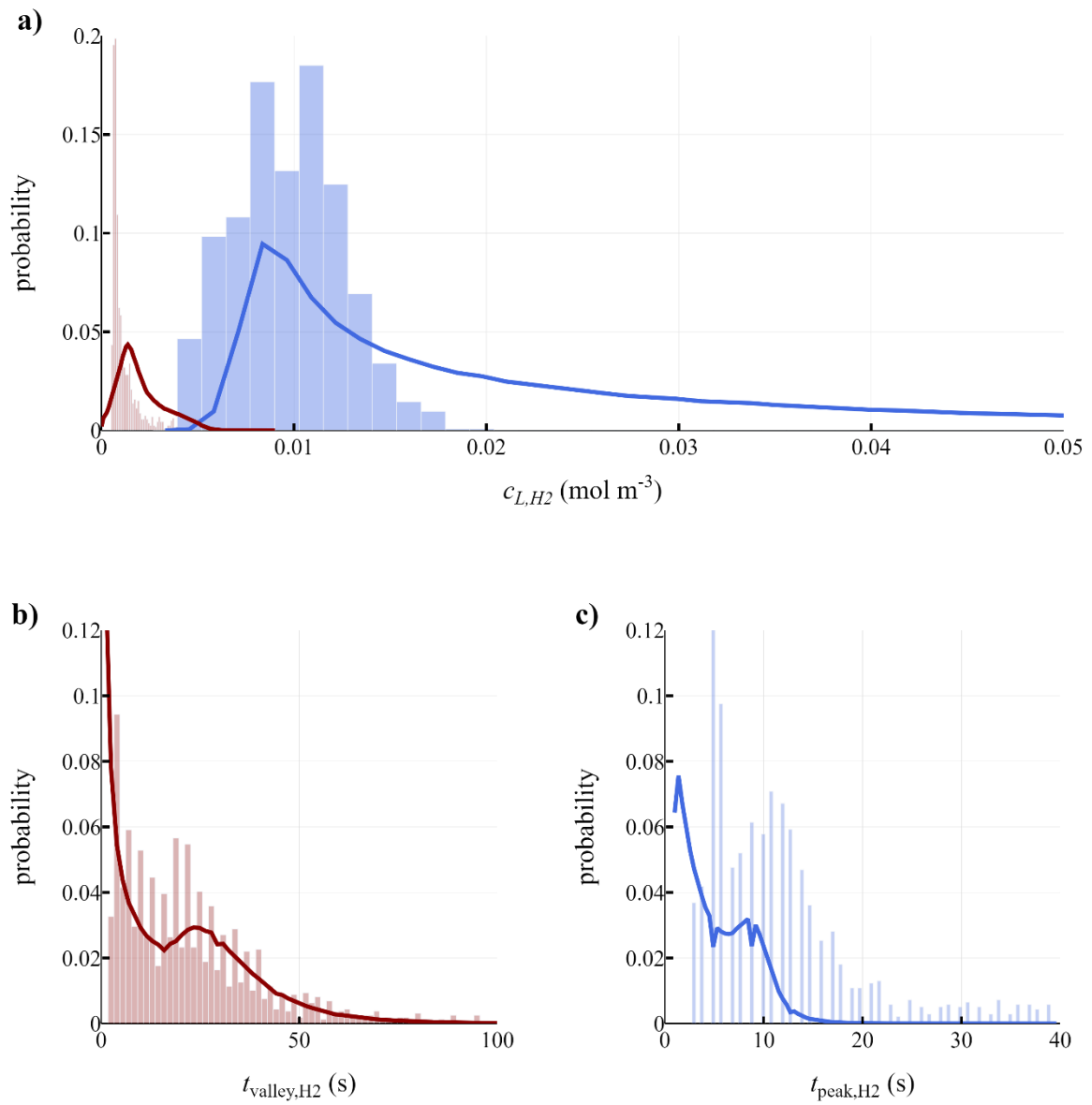


Figure S13. Comparison of the probability density functions obtained by the scale-down simulator (bars) with the CFD results (lines). Probability density functions for a) the concentration of dissolved H_2 during the peaks (blue) and the valleys (red), as well as the residence time in a b) valley or c) peak, obtained with 10 g L^{-1} biomass and simulating 2000 peaks.

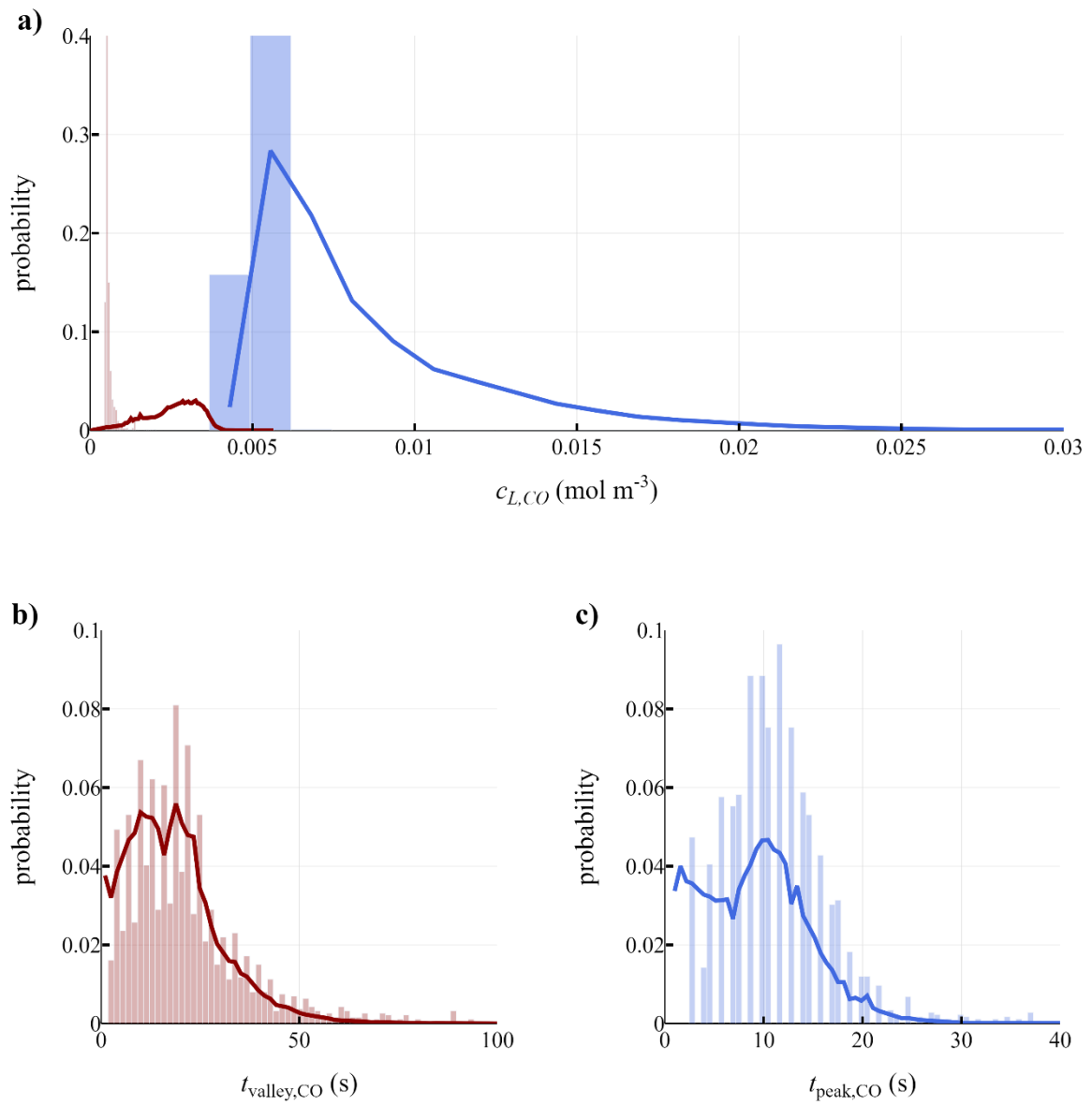


Figure S14. Comparison of the probability density functions obtained by the scale-down simulator (bars) with the CFD results (lines). Probability density functions for a) the concentration of dissolved CO during the peaks (blue) and the valleys (red), as well as the residence time in a b) valley or c) peak, obtained with 25 g L⁻¹ biomass and simulating 2000 peaks.

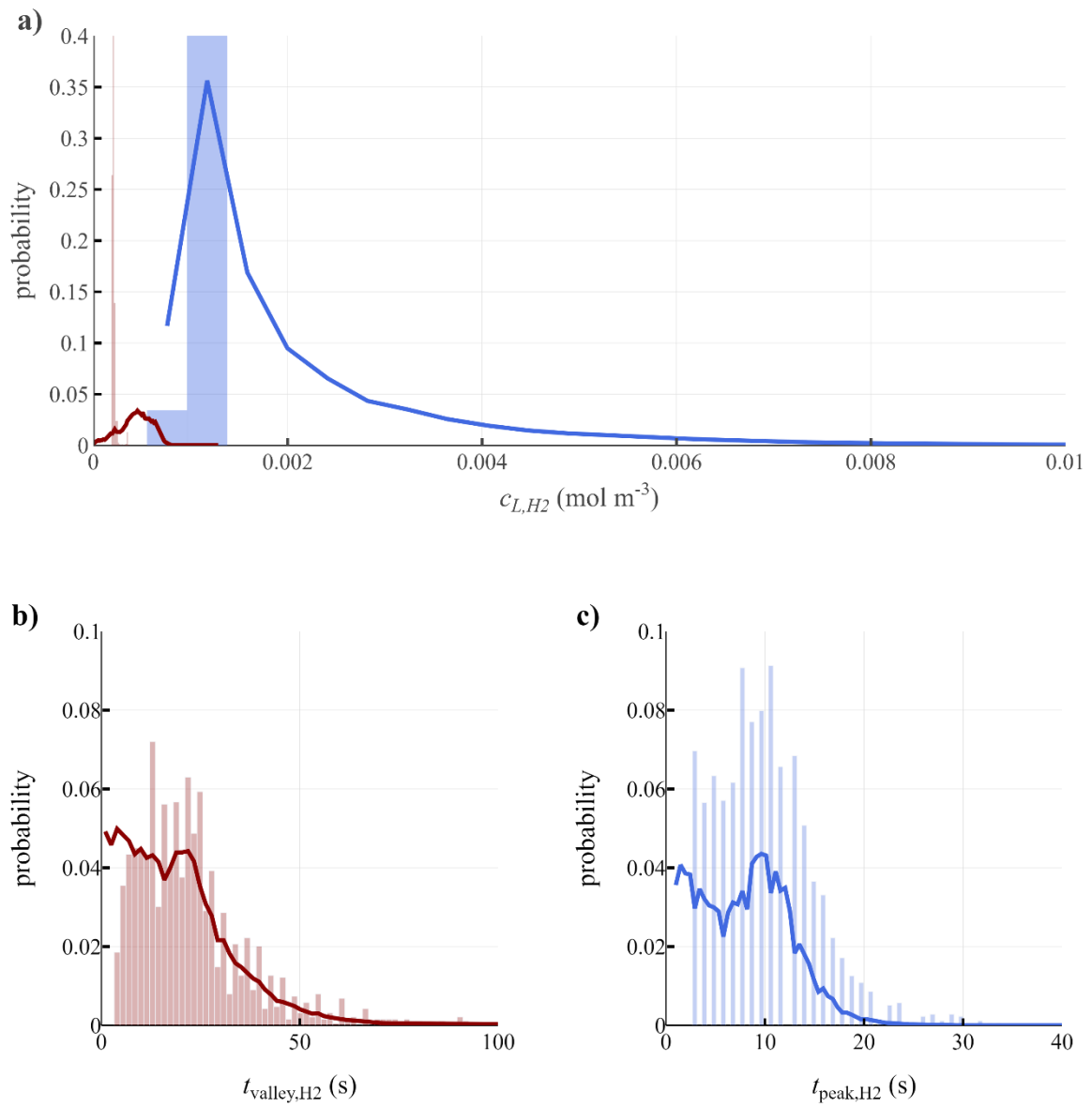


Figure S15. Comparison of the probability density functions obtained by the scale-down simulator (bars) with the CFD results (lines). Probability density functions for a) the concentration of dissolved H_2 during the peaks (blue) and the valleys (red), as well as the residence time in a b) valley or c) peak, obtained with 25 g L^{-1} biomass and simulating 2000 peaks.

Supplementary References

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