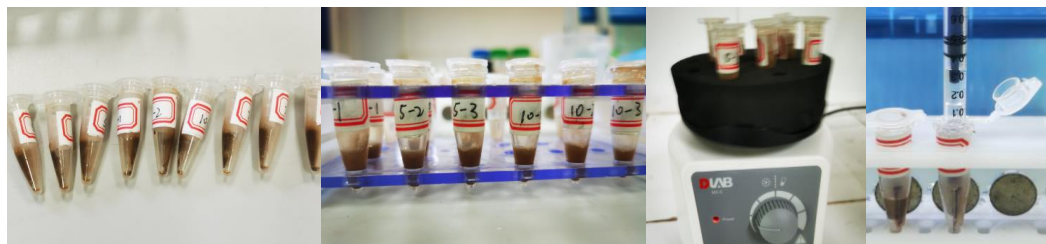


1. Detection of spiking sample by HPLC with nano-affinity cleaning up

To confirm the reliability of the immuno-separated assay, the collected IMAs should be eluted by 200 μ L 80% methanol solution (containing 0.5% acetic acid) for twice. The two elutes were combined and filtered by 0.1 μ m membrane for HPLC-FLD detection according to the reference (Campone et al. 2018).

1.1. Sample elution

(1) The collected IMAs were eluted with 200 μ L elution buffer by 10 mins shaking. The elution buffer were collected after separation.



(2) Another elution, separation and collection.



(3) Combine elutes for filter.

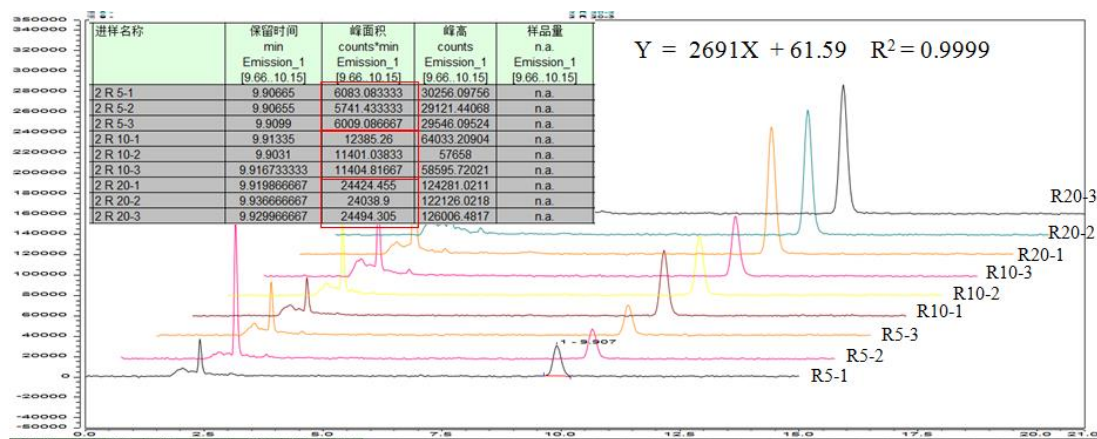


(4) Eluting solutions were detected by HPLC-FLD.

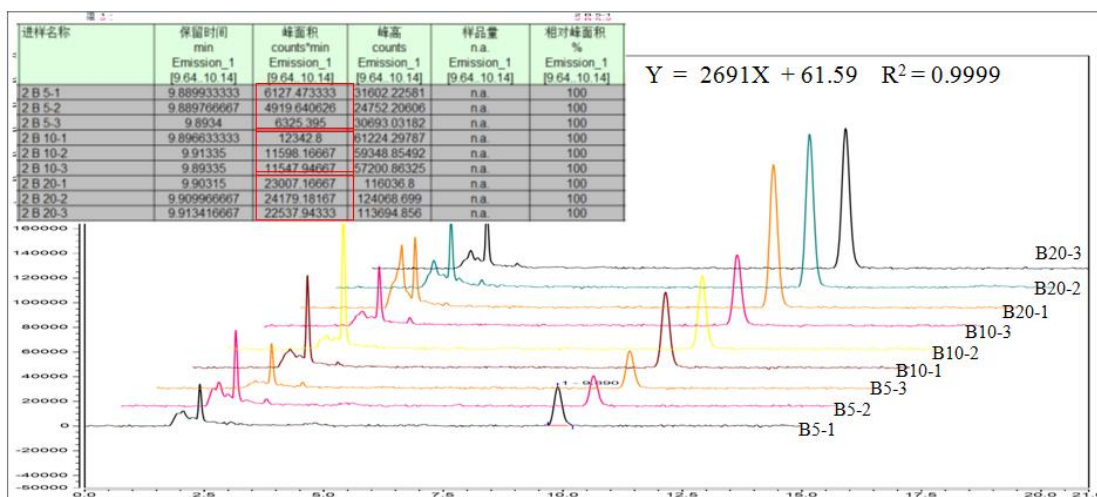


1.2. Detection results

(1) Detection results of rice samples by LC-FLD following nano-affinity cleaning up



(2) Detection results of beer samples by LC-FLD following nano-affinity cleaning up



2. Detection of spiking sample by HPLC purified by immuno-affinity column

2.1. Commercial immuno-affinity column



2.2. Sample elution

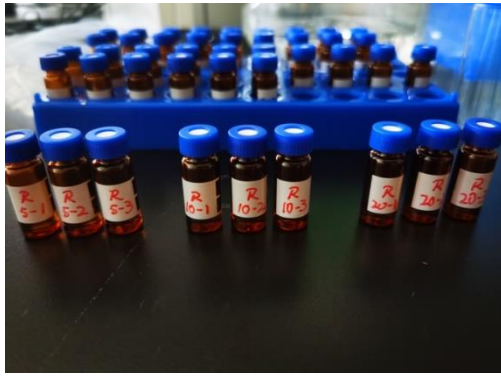
(1) 5.0g spiked rice samples were extracted by 10mL of 70% methanol.



(2) 4.0mL extracting solutions were diluted with 0.01M PBS, then were loaded on the IAC.



(3) The IAC were cleaned with 20mL 0.01M PBS and dried up. 2.0mL of 70% methanol (containing 0.5% formic acid) was used to eluted the IAC and collected the elutes.



Rice samples

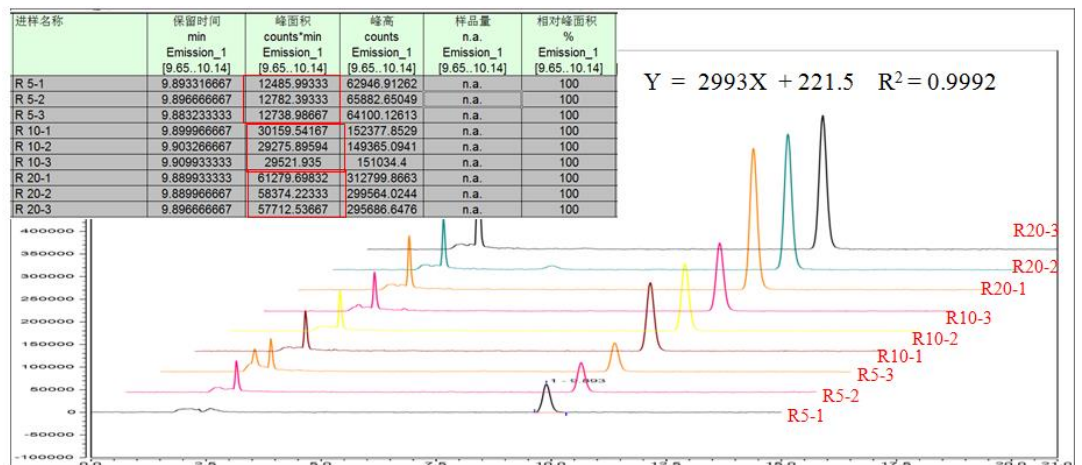


Beer samples

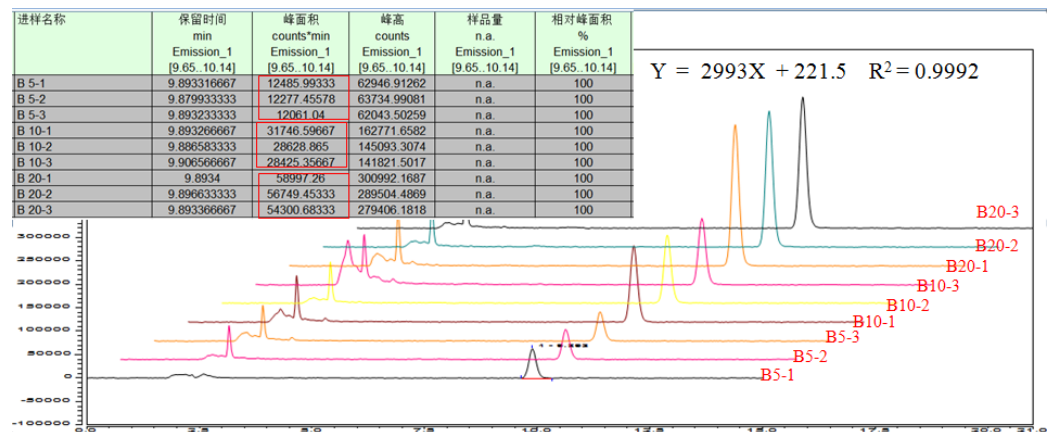
(4) The elutes were filtered for HPLC detection. The diluted ratios were 1:1.

2.3. Detection results

(1) Detection results of rice samples by LC-FLD following IAC cleaning up



(2) Detection results of beer samples by LC-FLD following IAC cleaning up



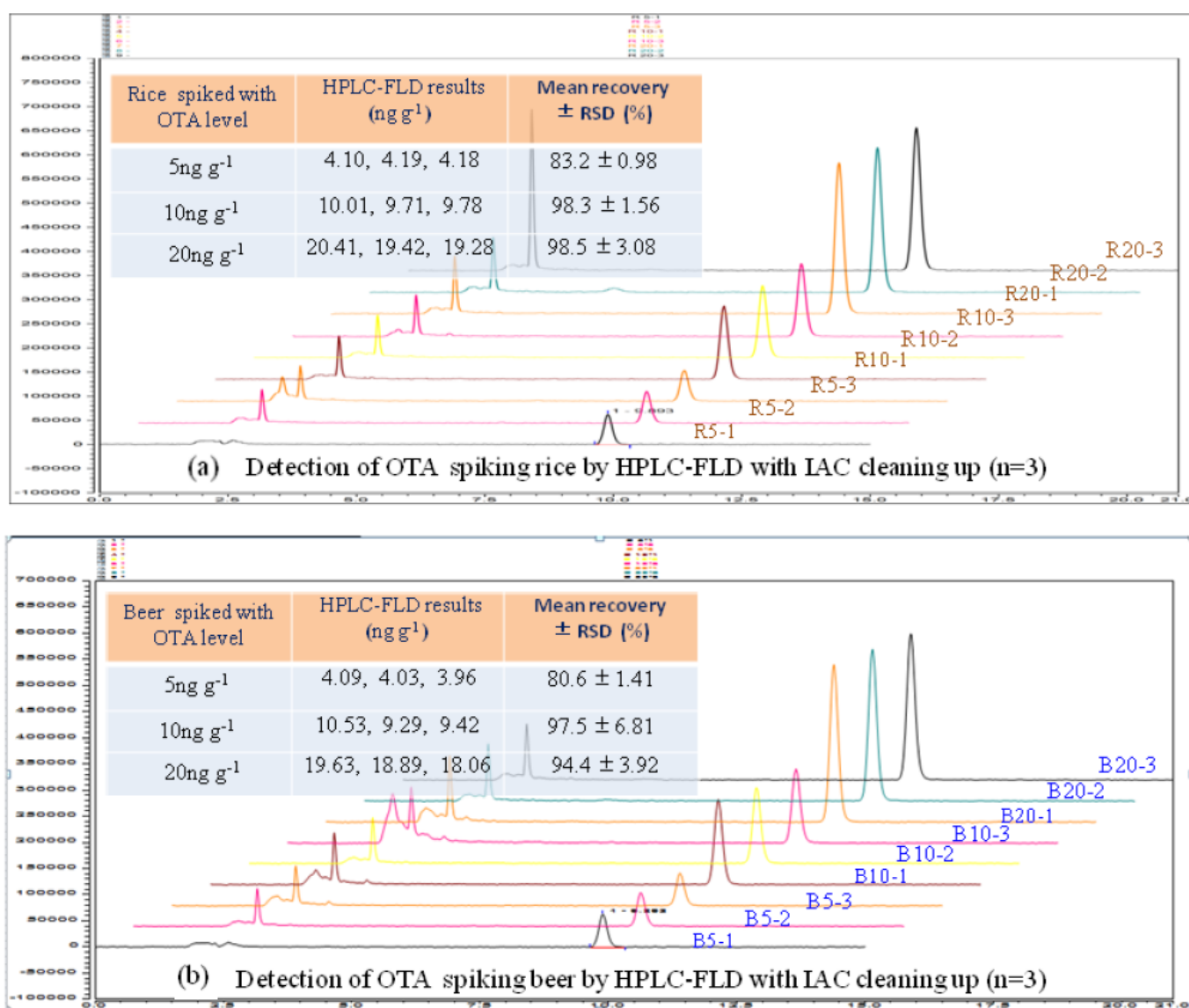


Figure S4. Detection of OTA spiking rice and beer samples by LC-FLD following immuno-affinity column cleaning up.