



Insights into Equilibrium and Adsorption Rate of Phenol on Activated Carbon Pellets Derived from Cigarette Butts

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Abstract: In the present work, the preparation of activated carbon pellets from cigarette butts by thermal treatment was evaluated. The morphological, textural, topological, and surface chemical properties were studied by SEM-EDX, N₂ adsorption, Raman, and FTIR spectroscopy. For adsorption assays, activated carbon was tested for the adsorption of phenol as a model molecule at different solution pH, temperature, and type of water. In addition, leaching tests before and after carbonization were conducted to evaluate the lixiviation of ions present in the solid. The results revealed a microporous material, composed of cylindrical fibers (thickness of 13 µm) with a microporous area of 713 m²/g and narrow and uniform slit-shaped pores (0.4–0.8 nm). The surface chemistry analysis evidenced the presence of oxygenated groups (carboxylic, esters, and phenolics). Activated carbon leaching tests indicated that the concentrations of the leached ions did not exceed the maximum permissible limit for drinking water. Phenol adsorption revealed an exothermic process with a maximum adsorption capacity of 272 mg/g at 10 °C. Finally, it was confirmed that phenol diffusion was drastically affected by hindered phenomena due to the similarity in the molecular size of phenol and the average size of micropores, and as a result an effective diffusion coefficient between 6.10×10^{-0} and 5.50×10^{-12} cm²/s and a maximum tortuosity value of 3.3 were obtained.

Keywords: cigarette butts; filter; activated carbon; adsorption; phenol

1. Introduction

At present, there are several environmental concerns related to waste generation derived from different anthropogenic activities [1]. One of the main issues in waste management is the treatment and disposal of cigarette butts (CBs), given that their toxic characteristics represent a risk to human health and environment. Previous reports showed that CBs have more than 4000 toxic chemical compounds, including nicotine, nicotine alkaloids, compounds specific to solanaceae, catechols, aromatic hydrocarbons, and polycyclic aromatic hydrocarbon (PAH), and these represent hazardous risks for living organisms and their environment [1,2]. The main components of CBs are a filter (generally made of cellulose acetate), tobacco residues, paper, and ashes [3]. In addition, CBs are considered one of the most abundant municipal waste materials [4]. The estimated amount of global discarded cigarette waste is about 340–680 million kg by year [1]. Furthermore, it was projected that smokers would consume approximately nine trillion cigarettes worldwide by



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Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). 2025 [3]. The most common alternatives for the management of CB wastes are incineration and landfilling; however, the emission of toxic compounds and the leaching of heavy metals can occur in the environment [5]. In this regard, different recycling methods and practices have been proposed to allow for the valorization of CB waste, such as mixing in construction materials, cellulose pulp production, corrosion inhibition, and preparation of carbonaceous materials, among others [2]. These valorization methods are effective and environmentally friendly alternatives to reduce the impact of CB waste [2,6]. Of this group of technologies, the development of carbonaceous materials stands out due to the fact that it be considered an economical and efficient alternative, since developing of commercial activated carbons implies a high-cost complex manufacturing process [7]. In this sense, CB waste transformation into activated carbons pellets is an alternative for obtaining low-cost adsorbents and for solving environmental problems, such as the presence of toxic pollutants in water and wastewater. Lima et al. [8] prepared activated carbon from CBs through a hydrothermal carbonization process with NaOH activation (S_{BET} $3.74 \text{ m}^2 \text{ g}^{-1}$, irregular morphology) and successfully tested for the adsorption of methylene blue, achieving a maximum adsorption capacity of 532.9 mg g^{-1} . On the other hand, Alhokbany et al. [4] obtained a highly porous functionalized carbon material from CBs prepared by means of a carbonization and hydrothermal process, and they studied the adsorption process of bisphenol; they found that the process is endothermic and spontaneous in nature, and that the adsorption mechanism was due to electrostatic and π - π interactions, achieving a bisphenol A removal efficiency of 92.47%. Finally, Manfrin et al. [9] and Conradi et al. [7] evaluated the adsorption of lead on activated carbon derived from CBs with a different chemical activation process (NaOH, ZnCl₂ and ZnCl₂ + CO₂); the materials presented a tubular structure and spongy aspects with broad surface functional groups (alkane, aldehyde, hydroxyl, carboxylic acid, carbonate, alcohol, and ether). The studies revealed that the adsorption process between adsorbate and adsorbent was due to chemical interactions between oxygenated groups and Pb2+. The maximum adsorption capacities obtained were in the order of 17.9 and 84.7 mg g^{-1} . Although the studies reported above represent an important contribution, it is necessary to describe the mass transfer mechanisms that govern the adsorption process in order to fully understand the phenomenon and establish the optimal conditions for the operation of the adsorption system.

The present work aims to prepare activated carbon pellets with CBs as raw material using a simple thermal method that allows the original monolithic structure of the precursor to be maintained, which is not a common characteristic accomplished in previous studies using CBs [4,7–9]. The efficiency of the activated carbon is evaluated by investigating the equilibrium and adsorption rate of phenol in aqueous solution. In addition, a deeper comprehension of the mass transfer mechanism governing the overall adsorption rate is attempted by applying diffusional models. Finally, leaching tests before and after carbonization demonstrate the advantages of this process in minimizing the release of secondary contaminants into the environment. In this study, the phenol molecule is used as a model pollutant, based on the consideration that it is a toxic organic pollutant that is widely found in industrial wastewater, its degradation is difficult, and it is reported to produce harmful effects on human health by inhalation and skin contact [10,11].

2. Materials and Methods

2.1. Materials

CBs of different trademarks were collected at the campus of the Autonomous University of San Luis Potosi, in San Luis Potosi, Mexico. CBs that presented defects on their cylindrical structure were discarded. As a first step, the unburned cigarette body of the cigarette was removed, and then the outer wrapping paper was stripped. Subsequently, the CBs were placed in an oven at 90 °C overnight to remove the excess of moisture. Finally, the dried CBs were placed in a sealed container for further processing.

2.2. Preparation of Hierarchical Activated Carbon

According to the thermogravimetric (TGA) and differential scanning calorimetry (DSC) analyses (see Figures S1 and S2 in Supplementary Materials), cellulose acetate fibers were thermally stable at temperatures below 200 °C. Two steps were observed in the TGA analysis. A weight loss around 115 °C was due to dehydration of the material (15 wt.%), while a significant second weight loss that was observed from 200 to 600 $^{\circ}$ C was associated with the decomposition of the lignocellulosic material (75 wt.%) [12]. A residue of 10 wt.% was obtained. From the DSC analysis, an exothermic melting process occurred at temperatures close to 300 °C where 90% of the initial mass was lost. Based on this information and to avoid losing the main structure exhibited by the CBs, the following procedure was performed to obtain activated carbon. A mass of 10 g of CBs was loaded in a crucible and placed in a furnace tube (Carbolite Gero model CTF 12/65/550, England) for carbonization at 200 °C for 2 h (1 °C min⁻¹) under N₂ flow of 100 mL min⁻¹. The heating rate of 1 $^{\circ}$ C min⁻¹ was chosen because higher values produce the melting of the fibers, resulting in the loss of the morphology. Subsequently, the furnace was turned off, and the N₂ stream was replaced by an air stream (100 mL min⁻¹) until room temperature was reached. This step was performed to incorporate oxygen functional groups to stabilize the carbon fibers. Afterwards, the sample was treated in a two-step thermal program under N₂ flow (100 mL min⁻¹), which was performed firstly at 300 °C for 2 h (5 °C min⁻¹) and then the temperature was raised to 550 °C for 1 h (10 °C min⁻¹). Finally, the sample was physically activated under CO₂ atmosphere at 850 °C for 2 h through a flow of 100 mL min⁻¹ at a heating rate of 10 °C min⁻¹.

2.3. Chemical and Textural Characterization

Textural properties of the activated carbon were determined by N₂ adsorption–desorption isotherms at -196 °C using a Micromeritics ASAP 2020 analyzer (Micromeritics Instrument Corporation, USA). Before measurements, the sample was degassed at 100 °C under vacuum (pressure $\approx 10^{-2}$ Pa) for 12 h as previously reported by other authors [13,14]. The pore volume was determined at P/P₀ \approx 0.98, and the micropore volume (W₀) was obtained by applying the Dubinin–Radushkevich method. On the other hand, the Stoeckli equation [15] was applied to estimate the surface area and the average micropore width (L₀). In addition, the pore size distribution of the sample was characterized by using nonlocal density functional theory methods (NLDFT) [16]. The surface and morphology of the particles were observed in an ESEM, i.e., model Quanta 250, FEI, Brno, Czech Republic. To show the distribution of chemical elements on the surface, an energy dispersive spectroscopy (EDS) elemental mapping was performed using an EDAX microprobe, model Octane Plus, NJ, USA.

The functional groups of CBs and activated carbon were identified by means of an FTIR spectrophotometer (Thermo Scientific, model Nicolet iS10, WI, USA). Infrared spectra were collected over a spectral range of 500 to 4000 cm⁻¹. The activated carbon samples were prepared in mixture with KBr in a ratio of 1:500.

The concentration of active sites of activated carbon was determined based on the method proposed by Boehm (1994) [17]. Additionally, the pH of zero charge point (pH_{PZC}) was determined by a titration method preparing various 0.01 N NaCl solutions with 0.004 N NaOH and 0.1 g of activated carbon. After 48 h, the solution pH was recorded, and the solution was titrated with a 0.1 N HCl solution. In addition, solutions were prepared without activated carbon. The pH_{PZC} was determined with the solution pH at which the titration curve with activated carbon intersects the titration curve without an adsorbent.

Finally, in order to study the topological structure of the activated carbon, Raman spectra were recorded using a RENISHAW InVía MicroRaman (Wotton-under-Edge, UK) with 532 nm laser operating at ca. 0.4 mW (1%) and 1800 lines per mm grating. Spectra were collected by averaging 5 acquisitions of 60 s duration. The Raman shift was calibrated using the Rayleigh peak and the 520.7 cm⁻¹.

2.4. CBs and Activated Carbon Leaching Test

The leaching of CBs and activated carbon in aqueous systems was evaluated by determining the content of heavy metals and other parameters for drinking water in 1 L of deionized water. The CBs were placed in a 15 cm³ acrylic column, and 1 L of water was recirculated using a peristaltic pump with a volumetric flow of 5 cm^3 min⁻¹ for 48 h. The same process was performed with the activated carbon. The resulting solution was analyzed by inductively coupled plasma optical emission spectrometry (ICP-OES) using a Varian 738 ES (Springvale, Australia) and according to the parameters established by the Mexican legislation and the values suggested by the WHO. Moreover, a gas chromatography analysis was performed to identify the main organic substances. This analysis was carried out by a direct injection of 1 µL of the sample into a gas chromatograph (Agilent 6890) equipped with a 30 m long, 0.25 mm internal diameter HP-5M (95%) polydimethylsiloxane (95%) capillary column. An initial temperature of 90 °C was used, which remained constant for 2 min, and then heated at a rate of 10 °C min⁻¹ to a final temperature of 230 °C. The temperature of the interface was maintained at 280 °C. The following conditions were used in the mass detector: temperature of the ionization chamber at 230 °C, helium as a carrier gas, and a flow rate of 0.9 mL min⁻¹.

2.5. Adsorbate

Phenol with a purity >99.9% was used as an adsorbate model molecule. Its physicochemical properties are presented in Table 1. The concentration of phenol in the aqueous solution was determined by UV–Vis spectroscopy. The absorbance of the phenol sample was determined using a double-beam spectrophotometer (Shimadzu, UV–160, Kyoto, Japan) at a wavelength of 269 nm.

Table 1. Physicochemical properties of phenol, adapted with permission from [18].

Compound	Molecular Structure	Molecular Formula	рК _а	* $D_{AB} \times 10^6~cm^2~s^{-1}$	Molecular Weight (g mol^{-1})	Size X, Y (nm)
Phenol		C ₆ H ₆ O	9.86	8.1	94.11	0.43 0.57

* Experimental molecular diffusivity of phenol in water at $T = 25 \degree C$, taken with permission from [19].

2.6. Equilibrium Experiments

Adsorption equilibrium experiments were carried out in a batch system using 50 mL plastic centrifuge tubes (adsorbers). For experiments, a specific mass of the activated carbon (0.1 g) was added together with phenol solutions (40 mL) of a known initial concentration (10 to 2000 mg L⁻¹). Deionized water with a conductivity <<0.055 μ S cm⁻¹ was used to prepare all solutions. The adsorbent–solution mix was left in contact until reaching equilibrium (21 days) under shaking, and the temperature was controlled by immersing the adsorbers in a constant-temperature bath. The effect of the solution pH was analyzed at values of 2, 5, 7, 10, and 11, using phenol solutions with a concentration of 500 mg L⁻¹. The solution pH was kept constant by adding drops of 0.01 N NaOH or 0.1 N HCl. After reaching equilibrium, the final phenol concentration was determined, and the mass of phenol adsorbed at equilibrium was calculated using the following equation:

$$q_e = \frac{V(C_{A0} - C_e)}{m} \tag{1}$$

where q_e is the mass of phenol adsorbed (mg g⁻¹), V corresponds to the volume of the solution (L), m is the mass of the adsorbent (g), C_{A0} represents the initial concentration (mg L⁻¹), and C_e denotes the equilibrium concentration (mg L⁻¹). The adsorption equilibrium

rium data of phenol from wastewater were obtained by preparing solutions of phenol using wastewater that exited a treatment plant located in San Luis Potosi, Mexico. The physicochemical characteristics of the wastewater were determined by the standard examination methods and are reported in Table S1 in the Supplementary Materials.

2.7. Obtaining the Concentration Decay Curves

The concentration decay curves of phenol on activated carbon were obtained using a rotating basket batch system that consisted of a three-necked reaction flask with two stainless steel mesh baskets coupled to a metal tube actuated for a variable-speed motor. The mechanical details of the rotating system can be found in a previous publication [20]. The adsorbent (1 g) was placed inside the two baskets, and 1 L of a phenol solution at pH 7 (concentration from 100 to 1500 mg L⁻¹) was added to the reactor. The system was stirred at 200 rpm. Previous studies reported that stirring speeds higher than 200 rpm do not affect the adsorption rate. The temperature was controlled through partial immersion of the reactor in a constant-temperature bath. The phenol concentration over time was monitored, sampling aliquots of 1 mL of the solution until the equilibrium condition; the total subtracted volume did not exceed 5% of the total volume, and so the total volume was assumed constant. The phenol concentration decay curve was constructed by plotting the dimensionless concentration (C_A/C_{A0}) against time (t).

3. Results

3.1. Characterization of CBs and Activated Carbon Pellets

The morphology of CBs before the carbonization process is presented in Figure 1a–c. Raw material consisted of a set of Y-shape cellulose acetate fibers interwoven in an axial direction with an average thickness of 25 μ m. After the carbonization process, the filter volume was drastically reduced up to 35 folds but its monolithic shape was maintained as illustrated in Figure 1d,g,h, indicating that the heating program to produce the activated carbon was optimal. The conservation of a monolithic shape is not a common characteristic, as found in previous studies using CBs [4,7,9,10]. In this sense, other experiments show that an increase in the heating rate or operation temperatures produced a complete melting of the fibers. On the other hand, the fibers suffered a visible morphological change (see Figure 1e,f), as it switched from a Y-type fiber to a cylindrical fiber with an average diameter of 13 μ m. This transition can be due to the polymerization of the cellulose chains to a more stable configuration due to the high calcination temperature. Finally, the overall yield for the activated carbon production was 11% due to the weight of the filters used as raw material.



Figure 1. Micrographs obtained by SEM for raw material (a-c) and activated carbon (d-i).

The EDS elemental mapping performed on the surface of the activated carbon is displayed in Figure 2. These results allowed for the identification of C, K, Ti, O, Al, Si, Ca, and Fe, with K and Ti standing out, since these can be derived from the additives added to tobacco, where K is added as potassium sorbate and Ti as titanium dioxide [21]. Moreover, it is notable that elemental carbon is the main chemical element in activated carbon with 89.5 wt.% while in the case of oxygen at about 8.5 wt.% and for the other constituents at 2 wt.%.



Figure 2. EDS mapping of C, O, Al, Si, K, Ca, Ti, and Fe at the surface of the activated carbon.

Figure 3 depicts the N₂ adsorption isotherm for the activated carbon prepared by activation with CO_2 . The isotherm presented a step shape with a high amount of N_2 adsorbed at low pressures ($V_{liq} = 0.20 \text{ cm}^3 \text{ g}^{-1}$), indicating a strong interaction between the N₂ molecules and the surface of the material. Moreover, an increase in the relative pressure slightly raised the amount of N₂ adsorbed by obtaining a value of 0.253 cm³ g⁻¹ at $P/P_0 = 0.98$. According to the International Union of Pure and Applied Chemistry, the adsorption isotherm showed a type 1A behavior, which is characteristic of microporous materials composed with narrow and uniform slit-shaped pores [22]. From the application of the Dubinin-Radushkevich method, the estimated microporous area was $S_{mic} = 713 \text{ m}^2 \text{ g}^{-1}$ with a micropore volume of $W_0 = 0.22 \text{ cm}^3 \text{ g}^{-1}$, indicating that 87% of the total porous structure corresponds to micropores. The obtained microporous area was higher than the reported values in the literature for the activated carbons prepared from lignocellulosic waste that used $ZnCl_2$ as an activating agent [23]. The pore size distribution obtained from the application of the NLDFT method is also observed in Figure 3. An unimodal distribution of narrow pores between 0.4 and 0.8 nm with a slight presence of wide micropores was determined. Moreover, an average micropore width of $L_0 = 0.62$ nm was obtained. These results suggest that despite the excellent material-specific area, its application as an adsorbent must be addressed to low-sized molecules (less than 8 nm) to promote diffusion into the narrow micropores.



Figure 3. Adsorption isotherm of N₂ at 77 K and pore size distribution obtained by NLDFT.

The FTIR spectra of CBs and activated carbon are shown in Figure S3 in the Supplementary Materials. The CBs spectrum has a band around 3400 cm⁻¹, which is attributable to the O-H stretching vibration of hydroxyl groups in cellulose and to the moisture in the material [9,24]. The bands observed from 3000 to 2850 cm^{-1} are attributed to the sp^2 and sp^3 C-H stretching vibration in aliphatic structures [24–26]. Bands associated with carbonyl groups (-C=O) and its hydroxyl (-OH) deformation vibration were observed at \approx 1720 cm⁻¹ and 1640 cm⁻¹, respectively [27]. The bands at 1430 cm⁻¹ and 1320 cm⁻¹ are relative to the C-H stretching of -CH₂ and -CH₃ groups, confirming the presence of cellulose [24]. In addition, at 1200 cm⁻¹, the band associated with C-O stretching of acetyl groups was observed, and at 1100 cm^{-1} , the band is attributed to C-O stretching of hydroxyl in cellulose was observed [24]. After activation, some similar bands were observed in the FTIR spectrum of the activated carbon, with a decrease in their intensity. This was due to the cellulose content of the cigarette filters diminishing because the aromaticity of the material increased during the thermal treatment [7]. Thus, the presence of carboxylic or phenolic groups can be evidenced with a band at 3310 cm⁻¹ related to -O-H stretching, and the presence of the band at 1450 cm⁻¹ could be associated with a complex arrangement of aromatic skeletal stretching vibration (C = C vibration) [28,29]. The band at around 1200 cm⁻¹ could be associated with the C-O stretching vibration in esters, carboxylic groups, or phenolic groups [12].

The concentrations of the acidic and basic sites obtained by Bohem's method were 0.052 and 0.492 meq g⁻¹, respectively. Thus, the surface of the activated carbon was basic, and this result is confirmed by the fact that the value of pH_{PZC} was 9.02. Finally, Figure 4 show the Raman spectra obtained for activated carbon, as seen in two well-defined peaks observed at 1336 cm⁻¹ and 1585 cm⁻¹, which can be assigned to defective band (D band) and graphitic band (G band), respectively. Moreover, the large estimated I_D/I_G value (0.92) confirms the rich structure nature of the material, similarly to that reported for other hierarchically ordered carbon materials.

3.2. Leaching Tests

The results of the CBs and activated carbon leaching tests are presented in Tables 2 and 3. For the elements determined by the ICP-OES (Table 2), it could be observed that when CBs or activated carbon were placed in contact with deionized water, there was an increase in the concentration of all the elements in comparison to them being placed the deionized water, with the increase in concentration of CBs being more noticeable. Moreover, possible variations in the concentration may be attributed to the uncertainty of the method of analysis <10% or to variations in the element concentrations of the deionized water used in the leaching tests or to the constituents of the chemicals used in the digestion

of the test samples. However, results suggest that the concentrations and types of elements determined are not significant enough to produce possible effects on human health.



Figure 4. Raman spectra of activated carbon prepared from CBs.

Table 2.	Elements	determined	by ICP-OES	in leaching te	sts.

Element	Deionized Water (mg L ⁻¹)	Cigarette Butt Leaching (mg L^{-1})	Activated Carbon Leaching (mg L^{-1})	
Ag	ND	< 0.003	ND	
Al	ND	0.121	ND	
As	ND	< 0.010	ND	
В	< 0.05	< 0.010	< 0.05	
Ва	3.153 *	< 0.010	5.900 *	
Be	ND	< 0.010	ND	
Ca	< 0.05	0.842	2.139	
Cd	ND	< 0.010	ND	
Со	ND	0.023	ND	
Cr	ND	< 0.010	ND	
Cu	ND	0.062	ND	
Fe	ND	< 0.010	ND	
K	0.147	3.199	0.667	
Li	3.929 *	0.569	6.340 *	
Mg	< 0.05	0.218	0.101	
Mn	0.830 *	< 0.010	3.233 *	
Мо	ND	< 0.010	ND	
Na	< 0.05	0.849	0.082	
Ni	ND	< 0.010	ND	
Р	ND	< 0.010	ND	
Pb	ND	< 0.010	ND	
Sb	ND	< 0.010	ND	
Se	ND	< 0.010	ND	
Si	<0.25	< 0.010	<0.25	
Sn	ND	< 0.010	ND	
Sr	1.353 *	< 0.010	5.526 *	
Ti	ND	< 0.010	ND	
T1	ND	< 0.010	ND	
V	ND	0.137	ND	
Zn	ND	<0.010	ND	

ND, not detectable; * $\mu g L^{-1.}$

Parameter

pН Color (Co/Pt) Odor Turbidity (NTU) Total hardness (mg CaCO₃ L^{-1}) Ca hardness (mg CaCO₃ L^{-1})

Mg hardness (mg CaCO₃ L^{-1})

Chlorides (mg L^{-1})

Fluorides (mg L^{-1})

Nitrites (mg L^{-1})

Nitrates (mg L^{-1})

Sulfates (mg L^{-1})

Mesophilic Aerobic

Microorganisms (UFC mL $^{-1}$) Total coliforms (NMP/100 mL)

Fecal coliforms (NMP/100 mL)

Total dissolved solids (mg L^{-1})

Chlorine (mg L^{-1})

	Action to d Carbon Las shines	Permissible Limits		
CBs Leaching	Activated Carbon Leaching	NOM	WHO	
5.12	7.8	6.5–8.5	NVP	
55.0	Colorless	20.0	Colorless	
Tabaco	Odorless	Odorless	Odorless	
1.0	0.2	10.0	<0.2	
2.0	2.0	500	NVP	

NVP

NVP

250

1.50

1.00

10.0

400

NVP

Absent

Absent

1000

0.10-0.50

2.0

0.0

2.0

0.10

Absent

0.15

4.00

Absent

_

-

_

Table 3. Parameter

2.0

0.0

4.0

0.12

Absent

0.11

4.50

Absent

Absent

Absent

10.5

Absent

Table 3 shows the parameters determined for the water generated in the leaching tests and the maximum permissible limit for each parameter measured according to the Mexican Official Standards (NOM) [30] and the World Health Organization Guidelines for Drinking Water Quality (WHO, 2017) [31]. Leaching tests for CBs show leaching with an acidic pH and a 55.0 Co/Pt color; moreover, a characteristic odor was produced, which caused the water to not comply with both Mexican legislation and what was suggested by the WHO. In the case of activated carbon, it can be observed that no concentration was above the maximum permissible limit in water for human consumption. Thus, it was demonstrated that the activated carbon prepared from CBs could be used to treat water for human consumption.

Gas chromatography (GC) analyses show the various organic compounds present in the leaching of CBs, such as nicotine, octanoic acid, trifluoroacetic acid, and 1,2,4trimethylcyclohexane which is considered a toxic compound; at the same time, carcinogenic substances such as dichloromethane, 2-ethylphenol, and pyridine were also found. These compounds were not detected in the leaching of activated carbons due to the pyrolysis and activation conditions used in their synthesis.

3.3. Effect of Temperature, Type of Water, and Solution pH on Adsorption Equilibrium

The efficiency of the adsorbent material for the removal of phenol was performed by obtaining the adsorption isotherms at 10, 25, and 40 °C, keeping the solution pH at 7. Figure 5a shows the adsorption isotherms at the three different temperatures, where an L-type behavior was observed, according to the Giles classification [32]. These results are in agreement with previous reports that the aromatic ring of phenol is adsorbed in parallel to the graphene layers of the carbon material [24,33]. It was also observed that phenol removal is gradually improved by decreasing the system temperature from 40 to 10 °C. For example, at an equilibrium concentration of 1200 mg L^{-1} , the adsorption capacities were approximately 200, 248, and 272 mg g^{-1} at 40, 25, and 10 °C, respectively.

NVP

NVP

NVP

1.50

3.00

50.0

NVP

NVP

Absent

Absent

NVP

5.00

These results indicate that a decrease in temperature from 40 to 25 °C and 40 to 10 °C improves the adsorption capacity of the material by 25% and 35%, respectively, showing the exothermic nature of the adsorption process. It has been reported that the phenol adsorption process on activated carbon shows different trends, i.e., some authors have reported an endothermic process [24,34], while other studies indicated that the adsorption process is exothermic [33,35]. This behavior could be related to the chemical nature of the surface material, since the number and the type of the surface functional groups present in the material can be influenced by the active carbon preparation conditions (chemical/physical treatment) and the precursor material [10].



Figure 5. Adsorption isotherms of phenol on activated carbon as a function of temperature and type of water: (**a**) effect of the temperature and type of water; (**b**) Van 't Hoff plot.

The experimental data were fitted using the Langmuir isotherm model represented by the following equation:

$$q_e = \frac{q_m K C_e}{1 + K C_e} \tag{2}$$

where q_e represents the equilibrium adsorption capacity in mg g⁻¹, C_e is the equilibrium concentration in mg L⁻¹, q_m is the maximum adsorption capacity in mg g⁻¹, and K is the equilibrium constant in L mg⁻¹. The q_m and K values were obtained by a nonlinear fit using Statistica 10.0 software, and their values are recorded in Table 4 together with the R² values.

Table 4. Langmuir isotherm fitting parameters of the phenol adsorption over activated carbon pellets.

Temperature (°C)	Type of Water	K (L mg ⁻¹)	Ke	q_m (mg g ⁻¹)	R ²
10	Deionized water	0.01542	1451.17	285.11	0.9756
25	Deionized water	0.01180	1110.49	268.9	0.9846
40	Deionized water	0.00965	908.16	211.45	0.9563
25	Wastewater	0.00530	498.78	56.58	0.9886

Table 4 shows that the q_m values are in the range of 211.45–285.11 mg g⁻¹ in the interval of temperatures studied. In addition, by the R² values, it was concluded that the Langmuir model satisfactorily fitted the experimental data as shown in Figure 5a. Although there are other isotherm models in the literature with more fit parameters that would lead to obtaining R² values close to 1, the Langmuir model was chosen because it physically interprets the phenomena, since the phenol molecule adsorbs in a monolayer on

the surface of the carbonaceous material because of the average width of the micropore and the dimensions of the adsorbate molecule.

The changes in Gibbs free energy (ΔG^0), enthalpy (ΔH^0), and entropy (ΔS^0) were estimated from the transformation of K values reported in Table 4 using the following equations [36]:

$$Ln(K_e) = \frac{\Delta S^{\circ}}{R} - \frac{\Delta H^{\circ}}{R} \frac{1}{T}$$
(3)

$$\Delta G^{\circ} = -RTLn(K_{e}) \tag{4}$$

$$K_{e} = \frac{1000 \times Mw \times [phenol]^{\circ} \times K}{\gamma}$$
(5)

where Mw is the molecular weight of phenol; [phenol]° is the standard concentration of phenol (1 mol L⁻¹); R is the universal gas constant in kJ mol⁻¹ K⁻¹; T is the absolute temperature; and γ is the coefficient of activity (dimensionless). It was assumed that the phenol solution is diluted to suppose that $\gamma = 1$. The relationship between Ln (K_e) vs. 1/T is known as the Van 't Hoff plot, and it is shown in Figure 5b along with the prediction of Equation (3), where it is evident that a linear behavior of the experimental values in the range of temperatures analyzed is found. From this plot, the values of Δ H° and Δ S° were -11,387.4 J mol⁻¹ and Δ S° 19.73 J mol⁻¹ K⁻¹, respectively. The Δ G° values were -17,138.2, -17,382.86, and -17,773.73 J mol⁻¹ at 283.15, 298.15, and 213.15 K, respectively. These results corroborated the feasibility and the exothermic nature of the adsorption process. In addition, a positive Δ S° value indicates an increase in the level of disorder and randomness at the solid–solution interface during the phenol adsorption process.

Figure 5a also presents the phenol adsorption isotherm obtained with wastewater at 25 °C and pH 7. The results indicate that the adsorption capacity of the activated carbon is drastically reduced with this type of water, reaching a maximum value of 56 mg g⁻¹, showing an approximate reduction of 80% compared with the adsorption capacity obtained using ultrapure water. A physicochemical analysis of the water revealed a high presence of metal ions, which can block the pores of the adsorbent, preventing the diffusion of phenol towards the active sites. Similarly, the presence of organic matter contributes to a decrease in the adsorption capacity of activated carbons as reported in the literature [37]. Hence, the adsorption capacity of the material was reduced using wastewater. Figure 6 displays a micrograph of activated carbon exposed to wastewater. The micrograph shows a large number of crystals deposited on the surface of the adsorbent. Additionally, the EDS analysis confirmed the presence of some species such as Al, C, Ca, Cl, Fe, K, Mg, Na, O, P S, Si, and Ti.



Figure 6. Micrographs obtained by SEM for activated carbon after adsorption with wastewater.

The solution pH is one of the most relevant factors in the study of adsorption processes since it directly affects the surface charge of the material and the speciation of the solute. To evaluate this effect, the adsorption capacities were obtained at pH 2, 5, 7, 10, and 11. The results are illustrated in Figure 7, where it can be observed that the adsorption capacity remained almost constant ($q \approx 215 \text{ mg g}^{-1}$) in a pH range between 2 and 7, but it decreased linearly when the pH increased from 7 to 11. Thus, the adsorption capacity decreased by 43%, thereby increasing the pH of the solution from 7 to 11. These results are explained by considering that the activated carbon presented a pH_{PZC} = 9.02, so that when the solution pH > pH_{PZC}, the surface of the material is negatively charged. Thus, considering that the pKa value of phenol is 9.86, it can be assumed that at pH values higher than 9.86, the phenol molecule is negatively charged. This generates an increase in the repulsive electrostatic interactions as the solution pH is augmented, causing a progressive decrease in the adsorption capacity. On the other hand, at pH values from 2 to 7, the phenol molecule is neutral, indicating that in this pH range, the adsorption mechanism is governed by π - π interactions.



Figure 7. Effect of the solution pH on the adsorption capacity.

Table 5 shows maximal adsorption capacities obtained in previous research for phenol removal using activated carbon prepared from different agroindustrial waste. The activated carbon prepared in this study shows a high adsorption capacity compared to most of the other activated carbons obtained from agroindustrial waste. In this sense, the superficial functional groups and microporosity developed in the activated carbon obtained from CBs (under the preparation conditions used) are responsible for having a good material for removing organic low-sized molecules such as phenol.

Activated Carbon Obtained from Different Raw Materials	T (°C)	q_{max} (mg g ⁻¹)	Ref.
Black wattle bark waste	25	85.75	[24]
Toona sinensis leaves	25	325.0	[35]
Tithonia diversifolia	25	50.55	[34]
Rise husk-KOH Rise husk-KOH/EDTA—4Na	25	194.2 215.3	[10]
Coconut shell	25	144.9	[33]
Avocado kernel seeds	25	90.0	[38]
Cigarette filters	25	268.9	Present study

Table 5. Maximum adsorption capacities for phenol adsorption using activated carbon produced from different agroindustrial wastes.

3.4. Adsorption Rate

The adsorption rate is a crucial parameter for scaling-up fixed-bed adsorbers since it directly influences their size. To investigate this aspect, decay curves were obtained at a different initial concentration (102.7–1520.1 mg L^{-1}), keeping the adsorbent mass and the solution volume constant. Consequently, each kinetic curve presented a different equilibrium adsorption capacity, qe. The operating conditions for each adsorption kinetic are reported in Table 6, while the experimental data are presented in Figure 8a,b. In both figures, the time to reach equilibrium increased considerably with the rising of the adsorbed mass at equilibrium. For example, q_e values of 73.79, 196.7, and 240 mg g⁻¹ were obtained for the equilibrium times of 6000, 20,000, and 30,000 min, respectively. These results show an exponential increase in the equilibrium time as qe rises. Leyva-Ramos et al. [19] investigated the adsorption of phenol on activated carbon cloth (fibril diameter of 5 mm, an average pore diameter of 1.95 nm); they found that the time to achieve equilibrium was only 10 min [19], which is much less than that found in this present work. On the other hand, Lorenc-Grabowska et al. [18] used six microporous activated carbons to evaluate the phenol adsorption rate, and they found that the time to reach equilibrium was 1080, 1440, 1440, 2700, 3120, and 7200 min for the CS, CWZ, ASN, PFA, PET, and PAN samples, respectively [18]. It is essential to mention that the six samples showed a maximum pore size distribution between 1 and 1.4 nm. From the results, it can be inferred that the adsorption rate of phenol in microporous materials depends drastically on the average width of the micropores. Therefore, the high equilibrium times obtained in this work are attributed to the narrow microporosity (0.4–0.8 nm) of the material, which causes the diffusion phenomena inside the pores. This performance becomes more evident as the number of adsorbed molecules increases, which induces partial blockage of the porous structure of the material, increasing the equilibrium time.

Table 6. Experimental conditions and mass transport parameters estimated by applying the diffusional model with $\varepsilon_p = 0.6$, $\rho_p = 1.1$ g cm⁻³, and $R_p = 6.5$ µm.

Exp. Number	C_{A0} (mg L^{-1})	q_e (mg g ⁻¹)	$k_L \times 10^5$ (cm s^{-1})	D_{ep} (cm ² s ⁻¹)	τ
1	102.7	73.79	3.51	$6.10 imes10^{-10}$	7967
2	201.3	119.1	3.85	$1.55 imes10^{-10}$	31,354
3	310.2	169.4	3.76	$9.10 imes10^{-11}$	53,406
4	405.2	196.7	4.23	$8.35 imes10^{-11}$	58,203
5	500.5	210.3	3.57	$4.02 imes 10^{-11}$	120,895
6	726.3	181.2	5.56	$8.11 imes10^{-12}$	599,260
7	1017.3	235	5.47	$6.84 imes 10^{-12}$	710,526
8	1520.1	240	5.66	$5.50 imes 10^{-12}$	883,636



Figure 8. Concentration decay curves of phenol on activated carbon pellets for (**a**) Exp. 1–4 and (**b**) Exp. 5–8. The lines represent the prediction of the diffusional model.

In the literature, different mathematical models were reported to interpret the adsorption rate. These models are based on one or more mass transfer resistances involved during the process (external mass transfer, intraparticle diffusion, or adsorption at an active site). The most widely used mathematical models are kinetic models (first order and second order), which ignore both the external mass transfer as the intraparticle diffusion, and they only consider the adsorption on an active site. These models would adequately interpret the experimental data. However, their application would have no physical meaning since intraparticle diffusion plays an important role in this system. For this reason, the experimental data were interpreted using a diffusional model based on the pore volume diffusion. This model was developed in a previous work [19] and is given by the following set of equations.

$$V\frac{dC_{A}}{dt} = -mSk_{L}\left(C_{A} - C_{Ar}\big|_{r=Rp}\right)$$
(6)

$$t = 0 C_A = C_{A0}$$
 (7)

$$\varepsilon_{p}\frac{\partial C_{Ar}}{\partial t} + \rho_{p}\frac{\partial q}{\partial t} = \frac{1}{r}\frac{\partial}{\partial r}\left[r\left(D_{ep}\frac{\partial C_{Ar}}{\partial r}\right)\right]$$
(8)

$$C_{Ar} = 0 \quad t = 0 \quad 0 \leq r \leq R_p \tag{9}$$

$$\left. \frac{\partial C_{\rm Ar}}{\partial r} \right|_{r=0} = 0 \tag{10}$$

$$D_{ep} \frac{\partial C_{Ar}}{\partial r} \Big|_{r=Rp} = k_L (C_A - C_{Ar}|_{r=R})$$
(11)

where V is volume of the solution in L; S is the external surface area per mass of adsorbent in cm² g⁻¹; C_{Ar} is the concentration of phenol within the particle at distance r in mg L⁻¹; C_{Ar|Rp} is the concentration of phenol at the external surface of the particle at $r = R_p$ in mg L⁻¹; k_L is the external mass transfer coefficient in liquid phase in cm s⁻¹; D_{ep} is the effective pore volume diffusion coefficient in cm² s⁻¹; ε_p is the void fraction of adsorbent particles; ρ_p is the density of adsorbent particles in g cm⁻³; R_p is the radio of the fiber in cm; and q is the mass of phenol adsorbed in mg g⁻¹. Equations (6) and (8) represent the mass balance in the solution and inside the particle, while the initial conditions are given by Equations (7) and (9). Equation (10) is a boundary condition that represents the absence of mass flux at the center of the particle, whereas Equation (11) establishes the continuity of fluxes at the external surface of the particle. Finally, if the adsorption rate of phenol on an active site is instantaneous, it is appropriate to consider a local equilibrium to be taking place between the phenol concentration in the solution inside the pore and the mass of phenol adsorbed on the pore surface. This equilibrium is given by Equation (2), which is the mathematical relationship between q and C_{Ar} .

To solve the mathematical model denoted by Equations (6)–(11), it is necessary to know the values of k_L and D_{ep} . The k_L value was assessed by the procedure suggested by Furusawa and Smith [39]. This method is based on the fact that when t = 0 min, then $C_{Ar} = 0 \text{ mg } L^{-1}$ and $C_A \rightarrow C_{A0}$. Substituting these conditions in Equation (6), the following equation can be obtained:

$$k_{\rm L} = -\frac{V}{mS} \left(\frac{dC_{\rm A}}{dt} \right)_{t \to 0} \tag{12}$$

The term at the left of Equation (12) is the slope of the concentration decay curve estimated by using the first two data points at t = 0 and t = 5 min. The values of k_L are summarized in Table 6 and are ranged from 3.51×10^{-5} to 5.66×10^{-5} cm s⁻¹, which are lower than those reported in the literature for the adsorption of aromatic compounds on carbon materials [40–42]. This difference is mainly attributed to the long times required to achieve equilibrium in this system.

The optimal D_{ep} value for each experiment was obtained by fitting the numerical solution of the diffusional model to the experimental data, minimizing the following objective function:

$$\operatorname{Error} = \int_{t=0}^{t \text{ final}} (C_{A, exp} - C_{A, Pred})^2 dt$$
(13)

where $C_{A,exp}$ is the experimental concentration of phenol in mg L⁻¹ and $C_{A,Pred}$ is the concentration of phenol predicted with the diffusional model in mg L⁻¹.

As an example, Figure 9 shows the experimental data for Exp. 1 with the prediction of the diffusional model using different values of D_{ep} . This figure indicates that the prediction of the model is highly influenced by the value of D_{ep} . For instance, for values of $D_{ep} = 4.86 \times 10^{-7}$ cm² s, the model predicts an adsorption rate almost ten times faster than those obtained experimentally, indicating that smaller values of $D_{ep\lambda}$ are necessary for improving the prediction of the experimental data. After performing an optimization process, the best value of D_{ep} that matched the experimental data was $D_{ep} = 6.10 \times 10^{-10}$ cm² s⁻¹, whose prediction (solid line) is illustrated in Figure 9.



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Figure 9. Concentration decay curve during the adsorption of phenol on activated carbon. The lines represent the prediction of diffusional model with different values of D_{ep}.

A similar procedure was used to interpret the data for Exp. 2–8 from Table 6. The best fit of the experimental data is illustrated in Figure 8a,b, while the optimal D_{ep} values are summarized in Table 6. From both figures, it can be inferred that the model adequately predicts the experimental data for all experimental conditions. Furthermore, it is observed in Table 6 that the D_{ep} values decreased exponentially from 6.10×10^{-10} to 5.50×10^{-12} cm² s⁻¹ as the adsorbed mass at equilibrium increased from 73.79 to 240 mg g⁻¹. From the optimized values of D_{ep} , the material's tortuosity factor, τ , can be estimated using the following equation [43]:

$$\tau = -\frac{D_{AB}\varepsilon_p}{D_{ep}} \tag{14}$$

The estimated values of τ are also collected in Table 6, and they present a contrary trend to D_{ep} , that is, the value of τ increased exponentially with the rising of q_e . Tortuosity is defined as the relationship between the effective distance traveled by a species that moves by molecular diffusion per unit length of the medium [44]. For activated carbon, it was estimated that the value of τ varies between 2 and 6 due to its macro, meso, and microporous structure. Therefore, as the obtained activated carbon has micropores exclusively, it would be expected to obtain a τ value of less than 3. However, the calculated τ values are above the values previously reported for activated carbons [43–45]. In addition, the value of τ is an intrinsic property of the material, and therefore its value must remain constant with different operating conditions. Thus, the values of τ calculated with Equation (14) have no physical meaning. A possible explanation for this behavior could be that the diffusion of phenol drastically decreases inside the pores of the material, because of its molecular size. Since it is quite similar to the size of the micropores, it can lead to hindered diffusion on the material. Saterfield et al. [46] modified Equation (14) to consider the effects of restrictive diffusion and in turn proposed the following equation:

$$D_{ep} = \frac{D_{AB}\varepsilon_p K_p K_r}{\tau}$$
(15)

where K_p and K_r are the restriction factors for exclusion and friction, respectively. Several correlations have been developed to estimate K_p and K_r , based on the relationship of the molecule size (d_m), and the pore diameter (L_0), under the consideration that the pores are cylindrical and that the molecules are rigid spheres. The most common mathematical expressions are the Renkin equation (Equation (16)) and the Bungay–Brenner equation (Equation (17)) [47,48] described as:

$$K_{\rm p} = (1 - \zeta)^2 \quad K_{\rm r} = \frac{6\pi}{K_{\rm t}} \tag{16}$$

$$K_{t} = \frac{9\pi^{2}\sqrt{2}}{4}(1 - \zeta)^{-5/2} \left[1 + \sum_{n=1}^{2} a_{n}(1 - \zeta)^{n}\right] + \sum_{n=0}^{4} (a_{n+3}) \zeta^{n}$$
(17)

where $a_1 = -73/60$; $a_2 = 77,293/50,400$; $a_3 = -22.5083$; $a_4 = -5.6117$; $a_5 = -0.3363$; $a_6 = -1.216$; $a_7 = 1.64$; and ζ represents the relationship between the phenol molecular size and the average width of the micropores. In this system, the value of ζ varies between 0.694 (0.43/0.62) and 0.919 (0.57/0.62), depending on the characteristic length of the phenol molecule (see Table 1). Substituting these values of ζ in Equations (16) and (17), K_p and K_r values are obtained, i.e., 4×10^{-3} and 7.92×10^{-6} for $\zeta = 0.694$ and $\zeta = 0.919$, correspondingly. These values indicate that the phenol adsorption rate is reduced between 248 and 125,000 folds due to the presence of restrictive phenomena. Therefore, this explains the high equilibrium time and consequently the low D_{ep} values obtained.

Saterfield et al. [46] also modified Equation (15) to include an additional parameter to take into account the obstruction effects due to adsorbed molecules on the surface of the adsorbent, in turn obtaining the following equation:

$$D_{ep} = \frac{D_{AB}\varepsilon_p K_p K_r K_a}{\tau}$$
(18)

where K_a is the correction factor for the restrictive effects caused by the adsorbed molecules. Leyva-Ramos et al. [48] proposed that Equation (18) can be expressed as follows:

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$$\frac{D_{ep}}{D_{AB}\varepsilon_p K_p K_r} = \frac{K_a}{\tau} = D_{corr}$$
(19)

where D_{corr} is a dimensionless diffusion coefficient that incorporates the constraint corrections. An important feature of Equation (19) is that at the limit when $q_e \rightarrow 0$ and $K_a \rightarrow 1$, then $1/D_{corr} \rightarrow \tau$.

The $1/D_{corr}$ values were calculated using Equation (19), and the obtained values are presented in Figure 10a,b for the two representative phenol lengths. Additionally, the experimental data were correlated using the next correlation [48]:

$$\frac{1}{D_{corr}} = \tau e^{a \times q_e} \tag{20}$$

where a is a positive constant (g mg⁻¹). This equation also meets the requirement that $1/D_{corr} \rightarrow \tau$ as $q_e \rightarrow 0$. The values of τ were obtained by a nonlinear estimation, using the Statistica software being $\tau = 3.3$ and $\tau = 0.081$ for the phenol characteristic lengths of 0.43 and 0.57 nm, respectively (see Figure 10a,b). It is evident that the second value of τ lacks physical meaning since for the straight and uniform pores, $\tau = 1$. Therefore, the maximum value of τ for the carbon sample used in this work is the lower limit of reported values for activated carbons.



Figure 10. Effect of the adsorbed mass at equilibrium on the dimensionless effective diffusion coefficient.

4. Conclusions

In this work, activated carbon pellets were obtained from CBs, with an excellent specific area and wide applications as an adsorbent. This material was successfully used as an adsorbent material for phenol concentration ($q_{max} = 268.9 \text{ mg g}^{-1}$).

Adsorption isotherms indicated that the aromatic ring of phenol is adsorbed parallel to the graphitic layer of activated carbon. In addition, the adsorption capacity improved by a decrease in temperature, showing the exothermic nature of the adsorption process. Phenol adsorption was favored at pH values from 2 to 7 since basic pH values led to an

important decrease of adsorption capacities due to the repulsive electrostatic interactions between phenol and the negatively charged surface of the activated carbon.

The microporosity of the material is a crucial parameter; therefore, high equilibrium times were required, and activated carbon must be useful for adsorption of low-sized molecules (less than 8 nm) to promote diffusion into the narrow micropores. It can thus be seen that intraparticle diffusion plays an important role in this system. Experimental data were fitted to a pore volume diffusion model that adequately predicted the experimental data to all conditions used. Finally, it was found that the tortuosity values obtained from this model for the activated carbon were $\tau = 3.3$.

Perspectives

The obtention of an adsorbent material from cigarette waste makes it necessary to think about its adequate separation from the total municipal waste. Hence, its treatment implies taking action from its generation, e.g., the implementation of recollection points in many local places such as parks, restaurants, supermarkets, coffee shops, universities, among others. Currently, there are social organizations or movements in several cities around the world that are taking the initiative to collect this type of waste. Finding a use for this waste allows for the minimization of its negative impact on the environment and gives added value to this waste material. Therefore, the use of cigarette butts as a precursor of an adsorbent material can be a feasible and justified business option in the future.

Supplementary Materials: The following are available online at https://www.mdpi.com/article/10 .3390/pr9060934/s1, Figure S1: Thermogravimetric (TGA) analysis of CBs, Figure S2: Differential scanning calorimetry (DSC) analysis of CBs, Figure S3: FTIR spectra of CBs and activated carbon, [S1: Physicochemical characteristics of the wastewater exiting a treatment plant located in San Luis Potosi, Mexico].

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