Supporting Information

A rationally designed, spiropyran-based chemosensor for magnesium

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Figure S1. ¹H NMR spectrum of chemosensor 4 recorded in DMSO-D₆ at 500 MHz.



Figure S2. ¹³C NMR spectrum of chemosensor 4 recorded in DMSO-D₆ at 500 MHz.



Figure S3. ¹H NMR spectrum of chemosensor 3 recorded in DMSO-D₆ at 500 MHz.



Figure S4. ¹³C NMR spectrum of chemosensor **3** recorded in DMSO-D₆ at 500 MHz.



Figure S5. ¹H NMR spectrum of chemosensor **2** recorded in CD₃CN and d_6 -DMSO at 500 MHz. The sensor is present in both the merocyanine and spiropyran forms in the spectrum. H₂O signal at δ 3.1 ppm supressed.



Figure S6. ¹H NMR spectrum of chemosensor 1 recorded in CD₃CN at 500 MHz.



Figure S7. ¹³C NMR spectrum of chemosensor 1 recorded in CD₃CN at 500 MHz.



Figure S8. ESI-HRMS spectra of chemosensors (a) 4, (b) 3, (c) 2 and (d) 1.

DFT Calculations

Reaction energies were calculated based on the following equation, where 'M' denotes Mg^{2+} or Ca^{2+} , and 'MC' denotes the merocyanine form of chemosensor 1 or 2.

$$2(MC) + M [(H_2O)_6]^{2+} \rightarrow M [MC_2(H_2O)_2] + 4(H_2O)$$



Figure S9. B3LYP/6-311G^{**} optimized structure of chemosensor **2** bound to (A) Mg²⁺ (yellow) and (B) Ca²⁺ (gold), respectively, in a 2:1 ratio, showing oxygen atoms (red) chelating to the metal ion. Hydrogen atoms are omitted for clarity. The Mg²⁺...O distances are between 2.08 and 2.12 Å, and Ca²⁺...O distances are between 2.36 and 2.45 Å. Reaction energies for the formation of the M[**2**(MC)₂(H₂O)₂] species were calculated to be - 315.7 kJ/mol and -151.2 kJ/mol for Mg²⁺ and Ca²⁺, respectively.

 $Mg[(1MC)_2(OH_2)_2]^{2+}$ structural coordinates

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С	-7.08145	-1.73431	-0.88255	0	-1.28239	-1.83632	-2.66599
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С	-7.04168	-2.91953	-1.91309	Н	5.28631	3.33515	0.09252
С	-6.79695	-1.66695	-1.35269	Н	2.94475	4.1346	0.22628
С	-7.82695	-0.7621	-1.08604	Н	3.42554	4.04802	-1.48713
С	-9.14147	-1.10265	-1.36885	Н	-10.4358	-2.6409	-2.15155
Ν	-5.56287	-1.09148	-0.98836	Н	-8.59228	-4.22041	-2.63136
С	-5.72592	0.17428	-0.50037	Н	-6.24123	-3.61823	-2.12574
С	-7.2325	0.51333	-0.51415	Н	-9.95254	-0.40843	-1.16742
0	-1.91833	1.69596	0.31979	Н	-5.77661	2.69963	0.46846
С	-2.32902	2.8365	0.69344	Н	-3.6555	0.57098	-0.14199
С	-3.75509	3.17305	0.75162	Н	-0.33794	3.61634	1.01586
С	-4.7774	2.28311	0.37721	Н	-1.10625	5.87076	1.79147
С	-4.65871	0.97074	-0.09314	Н	-5.19279	4.74598	1.25293
С	-1.39234	3.86725	1.08005	Н	-8.63285	1.82988	-1.52962
С	-1.81316	5.09981	1.49937	Н	-7.12111	2.63804	-1.09049
С	-3.19937	5.3951	1.55771	Н	-7.15914	1.51878	-2.45954

Н	-7.55701	-0.085	1.56843	Н	-3.86905	-3.25597	2.01442
Н	-7.35034	1.65952	1.36414	Н	-4.62234	-1.73396	1.62883
Н	-8.86427	0.88512	0.87309	Н	0.66061	-0.48721	3.21814
Н	-3.46971	-1.11439	-1.14072	Н	-0.87994	-0.56414	2.85308
Н	-4.32641	-2.25646	-2.16897	Н	0.86595	0.86191	-2.19686
Н	-3.18888	-3.48377	-0.45122	Н	0.96159	-0.68592	-2.07069
Н	-4.91492	-3.61158	-0.12036				

Sensor 1(MC) structural coordinates

С	5.27391	-2.4027	-0.46016	0	1.61985	1.21483	2.80677
С	5.62874	-1.16485	-1.00067	Ν	-6.0465	-1.04299	0.64108
С	4.6906	-0.13544	-1.13021	0	-5.72276	-1.97821	1.39201
С	3.39042	-0.38943	-0.69761	0	-7.23036	-0.73717	0.41515
С	3.01521	-1.63123	-0.17694	Н	6.02032	-3.18583	-0.36529
С	3.95628	-2.64122	-0.04575	Н	6.65036	-0.99267	-1.3274
Ν	2.26593	0.45748	-0.73396	Н	4.9708	0.82138	-1.55261
С	1.13931	-0.20969	-0.367	Н	3.67806	-3.60745	0.36632
С	1.52982	-1.61182	0.13587	Н	-1.23349	-1.12223	0.5672
0	-2.11432	1.85488	-1.98539	Н	-0.2555	1.23287	-1.12414
С	-2.97096	1.2165	-1.34748	Н	-4.64149	2.30743	-2.19444
С	-2.64148	0.12406	-0.39614	Н	-6.42254	0.99849	-1.0296
С	-1.31277	-0.2709	-0.10378	Н	-3.45801	-1.39872	0.91313
С	-0.14187	0.31889	-0.55687	Н	1.19902	-3.71178	-0.30702
С	-4.39699	1.49527	-1.51684	Н	-0.261	-2.72671	-0.47401
С	-5.37014	0.78461	-0.88485	Н	0.99622	-2.65822	-1.71749
С	-5.01359	-0.28076	-0.00376	Н	1.64567	-0.83115	2.19359
С	-3.68676	-0.5886	0.22862	Н	0.22865	-1.84521	1.88829
С	0.81752	-2.74146	-0.64113	Н	1.81298	-2.61912	2.03952
С	1.29337	-1.72867	1.66815	Н	1.36671	2.31867	-0.7776
С	2.34917	1.90772	-0.9962	Н	2.59137	2.05823	-2.05583
С	3.33904	2.6548	-0.07371	Н	3.02408	3.70182	-0.12396
С	3.28493	2.19672	1.38841	Н	4.36348	2.60168	-0.46456
С	1.8216	2.11555	1.95165	Н	3.84863	2.91685	1.99734
0	1.02096	2.95906	1.4759	Н	3.78048	1.22911	1.52155

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Sensor 2(MC) structural coordinates

С	4.89339	-2.27376	-0.35189
С	5.13107	-1.11982	-1.10065
С	4.12441	-0.17131	-1.3121
С	2.87322	-0.41403	-0.74556
С	2.61478	-1.58035	-0.01515
С	3.62313	-2.50762	0.19501
Ν	1.70321	0.3573	-0.82063

0.63545	-0.32058	-0.29162
1.15613	-1.58877	0.41121
-2.82145	1.19375	-2.0613
-3.59625	0.62389	-1.26534
-3.14832	-0.24615	-0.15231
-1.79527	-0.49089	0.12708
-0.67795	0.08343	-0.49027

С	-5.04467	0.77387	-1.40679	Н	-1.61787	-1.1913	0.93982
С	-5.93547	0.1381	-0.5908	Н	-0.88199	0.88327	-1.19026
С	-5.45984	-0.70325	0.45015	Н	-5.37728	1.42265	-2.21123
С	-4.13077	-0.89345	0.66859	Н	-7.00925	0.25627	-0.71212
С	0.47081	-2.87833	-0.08915	Н	-3.80999	-1.53651	1.4838
С	1.03932	-1.43586	1.95366	Н	0.93843	-3.75054	0.38029
С	1.66465	1.76589	-1.25466	Н	-0.59247	-2.8912	0.15761
F	-6.38742	-1.32282	1.23642	Н	0.56767	-2.98317	-1.17409
С	2.70648	2.67248	-0.56393	Н	1.37895	-0.44479	2.28465
С	2.83047	2.43316	0.94458	Н	0.0001	-1.55271	2.27523
С	1.44537	2.46059	1.68491	Н	1.63222	-2.22117	2.43708
0	0.60741	3.26861	1.21247	Н	0.69014	2.14977	-0.95852
0	1.33825	1.67678	2.66428	Н	1.75804	1.80842	-2.3479
Н	5.69118	-2.99431	-0.19664	Н	2.33173	3.68943	-0.71833
Н	6.11465	-0.94948	-1.53008	Н	3.6848	2.60351	-1.05756
Н	4.31605	0.71854	-1.89858	Н	3.4582	3.23017	1.36779
Н	3.43317	-3.40955	0.77116	Н	3.34337	1.49036	1.16104

Fluorescence Experiments

Job's Plots

Stock solutions of spiropyran chemosensor (SP, 100 μ M) and metal ion salts (M, 100 μ M) were prepared separately in HPLC-grade acetonitrile. Solutions were prepared (in triplicate) in the same clear-bottom microplate tray from varying volumes of the spiropyran stock solution and of the ion stock solutions, respectively, until the total volume of each replicate was 200 μ L. The concentration ratios of ([SP], [M]) in μ M were (100, 0), (95, 5), (80, 20), (70, 30), (50, 50), (40, 60), (30, 70), (20, 80), (5, 95) and (0, 100). As such, each solution contained a constant total combined concentration of spiropyran and metal ion ([SP]+ [M] = 100 μ M). Fluorescence emission spectra were recorded between 555 and 800 nm, respectively, at 25 °C using a BioTek Synergy H4 Hybrid Multi-Mode Microplate Reader. Fluorescence excitation was at 532 nm, with 100 gain setting and scanning resolution was 5nm, with band pass of 9 nm. Job's plots were derived by plotting the mean fluorescence at the maximum emission wavelength for each concentration ratio of (SP], [M]).

Dissociation Constants (Kd)

Stock solutions of metal ion salts (0.02-2 mM) were prepared separately in HPLC-grade acetonitrile. Replicate solutions were prepared (in triplicate) in the same clear-bottom microplate tray from 2 μ L of spiropyran stock (5 mM) and 10 μ L of the respective ion stock solutions. 188 μ L of HPLC grade acetonitrile was then added to dilute each replicate, such that the final concentrations of spiropyran and metal ions in each solution were 50 μ M and 1-100 μ M, respectively. Fluorescence emission spectra were recorded between 555 and 800 nm, at 25 °C using a BioTek Synergy H4 Hybrid Multi-Mode Microplate Reader. Fluorescence excitation was at 532 nm, with 100 gain setting and scanning resolution was 5nm, with band pass of 9 nm. Concentration curves were prepared from the fluorescence emission maxima for each ion concentration. The apparent dissociation constants (K_d) of spiropyrans for metal ions were then calculated by fitting an appropriate non-linear regression in GraphPad Prism version 7.02. The 'Hill Plot with Specific Binding' model was selected, as it represents a saturation binding experiment, where concentration of 'radioligand' (i.e. metal ion) is varied and binding to the 'recepter' (i.e. merocyanine isomer) is measured, in this case as a fluorescence emission. The model uses the following equation, $Y = Bmax^*X^h/(K_d^h + X^h)$

Where Y is the relative fluorescence intensity at any given concentration of metal ion, Bmax is the maximum specific binding in the same units as Y (i.e. in this case the fluorescence at sensor saturation), X is the concentration of metal ion and K_d is the metal ion concentration needed to achieve the half-maximum binding at equilibrium, expressed in the same units as X. The parameter h is the Hill slope, and h = 1.0 when a monomer binds with no cooperativity to one site. If h > 0, i.e. the 'receptor' (merocyanine) or 'radioligand' (metal ion) has multiple binding sites with positive cooperativity, and the graph will have a sigmoidal appearance [1].

Quantum Yield (Φ)

A stock solution containing spiropyran chemosensor (SP, 50 μ M) and metal ion salt (M, 100 μ M) was prepared in HPLC-grade acetonitrile. Solutions were prepared (in triplicate) on the same clear-bottom microplate tray from varying volumes of the combined spiropyran/metal ion stock solution and HPLC-grade acetonitrile, until the volume of each replicate was 200 μ L. The concentration ratios of ([SP], [M]) in μ M were (10, 20), (20, 40), (30, 60), (40, 80), (50, 100), (60, 120), (80, 160) and (100, 200). Absorbance (400-600 nm, 2nm band pass) and fluorescence emission spectra (535-800 nm, 5nm band pass, excitation 514 nm, 80 gain) were recorded at 25 °C using a BioTek Synergy H4 Plate Reader. These absorbance and fluorescence measurements was repeated for Rhodamine B (0, 1, 2, 3, 4, 5 μ M), and a graph of integrated fluorescence *vs* absorbance at 514 nm (up to an approximate absorbance value of 0.1) was obtained. Quantum yield was calculated using the following equation:

$\Phi_x = \Phi_{ST} (Grad_x/Grad_{ST})(\eta^2_x/\eta^2_{ST})$

Where subscripts ST and X denote standard (Rhodamine B) and the unknown (spiropyran), respectively. Φ is fluorescence quantum yield, Grad is the gradient from the plot of integrated fluorescence intensity vs absorbance and η is the refractive index of the solvent ($\eta_{(water)} = 1.330$, $\eta_{(acetonitrile)} = 1.344$). The fluorescence quantum yield of Rhodamine B in water at $\lambda_{ex} = 514$ nm is 0.31, as reported in the literature [2].



Figure S10. Absorbance spectra of (a) chemosensor **1** (50 μ M) and (b) chemosensor **2** (50 μ M) in the presence of an excess of biologically relevant metal ions (100 μ M). Experiments were performed under ambient light conditions in acetonitrile solvent.



Figure S11. Job's plot of chemosensor **1** (50 μ M) in the presence of Ca²⁺ ions (100 μ M). Fluorescence excitation was at 532 nm, with experiments performed under ambient light conditions in acetonitrile solvent.



Figure S12. Metal ion titration curves of chemosensor **1** (50 μ M) with (a) Mg²⁺ (Kd (Mg²⁺) = 6.0 \pm 0.3 μ M, h = 4.3 \pm 0.9) and (b) Ca²⁺ (Kd (Ca²⁺) = 18.7 \pm 0.6 μ M, h = 4.0 \pm 0.5) measured from their respective maximum emissions. Experiments were recorded in HPLC-grade acetonitrile, with excitation at 532 nm under ambient light conditions.



Figure S13. Integrated fluorescence spectra after excitation at 514 nm versus the absorbance at 514 nm for Rhodamine B (0 – 5 μ M, water); (1)MC-Mg²⁺ (0-100 μ M, acetonitrile) in the presence of a 2-fold excess of Mg²⁺; and (1)MC-Ca²⁺ (0-100 μ M, acetonitrile) in the presence of a 2-fold excess of Ca²⁺. Spectra were recorded under ambient light conditions, in acetonitrile solvent. The quantum yield of (1)MC-Mg²⁺ was calculated to be $\Phi = 0.20$, and the quantum yield of (1)MC-Ca²⁺ was calculated to be $\Phi = 0.06$.



Figure S14. Fluorescence emission spectra of chemosensor **2** (50 μ M) in the absence (black) and presence of excess Mg²⁺ (red, 100 μ M) and Ca²⁺ (blue, 100 μ M), respectively. Inset: Selectivity profile of chemosensor **2** in the presence of various biologically relevant metal ions (100 μ M), with λ_{max} at 560 nm. Excitation was at 532 nm, and all experiments were performed under ambient light conditions, in HPLC-grade acetonitrile solvent.



Figure S15. Job's plot of chemosensor **2** (50 μ M) in the presence of Mg²⁺ ions (100 μ M), with λ_{max} at 560 nm. Fluorescence excitation was at 532 nm, with experiments performed under ambient light conditions in acetonitrile solvent. The plot suggests a more complex binding stoichiometry for chemosensor **2**.



Figure S16. Selectivity profiles of (a) compound **3** (50 μ M) and (b) compound **4** (50 μ M) in the presence of an excess of biologically relevant metal ions (100 μ M) in HPLC-grade acetonitrile. Fluorescence excitation was at 532 nm.

References

- "Equation: Specific binding with Hill slope", GraphPad Curve Fitting Guide. Accessed 10 April 2018. <u>https://www.graphpad.com/guides/prism/7/curve-fitting/index.htm?reg_specific_hill.htm</u>.
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