

Table S1. AP2 and RAV genes identified in Carnation DB.

Gene Name	ORF ID	Scaffold ID	CDS (bp)	Amino Acid Residue Number	Information on Expression
<i>DcAP2-1</i>	Dca57571.1	scaffold83	1782	594	EST [47], TSA [48]
<i>DcAP2-2</i>	Dca21030.1	scaffold2131	1071	357	-
<i>DcAP2-3</i>	Dca22443.1	scaffold225	1662	554	TSA [48]
<i>DcAP2-4</i>	Dca44633.1	scaffold515	1365	455	TSA [48]
<i>DcAP2-5</i>	Dca3403.1	scaffold1120	1884	628	TSA [48]
<i>DcAP2-6</i>	Dca28099.1	scaffold28	1491	497	TSA [48]
<i>DcAP2-7</i>	Dca24904.1	scaffold248	1716	572	-
<i>DcAP2-8</i>	Dca20778.1	scaffold211	1431	477	-
<i>DcAP2-9</i>	Dca29759.1	scaffold297	1887	629	-
<i>DcRAV1</i>	Dca18854.1	scaffold1983	1086	362	EST [47], TSA [48]

Table S2. ERF genes identified in Carnation DB.

Gene Name	ORF ID	Scaffold ID	CDS (bp)	Amino Acid Residue Number	Information on Expression
<i>DcERF1</i>	Dca51601.1	scaffold67	813	271	Nucleotide [26], EST [47], TSA [48]
<i>DcERF2</i>	Dca17620.1	scaffold19	963	321	EST [47]
<i>DcERF3</i>	Dca2902.1	scaffold110	894	298	EST [47], TSA [48]
<i>DcERF4</i>	Dca38307.1	scaffold41	864	288	-
<i>DcERF5</i>	Dca23859.1	scaffold2381	1218	406	-
<i>DcERF6</i>	Dca62148.1	scaffold9805	552	184	EST [47]
<i>DcERF7</i>	Dca42253.1	scaffold4737	816	272	-
<i>DcERF8</i>	Dca61162.1	scaffold9463	915	305	-
<i>DcERF9</i>	Dca21337.1	scaffold216	723	241	-
<i>DcERF10</i>	Dca57583.1	scaffold830	738	246	EST [47], TSA [48]
<i>DcERF11</i>	Dca57446.1	scaffold8257	576	192	-
<i>DcERF12</i>	Dca32515.1	scaffold33	585	195	-
<i>DcERF13</i>	Dca39896.1	scaffold434	939	313	EST [47]
<i>DcERF14</i>	Dca43835.1	scaffold50	951	317	TSA [48]
<i>DcERF15</i>	Dca35739.1	scaffold373	1062	354	EST [47], TSA [48]
<i>DcERF16</i>	Dca48192.1	scaffold592	1071	357	EST [47], TSA [48]
<i>DcERF17</i>	Dca56998.1	scaffold813	1002	334	TSA [48]
<i>DcERF18</i>	Dca57004.1	scaffold813	996	332	EST [47], TSA [48]
<i>DcERF19</i>	Dca55626.1	scaffold777	804*	268*	-
<i>DCERF20</i>	Dca10360.1	scaffold14520	645	215	EST [47]
<i>DcERF21</i>	Dca39736.1	scaffold4310	612	204	EST [47], TSA [48]
<i>DcERF22</i>	Dca21114.1	scaffold214	789	263	EST [47], TSA [48]
<i>DcERF23</i>	Dca11198.1	scaffold15	945	315	EST [47], TSA [48]
<i>DcERF24</i>	Dca11199.1	scaffold15	801	267	-
<i>DcERF25</i>	Dca52842.1	scaffold7	1557	519	TSA [48]
<i>DcERF26</i>	Dca3623.1	scaffold113	1221	407	EST [47]
<i>DcERF27</i>	Dca12450.1	scaffold157	978	326	EST [47], TSA [48]
<i>DcERF28</i>	Dca17458.1	scaffold189	1005	335	TSA [48]
<i>DcERF29</i>	Dca4734.1	scaffold1179	1017	339	EST [47], TSA [48]
<i>DcERF30</i>	Dca61124.1	scaffold945	915	305	-
<i>DcERF31</i>	Dca16525.1	scaffold1819	1938	646	TSA [48]
<i>DcERF32</i>	Dca49218.1	scaffold6121	1479	493	-

* Data were obtained from the cDNA sequences cloned from ‘West Diamond’ (LC659678) and ‘Ekubo’ (LC659682).

Table S3. Primers used for cDNA cloning and real-time RT-PCR.

Gene	Forward Primer (5' to 3')	Reverse Primer (5' to 3')
cDNA cloning (3' RACE)		
<i>DcERF19</i>	CCCGAAAGCCATTACAACAGC	GTTTCCAGTCACGAC*
cDNA cloning (whole CDS)		
<i>DcSUS2</i>	CAGAAAATGGCAAGTCGTTTGAC	CGACCAGCGGAGAACACGTA
<i>DcADH1</i>	TAATGTCGAGTACCGCCGGACAAG	GATGTCGATTCAAGCGTCC
<i>DcERF19</i>	CATGTGTGGTGGTGCAATTTTAGCCA	GCAGCTTATCTTTGTTGCAAACA
<i>DcPGB1</i>	GAGGGAAGCAAATAACATGG	CCGGAGATGAACAAGCAGAA
Real-time RT-PCR		
<i>DcERF15</i>	GGATGGTCTGACCAATGTGG	CGAACAGATGACCCTGCGAA
<i>DcERF16</i>	GCTAGCGAGTTTTCTCAGGA	GCTCGGAAATCCATTGCCAA
<i>DcERF17</i>	GAGACGTCGTTCGTGGACGA	GCGATAACAGATACATGTCTGTAGAC
<i>DcERF18</i>	GAGACGCCATTCGTGGACAA	ACGATAACAGATACACGTCCATC
<i>DcERF19</i>	GCTGCGTAAAACTGTTAGG	GCAGCTTATCTTTGTTGCAAACA
<i>DcSUS2</i>	GGCTGCTAACTCTTGCGGGT	CGACCAGCGGAGAACACGTA
<i>DcADH1</i>	CAAGCCTAGAACCGACATAC	GATGTCGATTCAAGCGTCC
<i>DcPGB1</i>	CTGCGATTCAACTGCGAGAG	GCAGTGCTTCTTTCACAACC
<i>DcUbq3-7</i>	GTTGTTGGTTTCAGGGCTGGTTTG	CTACGGTAATTGAGAATTCACACCGAAATG

* M13PrimerM4

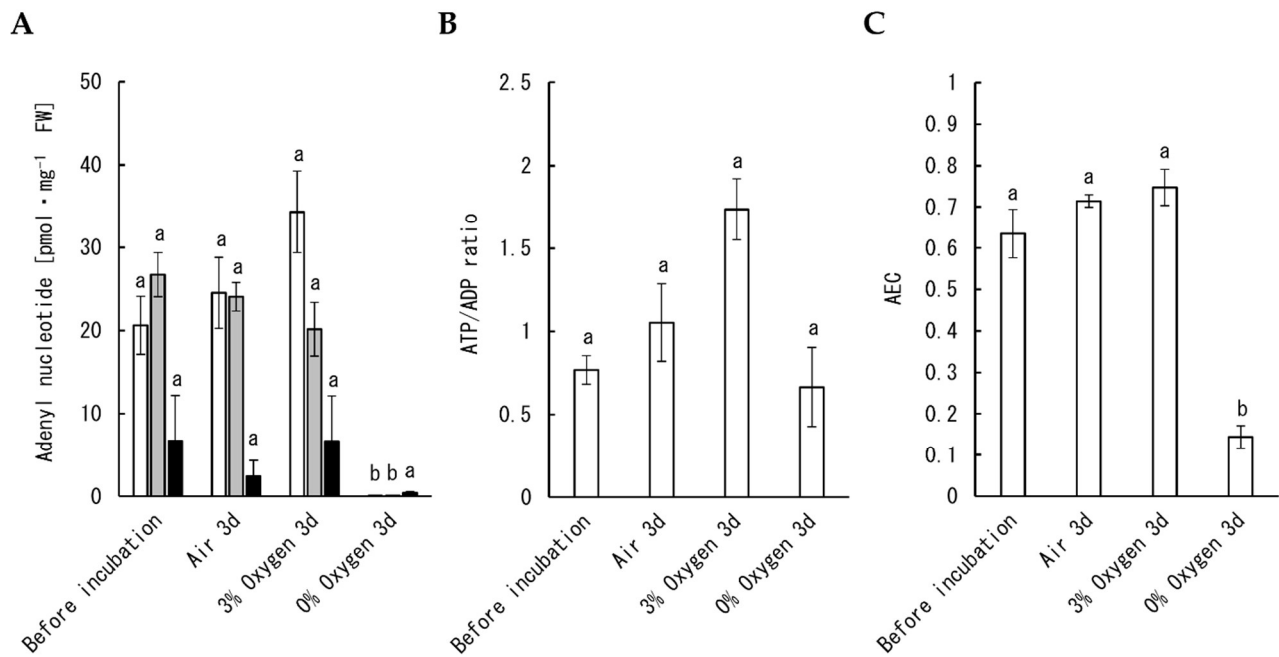


Figure S1. Effects of hypoxia and anoxia on adenyly nucleotide contents, ATP/ADP ratio, and AEC in carnation petals. Contents of ATP (white bars), ADP (gray bars), and AMP (black bars) in petals before and after incubation in air, and under hypoxia (3% oxygen) and anoxia (0% oxygen) for three days were determined using a luminometric method (**A**), and used for calculation of ATP/ADP ratio (**B**) and AEC (**C**). Data are expressed as the mean \pm SE of three separate samples. Significant differences ($p < 0.05$) detected using Tukey's multiple comparison test are indicated by different letters above the bars.



Figure S2. Multiple alignment of deduced amino acid sequences of AP2 and RAV from carnation. The sequences were aligned using ClustalW and BioEdit software. Identical or similar amino acids are indicated by a black or gray background, respectively, and gaps are indicated by dashes. The two AP2 (or ANT) domains, the EAR motif-like sequences, a putative nuclear localization signal (NLS) motif, a liker domain and the sequence derived from the target site for miR172 binding described in previous studies [33,34] are indicated by color-coded boxes. The B3-like domain conserved in RAV is indicated by blue underlines.

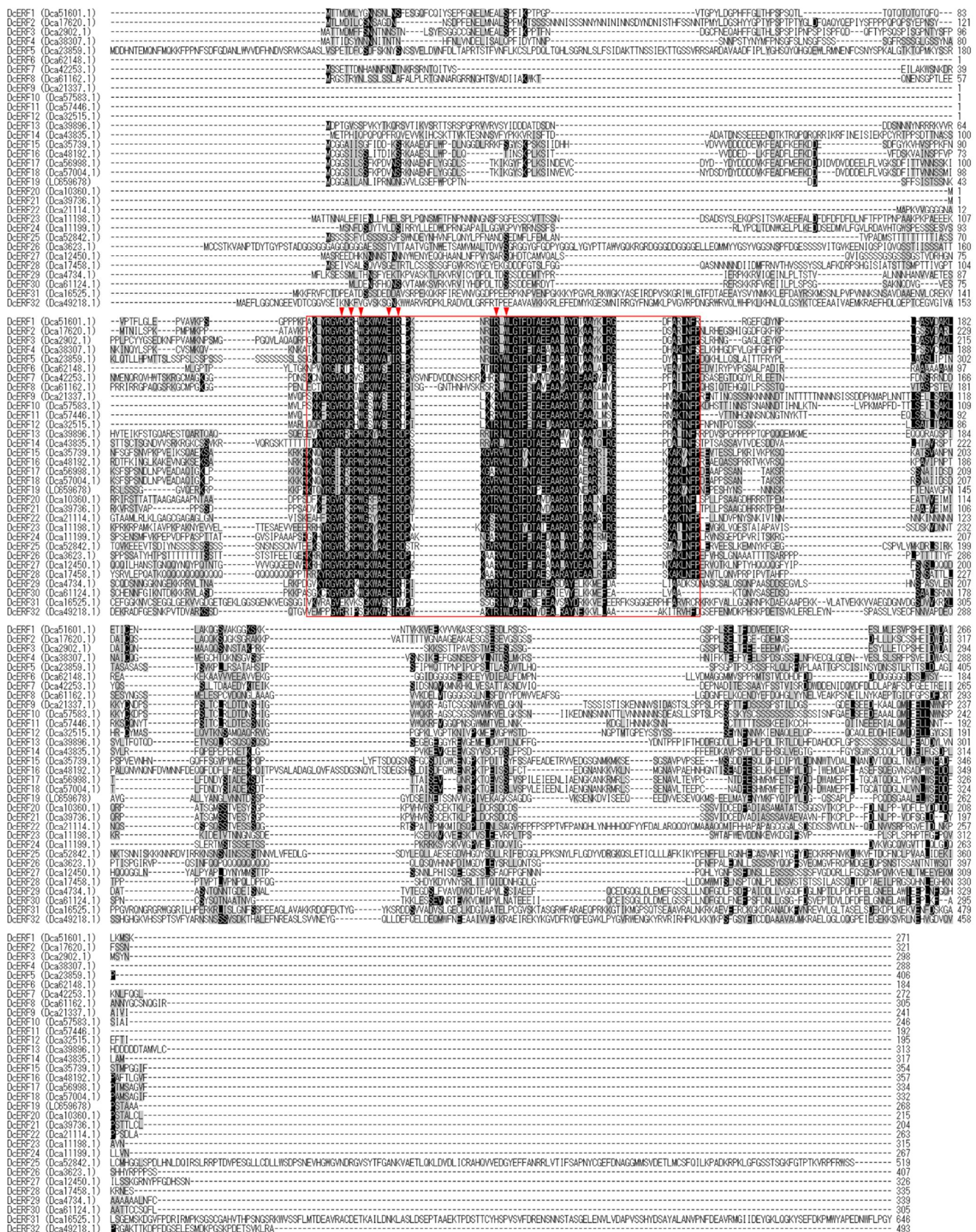


Figure S3. Multiple alignment of deduced amino acid sequences of ERF from carnation. The sequences were aligned using ClustalW and BioEdit software. Identical or similar amino acids are indicated by a black or gray background, respectively, and gaps are indicated by dashes. AP2/ERF domain and amino acid residues directly contact with DNA [18] are indicated by a box and arrowheads, respectively.

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PGB1 (At2g16060) MESEGKIVFTEEQEALVWKSWSVMKKNSAELGKLFIKLFEIAPITTKMFSLRDSPIPA 60
DcPGB1 (Dca3435.1) -----MVETEKEESLVKESWEILKLNIPENSLRFITILLIETAPAAKDLFSFLRDSQVPS 54

PGB1 (At2g16060) EQNPKLPHAMSVFVMCDESANQLRKTGKVTIETITLKR LGASHSKYGVVDEHFEVAKYA 120
DcPGB1 (Dca3435.1) QNNPKLKAHAMKVFKITDESATQLRECEMVVGDSTLKY LGAIHSNSGVVGPVFEVWKEA 114

PGB1 (At2g16060) LETIKAEVPEMWSPKVMVWGQAYDHLVAATKAEMLSN- 160
DcPGB1 (Dca3435.1) LLKTIQEAVGDKWNQQMSCAWAAAYDQAAATKSEMNHPTS 155

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Figure S4. Multiple alignment of deduced amino acid sequences of PGB from *Arabidopsis* and carnation. The sequences were aligned using ClustalW and BioEdit software. Identical or similar amino acids are indicated by a black or gray background, respectively, and gaps are indicated by dashes. The conserved amino acid residues involved in heme and ligand binding are indicated by arrowheads.

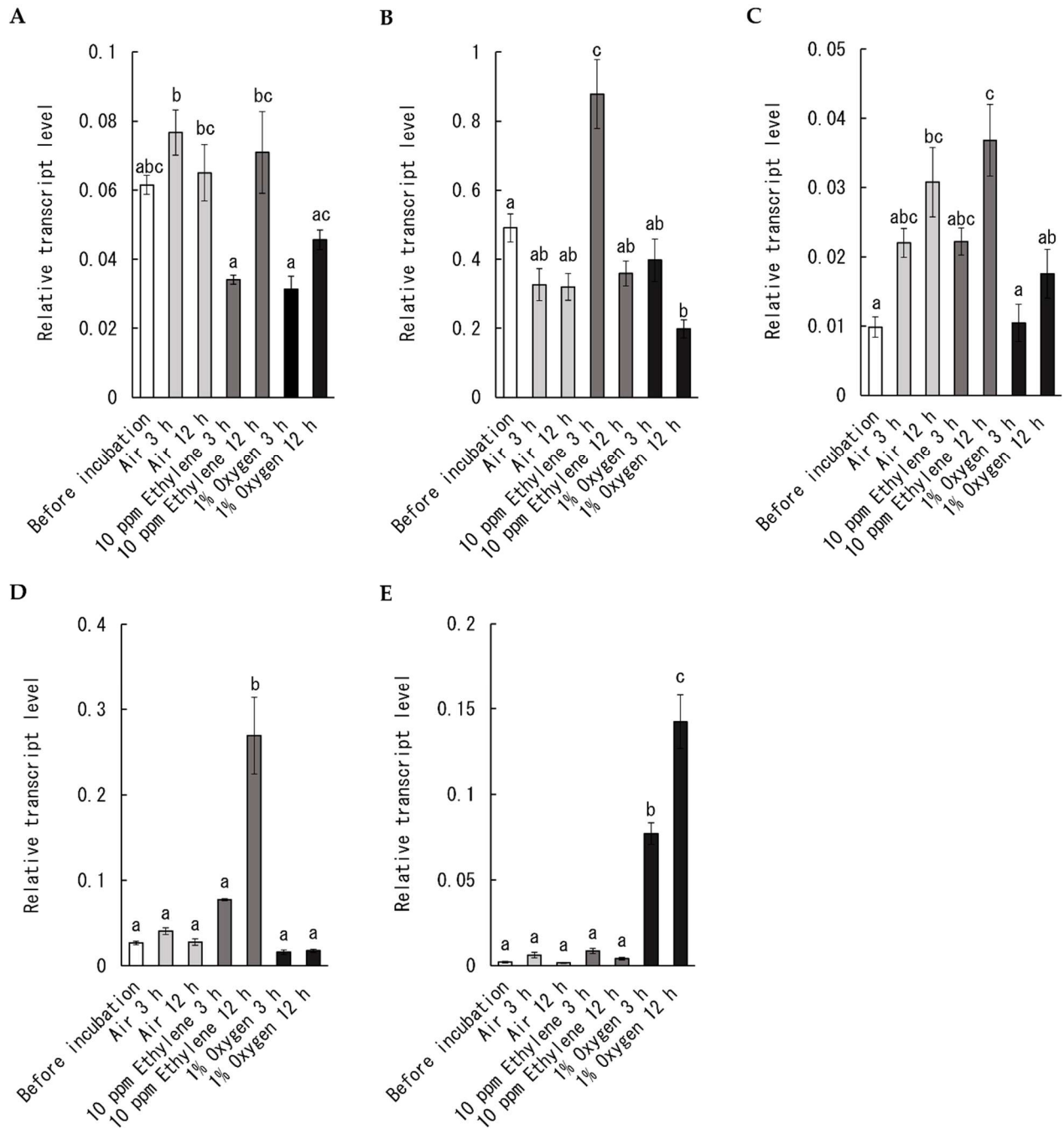


Figure S5. Effects of ethylene and hypoxia on transcript levels of group VII ERF genes in carnation ('West Diamond') petals. Relative transcript levels of *DcERF15* (A), *DcERF16* (B), *DcERF17* (C), *DcERF18* (D), and *DcERF19* (E) in petals before and after incubation in air, and under 10 ppm ethylene and hypoxia (1% oxygen) for 3 and 12 h were determined using real-time RT-PCR with *DcUbg3-7* as a standard. Data are expressed as the mean \pm SE of three separate samples. Significant differences ($p < 0.05$) detected using Tukey's multiple comparison test are indicated by different letters above the bars.

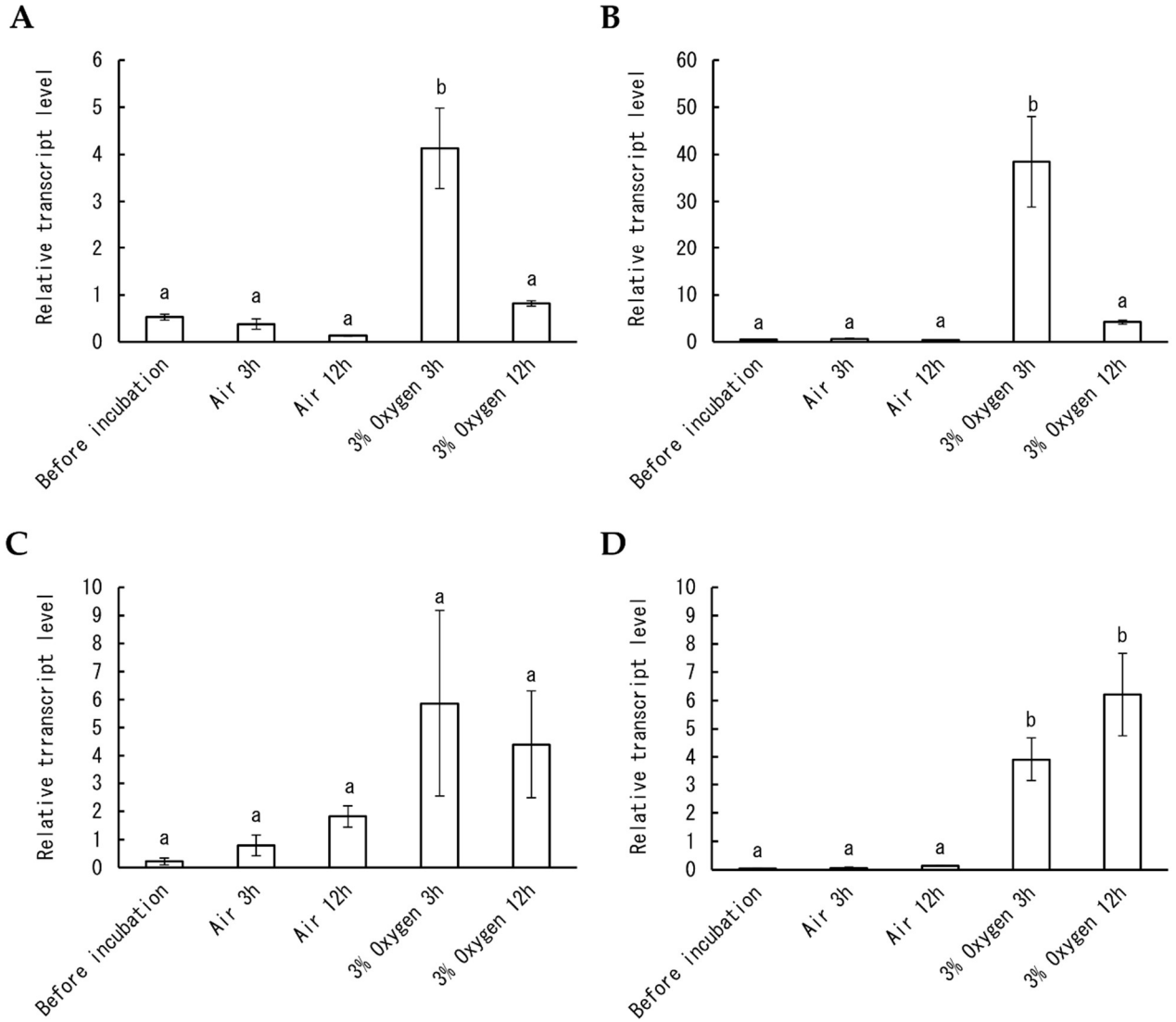


Figure S6. Effects of hypoxia on transcript levels of hypoxia-related genes in carnation petals. Relative transcript levels of *DcSUS2* (A), *DcADH1* (B), *DcERF19* (C), and *DcPGB1* (D) in petals before and after incubation in air, and under hypoxia (3% oxygen) for 3 and 12 h were determined using real-time RT-PCR with *DcUbq3-7* as a standard. Data are expressed as the mean \pm SE of three separate samples. Significant differences ($p < 0.05$) detected using Tukey's multiple comparison test are indicated by different letters above the bars.