

# Supplemental Material

## Emergence and Enhancement of Ultrasensitivity through Posttranslational Modulation of Protein Stability

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### Ordinary differential equations for the full model

$$\frac{dR}{dt} = k_0 - k_3 * R - k_{1f} * X * R + k_{1b} * RX + k_{2c} * R_p Y, \quad (S1)$$

$$\frac{dR_p}{dt} = k_{1c} * RX - k_4 * R_p - k_{2f} * R_p * Y + k_{2b} * R_p Y, \quad (S2)$$

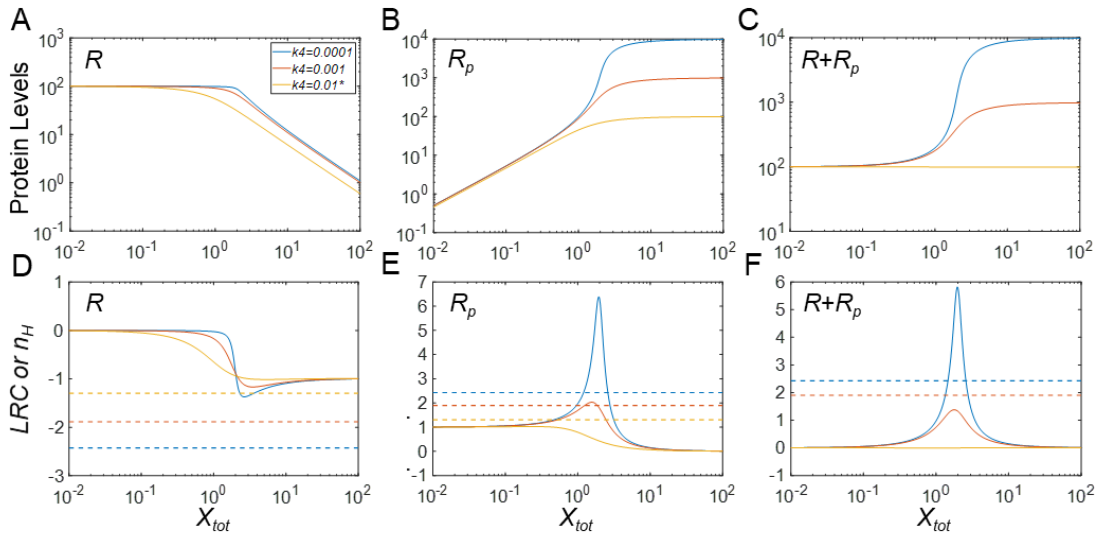
$$\frac{dRX}{dt} = k_{1f} * X * R - k_{1b} * RX - k_{1c} * RX - k_3 * RX, \quad (S3)$$

$$\frac{dR_p Y}{dt} = k_{2f} * R_p * Y - k_{2b} * R_p Y - k_{2c} * R_p Y - k_4 * R_p Y, \quad (S4)$$

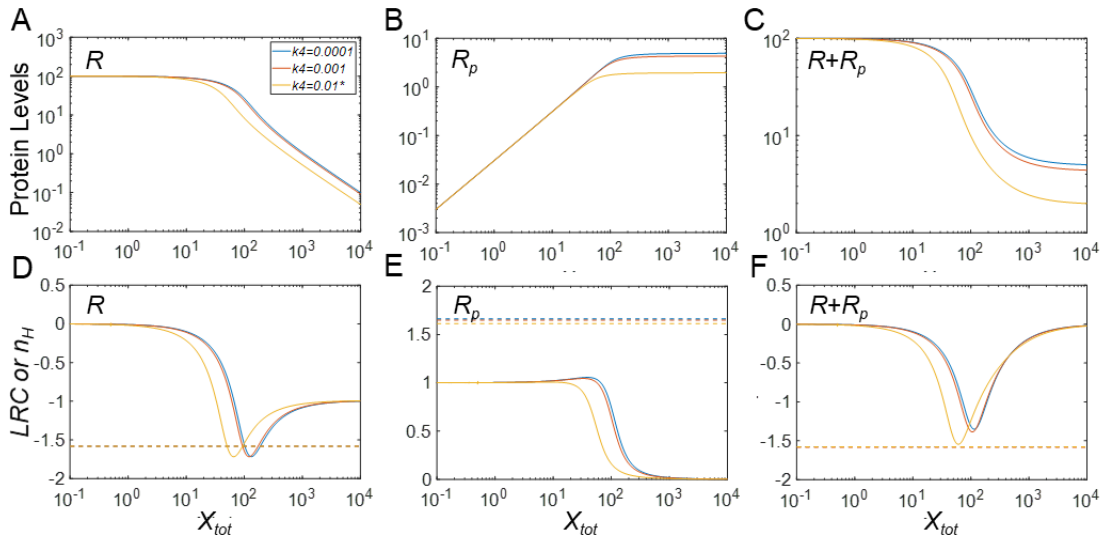
where  $X = X_{tot} - RX$  and  $Y = Y_{tot} - R_p Y$ .

### Additional Figures and Explanations

#### 1. Emergence of ultrasensitivity in the full model

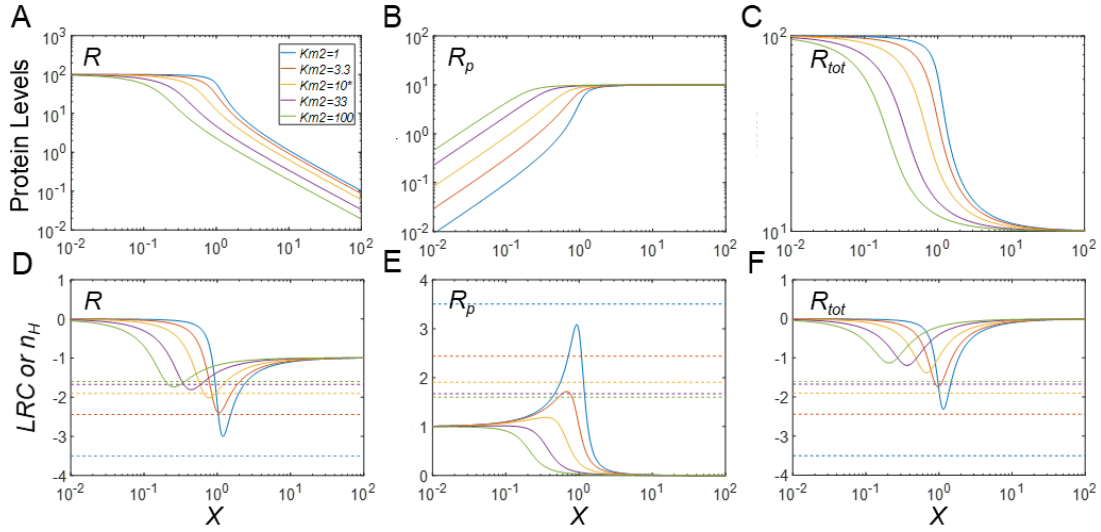


**Figure S1. Emergence of ultrasensitivity through phosphorylation-induced protein stabilization in the full model (with increased apparent  $K_m$  values).** (A-C) Steady-state DR curves for  $R$  vs.  $X_{tot}$ ,  $R_p$  vs.  $X_{tot}$ , and free  $R_{tot}$  ( $R+R_p$ ) vs.  $X_{tot}$ , respectively, for different values of  $k_4$ , as indicated in panel A. The same color-scheme for  $k_4$  values holds for all panels. As  $k_4$  decreases, ultrasensitivity emerges for  $R_p$  and free  $R_{tot}$ . (D-F) LRC (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and free  $R_{tot}$ , respectively, for different values of  $k_4$ . \* $k_4=0.01$  is the default value. For these simulations,  $k_{1b}=k_{2b}=990$  such that the apparent  $K_{m1}=K_{m2}=100$ .



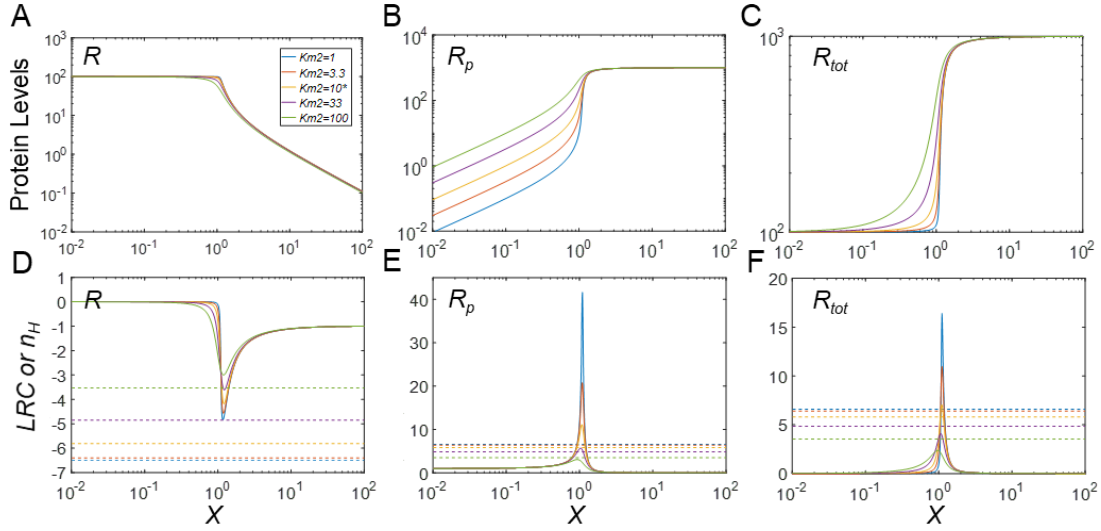
**Figure S2. Emergence of ultrasensitivity through phosphorylation-induced protein stabilization in the full model (with increased enzyme  $Y$  level).** (A-C) Steady-state DR curves for  $R$  vs.  $X_{tot}$ ,  $R_p$  vs.  $X_{tot}$ , and free  $R_{tot}$  ( $R+R_p$ ) vs.  $X_{tot}$ , respectively, for different values of  $k_4$ , as indicated in panel A. The same color-scheme for  $k_4$  values holds for all panels. As  $k_4$  decreases, ultrasensitivity emerges for  $R_p$  and free  $R_{tot}$ . (D-F) LRC (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and free  $R_{tot}$ , respectively, for different values of  $k_4$ . \* $k_4=0.01$  is the default value. For these simulations,  $Y_{tot}=300$ .

## 2. Effects of Michaelis constant $K_{m2}$



**Figure S3. Effects of  $K_{m2}$  on ultrasensitivity with phosphorylation-induced protein destabilization in the MM model ( $k_4 = 0.1$ ).** (A-C) Steady-state DR curves for  $R$  vs.  $X$ ,  $R_p$  vs.  $X$ , and  $R_{tot}$  vs.  $X$ , respectively, for different values of  $K_{m2}$ , as indicated in panel A. The same color-scheme for  $K_{m2}$  is used for all panels. (D-F) LRC (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and  $R_{tot}$ , respectively. \*  $K_{m2}=10$  is the default value.

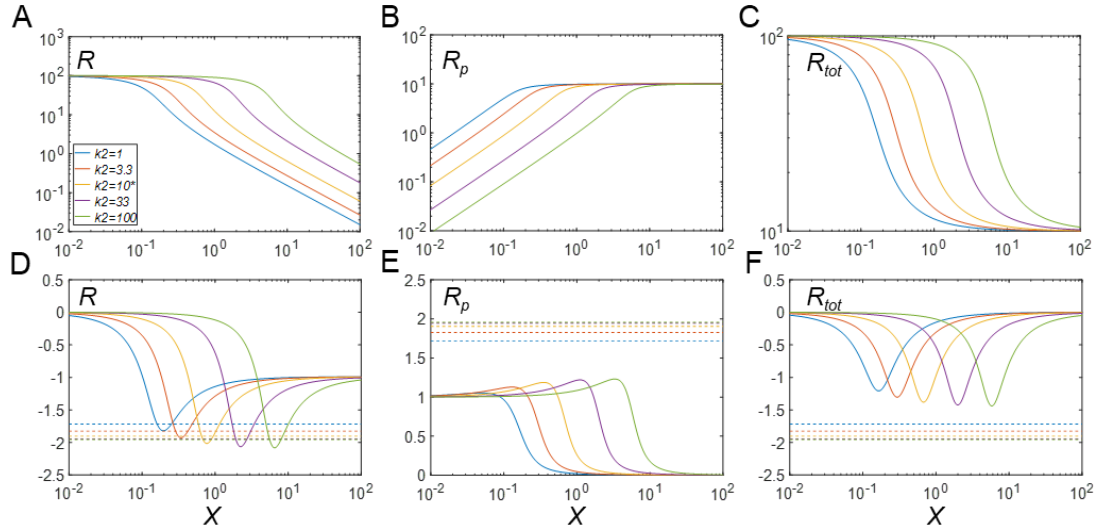
In the analysis leading to Fig. S3, we explore the role of the Michaelis constant  $K_{m2}$  and examine the case where phosphorylation of  $R$  to  $R_p$  results in destabilization; as the baseline, we set  $k_4=0.1$ , which is 10-fold greater than  $k_3$ . As  $K_{m2}$  decreases, the DR curves for  $R$ ,  $R_p$ , and  $R_{tot}$  become increasingly steeper (Figs. S3A-S3C). For low  $K_{m2}$  values,  $|LRC|_{max}$  can be much greater than  $|n_H|$ , whereas for high  $K_{m1}$  values,  $|LRC|_{max}$  approaches 1 for  $R_p$ , indicating loss of ultrasensitivity (Figs. S3B and S3E). For the  $R$  response, increasing  $K_{m1}$  reduces the steepness of the DR curve without complete loss of ultrasensitivity (Figs. S3A and S3D). Lastly, increasing  $K_{m1}$  reduces the steepness of the DR curve for  $R_{tot}$  with  $|n_H|$  approaching a fixed value slightly below 2, and ultrasensitivity is lost for high  $K_{m1}$  values as indicated by  $|LRC|$  (Figs. S3C and S3F).



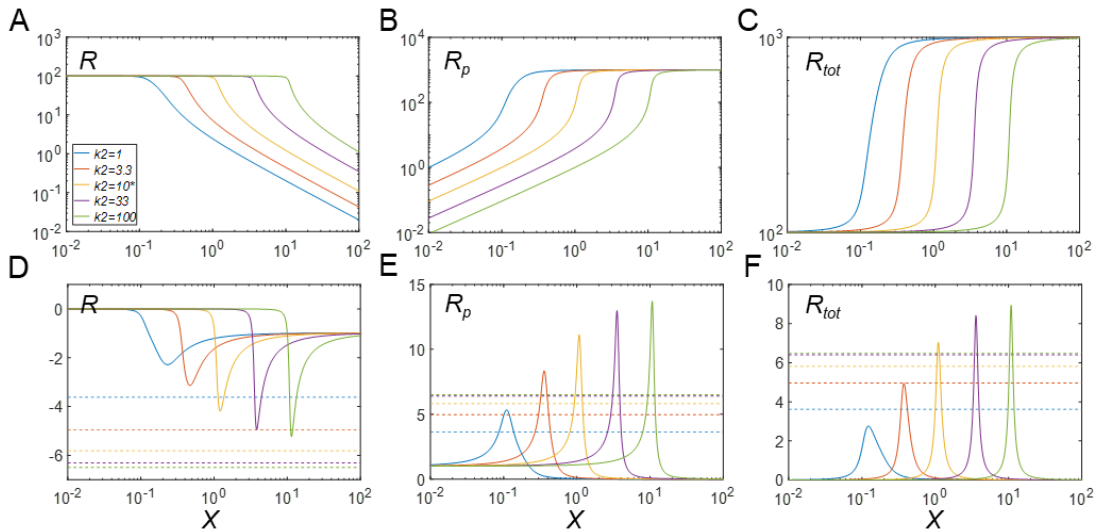
**Figure S4. Effects of  $K_{m2}$  on ultrasensitivity with phosphorylation-induced protein stabilization in the MM model ( $k_4 = 0.001$ ).** (A-C) Steady-state DR curves for  $R$  vs.  $X$ ,  $R_p$  vs.  $X$ , and  $R_{tot}$  vs.  $X$ , respectively, for different values of  $K_{m2}$ , as indicated in panel A. The same color-scheme for  $K_{m2}$  is used for all panels. (D-F) LRC (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and  $R_{tot}$ , respectively. \*  $K_{m2}=10$  is the default value.

The rationale for the second analysis is a situation where phosphorylation of  $R$  into  $R_p$  results in strong protein stabilization ( $k_4=0.001$ , 10-fold lower than  $k_3$ ). When  $K_{m2}$  decreases below its baseline value of 10, the DR curves for  $R_p$  and  $R_{tot}$  become increasingly steeper with minimal changes in the steepness for  $R$  (Fig. S4A-S4C). For the response of  $R$ ,  $|n_H|$  obviously overestimates the degree of ultrasensitivity as evaluated by  $|LRC|_{max}$  (Fig. S4A and S4D). For the  $R_p$  response, increasing  $K_{m2}$  reduces the steepness of the DR curve with  $|LRC|_{max}$  consistently greater than  $|n_H|$  (Fig. S4B and S4E). Lastly, increasing  $K_{m1}$  reduces the steepness of the DR curve for  $R_p$  with  $|n_H|$  overestimating the degree of ultrasensitivity as represented by  $|LRC|_{max}$  for large  $K_{m2}$  values, and underestimating for small  $K_{m2}$  values (Fig. S4C and S4F).

### 3. Effects of catalytic constant $k_2$



**Figure S5. Effects of  $k_2$  on ultrasensitivity with phosphorylation-induced protein destabilization ( $k_4 = 0.1$ ) in the MM model. (A-C)** Steady-state DR curves for  $R$  vs.  $X$ ,  $R_p$  vs.  $X$ , and  $R_{tot}$  vs.  $X$ , respectively, for different values of  $k_2$ , as indicated in panel A. The same color-scheme for  $k_2$  is used for all panels. **(D-F)** LRC (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and  $R_{tot}$ , respectively. \*  $k_2=10$  is the default value.



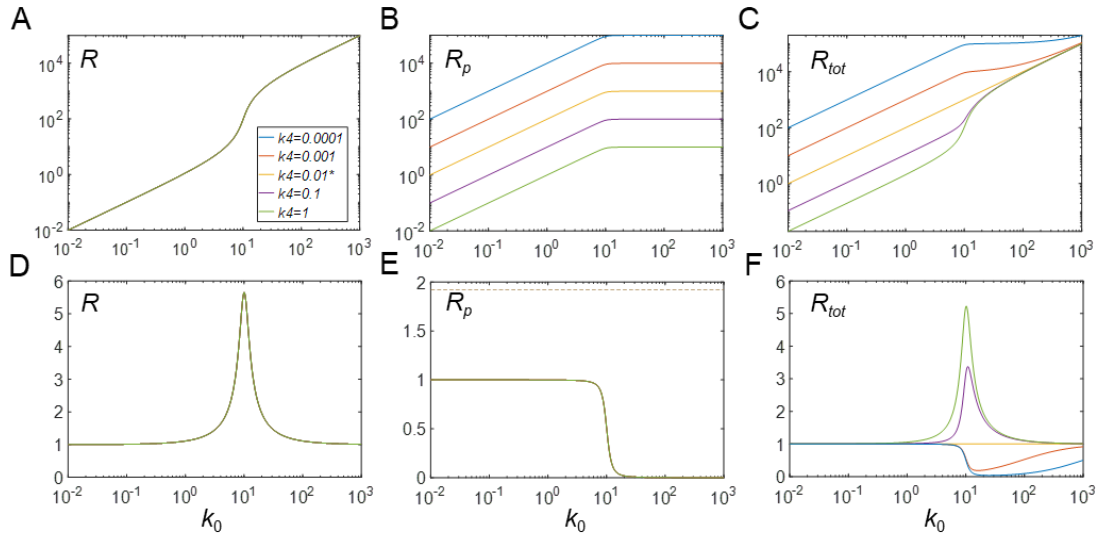
**Figure S6. Effects of  $k_2$  on ultrasensitivity with phosphorylation-induced protein stabilization in the MM model ( $k_4 = 0.001$ ). (A-C)** Steady-state DR curves for  $R$  vs.  $X$ ,  $R_p$  vs.  $X$ , and  $R_{tot}$  vs.  $X$ , respectively,

for different values of  $k_2$ , as indicated in panel A. The same color-scheme for  $k_2$  values is used for all panels.

**(D-F)**  $LRC$  (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and  $R_{tot}$ , respectively. \*  $k_2=10$  is the default value.

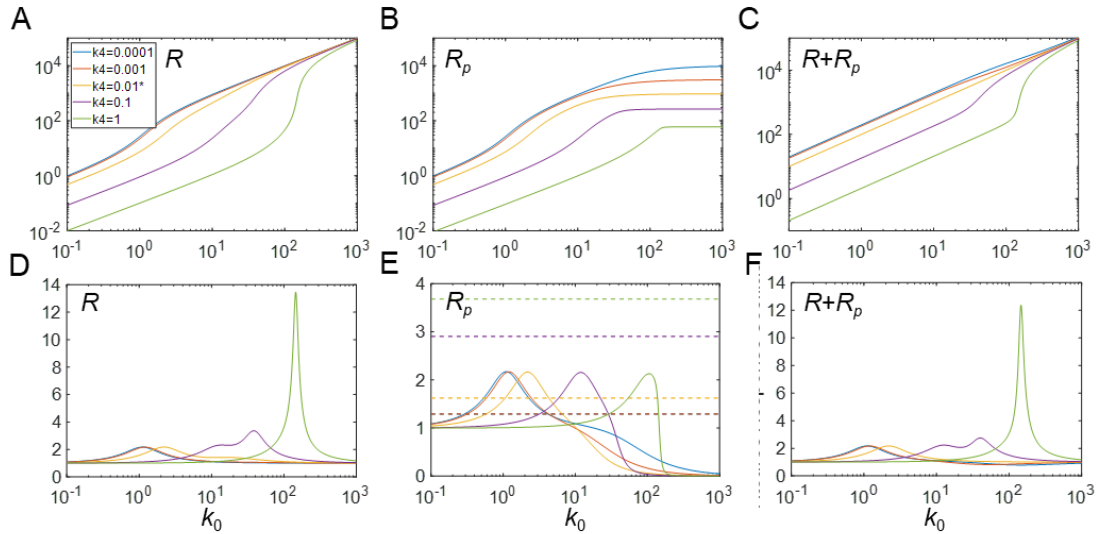
Instead of focusing on alterations in  $K_m$  values, one may explore here the effects of varying  $k_2$ , the catalytic rate of the dephosphorylation step, on ultrasensitive behavior when changes in protein stability are induced by PTMs. We first examine the case where phosphorylation of  $R$  to  $R_p$  results in destabilization ( $k_4=0.1$ , 10-fold greater than  $k_3$ ). As shown in Fig. S5,  $k_2$  only has a marginal effect on the degree of ultrasensitivity as assessed by either  $n_H$  or  $LRC$ , while decreasing  $k_2$  generally sensitizes all responses, with the DR curves shifting to the left. Increasing  $k_2$  appears to increase  $|n_H|$  and  $|LRC|_{max}$  only slightly, and all DR curves approach a fixed value when  $k_2$  becomes very large. For the case where phosphorylation of  $R$  into  $R_p$  results in protein stabilization ( $k_4=0.001$ , 10-fold lower than  $k_3$ ), as shown in Fig. S6, increasing  $k_2$  desensitizes all responses and shifts the DR curves to the right, thereby increasing the degree of ultrasensitivity. However, for very high  $k_2$  values, both  $|n_H|$  and  $|LRC|_{max}$  seem to reach some maximal values.

#### 4. Ultrasensitivity in response to changes in protein synthesis



**Figure S7.  $k_0$ -driven ultrasensitivity with phosphorylation-induced changes in protein stability in the MM model ( $k_2=0$ ).** (A-C) Steady-state DR curves for  $R$  vs.  $k_0$ ,  $R_p$  v.s  $k_0$ , and  $R_{tot}$  vs.  $k_0$ , respectively, for different values of  $k_4$ , as indicated in A. The same color-scheme for  $k_4$  values is used for all panels. (D-F) LRC (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and  $R_{tot}$ . \*  $k_4=0.01$  is the default value.  $X=1$  for all conditions. Note that no  $n_H$  was evaluated for  $R$  and  $R_{tot}$  because the responses do not saturate. Note: in panels A, D and E, all five curves overlap.

To further explore the mechanism of ultrasensitivity driven by the synthesis rate  $k_0$ , the dephosphorylation step is disabled by setting  $k_2=0$ . It is clear for the DR curve of  $R$  vs  $k_0$  that ultrasensitivity remains regardless the direction in which phosphorylation alters the stability of  $R_p$  (Figs. S7A and S7D), suggesting that it is the saturation of the phosphorylation that is responsible for the ultrasensitivity. For the DR curve of  $R_{tot}$  vs  $k_0$ , ultrasensitivity is only present when  $k_4 > k_3$ , i.e., phosphorylation results in destabilization of  $R_p$  (Figs. S7C and S7E), where  $R$  dominates the fraction of  $R_{tot}$ . No ultrasensitivity exists for  $R_p$  (Figs. S7C and S7F).



**Figure S8.  $k_0$ -driven ultrasensitivity with phosphorylation-induced changes in protein stability in the full model ( $X_{tot} = Y_{tot} = 100$ ).** (A-C) Steady-state DR curves for  $R$  vs.  $k_0$ ,  $R_p$  v.s  $k_0$ , and free  $R_{tot}$  ( $R+R_p$ ) vs.  $k_0$ , respectively, for different values of  $k_4$ , as indicated in (A). The same color-scheme for  $k_4$  values is used

for the other panels. **(D-F)**  $LRC$  (solid lines) and  $n_H$  (dashed horizontal lines) for  $R$ ,  $R_p$ , and free  $R_{tot}$ . \*  $k_4=0.01$  is the default value. Note that no  $n_H$  was evaluated for  $R$  and  $R_{tot}$  because the responses do not saturate.