

Supplementary Material

Atlantinone A, a Meroterpenoid Produced by *Penicillium ribeum* and Several Cheese Associated *Penicillium* Species

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Table 1. NMR data of atlantinone A ^a (2).

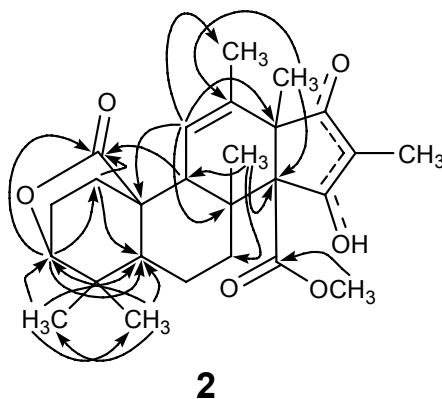
Position	¹ H	¹³ C
1 α	1.33 1H, m	31.5
2 α	1.73 1H, m	21.2
3	4.07 1H, s	83.0
4		36.4
5	1.32 1H, m	50.7
6 α	1.16 1H, m	21.2
7 α	1.93 1H, m	30.9
8		40.7
9	1.80 1H, s	45.4
10		44.8
11	5.35 1H, s	124.4
12		131.8 br.s
13		55.3
14		67.5 br.s
15 ^b		
16		112.0 br.s
17 ^b		
18	1.47 3H, s	6.9
19		170.6
20	1.09 3H, s	15.8
21	1.71 3H, s	19.5
22	1.25 3H, s	16.5
23		174.4
24	1.05 3H, s	27.2
25	0.92 3H, s	21.8
19-OMe	3.48 3H, s	51.1

^a Measured in DMSO-*d*₆ (¹H at 500 MHz, ¹³C at 125 MHz) and referenced to solvent residual signals and solvent signals at 2.50 ppm (¹H-NMR) and 39.50 ppm (¹³C-NMR). Numbering is arbitrary and follows the system used for citreohybridone derivatives (4); ^b Signals not observed.

Structural elucidation. The ¹H-NMR spectrum (Table 1) of **2** showed the presence of an olefinic proton (δ_{H} 5.35, C₁₁-H), one methoxy group (δ_{H} 3.48, C₁₉-OMe), one methyl group attached to a double bond (δ_{H} 1.71, H₃-21), and four methyl groups connected to quaternary carbons [δ_{H} 0.92 (H₃-25), 1.05 (H₃-24), δ_{H} 1.09 (H₃-20), 1.25 (H₃-22)]. The ¹³C-NMR exhibits 24 carbon signals including signals assigned to two carbonyl carbons [δ_{C} 170.6 (C-19), δ_{C} 174.4 (C-23)], two olefinic carbons [δ_{C} 124.4 (C-11), δ_{C} 131.8 (C-12)], two oxygenated carbons [one methine (δ_{C} 83.0, C-3) and one methoxycarbonyl (δ_{C} 51.1, C₁₉-OMe)]. The signals originating from C-15 and C-17 was not observed owing to the fast equilibrium between the C-17-oxo-C-15-hydroxy, and C-17-hydroxy-C-15-oxo tautomeres. Analogue findings have been reported for other andrastins (1, 3). The gross structure of **2**

was determined by detailed analyses of one and two dimensional NMR spectra. From COSY data we inferred the presence of three partial structures; X-CH₂-CH₂-CH-X; X-CH₂-CH₂-CH-O-X and -CH-CH= . The HMBC cross peaks revealed these fragments to form part of the skeleton of ring A, B and C. The ³J_{H-C} established the presence of geminal methyl groups (C-24 and C-25) on the A ring together with a lactone bridge.

Supplementary Figure 1. Important HMBC connectivities in (**2**).



The low field shift of H-3 (δ 4.07) placed the oxygen on C-3. The HMBC connectivities between H-1b, H-3, H-5 and C-23 established the γ -lactone ring bridging C-3 and C-10. Broad carbon signals (C-14 and C-16) and the high field shifted carbon at δ 6.9 (C-18) are indicative of the presence of keto-enol tautomerism at the cyclopentane ring (ring D) as in andrastin A, B, C and D (1, 3). The remaining elements of **2** were connected as shown in Figure 2 in accordance with the HMBC connectivities observed.