# Supplementary Materials: Developmental Control of NRAMP1 (SLC11A1) Expression in Professional Phagocytes

#### Mathieu F. M. Cellier

#### Correlating CAGE signals and other marks of NRAMP1 expression in AMLs.

The area spanning *NRAMP1* exons XIII-XV that comprises DHS F10 and F3 [11] is heavily decorated with K9/27ac and K4me2/3 in K562 cells, whereas CD14<sup>+</sup>MNs show strong marking with K4me1 only (Figure S2B). Histone modifications thus appear consistent with CAGE data (Figure S1B), suggesting that RNA Pol II activity at the 3' end of *NRAMP1* Orf may coincide with absence of full-length gene expression in myeloid cell types either divergent from the myelo-monocytic fate (such as K562 cells) or representing some transformed, aberrant progenitor stage (e.g., NB4 APL, Figure 1). In contrast, RNA Pol II activity at *NRAMP1* TSS characterizes cells either expressing or prone to express *NRAMP1*, such as CD14<sup>+</sup>MNs and HL-60 cells, respectively.

Along these lines, the area of *NRAMP1* exon XV comprises DNAse footprint F3 found in both megakaryocytic progenitors and CMPs that matches a potential binding site for ELF1 revealed by ChIP-seq (see text section 2.3.1.7.). Given ELF1 involvement in gene regulation in both erythrocytes and megakaryocytes [80, 81, 91], its peak of expression in neutrophil precursors from human bone marrow [84] and its ability to either activate transcription [86,89] or mediate transcriptional repression [79], including in MCs [82], data suggest possible cooperation between F3 element overlapping *NRAMP1* exon XV and F10 candidate alternative TSSs located in intron 12. As a result, full-length transcription of *NRAMP1* may be prevented in myeloid cell types that either represent early progenitors of microbicidal phagocytes (i.e., MNs, PMNs and PM eosinophils) or diverge toward different lineages (e.g., erythrocytic/megakaryocytic) or various cell types such as DC and MC.

The suspected *NRAMP1* alternative TSS was also detected in larger CAGE datasets: it is the sole cluster of tags detected in ENCODE CAGE dataset (82 samples, including K562 cells but no other myeloid sample) [92] while it is apparently absent from two other sets that contain data from the monocytic lineage: (FANTOM3: 120 samples representing various human tissues, hence expected to contain resident populations of macrophages, including lung which is a major site of *NRAMP1* expression, [112]) [94] and (FANTOM4: THP1 CAGE, 36 samples including both untreated and phorblol-ester stimulated cells) [93]; and it was found in FANTOM5 (1829 sample-set that includes BloodCAGE data presented in Figure 1 and Figures S3-S7) [16,17].

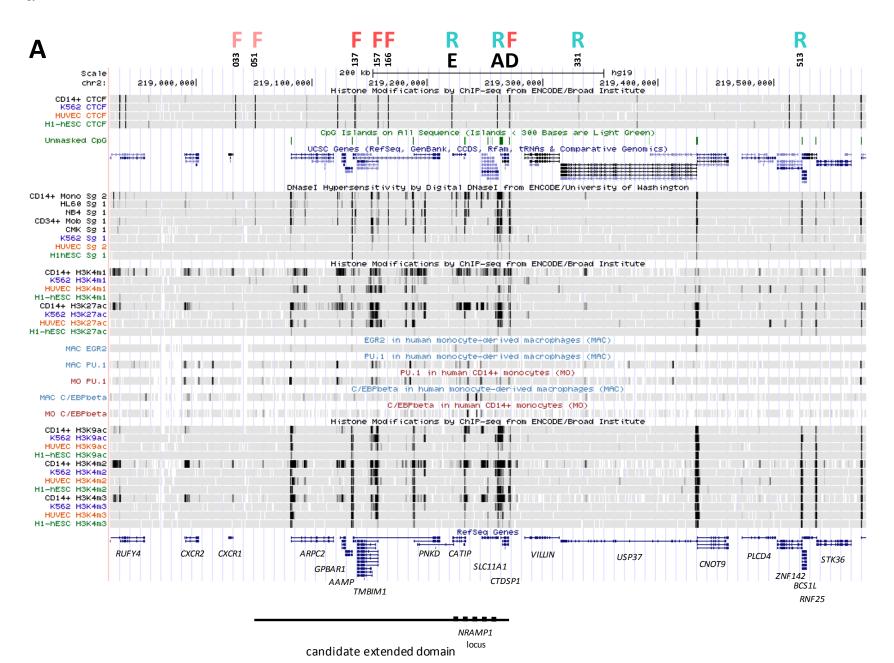
Further examination of *NRAMP1* expression data in THP1 monocytic cells (FANTOM4) shows that the gene is transcribed at low level in untreated cells. Gene expression is significantly induced after 24h treatment with phorblol-ester, to a level that is maintained slightly increased after 96h. These data confirm previous results obtained with HL-60 cells showing that *NRAMP1* expression is not induced rapidly during macrophage-like differentiation stimulated with phorbol ester (maximal induction attained after 48h treatment; [74]). In addition, examination of *NRAMP1* transcript level after siRNA perturbation against 53 TFs showed an up-regulation trend (twofold or more) in three instances: CTCF, IRF7 and MYC, suggesting that these factors may interfere with *NRAMP1* expression [93]. However since transcript expression was probed using the last exon (XV) the nature of the TSS involved (5' or "3 'alternate") is not known.

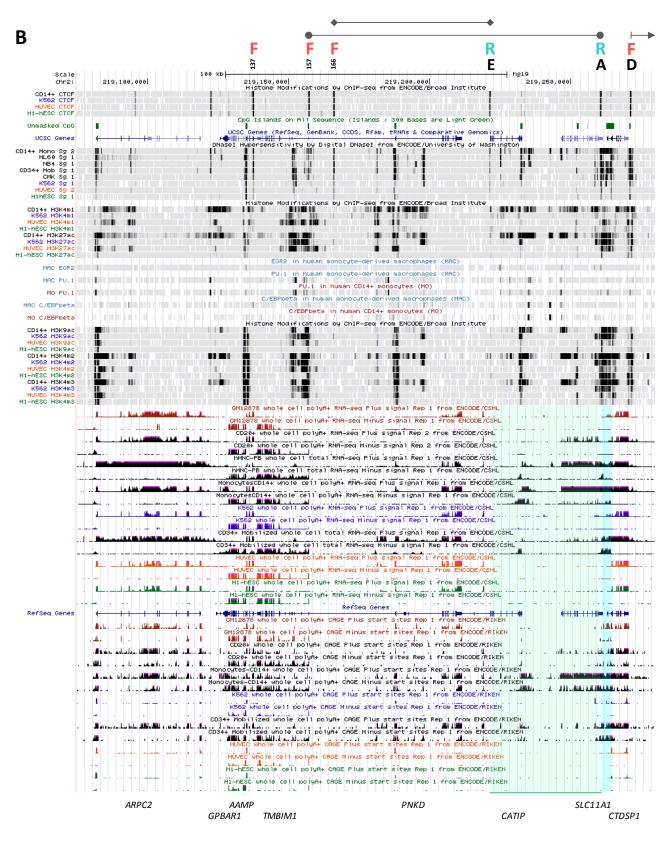
Lastly, ChIP-seq data from untreated THP1 cells [93] associated the TFs SP1 and PU.1 with both the 5' and 3' parts of *NRAMP1* Orf, in agreement with other results implicating SP1 in *NRAMP1* expression in VitD-differentiated HL-60 cells [12] and demonstrating PU.1 binding at *NRAMP1* locus both in MNs and MDMs [13], whereas K9ac mark and RNA Pol II were found spread along *NRAMP1* locus including the 5' enhancer determinants. The negative element suspected in *NRAMP1* intron 12 (F10) corresponds to a peak in all four

ChIP-seq analyses, which is more prominent in the case of SP1 compared to PU.1. In contrast, CAGE tags were few at this site while tag clustering at *NRAMP1* TSS was comparatively inferior to *CTDSP1* TSS, consistent with low level *NRAMP1* expression in untreated THP1 cells.

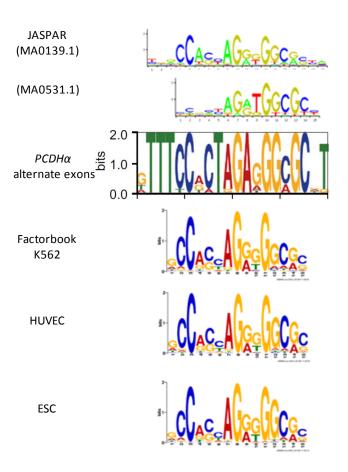
#### Legend to Figures S1-S28

Figure S1. CTCF-dependent topological organization of NRAMP1 locus. A. View of a 700 kb chromosome 2 segment centered on NRAMP1 (SLC11A1) gene [9, 10, 48, 78]. From top to bottom: Orientation of CTCF sites identified by ENCODE project, forward (F) and reverse (R), and labelled with numbers or letters; Chromosome 2 scale; CTCF ChIP-seq data from CD14<sup>+</sup> MNs, K562, HUVEC and ESC; CpG islands; UCSC gene descriptions; DHS mapped in CD14<sup>+</sup> MN, HL-60, NB4, G-CSF mobilized CD34<sup>+</sup> precursors (mCD34/CMP), CMK, K562, HUVEC and ESC; Histone marks of activation (K4me1 and K27ac) in CD14<sup>+</sup> MNs, K562, HUVEC and ESC; ChIP-seq data for EGR2 (MF/MAC), PU.1 and C/EBPb (MAC and MN/MO; [13]); Histone marks of transcriptional activity (K9ac, K4me2, K4me3) in CD14<sup>+</sup> MN, K562, HUVEC and ESC; RefSeq gene descriptions. A candidate super-domain encompassing NRAMP1 locus is indicated by a thick black line. B. Close-up view of CTCF-dependent looping around NRAMP1 locus. From top to bottom: CTCF-dependent loops revealed by ChIA-PET analyses in both K562 and MCF7 cells; Orientation of the corresponding CTCF sites, F or R, identified by their genomic location; Chromosome 2 scale; CTCF ChIP-seq data from CD14<sup>+</sup> MN, K562, HUVEC and ESC; CpG islands; UCSC gene descriptions; DHS mapped in CD14<sup>+</sup> MN, HL-60, NB4, mCD34, CMK, K562, HUVEC and ESC; Histone marks of activation (K4me1 and K27ac) in CD14+ MN, K562, HUVEC and ESC; ChIP-seq data for EGR2 (MF), PU.1 and C/EBPb (MF and MN); Histone marks of transcriptional activity (K9ac, K4me2, K4me3) in CD14+ MN, K562, HUVEC and ESC; RNA-seq data for both strands from GM12878 lymphoblasts, CD20+ B lymphocytes, mononuclear cells from peripheral blood, CD14<sup>+</sup> MN, K562, mCD34, HUVEC and ESC; RefSeq gene descriptions; whole cell CAGE for both strands from GM12878 lymphoblasts, CD20+ B lymphocytes, CD14+ MN, K562, mCD34, HUVEC and ESC. Green highlighting underlines myelo-monocytic specific transcription that typifies NRAMP1 locus; a small contiguous area (blue highlight) delineates hematopoietic-specific transcription signals. C. Sequence of CTCF sites. Top: CTCF consensus sequence from various sources: JASPAR TF database [63], Factorbook database [61] and Guo et al. study [58]; Bottom: sequence and orientation of CTCF sites identified by ENCODE around NRAMP1; the three most conserved positions are underlined; # refers to CTCF site labels in panel A. D. Both NRAMP1 locus CTCF boundaries comprise predicted eQTLs [54]. a. Immunpop server [54] display of 416 local eQTLs associated with NRAMP1, based on gene expression level variations measured in M2 MF/immature DC (CD14<sup>+</sup>, CD1a<sup>+</sup>, CD83<sup>low</sup>, HLA-DR<sup>low</sup>), either resting or infected for two hours with Listeria or Salmonella, from individuals of European and/or African ancestry. b. Blow-up of NRAMP1 locus area (~70kb) highlighting the location of two SNPs widely used for NRAMP1 genotyping which were previously associated with susceptibility to infectious diseases, including tuberculosis, and/or autoimmune pathologies (e.g., inflammatory bowel disease, rheumatoid arthritis [111]). Horizontal green arrows indicate eQTL haplotypes that were co-detected in independent studies [54,50]; the corresponding individual eQTLs are pointed with green arrowheads. c. UCSC genome browser display of the CTCF topologically associating domain (TAD) delineating NRAMP1 locus. From top to bottom: Chromosome 2 scale; CTCF binding sites revealed by ChIP-seq assays in CD14<sup>+</sup> MNs and K562 cells; CpG islands; RefSeq genes; DHS in CD14<sup>+</sup> MN (x2); ENCODE TF binding sites detected by ChIP-seq assays; UCSC genes; Chromosome 2 regions (i-v) part of NRAMP1 locus (CTCF TAD) carrying sets of transcriptional regulatory elements (F1-F14) predicted to control gene expression.





# C CTCF consensus sites:



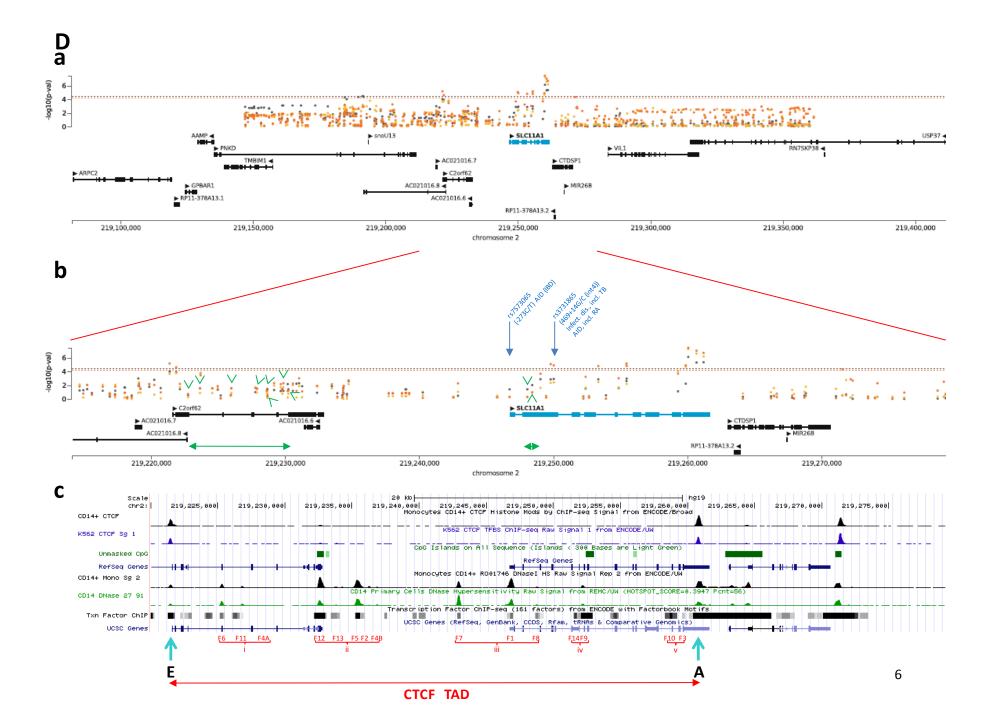
Mathelier, Fornes, Arenillas, Chen et al., 2016

Guo, Monahan, Wu et al., 2012

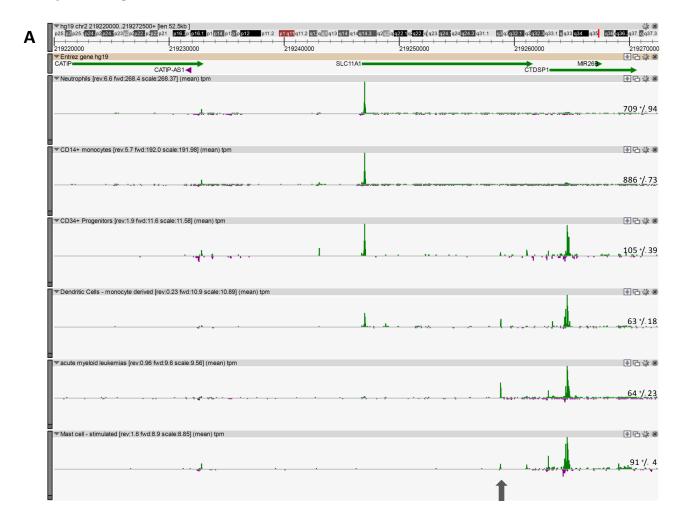
Wang, Zhuang, Iyer, Lin, Whitfield et al., 2012

# CTCF sites around NRAMP1 locus:

	#	Strand	Orientation
GCCACAAGGGGGGCAG	033	+	F
TCCAGGAGAGGGCAG	051	+	F
TCCACCAGAGGGCGC	137	+	F
GCCGCCAGGGGGCGC	157	+	F
ACCACTAGGGGGCAG	166	+	F
TCCAGCAGAGGGAGC	Е	-	R
GCCCCTAGGGGGAGC	А	-	R
GCCGCTAGAGGCCTC	D	+	F
AACAGTAGGTGTCAC	331	-	R
CCCACAAGAGGGTGC	513	-	R

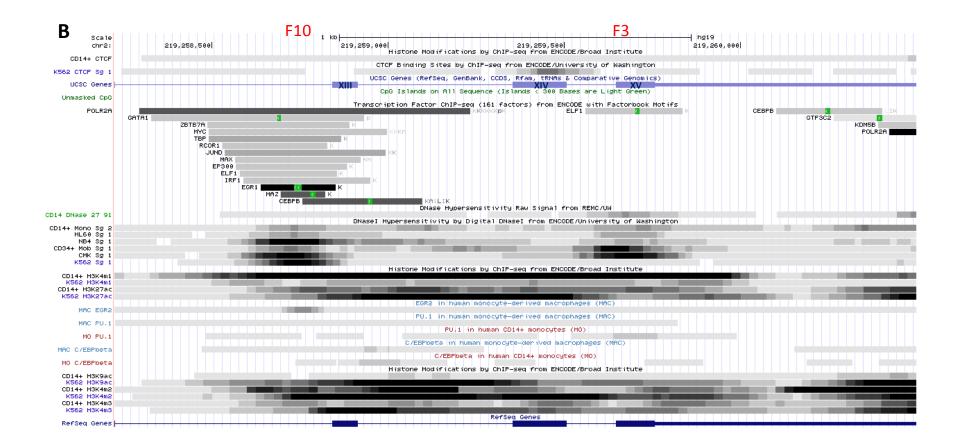


**Figure S2.** Regulatory element in *NRAMP1* intron 12 having negative impact on gene expression. **A.** Cap analysis of gene expression (CAGE) at *NRAMP1* locus showing the transcription start sites (TSS) of *NRAMP1* and downstream gene *CDTSP1* whereas the upstream gene *CATIP* is not expressed in myeloid cells [68]. CAGE for representative examples of cell types that express *NRAMP1* at high level (neutrophils, CD14<sup>+</sup> MNs), at moderate level similar to *CDTSP1* (CD34<sup>+</sup> progenitors, dendritic cells) or that show negligible *NRAMP1* expression (acute myeloid leukemias, mast cells). The mean of tags per million (tpm) detected over the 50kb analyzed is given for each cell type to illustrate the predominance of *NRAMP1* expression in mature phagocytes. A vertical arrow indicates a putative *NRAMP1* internal TSS that is detected in cells producing little full-length transcript.



#### Biology 2017, 6, 28

and DNAse 1 footprints. *From top to bottom*: F10 and F3 denote DNAse 1 footprints previously described [11]; chromosome 2 scale; ENCODE ChIP-seq for CTCF (CD14<sup>+</sup> MNs, K562 cells), UCSC browser description of *NRAMP1* structure; CpG islands; ENCODE ChIP-seq data for various TFs; footprints of areas sensitive to DNase1 digestion (CD14<sup>+</sup> MNs, HL-60 and NB4 cells, mCD34, CMK and K562 cells); ChIP-seq data for select histone marks of regulatory element priming (K4me1, K27ac; CD14<sup>+</sup> MNs, K562 cells); ChIP-seq data for the TFs EGR2, PU.1 and C/EBPb in MO and MAC [13]; ChIP-seq data for select histone marks of transcriptional activation (K9ac, K4me2, K4me3) in CD14<sup>+</sup> MNs and K562 cells; RefSeq description of *NRAMP1* structure.



S8 of S63

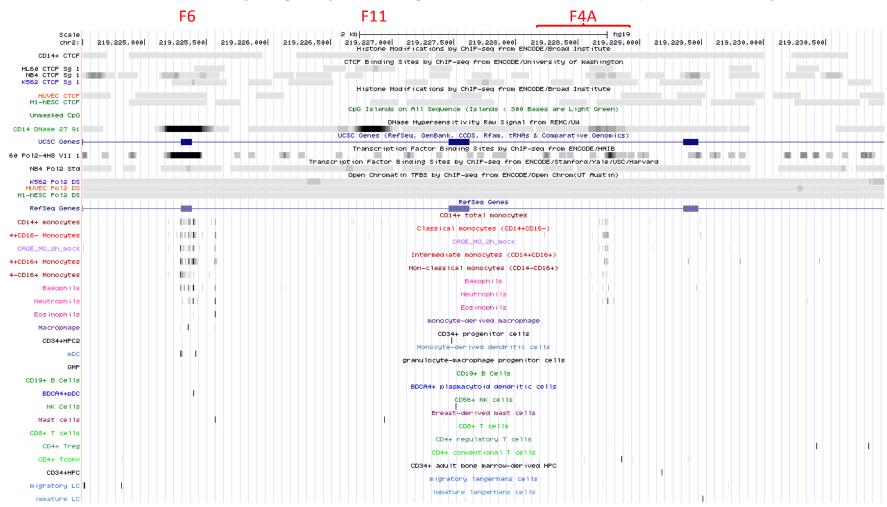
**Figure S3.** Detail of CAGE at *NRAMP1* region ii spanning DNASe1 footprints (DHS) F12, F13, F5, F2 and F4B. *From top to bottom*: Location of DNASe1 footprints (DHS, ENCODE); chromosome 2 scale; ENCODE CTCF ChIP-seq data for CD14<sup>+</sup> MNs, HL-60, NB4 and K562 cells, HUVEC and ESC; CpG islands; DNAse1 footprints in CD14<sup>+</sup> MNs; UCSC browser gene descriptions; RNA Pol II ChIP-seq data for HL-60, NB4, K562, HUVEC and ESC; RefSeq genes; CAGE data for 22 blood cell types [68].



# Figure S4. Detail of CAGE at NRAMP1 region iii spanning DNASe1 footprints (DHS) F7, F1 (TSS) and F8. From top to bottom: same as Figure S2.

	F7	F1	F8
Scale chr2:   2:	19,242,500  219,243,000  219,243,500  219,244,000  219,244,50	2 kb	248,500  219,249,000
CD14+ CTCF		TCF Binding Sites by ChIP-seq from ENCODE/University of Washington	
HL60 CTCF Sg 1 NB4 CTCF Sg 1 K562 CTCF Sg 1		Histone Modifications by ChIP-seq from ENCODE/Broad Institute	
HUVEC CTCF		Histore Mourritations by Chir-seq from Encode/Broad Institute	
Unmasked CpG		CpG Islands on All Sequence (Islands < 300 Bases are Light Green)	
CD14 DNase 27 91		DNase Hypersensitivity Raw Signal from REMC/UW	
UCSC Genes	UCS	C Genes (RefSeq, GenBank, CCDS, Rfam, tRNAs & Comparative Genomics)	
60 Po12-4H8 V11 1		Transcription Factor Binding Sites by ChIP-seq from ENCODE/HRIB	
NB4 Po12 Std		Open Chromatin TFBS by ChIP-seq from ENCODE/Open Chrom(UT Austin)	
K562 Po12 DS HUVEC Po12 DS H1-hESC Po12 DS			
RefSeq Genes		RefSeq Genes	
CD14+ monocytes		CD14+ total monocytes	
+CD16- Monocytes		Classical monocytes (CD14+CD16-)	
CAGE_MO_2h_mock		CRGE_MO_2h_mock	
+CD16+ Monocytes	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	Intermediate monocytes (CD14+CD16+)	
-CD16+ Monocytes		Non-classical monocytes (CD14-CD16+)	
Basoph i 1s		Basoph i Is	10 11 11 11 1
Neutroph i 1s		Neutroph i 1s Eosinoph i 1s	
Eos inoph i 1s		monocyte-derived macrophage	
Macrophage		CD34+ progenitor cells	11 11 11 11
CD34+HPC2		Monocyte-derived dendritic cells	
mDC		granulocyte-macrophage progenitor cells	
GMP		CD19+ B Cells	
CD19+ B Cells		BDCA4+ plasmacytoid dendritic cells	
BDCR4+pDC		CD56+ NK cells	
NK Cells		Breast-derived mast cells	
Mast cells		CD8+ T cells	
CD8+ T cells		CD4+ regulatory T cells	
CD4+ Treg		CD4+ conventional T cells	
CD4+ Tconv		CD34+ adult bone marrow-derived HPC	
CD34+HPC		migratory langerhans cells	
migratory LC immature LC		immature langerhans cells	

## Figure S5. Detail of CAGE at NRAMP1 region i spanning DNASe1 footprints (DHS) F6, F11 and F4A. From top to bottom: same as Figure S2.



## Figure S6. Detail of CAGE at NRAMP1 region iv spanning DNASe1 footprints (DHS) F14 and F9. From top to bottom: same as Figure S2.



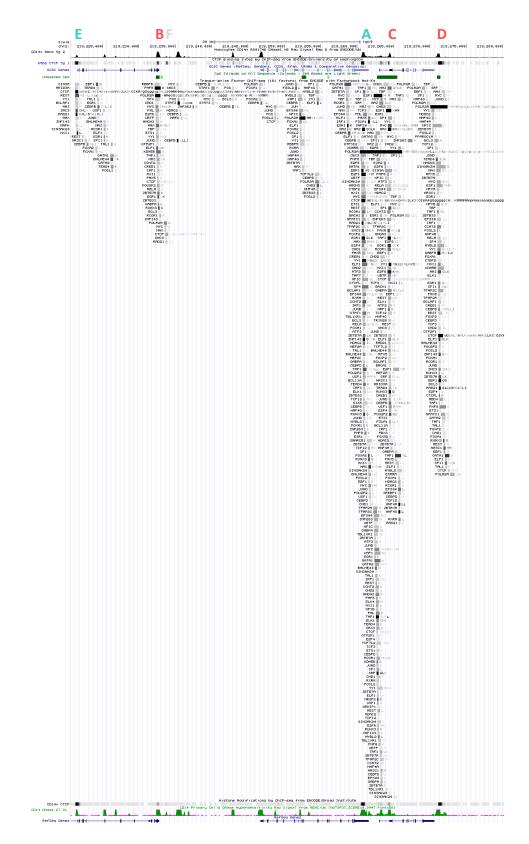
## Figure S7. Detail of CAGE at NRAMP1 region v spanning DNASe1 footprints (DHS) F10 and F3. From top to bottom: same as Figure S2.



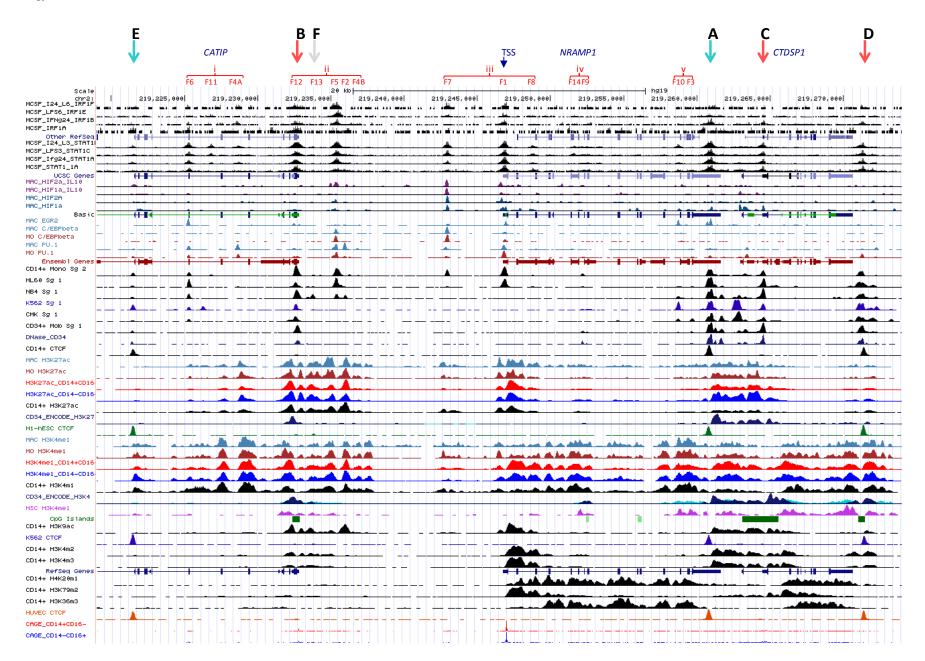
**Figure S8.** ChIP-seq data for histone marks of de-activation: K27me3 (transcriptionally silent chromatin) and K9me3 (heterochromatin). *NRAMP1* locus is examined at two different scales: **upper panel**, 6Mb, a vertical arrow localizes the gene; **lower panel**, 55kb, the gene is underlined. For each panel, *from top to bottom*: chromosome 2 banding pattern; local chromosome 2 scale; CpG islands; ENCODE CTCF ChIP-seq data for ESC, HUVEC, CD14<sup>+</sup> MNs and K562 cells; UCSC gene descriptions; DNAse1 footprints for CD14<sup>+</sup> MNs, HL-60 and NB4 cells, mCD34, CMK and K562 cells; ChIP-seq data for the TFs EGR2, PU.1 and C/EBPb in MN and MF [13]; ChIP-seq data for select histone marks of chromatin de-activation (K27me3 and K9me3; ESC, HUVEC, CD14<sup>+</sup> MNs and K562 cells); RefSeq gene descriptions.

chr2 (q35) [	
Scale chr2:	2 Mb
Unmasked CpG	CpG Islands on All Sequence (Islands < 300 Bases are Light Green) Histone Modifications by ChIP-seq from ENCODE/Broad Institute
H1-hESC CTCF HUVEC CTCF CD14+ CTCF K562 CTCF	
UCSC Genes	
DNase 27 91	DNase Hypersensitivity Raw Signal from REMC/UW DNaseI Hypersensitivity by Digital DNaseI from ENCODE/University of Washington
.4+ Mono Sg 2 HL60 Sg 1 NB4 Sg 1 34+ Mob Sg 1 CMK Sg 1 K562 Sg 1	
MAC EGR2	EGR2 in human monocyte+derived macrophages (MRC) PU.1 in human monocyte+derived macrophages (MRC)
MAC PU.1	PU.1 in human CD14+ monocytes (MO)
MO PU.1	C/EBPbeta in human monocyte-derived macrophages (NRC)
AC C/EBPbeta	C/EBPbeta in human CD14+ monocytes (MO)
MO C/EBPbeta -hESC H3K9m3	Histone Modifications by ChIP-seq from ENCODE/Broad Institute
HUVEC H3K9m3 CD14+ H3K9m3 K562 H3K9m3 hESC H3K27m3	
UVEC H3K27m3	
	n n n n n n n n n n n n n n n n n n n
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chr2 (q35) Scale chr2 (q35) Scale chr2: Unmasked CpG H1-hESC CTCP K862 CTCP K862 CTCP UCSC Genes I DNase 27 91 HL69 S 2 1 HL69 S 2	P21       14       ED12       13       31.1       Q34QS       NRAN         219,225,000       219,235,000       219,245,000       219,245,000       219,255,000       <
chr2 (q35) Scale chr2 (q35) Unmasked CpG H1-hESC CTCP K184 CTCP K184 CTCP UCSC Genes DNase 27 91 HL69 S 2 HL69 S 2 HL69 S 1	Refseq Cenes         Image: Construction of the second of
chr2 (q35) scale chr2 (q35) Scale chr2; Unmasked CpG HU-hESC CTOF HUVEC CTOF HUVEC CTOF HUVEC CTOF HUVEC CTOF UCSC Genes P Dhase 27 91 I-H Mono 52 HL69 82 11 NB4 82 11	Image: Section of the section of th
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RefSeq Genes           chr2 (q35)           Scale           chr2;           Unmasked CpG           HLVVEC CTOF           CD14+ CTOF           CD25C Genes           4 DNase 27 91           14+ MON 58 2           HD45 331           CMH 583           NH5 381           CMM 583           MAC 289 1           MAC EGR2           MAC EOR2           MAC PU.1	Prefseq Genes         Prefseq Genes <td< td=""></td<>
RefSeq Genes           chr2 (q35)           Scale           chr2:           Unmasked CpG           CD14:           CO14:           CO25:           USC Genes           4 Dhase 27 91           14:           NB4 Sg 1           CMK Sg 1           K562 S1           NB4 Sg 1           NB4 Sg 1           CMK Sg 1           MAC EOR2           MAC PU.1	Refseq Denes
RefSeq Genes           chr2 (q35)           Scale           chr2:           Unmasked CpG           CD14:           CO14:           CO25:           USC Genes           4 Dhase 27 91           14:           NB4 Sg 1           CMK Sg 1           K562 S1           NB4 Sg 1           NB4 Sg 1           CMK Sg 1           MAC EOR2           MAC PU.1	Image: Information in the information of the information o
Scale chr2: Unmasked CpG H1-hESC CTCF UUSC CTCF CD14+ CTCF KS52 CTCF UCSC Genes 4 DNase 27 91 14+ Mono Sg 1 UCSC Genes HL56 Sg 1 N64 Mob Sg 1 CMK Sg 1 K552 Sg 1 N64 Sg 1 K552 Sg 1 MAC C/EBFbeta MO C/EBFbeta MO C/EBFbeta MO C/EBFbeta St52 H3K9m3 -HESC H3K9m3 -HESC H3K9m3 -HESC H3K2m3	P21       14       201       13       31.1       05402       NRAA         219,225,000       219,230,000       219,240,000       219,245,000       219,245,000       219,245,000       219,245,000       219,245,000       219,245,000       219,255,000       219,265,000       219,270,000         219,225,000       219,230,000       219,246,000       219,245,000       219,245,000       219,255,000       219,270,000         219,225,000       219,230,000       219,246,000       219,245,000       219,245,000       219,255,000       219,270,000         Histone Modifications by ChIP-seq from ENCODE/Broad Institute       109,265,000       219,270,000       119,270,000         Histone Modifications by ChIP-seq from ENCODE/Broad Institute       100,250,000       219,270,000       119,270,000         Histone Modifications by ChIP-seq from ENCODE/Broad Institute       100,000       100,000       100,000       100,000         Mase Hypersensitivity Raw Signal from ENCODE/University of Hashington       100,000       100,000       100,000       100,000         E062 in human monocyte-derived macrophages (MRC)       PU.1 in human monocyte-derived macrophages (MRC)       100,000       100,000       100,000       100,000       100,000       100,000       100,000       100,000       100,000       100,000       100,00
chr2 (q35)           scale           chr2 (q35)           scale           chr2 (q35)           Unmasked CpG           HUVEC CTCF           CD14+ CTCF           HUVEC CTCF           UCSC Genes           4 DNase 27 91           14+ MON 59 2           HL66 59 1           MRC EGR2           MRC EGR2           MRC C/EEPbeta           MO C/LEPbeta           MO C14HSK083           HUVEC CTCF           L14+ MON 59 1           CK562 91           MRC C/EEPbeta           MO C/LEPbeta           MO CULH HSK083           K562 HSK083           HUVEC CTCF	Perfect Highligh High High High High High High High H

**Figure S9.** UCSC browser display of ENCODE TF-specific ChIP-seq data for *NRAMP1* locus shows TF association with multiple DHS, including several CTCF sites. *From top to bottom*: position and orientation of CTCF sites, indicated by colored letters: red (forward), blue (reverse) and grey (undetermined); chromosome 2 scale bar; CD14<sup>+</sup> MN DHS; CTCF-specific ChIP-seq (K562 cells); UCSC gene descriptions; CpG Islands; ENCODE TF-specific ChIP-seq data shows variable enrichment at CTCF sites; CTCF-specific ChIP-seq (CD14<sup>+</sup> MNs); CD14<sup>+</sup> MN DHS; Refseq gene descriptions.



**Figure S10.** Association of TF mediating inflammatory responses with *NRAMP1* locus. *From top to bottom*: major CTCF binding sites E, A and D; gene names and position of *NRAMP1* TSS; chromosome 2 regions carrying predicted transcriptional regulatory elements (F1-F14); chromosome 2 scale; IRF1 ChIP-seq data for MDM (in presence of M-CSF), stimulated for 6h with or without LPS and with or without 24h IFN-g priming [99]; Other RefSeq gene descriptions; STAT1 ChIP-seq data for MDM stimulated for 6h with or without 24h IFN-g priming [99]; UCSC gene descriptions; HIF1a and HIF2a association with *NRAMP1* locus in MDM (RPMI medium alone) exposed to hypoxia for 4h at 37°C after priming or not with IL-10 for 16h [103]; Basic gene depictions; ChIP-seq for EGR2 (MF), C/EBPb and PU.1 (MN and MF) [13]; Ensembl Genes; ENCODE DHS for CD14<sup>+</sup> MNs, HL-60, NB4, K562 and CMK cells, mCD34, CD34<sup>+</sup> progenitors; CTCF specific ChIP-seq in CD14<sup>+</sup> MNs; K27ac specific ChIP-seq data for MN and MF [13], CD14<sup>+</sup> CD16<sup>-</sup> MNs and CD14<sup>+</sup> CD16<sup>+</sup> MNs [152], CD14<sup>+</sup> MNs, CD34<sup>+</sup> progenitors and HSC (ENCODE); CPG islands; K9ac ChIP-seq data for MO and MAC [13], CD14<sup>+</sup> MNs, CD34<sup>+</sup> progenitors and HSC (ENCODE); CpG islands; K9ac ChIP-seq data (CD14<sup>+</sup> MNs); RefSeq genes; ChIP-seq in CD14<sup>+</sup> MNs (H4K20me1, K79me2, K36me3); CTCF specific ChIP-seq in HUVEC; CAGE data for CD14<sup>+</sup> CD16<sup>-</sup> MNs and CD14<sup>-</sup> CD16<sup>+</sup> MNs [152].

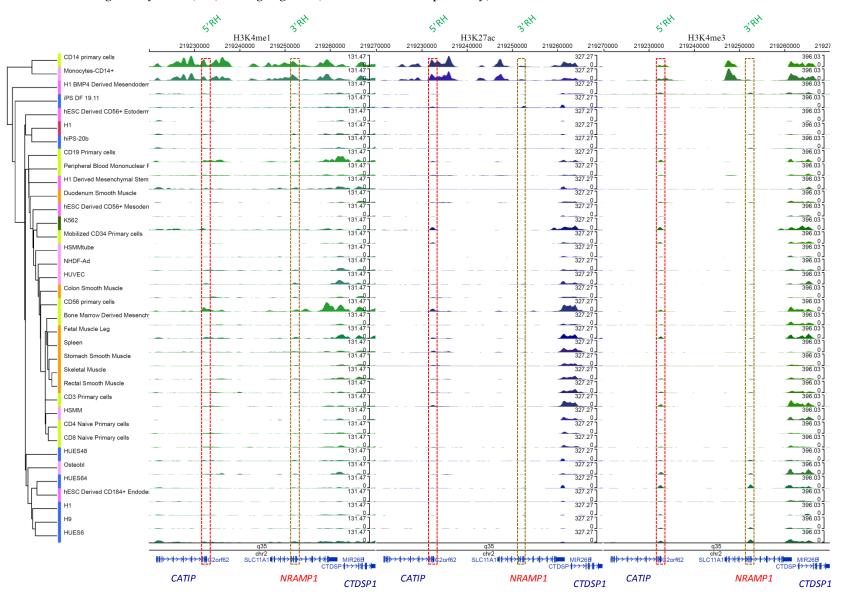


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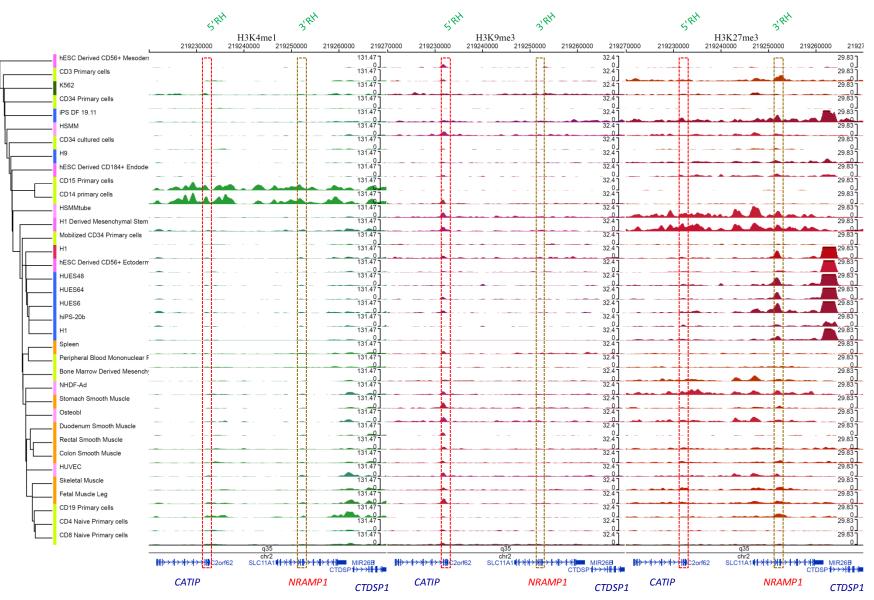
**Figure S11.** Predicted *NRAMP1* super-enhancer (S-E) determinant. *From top to bottom*: Chromosome 2 scale; ChIP-seq for CTCF (CD14<sup>+</sup> MNs, K562 cells, ESC); CpG islands; UCSC gene description; DHS (CD14<sup>+</sup> MNs, HL-60 and NB4 cells, mCD34, CMK and K562 cells); ChIP-seq data (K27ac, K4me1 and the TFs PU.1 and C/EBPb in MN and MF [13]; polyA+ CAGE tags for CD14<sup>+</sup> MNs, mCD34, K562 cells, ESC [16,17]; RefSeq genes; dbSUPER prediction of S-E determinants in CD14<sup>+</sup> MNs, K562 cells and CD34<sup>+</sup> precursors [101]



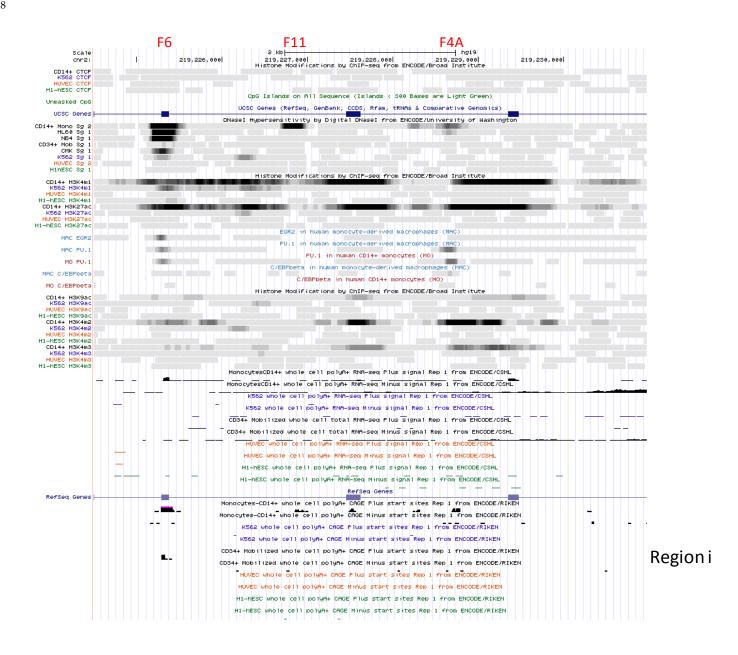
**Figure S12.** Comparison of ChIP-seq data at *NRAMP1* locus for histone marks of chromatin activation (K4me1, K27ac and K4me3) from select cell types (ESCs, ESC-derived cultured cells) and various tissues of mesodermal origin [14]. Each panel is depicted using a common Y-scale. *NRAMP1* predicted 5' and 3' regulatory hubs (RH) are highlighted (red and brown, respectively).

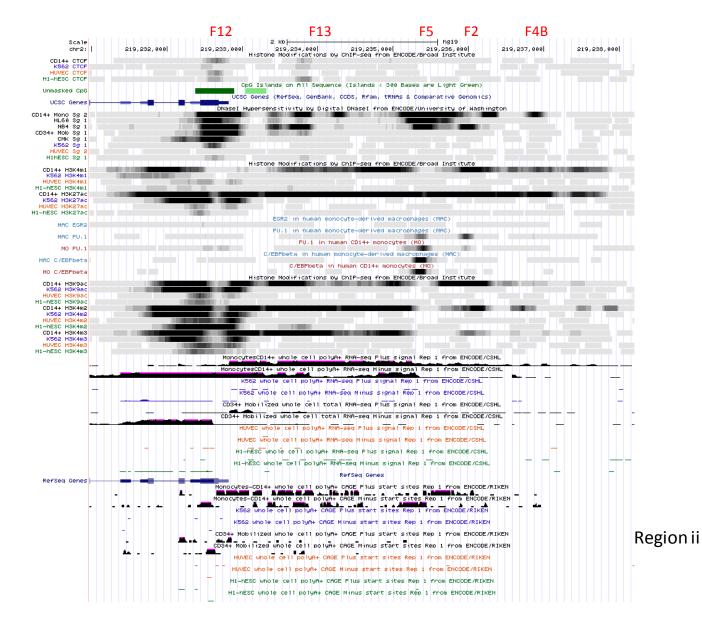


**Figure S13.** Comparison of ChIP-seq data at *NRAMP1* locus for histone marks of chromatin de-activation (K27me3 and K9me3) from select cell types (ESCs, ESC-derived cultured cells) and various tissues of mesodermal origin [14]. Each panel is depicted using a common Y-scale. *NRAMP1* predicted 5' and 3' regulatory hubs (RH) are highlighted (red and brown, respectively). K4me1 mark added for comparison with Figure S12.

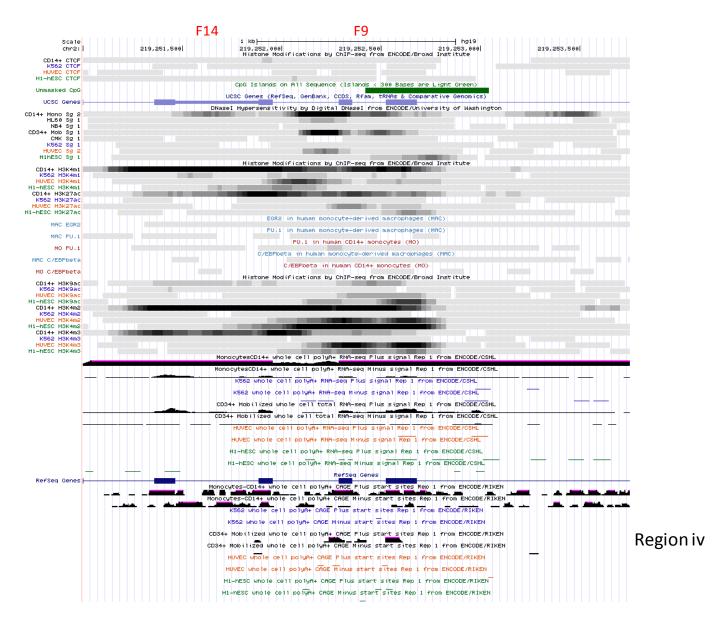


**Figure S14.** ENCODE CAGE and RNA-seq data corresponding to *NRAMP1* locus. Results are presented separately for five regions (i-v). *From top to bottom*: Location of DHSs; chromosome 2 scale; CTCF ChIP-seq for CD14<sup>+</sup> MNs, K562 cells, HUVEC and ESC; CpG islands; UCSC gene description; DHSs in CD14<sup>+</sup> MNs, HL-60, NB4, mCD34/CMP, CMK and K562 cells, HUVEC and ESC; ChIP-seq for histone marks of priming (K27ac, K4me1; CD14<sup>+</sup> MNs, K562 cells, HUVECs and ESCs); ChIP-seq for the TFs PU.1 and C/EBPb in MN and MF; ChIP-seq for histone marks of activation (K9ac, K4me2 and K4me3); RNA-seq data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC); RefSeq gene description; CAGE data (CD14<sup>+</sup> MNs, K562 cells, mCD34, HUVEC and ESC).











**Figure S15.** Chromatin status at *NRAMP1* locus in hematopoietic cells. *From top to bottom*: position and orientation of CTCF sites indicated by arrows, red (forward), blue (reverse) and grey (undetermined); chromosome 2 regions carrying predicted transcriptional regulatory elements (F1-F14); chromosome 2 scale bar; ENCODE CTCF-specific ChIP-seq (CD14<sup>+</sup> MNs, K562 cells); UCSC gene descriptions; CpG Islands; ENCODE TF-specific ChIP-seq data; ENCODE DHS for CD14<sup>+</sup> MNs (x2), HL-60, NB4, mCD34, CMK and K562 cells; ENCODE ChIP-seq data for select histone marks (K4me1 and K27ac) in CD14<sup>+</sup> MNs and K562 cells; ChIP-seq data for the TFs EGR2, PU.1 and C/EBPb in MO and MAC [13]; ENCODE ChIP-seq data for histone marks K9ac, K4me2 and K4me3 in CD14<sup>+</sup> MNs and K562 cells; RefSeq gene descriptions; NIH Roadmap epigenomic data for hematopoietic cells: DHS and ChIP-seq data for histone marks of priming, K4me1 and K27ac (panel **A**); ChIP-seq data for histone marks of transcriptional activity, K9ac, K4me3 and K36me3 (panel **B**); RNA-seq profile and ChIP-seq data for histone marks of repression, K9me3 and K27me3 (panel **C**)

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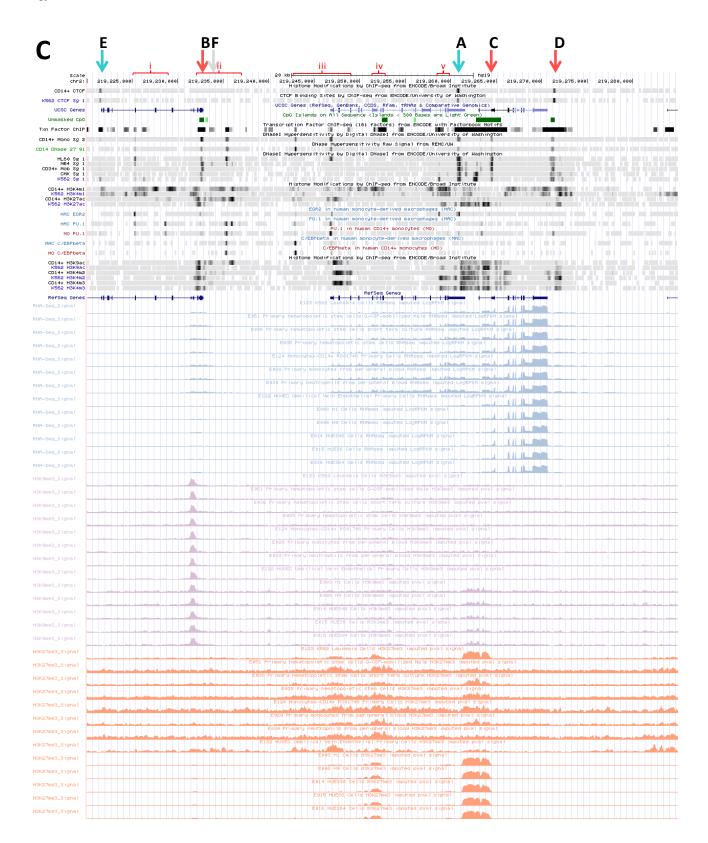
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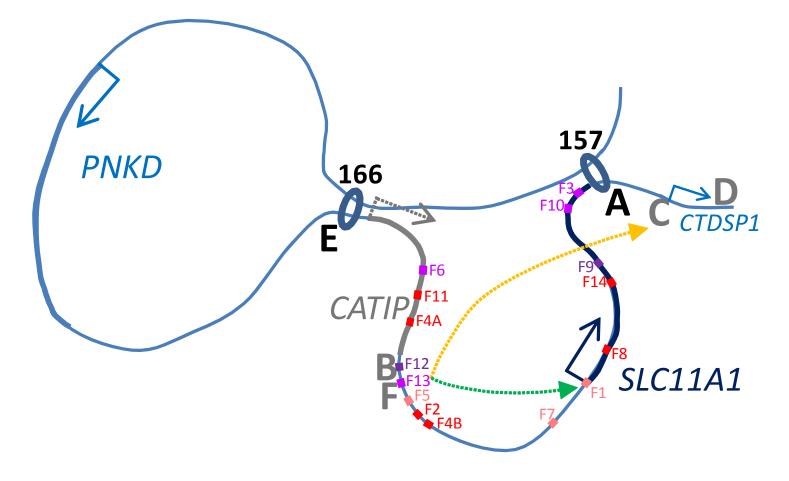
**Figure S16.** Chromatin status at *NRAMP1* locus in myeloid cells, HUVEC and ESC. *From top to bottom*: position and orientation of CTCF sites indicated by arrows, red (forward), blue (reverse) and grey (undetermined); chromosome 2 regions carrying predicted transcriptional regulatory elements (F1-F14); chromosome 2 scale bar; ENCODE CTCF-specific ChIP-seq (CD14<sup>+</sup> MNs, K562 cells); UCSC gene descriptions; CpG Islands; ENCODE TF-specific ChIP-seq data; ENCODE DHS for CD14<sup>+</sup> MNs (x2), HL-60, NB4, mCD34, CMK and K562 cells; ENCODE ChIP-seq data for select histone marks (K4me1 and K27ac) in CD14<sup>+</sup> MNs and K562 cells; ChIP-seq data for the TFs EGR2, PU.1 and C/EBPb in MO and MAC [13]; ENCODE ChIP-seq data for histone marks K9ac, K4me2 and K4me3 in CD14<sup>+</sup> MNs and K562 cells; RefSeq gene descriptions; NIH Roadmap epigenomic data for mesodermal cell types and ESCs: DHS and ChIP-seq data for histone marks of priming, K4me1 and K27ac (panel **A**); ChIP-seq data for histone marks of transcriptional activity, K9ac, K4me3 and K36me3 (panel **B**); RNA-seq profile and ChIP-seq data for histone marks of repression, K9me3 and K27me3 (panel **C**).

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K562 CTCF Sg 1 UCSC Genes Unmasked CpG			UCSC Genes (RefSeq, GenBank, CCDS, Rfam, tRNMs & Comparative Genomics) CpG Islands on R11 Sequence (Islands < 300 Bases are Light Green)
Txn Factor ChIP			Transcription Factor ChIP-seq (161 factors) from ENCODE with Factorbook Motifs DNaseI Hypersensitivity by Digital DNaseI from ENCODE/University of Washington
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CD14+ H3K9ac K562 H3K9ac CD14+ H3K4m2			Histone Modifications by ChIP-seq from ENCODE/Broad Institute
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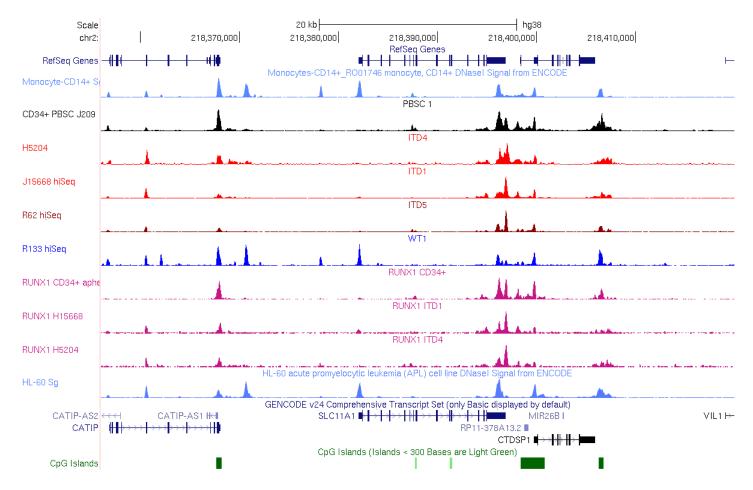
**Figure S17.** Hypothetical CTCF-dependent topology of *NRAMP1* locus and hematopoietic determinants of gene expression (see text for details, sections 2.3.3. and 2.3.4.). *NRAMP1* locus boundaries (CTCF sites A and E, "R" orientation, shown in black) are represented interacting with compatible and preferred upstream CTCF sites ("F" orientation, indicated as 157 and 166, respectively). Other CTCF sites are indicated in grey ("F" or undetermined orientation: B-D and F, respectively). DHS F1-F14 are colored depending on temporal order of mobilization: purple, light purple, pink and red. It is proposed that *NRAMP1* (*SLC11A1*) locus activation results from successive steps: i) F12/5'RH (CTCF\_B) and F9/3'RH are partially mobilized in nonhematopoietic cell types; ii) initiation of hematopoiesis may erase negative histone marks leading to increased *CTDSP1* transcription -a process that may result from formation of a CTCF-dependent loop involving sites F and C (orange dotted arrow); iii) myeloid-specific TF-dependent mobilization of F12/5' RH, F13 and co-activation of sites F6, F10-F3 may prime the locus while preventing gene expression; iv) as differentiation progresses, specific myelomocytic TFs activate elements F5, F7 and F9 leading to disruption of CTCF\_F-C loop and allowing contacts between distal regulatory elements and NR1 TSS (F1), through Mediator based interactions for instance, which activate *NRAMP1* transcription (green dotted arrow); and v) terminal differentiation into mature phagocytes leads to extensive priming/activation of the locus, including elements F11, F4A, F2, F4B, F8 and F14, and further modulation by signal-dependent TFs (e.g., tissue microenvironment, infections...). According to this hypothesis, CTCF determinants define *NRAMP1* locus priming for subsequent activation while myelo-monocytic TFs mediate *NRAMP1* expression and/or signal-dependent regulation.



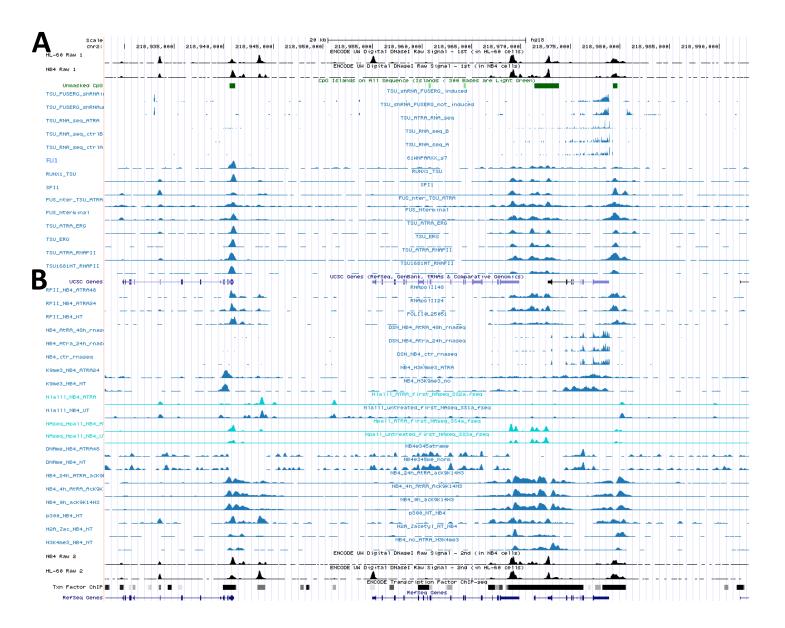
**Figure S18.** *NRAMP1* locus region i and ii represent candidate transcriptional regulatory determinants mediating innate memory in response to microbial stimuli from various origins (*Saccharomyces cerevisiae*  $\beta(1,3)D$ -glucan, BG, and *Escherichia coli* lipopolysaccharide, LPS) [128]. *From top to bottom*: Major CTCF sites E, A and D (blue and red arrows correspond to reverse and forward orientations, respectively); gene names and position of *NRAMP1* TSS; candidate *NRAMP1* regulatory regions i-v, including DHS F1-F14; chromosome 2 scale; CpG islands; ENCODE CTCF-specific ChIP-seq data in CD14<sup>+</sup> MNs; histone modification-specific ChIP-seq (K4me1, K27ac, K4me3) and Tn5 transposase accessibility (ATAC-seq) data for MNs stimulated with: a) LPS, for 24h and analyzed five days after (LPS\_D6), b) BG, for 24h and analyzed five days after (BG\_D6, c) no additive, for 24h and analyzed five days after (RPMI\_D6), d) LPS, for 24h (LPS\_D1), e) BG, for 24h (BG\_D1) and f) no additive, for 24h (RPMI\_D1), as well as for freshly explanted MNs (RPMI\_0); ENCODE DHS (x2) and ChIP-seq analyses for similar histone modifications in CD14<sup>+</sup> MNs (K4me1, K27ac, K4me3); UCSC genes; ChIP-seq data for the TFs EGR2, PU.1 and C/EBPb in MO and MAC [13]; ENCODE CTCF ChIP-seq data in K562 cells; ENCODE TF-specific ChIP-seq data; delineation of *NRAMP1* regions i-v.

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BG_D6_H3K4me3			GSM2262980_5640.H3K4me3.Normalized			
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ATAC_RPMI_d6			GSM2325691_ATAC_RPMI_d6_8454			<del></del>
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LPS_D1_H3K4me1			GSM2262950_6289.H3K4me1.Normalized			
LPS_D1_H3K27Ac			GSM2262933_6147.H3K27Ac.Normalized			
LPS_D1_H3K4me3		<b></b>	GSM2262934_6617.H3K4me3.Normalized			<b></b> -
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RPMI_0_H3K27ac			GSM2263036_6137.H3K27ac.Normalized			
RPMI_0_H3K4me3			GSM2262965_5631.H3K4me3.Normalized			
ATAC_Mono		A	GSM2325687_ATAC_Mono_8444		••••	•
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CD14+ H3K27ac		Monocytes CD14	+ H3K27ac Histone Mods by ChIP-seq Signal from ENCODE/Broad			
 CD14+ H3K4m3		Monocytes CD14	+ H3K4me3 Histone Mods by ChIP-seq Signal from ENCODE/Broad			
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**Figure S19.** DHS and binding of RUNX1 at *NRAMP1* locus in FLT3-ITD AML patients [139]. *From top to bottom*: chromosome 2 scale; RefSeq gene descriptions; DHS in CD14<sup>+</sup> MNs, CD34<sup>+</sup> peripheral blood stem cells (PBSC), FLT3-ITD AMLs (x3) and MNs; RUNX1-specific ChIP-seq in CD34<sup>+</sup> cells and FLT3-ITD AMLs (x2); DHS in HL-60 cells; GENCODE gene descriptions; CpG islands.



**Figure S20.** Effect of all *trans* retinoic acid ATRA on *NRAMP1* locus activity in AML (TSU1681MT) and APL (NB4). **A**. Gene expression and TF-specific ChIP-seq data for TSU-1621-MT AML [141]. *From top to bottom*: Chromosome 2 scale; DHS for HL-60 (M2 AML) and NB4 (M3 AML/APL); CpG islands; RNA-seq data in cells expressing ERG or not; ChIP-seq data for FLi1, RUNX1/AML1, SPI1/PU.1; FUS, ERG and RNA Pol II in cells treated with ATRA or not; UCSC gene descriptions. **B**. *NRAMP1* activity in NB4 cells ATRA-treated or not [146]. *From top to bottom*: ChIP-seq for RNA Pol II; RNA-seq data; ChIP-seq for K9me3; nuclease accessibility (*Nla* III; *Hpa* II, sensitive to DNA methylation); DNA methylation; ChIP-seq for K9/14ac; ChIP-seq data in untreated NB4 cells: p300 (HAT), H2A.Zac and K4me3. DHS in NB4 and HL-60 cells; ENCODE TF-ChIP-seq data; RefSeq gene depictions.



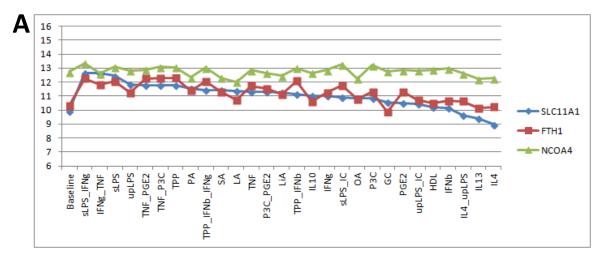
**Figure S21.** *NRAMP1* activity in Kasumi AML [142–44]. *From top to bottom*: chromosome 2 scale; CTCF-specific ChIP-seq (CD14<sup>+</sup> MNs); RNA-seq for Kasumi cells expressing ERG or not; UCSC gene descriptions; acetylated histone-specific ChIP-seq data: K9/14ac in cells treated with MS275 (HDAC inhibitor; for 24, 4, or 0h); H4ac (untreated cells); ENCODE TF-specific ChIP-seq data; ChIP-seq for HDAC1 and HDAC2, N-CoR, HAT p300 (x2), GATA, LYL1, FLi1, TAL1, RUNX1/AML1 (AML blasts, Kasumi (x2) and NB4 cells; CpG islands; ChIP-seq for AML1-ETO fusion (x5); ENCODE DHS in CD14<sup>+</sup> MNs (x2), HL-60 and NB4 AMLs, AML patients (2), mCD34, normal CD34<sup>+</sup> HSPCs, CMK and K562 cells; RefSeq genes; CTCF-specific ChIP-seq (K562 cells); K4me1 and K27ac ChIP-seq data (CD14<sup>+</sup> MNs and K562 cells); ChIP-seq for the TFs EGR2, PU.1 and C/EBPb in MN and MF [13]; K9ac, K4me2 and K4me3 ChIP-seq data (CD14<sup>+</sup> MNs and K562 cells).

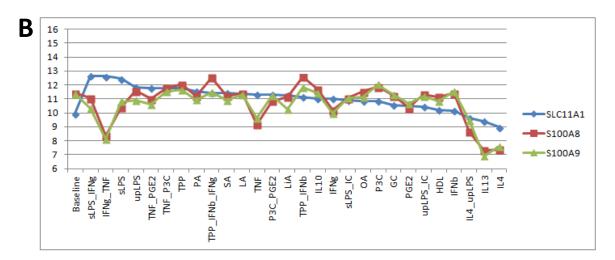
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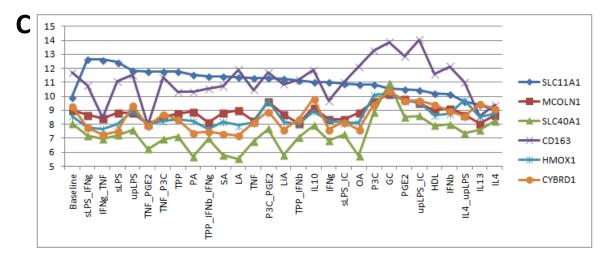
**Figure S22.** TF-specific ChIP-seq data for ME-1 AML [144]. *From top to bottom*: chromosome 2 scale; RefSeq genes; ENCODE TF-specific ChIP-seq data; DHS for HL-60 and NB4 APLs; ChIP-seq data specific for ERG, FLi1, TAL1, HEB, ELF1, PU.1, HDAC1, K9K14ac, HAT p300, TBP, RNA Pol II; DHS for NB4 and HL-60 APLs; CpG islands; RefSeq gene depictions.

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Txn Factor ChIP		
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NB4 Raw 1	- <b>• •</b> •	ENCODE UW Digital DNasel Raw Signal - 1st (in NB4 cells)
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ME_1_FLi1		
ME_1_TAL_1	-	ME_1_TAL_1
ME_1_HEB	—	
ME_1_ELF_1		
ME_1_PU1		ME_1_PU1
ME_HDAC1	+	ME_HDAC1
H3K9K14ac_ME_1		H3K9K14ac_ME_1
ME_p300chip		ME_p300chip
ME_1_TBP		ME_1_TBP
ME_1_RNAPH		ME_1_RNAPII
NB4 Raw 2		ENCODE UW Digital DNasel Raw Signal - 2nd (in NB4 cells)
HL-60 Raw 2		ENCODE UW Digital DNasel Raw Signal - 2nd (in HL-60 cells)
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**Figure S23.** mRNA levels induced by 28 stimulation conditions producing MF phenotypes distributing across M1-M2 spectrum and ranked according to *NRAMP1* mRNA level (log2 transformed values, blue), decreasing from left to right, except for basal conditions (M-CSF or GM-CSF), at most left. **A.** Expression values are represented for *NRAMP1* and two other genes: *FTH1* (red) and *NCOA4* (green). **B.** Superimposition on *NRAMP1* profile of expression values for *S100A8* (red) and *S100A9* (green). **C.** Comparison of multiple profiles: *NRAMP1* (blue), *CD163* (purple), *MCOLN1* (red), *HMOX1* (cyan), *CYBRD1* (orange), *SLC40A1* (green) [132].



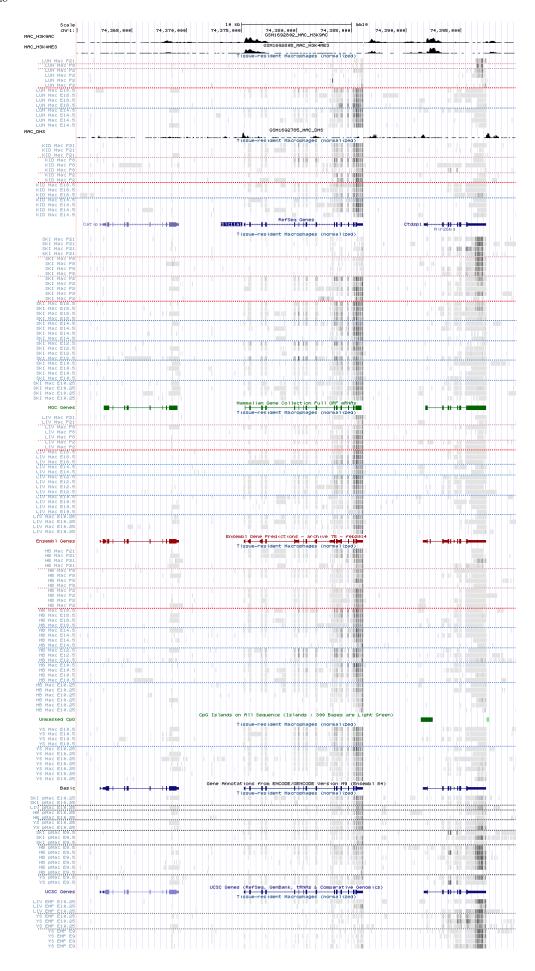




### (next three pages)

**Figure S24.** RNA-seq analysis of gene expression during mouse embryogenesis and organogenesis: *Nramp1* (**A**), *Emr1* (*F4/80*; **B**) and *Cx3cr1* (**C**) [118]. Transcript levels for the genes *Catip*, *Nramp1/Slc11a1* and *Ctdsp1*, across several developmental steps, from embryo to adult, including i) tissue MFs (Mac E10.25 and E10.5 (YS, HB, LIV, SKI); Mac E12.5 (HB, LIV, SKI); Mac E14.5 and E18.5 (HB, LIV, SKI, KID, LUN); Mac P2, P8 and P21 (HB, LIV, SKI, KID, LUN)) and ii) their precursors (YS EMP E9 and E10.25; LIV EMP 10.25; pMac E9.5 (YS, HB, SKI); pMac E10.25 (YS, HB, LIV, SKI)). *Cx3cr1* expression is the most precocious (starting in EMP), followed by *Nramp1* (beginning in pMac) and *Emr1* (*F4/80*; first detected in tissue Mac). Also, *Nramp1* expression is specifically down regulated in MFs of adult skin and lung (P8, P21 and P2, P8 and P21, respectively).

Α



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Figure S25. Epigenetic activity of *Nramp1* locus during primitive or definitive hematopoiesis. A. In vitro embryonic blood cell development: hematopoietic specification and differentiation starting from ESC [172]. Five successive stages of in vitro cell differentiation of ESC-derived embryoid bodies include: mesoderm cell (MES, day 1 of culture), hemangioblast (HB, day 2-3; smooth muscle, endothelial and hematopoietic potential), hemogenic endothelium (HE, day 4-5; endothelial and hematopoietic potential), CD41+ hematogenic precursor (HP, day 7) and Cd11b<sup>+</sup> MFs (MAC). Reported epigenetic modifications comprise ChIP-seq analyses directed either at histone modifications or transcription factors, and DNAse 1 hypersensitive sites (DHS), which together inform on chromatin activity. From top to bottom: candidate Nramp1 regulatory elements, labeled by analogy with NRAMP1 locus (cf Figure 7); mouse chromosome 1 scale; K27me3 (silent chromatin); UCSC genes; DHS (accessible chromatin); RefSeq genes; K27ac (activated chromatin); Ensembl genes; K9ac (transcriptionally active chromatin); Other RefSeq genes; K4me3 (transcriptionally active chromatin); CpG islands; association with TFs (Lmo2, Fli1, Tal1, Runx1, C/ebpb and Pu.1 (Spi1)). Note the presence of a cell-type-specific DHS spanning *Nramp1* TSS in MAC, which correlates with significant binding of few TFs in these cells only, including Pu.1, C/ebpb, Runx1, Tal1 but not Fli1 nor Lmo2 (Pu.1 binding was also detected in HP, with low intensity, data not shown). Nramp1 TSS and downstream area is also prominently decorated with K27ac, K9ac and K4me3 marks, which indicate gene transcription in MAC. Nramp1 TSS is labeled F1-like by analogy with human NRAMP1 regulation; other predicted regulatory areas that appear similarly positioned to human cis-elements are labeled accordingly (F12-like, F5-like, F7-like and F9-like) as well as Ctcf sites that either delimitate (E- and A-like) or flank Nramp1 locus in downstream (C- and D-like). See text for details. DHS and histone marks also suggest limited activity at F12-, F7- and F9-like predicted determinants in MAC (and HB, F12-like). Epigenetic marks at *Ctdsp1* appear unrelated or inversely related to *Nramp1* activity. The above data indicate, as observed with NRAMP1, that Nramp1 expression is restricted to late stages of myelo-monocytic differentiation and controlled by Pu.1 and C/ebpb TFs, consistent with the myeloid lineage determining roles of these factors. However, Nramp1 TF binding profiles differ from what was observed with NRAMP1 (e.g., Pu.1, C/ebpb, Runx1 and Tal1, compared to Figure 7 and Figures S19-22), implying regulatory divergence between mouse and human orthologous loci. **B.** Nramp1 gene activity in vivo in bone marrow monocyte and progenitor subsets [150]. Cells were sorted using surface antigens (Cd155<sup>hi</sup>, Cd117<sup>hi</sup> Cd135<sup>+</sup> and Ly6C<sup>hi</sup> or Ly6C<sup>lo</sup> for cMOP and MDP, respectively; Cd115<sup>hi</sup>, Cd117<sup>lo</sup>, Cd11c<sup>+</sup> and Ly6C<sup>hi</sup> or Ly6C<sup>hi</sup> for Ly6C<sup>hi</sup> and Ly6C<sup>lo</sup> MNs, respectively). ChIP-seq for Pu.1 (Ly6Chi and Ly6Clo MNs) and K27ac (MDP, cMOP, Ly6Chi and Ly6Clo MNs).

	Ctcf E-like	F <sub>12-like</sub> F <sub>5-like</sub>	F <sub>7-like</sub> F <sub>1-like</sub>	F 9-like	Ctcf A-like	Ctcf C-like	Ctcf D-like
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#### (next two pages)

Figure S26. De novo activation of *Nramp1* promoter in the myelo-monocytic lineage during definitive hematopoiesis in vivo (bone marrow) [25,173,177]. A. Myelopoiesis (long term HSC, LT-HSC; short-term HSC, ST-HSC; multipotent progenitor, MPP; CMP; GMP; GN; MN; MF). From top to bottom: candidate Nramp1 regulatory elements labeled by analogy with NRAMP1 locus (cf Figure 7); mouse chromosome 1 scale; Ctcf sites (murine erythroleukemia cells); UCSC genes; ChIP-seq for K27ac in B6, BXH2 (Irf8 R294C) and Irf1<sup>-/-</sup> BMDM, either naive of treated with Ifn-g; RefSeq genes; K27ac mark along myelopoiesis and in naive BMDM; other RefSeq genes; K4me1 mark along myelopoiesis and in naive BMDM (x2); super-enhancer tracks (MFs and pro-B cells); K4me2 mark along myelopoiesis; Ensembl genes; K4me3 mark along myelopoiesis and in naive BMDM; CpG islands; transposase accessible chromatin subjected to sequencing (ATAC-seq; Lsk (Lin<sup>-</sup>, Csa1<sup>hi</sup>, Kit<sup>hi</sup>, representing HSC), CMP, GMP, MN, GN); Pfam domains in UCSC genes. B. Erythropoiesis (MEPs; erythrocytic progenitors A, EryA (Ter119<sup>+</sup>, CD71<sup>+</sup>, high FSC); EryB (Ter119<sup>+</sup>, CD71<sup>+</sup>, low FSC) and lymphopoiesis (common lymphocytic progenitors, CLP; T lymphocytes Cd8+, CD8 and Cd4+, CD4; B lymphocytes, B; NK cells, NK). From top to bottom: candidate Nramp1 regulatory elements labeled by analogy with NRAMP1 locus (cf Figure 7); mouse chromosome 1 scale; Ctcf sites (murine erythroleukemia cells); RefSeq genes; naive BMDM RNA-seq data; ChIP-seq for K27ac in B6, BXH2 (Irf8 R294C) and Irf1+ BMDM, either naive of treated with Ifn-g; Other RefSeq genes; K27ac mark in naive BMDM and along erythropoiesis and lymphopoiesis; UCSC genes; K4me1 mark in naive BMDM (2) and along erythropoiesis and lymphopoiesis; super-enhancer (macrophage track); K4me2 mark along erythropoiesis and lymphopoiesis; super-enhancer (pro-B cell track); Ensembl genes; K4me3 mark in naive BMDM and along erythropoiesis and lymphopoiesis; CpG islands; ATAC-seq (MEP, CD4, CD8, B, NK); Pfam domains in UCSC genes. Note Nramp1 displays the stereotypical behavior of a de novo myelomonocytic gene (similar to S100A8 and S100A9, for instance) with stepwise acquisition of histone marks during differentiation: Nramp1 promoter becomes marked at the root of the myeloid commitment point (during the MPP to CMP transition; K4me1 and K4me2 marks), then primed in GMP (K27ac) and activated in mature phagocytes (K4me3; GN, Mono and MF). ATAC-seq areas were detected at F7- and F1-like elements, both starting in CMP and either maintained in mature phagocytes (F1-like) or reduced after GMP (F7-like), suggesting that co-activation of F7- and F1-like elements represent an intermediate state. Absence of epigenetic signal along erythropoiesis and lymphopoiesis demonstrates the myelo-monocytic specificity of Nramp1 expression.

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MEP_K27Ac				27Ac_EryA.ucsc	📥			
EryB_K27Ac			GSM1441276_K2	7Ac_EryB.ucsc	- +-			•
	<del>*</del>   *  -  -  +  -	+   +   +	GSM1441284_K	27AC_CLP.ucsc			··┼─│ │ <b>॑॓ऄॏ॓धी॑</b> वित्सच │ ┍┼ │	•
CD8_K27Ac				27Ac_CD8.ucsc				
CD4_K27Ac				27Ac_CD4.ucsc				
B_K27AC				K27Ac_B.ucsc				ТЦТ
NK_K27Ac	<b>*</b> _			27Ac_NK.ucsc	_ 🔺			
UCSC Genes I B6_K4me1	-	GSM	: (RefSeq, GenBank,	3S_frag200_res1_n	orm1e7		H+ <b>H</b> }====	
BMDM H3K4m1		BMDM 80 H3K4m	el Histone Mods by	ChIP-seq Signal	From ENCODE/LI	CR		
MEP_K4me1			GSM1441290_H3	3K4me1MEP.ucsc				
EryA_K4me1			GSM1441291_H3	K4me1EryA.ucsc				
EryB_K4me1		• +		K4me1EryB.ucsc				
CLP_K4me1				3K4me1CLP.ucsc	_			
CD8_K4me1				3K4me1CD8.ucsc				<b>.</b>
CD4_K4me1		- <b></b>		H3K4me1B.ucsc				
B_K4me1 NK_K4me1				3K4me1NK.ucsc			• • • • • • • • • • • • • •	
			SuperEnhance	rs Macrophage			·	
ancers Macrophage MEP_K4me2			GSM1441306_H3	3K4me2MEP.ucsc				
EryA_K4me2				K4me2EryA.ucsc				
EryB_K4me2				K4me2EryB.ucsc		-		
CLP_K4me2				8K4me2CLP.ucsc		-		
CD8_K4me2  CD4_K4me2	· · · · · · · ·			K4me2_CD4,ucsc				
CD4_K4me2	• 📥 - 🛛			13K4me2B.ucsc			·	
NK_K4me2		<b></b>	GSM1441313_H	3K4me2NK.ucsc				
ncers pro-B Cells	<b>≁</b>	- **	SuperEnhancer	s pro-B Cells	-			-
Ensemb1 Genes	+ <del>NB                                      </del>		mbl Gene Prediction	s – archive 65 –			H•18	
BMDM H3K4m3		BMDM 8w H3K4m		ChIP-seq Signal	From ENCODE/LI			
MEP_K4me3				K4me3EryA.ucsc				
EryB_K4me3			GSM1441324_H3	K4me3ĒryB.ucsc	- 🔺			-
CLP_K4me3	-			3K4me3CLP.Tucsc	-	· · · · · · · · ·		
CD8_K4me3			GSM1441330_H3	3K4me3CD8.ucsc				
CD4_K4me3		••	GSM1441329_H3	3K4me3CD4.ucsc				
B_K4me3				13K4me3B.ucsc				
NK_K4me3				3K4me3NK,ucsc				
Unmasked CpG MEP_ATAC		CpG Islands or	n All Sequence (Isl GSM146318	ands < 300 Bases 1_MEP,ucsc	are Light Gre	en)		
MEP_HIHC CD4_ATAC	+	· · - + <b>-</b> -   -		5_CD4.ucsc -	·	<b></b>	·· <mark>· · · · · · · · · · · · · · · · · ·</mark>	
CD8_ATAC	·   ≠- +   ≠-   +   +   +   +   +   +   +   +   +	·	- GSM146317	8_CD8.ucsc <sup>*+</sup>	👫	- <b>*</b>	┥┅┿┝╋╇┥┝╌┝	·    <del> </del>
B_ATAC		· - + -   -   -		72_8,ucsc	· + · - · <b>*</b> · -		····	-  + +
NK_ATAC				77_NK.ucsc				
Pfam in UCSC Gene			Pfam Domains H	in UCSC Genes	HH			

# (next page)

Figure S27. In vitro and in vivo regulation of Nramp1 gene expression [120,174–176,178,181]. From top to bottom: candidate Nramp1 regulatory elements labeled by analogy with NRAMP1 locus (cf Figure 7); mouse chromosome 1 scale; Ctcf sites (murine erythroleukemia cells); H2BK20ac ChiP-seq analyses of microglial BV2 cells (naive or stimulated with LPS with or without Tgfb knockdown) and untreated ESC; super-enhancer track (macrophage); K27ac mark in microglial BV2 cells (naive or stimulated with LPS with or without Tgfb knockdown) and untreated ESC; naive BMDM RNA-seq data; RefSeq genes; K27ac mark in naive BMDM, tissue-resident MFs (peritoneum, small intestine) and blood MNs; Other RefSeq genes; naive BMDM K4me1 mark; CpG islands; ATAC-seq analyses of developmental stages of microglia (yolk sac EMP, early microglia (until E14), pre-microglia (few weeks post-partum); adult mciroglia); UCSC genes; K4me2 mark in developmental stages of microglia (yolk sac EMP, early microglia, pre- and adult microglia); super-enhancer track (pro-B cells); K4me2 mark in bone marrow CMP subtypes (CD41+; CD41-; Flt3+ Csf1r+; Irf8- GFP+ MHCII+; Flt3<sup>-</sup> Csf1r<sup>-</sup> Irf8<sup>-</sup> GFP<sup>-</sup> MHCII<sup>-</sup>); Ensembl genes; K4me2 mark along in vitro definitive myelopoiesis (fetal liver HSC; MEP, CMP, GMP); DHS and Pu.1-specific ChiP-seq in GNs; Pfam domains in UCSC genes; K4me2 mark in tissue-resident MFs (peritoneum, small intestine) and blood MNs; K4me3 mark from naive BMDM and BMDM stimulated with LPS (0, 4,12 and 24h). Note MF populations studied in vitro display limited activation of F12-like area (e.g., K27ac in BMDM and BN2 cells vs tissue-resident MFs and blood MNs). In contrast, tissueresident MFs, including microglia, display high level of K27ac mark at F12-like element similar to TSS/F1-like; in addition, bone marrow myeloid progenitors show predominant K4me2 mark at F12-like element (four out of five CMP subtypes; HSC, MEP and CMP populations).

	Ctcf	12-like 5-like	7-like 1-like	9-like	Ctcf	Ctcf	Ctcf
	E-like			ь Ц	A-like	C-like	D-like
Scale chr1: 74,405,000	74,410,000  74,419	20 kb <b> </b> 5,000  74,42	0,000  74,425,000		74,435,000	mm9 74,440,000	74,445,000  74,450,000
MEL CTOF			L CTCF TFBS ChIP-set				
BV2LPS-TFGb_2BK20		ft	rack-BV2-LPS-TFGB-H				
BV2LPS_2BK20ac				K20ac-BV2-LPS.vste	p		
BV2_2BK20ac				28K20ac-BV2.vstep			
ESC_28K20ac		•		-H2BK20ac		<u></u>	
ancers Macrophage		f1	rack-BV2+LPS+TGFB-H	ers Macrophage 3K27ac-CMB018-mm9.	vstep		
BV2LPS-TGFb_K27ac  BV2LPS_K27ac				(27ac-BV2-LPS,vster			·
BV2_K27ac			ftrack-CRC001-	13k27ac-BV2.vstep		·	
ESC_K27ac				s–H3K27ac		<b></b>	· • • • • •
BMDM 8wk Sg 1		BMDM Adu	ilt 8 weeks RNA-seq	Level by the	ENCODE /LICR		
Prikd i	Cat ip • <b>•••</b> ••••••••••	→>> <b>  </b>	RefSe SIGUEI (++++++++++++++++++++++++++++++++++++	q Genes 	++ # <b>##</b>	Ctdsp1	<b>I-}</b>
Pnkd I BMDM H3K27a		вмом вы нака	27ac Histone Mods by	ChIP-seq Signal f	rom ENCODE/LI	Mir26b   DR	
monocyte_K27Ac_2			GSM1545993_merge	d_monocyte_K27Ac_2			
perit_mac_K27Ac_2			GSM1545988_merged_p	eritoneal_mac_K27A	c_2		
SI_Mac_K27Ac_2			GSM1545977_me	rged_SI_K27Ac_2			
Other RefSeq BMDM H3K4m1	HI I I	BMDM 8W H3K4	H H H	RefSeq Genes ChIP-seq Signal f	rom ENCODE/LI		
Unmasked CpG		CpG Islands	on All Sequence (Is		are Light Gree	n)	
Adult_microGl_ATA				dult_brain_ATAC			
Pre-microG1_ATAC	<b>.</b>	<u> </u>		87_NB_ATAC			
E12.5_microG1_ATA			<b>A</b>	RAIN_12_5_ATAC			<b></b>
YSp_12.5_ATAC	<b>b</b> a			d12_5_ys_ATAC			
UCSC Genes  Adult_microGl_k4m	+	UCSC Gene	es (RefSeq, GenBank, GSM2104282	tRNAs & Comparati	ve Genomics)	<b>₩     -  </b>	
Pre-microG1_k4me2			GSM2104284	_DAY_3_k4me2			
E12.5_microG1_k4m				_BRAIN_k4me2			
YSp_12.5_k4me2				_YS12.5_k4me2			
ncers pro-B Cells				rs pro-B Cells a kameo cmp coal m			
CMP_CD41_M_K4me2				d_K4ME2_CMP_CD41_M d_K4ME2_CMP_CD41_P	_ <b>_</b>	Antes des As	
CMP_CD41_P_K4me2	<b>_</b>	_ 🔺		erged_CMP_F1t3			
CMP_F1t3_K4me2				erged_CMP_MHCII		. <b></b>	· · · · · · · · · · · · · · · · · · ·
CMP_MHCII_K4me2	••			ged_CMP_tripleneg	<b></b>		····
CMP_trp1ng_K4me2 	· · • · ·	Ens			ec2011		· · · · · · · · · · · · · · · · · · ·
Ensembl Genes HSC_K4me2	+ <del>₩+ ++</del> +	sh	embl Gene Prediction	g counts for every	10 bp		
MEP_K4me2			ifted Merged MACS ta				
CMP_K4me2			ifted Merged MACS ta				
GMP_K4me2			ifted Merged MACS ta				
Gran2D_DNase_trea			ifted Merged MACS ta			<b>#</b>	
Gran2D_PU1			4 44	in UCSC Genes		· • • • • • • • • • • • • •	<b>_</b>
Pfam in UCSC Gene monocyte_K4Me2_2				d_monocyte_K4Me2_2	++	H-4	н
perit_mac_K4Me2_2	- +++  +++++	┥┿╸┼┼┼╴┝	GSM1545989_merged_p				╶┿╇╇╼┼┼│┽┾┾┾┼
SI_Mac_K4Me2_2		┼╼╾┥┤		rged_SI_K4Me2_2		·	
BMDM H3K4m3	· ┝ ┽	ВМОМ 80 НЗК-	Ame3 Histone Mods by	ChIP-seq Signal f	rom ENCODE/LI	DR	
BMDM_LPS0_K4me3	shi	fted Merged ta	g counts for every a	25 bp from chip-sed	ı data (by Wig	Maker3)	┼┼┼╦┽┿┤╎╴╎╎╎╎╎┤╎
		fted Merged ta	g counts for every :	25 bp from chip-sed	data (by Wig	Maker3)	
 BMDM_LPS12_K4me3		fted Merged ta	g counts for every :	25 bp from chip-sed	data (by Wig	Maker3)	┽╇╋╋┿┼┼║┽┼┥┥┥
BMDM_LPS24_K4me3		fted Merged ta	g counts for every a	25 bp from chip-sed	data (by Wig	Maker3)	
				- ! ! === + == == ! ! ! !			

#### (next page)

Figure S28. Regulation of *Nramp1* expression in vitro: association with TFs and response to infectious stimuli [126,180,182,184,185,187,188]. From top to bottom: candidate Nramp1 regulatory elements labeled by analogy with NRAMP1 locus (cf Figure 7); mouse chromosome 1 scale; ATAC-seq analyses of BMDM either naive or stimulated with lipid A; RefSeq genes; ChIP-seq analyses of Irf3 and RelA in BMDM stimulated with lipid A; Ctcf sites (murine erythroleukemia cells); RNA-seq of naive BMDC; co-ChIP-seq analyses in BMDC (Ctcf or C/ebpb or Pu.1 with H3, K27ac, K4me2 and K4me3); UCSC genes; ChIP-seq analyses of Irf8 and Pu.1 in Irf8+ Tot2 cells vs input; CpG islands; ChIP-seq analyses of Irf8 in BMDC (WT, Batf3<sup>-/-</sup> and Irf8<sup>-/-</sup>); Other RefSeq genes; RNA-seq of naive BMDM; ChIP-seq analyses of C/ebpa in macrophages, GMP and LSK (Lyn-, Sca1+, c-Kit-HSC population [186]); Ensembl genes; H4K8ac and K27ac activation marks in RN2 cells (MLL-AF9) and K27ac mark in BMDM; ChIP-seq analyses in RN2 cells (MLL-AF9) of C/ebpa, C/ebpb, Pu.1, Erg, Myb, Fli1 and p300. Note i) increased accessibility of F9-like element in response to lipid A treatment, which corresponds to timedependent chromatin association with both Irf3 and RelA; ii) association of F7-like element with C/ebpa and C/ebpb in cell types that do not express *Nramp1* (RN2 AML) or that express *Nramp1* at low level (BMDC, compare RNA-seq data between BMDC and BMDM, Nramp1 transcript abundance relative to Ctdsp1 mRNA levels) vs cells that express *Nramp1* at high level (e.g., C/ebpa in Mac; C/ebpb in ESCDM, Figure S25A); and iii) altered pattern of chromatin association of Irf8 in Batf3-/- BMDC.

Biology 2017, 6, 28

	Ctcf	F <sub>12-like</sub> F <sub>5-like</sub>	F <sub>7-like</sub> F <sub>1-like</sub>	9-like	Ctcf	Ctcf	Ctcf	
	E-like	<u>ч</u> ч 1	н н 1-	ъ Ъ	A-like	C-like	D-like	
Scale chr1: 74,405,00		20 kb) 5,000  74,420	,000 74,425,000	1 74,430,000	74,435,000	mm9 74,440,000	74,445,000	74,450,0
4A_120_rep3		Final_bedGr	aphs Total Tags =	1.66e+07, normalize			11 A. A. BALA	
1A_120_rep2		Final_bedGr		7.95e+06, normalize				
4A_0m_rep3		Final_bedGr			d to 1.00e+07		4	
4A_0m_rep2		Final_bedGr		d Genes	d to 1,00e+07		an an hàil m	
Pnkd I Pnkd I	Cat ip P <b>ali</b> () () ()		S160031 (>+>>> ++>>>	<del>╷╷╷╷╻╏╞╞╞╏╷╸╏╷╷╷╸╞┣╏</del>	÷₽	Ctdsp1 ++++++++++++++++++++++++++++++++++++		
RF3_120min_A		120min_f	A Total Tags = 1.99					
RF3_60min_A		60m in_A	A	e+07, normalized to	2.50e+07			
RF3_30min_A				olied Track				
RF3_15min_A		15min_A		e+07, normalized to	A			
RF3_0m in_A		0min_A	الما والعار والما والما		2.500+07			
te 1A_120v1				olied Track	·····			• • • • • • • •
e 1A_60v1				olied Track	<b></b>		<b></b>	
te 1A_30v1		<b></b>		olied Track				<b>.</b>
:e1A_15v1				olied Track	<b></b>			
e 1A_0v1		الالالة. المتاهنية. يبير		olied Track	ا. س.م.اه		مب _ م _ أستك فتقدر	بە بىد .
IEL CTCF		MEL	CICF IFBS ChiP-se	a signal from ENCOL	E/PSU	<u> </u>		
MDC_0hr		GSM1	620172_DC_0hr_th1		ips.bam			
OCTCF_H3.1					- 4 -	- +		
:oCTCF_K27ac.1				COCTCF_K27ac.1	_ ▲.	4		_
OCTCF_K4me2.1	- 4 -			coCTCF_K4me2.1 coCTCF_K4me3.1	<b>4</b> .			
CTCF_K4me3.1	••	-		_coCebpb_H3.1	<b>1</b> .		• <b>4</b> •	
©Cebpb_H3.1 				oCebpb_K27ac.1		<b> 8</b> 4		
:oCebpb_K27ac.1  :oCebpb_K4me2.1				OCebpb_K4me2.1			<b>.</b>	
	•- •- •- •			oCebpb_K4me3.1				÷ -
			- GSM223251	Ĩ_coPuĨ_H3.1	<b>م</b> د - م	<b>-</b> -	L	
 :oPu1_K27ac.1	- • • • • •		- GSM2232514	coPu1_K27ac.1	4	- 4		
:oPu1_K4me2.1		• •••	GSM2232517_	coPu1_K4me2.1	_ 🏎			
		·	GSM2232520_	.coPu1_K4me3.1		<b>.</b>		-
UCSC Genesı		UCSC Genes	s (RefSeq, GenBank,	tRNAs & Comparati	ve Genomics)			
Tot2-IRF8_IRF8		IRF8_ChIPseq	_Tot2 Total Tags =	2.45e+07, normaliz	ed to 1.00e+0	7		
Tot2-IRF8_PU1		PU1_ChIPseq_To	1 A A	= 3.69e+07, norma				
Tot2-IRF8_Input	Input	CDNA_ChIPseq_Tot	فرالمستح فراجه والع		normalized to			4
Unmasked CpG			n All Sequence (Is MSND Total Tags =					
(rf8_WT_DC			MSND TOCAT Tags =	- 1 540-87 poppa	ited to 1,888+87	.07		
rf8_Batf3K0_DC		IPIrf8onIrf8k	0_MSND Total Tags :	= 5.15e+06, normal	zed to 1.00e+	07		
(rf8_Irf8K0_DC		<u>a b -1</u>	Non-Mouse	RefSeq Genes	<b>6</b> • • • •		- <b></b>	
Other RefSeq SMDM 8wk Sg 2	F#I I	BMDM Adu	It 8 weeks RNA-seq		ENCODE/LICR	•{ 811		
lac_Cebpa		G	SM1223648_CEBPa_Mo	use_Macrophges_ChIf	'Seq		La receber.	
ас_серра 			GSM1187164_ChIP-se				· -··- <b>-</b> ·	
.SK_Cebpa		• <b> • •</b>	<b>A</b>	815_Cebpa				
·	····	Ense	mbl Gene Prediction	ns – archive 65 – o	lec2011	<u></u>	••• ••• •• •• •• ••	
Ensembl Genes N2_H4K8ac_1		╞──┼┼╋╋╸	GSM1614778_RN2	_H4K8ac_DMS0_rep1				
N2_H3K27ac_1	· ├ ┾ ╍┝╾┿╸┝╼┿╼┽┙╼┿╼┿╼┿╺┾		GSM1614774_RN2_	H3K27ac_DMS0_rep1				++++++
мом нзк27а		BMDM 8W H3K2	7ac Histone Mods by	ChIP-seq Signal f	rom ENCODE/LIC	R		-  - -  - -
N2_C_EBPa	╶┍┽┽┿╎┽┼┼┼┼╞╎┽		GSM1614791_R	N2_C_EBPa_rep1				
N2_C_EBPb			GSM1614793_R	N2_C_EBPb_rep1				
:N2_Pu1				_RN2_Pu1_rep1				
N2_Erg				RN2_Erg_rep1				
112_Myb			ar all an and a second	_RN2_Myb_rep1		ورجا ويعتدون أنفوهم ورور		
N2_F1i1			<b>A A</b>	RN2_Flii_repi				
N2_p300			GSM16147	34_RN2_p300			-	

Table S1. Expression of macrophage iron gen	nes
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<b>- - - - - - - - -</b>	Blood	Lung	Spleen	Mac <sup>#</sup> (max)	Ubq	TSS	Specificity <sup>¥</sup>	cS-E§
MN and PMN:								
SLC11A1	++	+	+	M1		no CpG no RPII NB4	MM	MN
SLC40A1	(+)	+	+	M2		CpG RPII NB4	(MM)	
FTH1	++	++	++	M1	+	CpG RPII NB4	(MM)	MN, Spl, Lng
NCOA4	++	++	++	++	+	CpG RPII NB4	(MM)	MN
FTL	++	++	++	++	+	CpG RPII NB4	(MM)	Spl
S100A8	++	++	++	M1		no CpG no RPII NB4	MM	
S100A9	++	++	++	+		no CpG no RPII NB4	MM	MN, Spl
MCOLN1	+	+	+	(M2)	low	CpG RPII NB4	(MM)	Spl
SLC11A2	+/-	(+)	(+)	+	low	CpG RPII NB4	(MM)	
TFRC	+/-	+	(+)	++	low	CpG RPII NB4	(MM)	
SLC49A1	+/-	+/-	+/-	(+/-)	vlo	CpG RPII NB4		
PCBP1	++	+	++	+	+	CpG RPII NB4	MM	MN, Spl, Lng
PCBP2	+	+	+	+	+	CpG RPII NB4		
CYBRD1	+/-	+	(+)	(M2)	+	CpG RPII NB4	(MM)	
GAPDH	++	++	++	(M1)	+	CpG RPII NB4	(MM)	MN, Spl, Lng
SLC48A1	+/-	+/-	+/-		vlo	CpG RPII NB4	(MM)	
MN and NOT in	PMN:							
CD91	+/-	+	+	(+/-)	+	CpG w/o RPII NB4	(MM)	
HMOX1	+	+	++	(M2)		CpG w/o RPII NB4	MM	
CD163	+	+	+	M2	~lo	no CpG no RPII NB4	MM	
SLC49A2	+/-	(+)	(+)	(M2)	vlo	CpG RPII NB4 (wk)	MM	
HAMP	+/-	+/-	+/-	(M2)		no CpG no RPII NB4	MM	
STEAP3	+/-	(+)	+/-	+	vlo	CpG RPII NB4	(MM)	
ITLN1	+/-	+/-	+/-	(M2)		no CpG no RPII NB4	MM	
SLC46A1	0	+/-	(+)	+/-	vlo	CpG w/o RPII NB4		
PMN and NOT i	n MN:							
LTF	+	+/-	(+)	+/-		CpG (wk) w/o RPII NB	34 MM	
LCN2	+	(+)	(+)	+/-		no CpG no RPII NB4		
NOT in PMN an								
SLC39A14	0	(+)	+/-	+		CpG RPII NB4		
TF	0	0	0	+/-		CpG w/o RPII NB4		
TFR2	+/-	0	0	+/-		CpG RPII NB4	(MM)	
LRP2	0	+/-	0			CpG w/o RPII NB4		
SCARA5	0	+/-	0	+/-		no CpG no RPII NB4		
СР	0	+/-	(+)		_	no CpG w/ RPII NB4		
HEPH	+/-	(+)	(+)	+/-	vlo	no CpG no RPII NB4		
PCBP3	+/-	+/-	+/-	+/-	vlo	no CpG no RPII NB4	(MM)	
PCBP4	+/-	(+)	(+)	(+/-)	low	CpG w/o RPII NB4	MM	

\*: compared to *SLC11A1* level; grey, no absolute values available

 $\ensuremath{\$}$  : candidate super-enhancer domain, MN: monocytes; Spl: spleen; Lng: lung

GTEx expression levels (rpkm) [50]:

++>200>+>20>(+)>5> +/-;

Ubq +>20>Ubq low>5>Ubq vlo

<sup>¥</sup> MM: myelo-monocytic

# Table S2. Macrophage iron gene mRNA expression levels [72].

Table S2. Macrop	hage iron gene mRNA	expression levels [72].				
Gene symbol	H1hESC	CD34	CD14	CD56	CD3	CD19
SLC11A1	1.47	47.82	7289.39	185.89	28.10	154.52
SLC40A1	1.70	7689.32	1681.30	1489.35	1621.83	709.28
FTH1	12478.75	52104.55	94947.65	27761.04	28520.66	14237.49
NCOA4	1944.77	15079.85	26725.16	8013.53	6585.10	6150.27
FTL	30226.82	36754.15	188509.89	32390.51	36014.26	36754.15
S100A8	0	3506.07	261343.64	2098.11	1047.96	3850.38
S100A9	0.60	4317.45	704402.40	3488.48	1958.10	4690.53
MCOLN1	1059.94	239.96	387.08	51.90	72.54	50.70
SLC11A2	1445.00	574.77	480.13	365.51	433.87	250.28
TFRC	2706.30	5399.45	752.67	967.00	1022.10	1378.73
SLC49A1	2125.13	230.89	328.92	286.20	298.91	321.54
PCBP1						
PCBP2	5600.73	17121.23	1967.30	3578.20	9041.77	2306.60
CYBRD1	466.81	1531.89	2657.30	711.21	73.03	138.75
GAPDH	630762.48	110615.15	61495.75	20351.54	25714.78	19192.00
SLC48A1	331.28	739.00	416.27	136.41	260.99	253.37
CD91	1814.69	49.40	2222.43	131.08	100.52	80.72
HMOX1	1690.77	43.28	8200.97	180.38	39.58	417.62
CD163	0.42	48.14	2233.87	91.48	12.53	35.68
SLC49A 2	471.22	30.48	884.00	38.14	52.87	31.47
HAMP	5.74	0	21.08	17.14	0	7.33
STEAP3	1866.10	228.60	193.95	46.93	1.76	45.01
ITLN1	0.41	6.06	139.99	5.36	0	1.74
SLC46A1	245.89	189.16	310.01	160.21	275.61	132.14
LTF	5.82	7.27	4.35	13.93	0	2.95
LCN2	11.24	22.64	5.11	88.17	0	40.74
SLC39A14	4531.96	212.85	46.63	130.75	224.29	148.33
TF	0.33	10.64	8.46	14.86	10.02	3.50
TFR2	75.49	1065.89	17.02	41.36	72.79	26.66
LRP2	82.61	3.59	1.23	0	1.01	0
SCARA5	0.63	0	8.06	17.55	0	30.25
СР	6.48	508.59	29.65	50.96	31.17	36.96
HEPHL1	9.82	6.35	17.26	59.32	6.05	40.86
PCBP3	34.13	92.03	0	18.24	20.22	15.29
PCBP4	893.95	132.47	29.58	195.63	299.66	264.89

Biology **2017**, 6, 28