



Review

# Point-of-Care-Testing in Acute Stroke Management: An Unmet Need Ripe for Technological Harvest

Dorin Harpaz <sup>1,2,3</sup> , Evgeni Eltzov <sup>4</sup>, Raymond C. S. Seet <sup>5</sup>, Robert S. Marks <sup>1,2,6,7,\*</sup> and Alfred I. Y. Tok <sup>2,3,\*</sup>

- Department of Biotechnology Engineering, Ben-Gurion University of the Negev, Beer-Sheva 84105, Israel; Harpaz.dorin@gmail.com
- School of Material Science & Engineering, Nanyang Technology University, 50 Nanyang Avenue, Singapore 639798, Singapore
- Institute for Sports Research (ISR), Nanyang Technology University and Loughborough University, Nanyang Avenue, Singapore 639798, Singapore
- <sup>4</sup> Agriculture Research Organization (ARO), Volcani Centre, Rishon LeTsiyon 15159, Israel; eltzov@gmail.com
- Department of Medicine, Yong Loo Lin School of Medicine, National University of Singapore, NUHS Tower Block, 1E Kent Ridge Road, Singapore 119228, Singapore; raymond\_seet@nuhs.edu.sg
- The National Institute for Biotechnology in the Negev, Ben-Gurion University of the Negev, Beer-Sheva 84105, Israel
- The Ilse Katz Centre for Meso and Nanoscale Science and Technology, Ben-Gurion University of the Negev, Beer-Sheva 84105, Israel
- \* Correspondence: rsmarks@bgu.ac.il (R.S.M.); MIYTok@ntu.edu.sg (A.I.Y.T.); Tel.: +65-6790-4935 (A.I.Y.T.)

Received: 4 July 2017; Accepted: 26 July 2017; Published: 3 August 2017

**Abstract:** Stroke, the second highest leading cause of death, is caused by an abrupt interruption of blood to the brain. Supply of blood needs to be promptly restored to salvage brain tissues from irreversible neuronal death. Existing assessment of stroke patients is based largely on detailed clinical evaluation that is complemented by neuroimaging methods. However, emerging data point to the potential use of blood-derived biomarkers in aiding clinical decision-making especially in the diagnosis of ischemic stroke, triaging patients for acute reperfusion therapies, and in informing stroke mechanisms and prognosis. The demand for newer techniques to deliver individualized information on-site for incorporation into a time-sensitive work-flow has become greater. In this review, we examine the roles of a portable and easy to use point-of-care-test (POCT) in shortening the time-to-treatment, classifying stroke subtypes and improving patient's outcome. We first examine the conventional stroke management workflow, then highlight situations where a bedside biomarker assessment might aid clinical decision-making. A novel stroke POCT approach is presented, which combines the use of quantitative and multiplex POCT platforms for the detection of specific stroke biomarkers, as well as data-mining tools to drive analytical processes. Further work is needed in the development of POCTs to fulfill an unmet need in acute stroke management.

**Keywords:** stroke; diagnostics; Point-of-Care-Test; biomarkers; time-dependent treatment; multiplex and quantitative detection; data-mining

# 1. Introduction

# 1.1. Stroke—A Leading Cause of Death

Approximately 15 million people worldwide suffer from stroke each year. More than 30% of stroke victims die every year; while over 30% suffer from permanent disability. Without intervention, the number of global deaths is projected to rise to 7.8 million in 2030 [1]. Stroke is the second most common cause of death and a major cause of disability. A common type of stroke is acute ischemic

Biosensors 2017, 7, 30 2 of 39

stroke (AIS) which is caused by an abrupt interruption of blood to the brain due to a thrombotic or embolic occlusion of the cerebral artery. If supply of blood is not promptly restored, this can lead to irreversible brain damage. AIS is contrasted with hemorrhagic stroke that is caused by rupture of blood vessels that are due to uncontrolled hypertension or underlying blood-vessel abnormalities such as cerebral aneurysms or arteriovenous malformations. Both the ischemic and hemorrhagic strokes can lead to permanent disabilities such as spasticity, depression, frequent falls and increased susceptibility to infections [2]. Although stroke frequently affects the elderly, young individuals are not spared by the threat of stroke [3]. In addition to traditional risk factors such as hypertension, dyslipidemia, diabetes mellitus, cigarette smoking, obesity and physical inactivity [2], individuals who suffer stroke during their youth harbor unique risk factors such as patent foramen ovale, Moya-moya disease, prothrombotic tendencies (e.g., anti-phospholipid antibodies and lupus anticoagulant) and mitochondrial disorders. Rehabilitation of stroke patients takes time–sometimes months or even years–and contributes to the rising healthcare costs in developed countries worldwide [4–6].

#### 1.2. Gaps in the Current System

The successful management of AIS lies with its timely recognition in the pre-hospital setting (ambulance) and a prompt diagnosis at the Emergency Department (ED). In clinical practice, several features, such as the sudden onset of neurologic symptoms (e.g., facial asymmetry, arm and leg weakness and slurring of speech) in the clinical history, point to a stroke diagnosis. Ischemic stroke is differentiated from hemorrhagic stroke by neuroimaging modalities such as a computed tomography (CT) of the brain and, in some centers, using magnetic resonance imaging (MRI) to provide a more accurate assessment of infarcted brain tissues and hemorrhages [7]. In patients with an AIS, recanalization of the occluded artery confers a high likelihood of clinical recovery. To date, intravenous recombinant tissue plasminogen activator (IV-tPA, Alteplase) is the only FDA-approved treatment for AIS, provided that it is administered within 4.5 h following stroke onset [8–10]. More recently, randomized trial data support the use of endovascular treatment (through mechanical clot retrieval and aspiration systems) to facilitate arterial recanalization [11–13]. Despite these advances, there exist critical gaps in the delivery of acute stroke care, namely insufficiently accurate diagnosis of AIS and its differentiation from other stroke mimics (seizures, migraine, brain space-occupying lesions and complicated migraine), need for an appropriate selection of patients for acute reperfusion treatment, better determination of stroke etiology and finally a critical triaging of patients for resource-intensive stroke units. Several blood-based biomarkers are now available to guide such clinical decision-making in stroke management that provides clinicians with an opportunity to confirm and personalize stroke diagnosis and risk [14]. With the growing incidence of stroke, there is an emerging need for timely bedside assessment of blood-based biomarkers. As such, the demand and adoption of these new technologies are likely to increase with time. A portable and easy to use POCT is desirable to enable time-sensitive treatment to be delivered promptly, preferably during the patient's first encounter with the paramedics or physicians. Stroke is a unique disease where treatment delays can cause significant repercussions in terms of long-term disability as a minute's delay in delivering acute stroke treatment can result in 1.9 million loss of brain neurons.

## 1.3. Rapid Diagnosis Improves Stroke Care

Brain cells die rapidly after stroke and any effective treatment must start as early as possible. In the clinical routine there is a "time-outcome" relationship, which continues to be the major limitation of therapeutic approaches [15,16]. The "time is brain" rational refers to the fact that brain cells die rapidly after the event of a stroke. The core of the focal ischemic lesion is irreversible and is destined to die, while the perilesional penumbra has lost its function due to reduced blood supply but maintains metabolic and structural integrity and, hence, may be salvaged. Nevertheless, there is strong evidence supporting the general association between time from onset and irreversible tissue damage—the so-called "time is brain" rationale [17,18]. Stroke has become a treatable disease: treatment with IV-tPA

Biosensors 2017, 7, 30 3 of 39

improves outcomes of AIS patients [12,18–20]. The effectiveness of most therapeutic options is time dependent and unfortunately, only a minority of eligible ischemic stroke patients receive recanalization therapies [21]. Time delays within the stroke survival chain are associated with worse outcomes [22]. Thrombolysis, IV-tPA treatment, rates remain low, with most patients being treated at the late end of the therapeutic window [17]. Current guidelines emphasize the need for early stroke care. However, significant delays occur during both the pre-hospital and in-hospital phases of care, making many patients ineligible for stroke therapies [23]. Time to thrombolysis is a critical determinant of favorable outcomes in AIS. It is a major clinical practice concern when patient outcomes are compromised due to out-of-hospital and in-hospital time delays [24]. Stroke outcome prediction mostly considers basic, most relevant, non-modifiable factors such as age and stroke severity. However, it is now well understood that stroke outcome can significantly worsen due to post-stroke complications, such as hemorrhagic transformations or post stroke infections [25]. In order to achieve the best outcome for the stroke patient, there is a need for an early diagnosis and identification of putative complications. For this on-going research, alternative-biological information, such as blood biomarkers and genetic polymorphisms, are examined in order to improve prognosis. This gap is currently addressed by barriers identification and solutions integration into the current practice. Effective reduction of time delays for AIS requires correct identification and targeted strategies [24,26,27]. The strategies to overcome those time barriers are categorized into pre-hospital, in-hospital and post-hospital strategies. Proposed pre-hospital strategies include public education on stroke symptoms awareness, prioritizing stroke by emergency medical services (EMS), increasing ease of access to medical records, pre-hospital notification and mobile CT scanning. In-hospital strategies include a streamlined code stroke system, CT scanner co-location with ED, 24/7 availability of stroke physicians, POCT or laboratory testing and access to expert neuroimaging interpretation. Post-hospital strategies include increasing availability of intravenous thrombolysis and simplification of informed consent procurement [24]. A POCT will enable a faster and better approach for stroke patients' care in their most critical state. POCT provides the relevant needed information on-site, without sample processing, central-laboratory long testing and results analysis procedures [28]. A POCT platform is a mobile laboratory device that is located directly at the site where the patient is treated, where the needed tests are performed by the same personnel who is treating the patient, thus potentially reducing interface times and examination times. POCT holds many advantages, such as simple measurement procedures, use of low sample volumes, automated data processing and off-course mobility [29]. The potential for POCT use in stroke care is therefore enormous.

## 2. Stroke Prognostic Care Shows Painful Needs

The stroke patient conventional prognostic care consists of multiple diagnostics, classifications and treatments steps, see Table 1 and Figure 1. It can be divided into 3 main categories in stroke prognostic care: (1) Pre-hospital care; (2) In-Hospital care (imaging, clinical tests, subtypes classification and therapeutic treatment) and (3) Post-Hospital care (recovery). The full scheme includes the stroke patient management from symptoms onset and up to the recovery phase. Pre-hospital management includes symptoms identification, dispatch alert and ambulance transportation to the hospital, while In-hospital management mainly includes brain imaging, nervous system evaluation through clinical tests, and most importantly stroke subtype classification followed by lifesaving therapeutic treatment. Post-hospital management mainly includes patient physical recovery, second stroke prevention and outcome improvement. This section presents the conventional stroke patient prognostic management scheme, while also focusing on the gaps in identification for outcome improvement.

Biosensors 2017, 7, 30

 Table 1. Stroke Conventional Prognostic Technologies.

Technology Name		Application	Clinical Value	Reference	
Imaging Technique which uses computerized x-ray imaging	Computed Tomography (CT)	3D scan of body tissues. Shows evidence of early ischemia and rules out haemorrhage. Can be performed with contrast agent for better visualization	Ischemic stroke diagnosis and admission of thrombolytic therapy	[2,30,31]	
	Multi-detector Computed Tomography (MDCT)	2D array of detector elements which enables multiple slices simultaneously, and faster image acquisition. High resolution and long range scans	Ischemic stroke diagnosis and admission of thrombolytic therapy	[2]	
	SPECT Computed Tomography (SPECT-CT)	This technology uses radioisotopes. Shows cross-sectional image of the target organ. The patient either swallows or is injected with a radioisotope, which travels to a target organ. The radioisotope emits radiation, which is detected.  Does not reliably distinguish between hemorrhage and infarction	Determine if a specific area of the body is receiving adequate blood flow	[32]	
	XENON-Contrast Computed Tomography (XENON-CT)	This technology uses the inert gas xenon to measure cerebral blood flow (CBF) in various brain regions. The patient inhales a mixture of xenon and oxygen over a period of a few minutes, allowing measurement of an increase in their density caused by their presence in the brain tissue	Determine local cerebral blood flow in small area	[32]	
	Positron Emission Tomography (PET)	Measures related changes in cortical function by tracking the chemical changes which occur in tissues. Detect biochemical changes in an organ or tissue that can identify the stroke onset before anatomical changes can be seen with other imaging processes	Guide decision making for brain surgical planning	[33]	
	Carotid Angiography (CA)	This technology uses dye to show the inside of human carotid arteries. A small tube (catheter) is put into an artery, usually in the groin (upper thigh), then moved upwards into one of the carotid arteries	Identification of extracranial vessel disease	[2]	
	Magnetic Resonance Imaging (MRI)	Show the slowing of water movement through the injured brain tissue, which is caused by ischemia and ruling out haemorrhage. Can detect a variety of changes in the brain and blood vessels and visualize blockages in the arteries	Ischemic stroke diagnosis and admission of thrombolytic therapy	[2,34–37]	
Imaging Technique which uses magnetic fields and radio waves	Magnetic Resonance Arteriogram (MRA)	Permits the visualization of blood flow in vessels and allows rapid characterization of the cervical and cephalic large vessels. Detects and grades cervical internal carotid stenosis with an accuracy of 85% to 96% compared to digital subtraction angiography  Identification of extracranial vessel disease		[34,35,38]	
	Diffusion-Weighted Imaging (DWI-MRI)	Can render ischemic fields visible within minutes of ischemia onset and detects the effects of stroke on caudal motor pathways in the recovery process.  Can also predict lasting motor impairment in stroke recovery	Early visualization of site and extent of ischemia	[35,38,39]	
	Perfusion-Weighted Imaging (PWI-MRI)	Used to assess cerebral blood flow and blood volume in various brain regions. Usually performed by injecting a contrast agent and then obtaining a rapid series of MRIs using an ultrafast technique	Early visualization of site and extent of ischemia	[35]	

*Biosensors* **2017**, 7, 30

 Table 1. Cont.

Technology	Name	Application	Clinical Value	Reference	
	Magnetic Resonance Spectroscopy (MRS)	Spectroscopy measurement, enabling measurement of ATP, lactate levels, and pH at discrete locations within the brain. Can distinguish areas that have no viable neurons from areas that may be salvageable	Identify areas in the brain that may be salvageable	[32]	
	Functional Magnetic Resonance Imaging (f-MRI)	Measurement of brain activity by detecting the changes in blood oxygenation and flow. Can measure differences in cognitive reserve	Monitoring patient recovery	[2,33,40–43]	
	Magnetic Resonance Imaging Diffusion Tensor (MRI-DTI)	Provide information on white matter damage. Correlate better with cognition than conventional MRI measures	Monitoring damage progression	[40,44-46]	
Imaging Technique which uses sound waves	Carotid Ultrasound (CU)	Show the inside of human carotid arteries and detect whether plaque has narrowed or blocked carotid arteries. Can include Doppler to show the speed and direction of blood flow through the blood vessels	Show the condition of the carotid arteries in the neck and/or intracranial vessels	[2]	
	B-Mode Carotid Ultrasound (B-CU)	Provides images of various levels, or planes, enabling the creation of a three-dimensional image of the carotid artery wall and surrounding structures. Provides information on the type and extent of arterial damage, though blood clots sometimes do not appear and the method cannot distinguish a narrowed from a completely occluded artery	dimensional image of the carotid artery wall and surrounding structures. des information on the type and extent of arterial damage, though blood sometimes do not appear and the method cannot distinguish a narrowed		
	Duplex-Carotid Ultrasound (D-CU)	Show the human carotid arteries and detect their condition. Combines B-mode imaging and pulsed Doppler ultrasound to provide more detail on the condition of arteries	Show the condition of the carotid arteries	[47]	
	Transcranial Doppler (TCD)	Probe is placed over areas on the head to detect blood velocity and pressure in certain arteries at various depths in the brain. Allows the assessment of the location and extent of occlusions or atheromatous plaques in extracranial carotid and large intracranial vessels	Show the condition of the carotid arteries and location of occlusions	[48]	
	Echo-Cardiography (ECHO)	Show images of the human heart, gives information on the size, shape and function of the organ. Can also detect possible blood clots inside the heart, and problems with the aorta		[2]	
Other	Electro-Cardiogram (ECG)	Records the heart's electrical activity, showing how fast the heart is beating, and its rhythm (steady or irregular). Can detect heart problems that may lead to a stroke and can also record the strength and timing of electrical signals which pass through the heart	Atrial fibrillation or previous heart attack diagnosis	[2]	
	Blood Tests (BT)	Includes: Glucose test, Platelets count and PT/PTT. Low glucose levels can cause symptoms similar to stroke and abnormal platelet levels can be a sign of bleeding/thrombotic disorder. Can also test whether the blood is clotting normally	Bleeding/Thrombotic disorders diagnosis	[2]	

Biosensors 2017, 7, 30 6 of 39

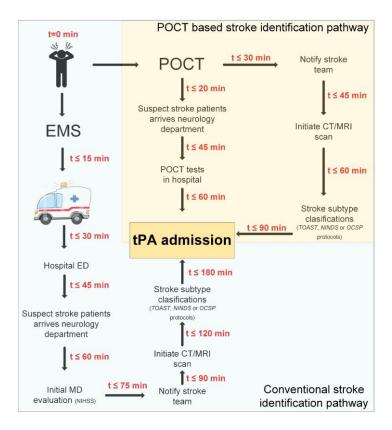


Figure 1. Stroke Patient Conventional Prognostic Management vs. Improved Using point-of-care-tests (POCTs). (1) In case of medical emergency (stroke symptoms onset), there is contact to emergency medical services (EMS) dispatch (t = 0). (2) EMS follows 'case entry protocol' and evaluate the patient medical status by specific parameters. (3) Ambulance patient-transportation to the hospital ( $t \le 15$  min). (4) Conventional stroke identification pathway: in the best scenario, the patients were already confirmed to be suffering from stroke by the EMS, so he is immediately transported to a stroke specialized unit. But in case he wasn't, the patient will go through the Emergency Department (ED), like any other patient suffering from a range of medical problems ( $t \le 30$  min). The suspected stroke patient will only then be evaluated by a neurologist ( $t \le 60$  min). The most common neurologic examination used is NIHSS, and followed by a CT/MRI scan for initial stroke diagnosis ( $t \le 90$  min). Subsequently, stroke subtype classification ( $t \le 120$  min) is conducted by either Trial of Org 10172 in Acute Stroke Treatment (TOAST), National Institute of Neurological Disorders and Stroke (NINDS) or Oxford Community Stroke Project (OCSP) classification schemes. Admission of (tissue plasminogen activator (tPA) will be on average less than 3hr from stroke-symptoms-onset (t  $\leq$  180 min). (5) POCT based stroke identification pathway: available pre-hospital POCT devices can shorten the time from stroke-symptoms-onset to neurologist examination (t  $\leq$  20 min). In addition, with the use of in-hospital POCT, the time to CT/MRI scan can be reduced ( $t \le 45 \text{ min}$ ) and, most importantly, the time to tPA admission can be reduced as well (60 min  $\leq$  t  $\leq$  90 min).

#### 2.1. Pre-Hospital: From Symptoms Onset to ED Admission

The risk of stroke is characterized by a combination of various factors, which include: previous stroke or transient ischemic attack, high blood pressure, physical inactivity, advanced age, diabetes, heart disease and smoking [2]. In most cases, stroke symptoms start suddenly (seconds to minutes), and depend on the area of the brain which is affected. If more areas of the brain are affected, more functions are likely to be lost. The major symptoms of stroke identified by the American Stroke Association include: (1) sudden weakness or numbness of arm, leg or face (especially on one side of the body); (2) trouble speaking/understanding and sudden confusion; (3) sudden vision trouble (one or both eyes); (4) sudden dizziness and loss of balance or co-ordination and (5) sudden severe

Biosensors 2017, 7, 30 7 of 39

headaches with no known cause [49]. Currently, the common term "Stroke chain of survival" is used to describe stroke patient diagnostic. This term refers to a rapid patient recognition and reaction to stroke warning signs. It is then followed by a rapid EMS dispatch, additional assessment [50], and rapid transportation with an urgent stroke unit/hospital pre-notification for a rapid in-hospital diagnosis and treatment [51–53]. Up to 70% of all stroke patients obtain first medical contact from the EMS [54]. Effective EMS systems can ultimately increase the number of patients reaching the hospital within the time window for thrombolytic therapy [55]. On the other hand, for more than 50% of all stroke cases, stroke clinical symptoms are sometimes not recognized by the EMS. The patient arrives to the hospital and is only upon arrival identified in the ED as a stroke patient. While going through a non-preprocessed patient admission, the critical survival rate for the patient significantly decreases [56]. Identification of pre-hospital stroke patients can be improved either by increasing access to EMS or by improving EMS/paramedics' patient evaluation and transport to hospital, while helping to target direct admission to the hospital as a stroke patient and thus avoid a general admission to the ED [57,58].

## 2.2. In-Hospital: Stroke Classification

Current stroke diagnosis remains largely a clinical diagnosis, with an addition of diagnostic and imaging tools. A common clinical technique conducts neurological evaluation using a scoring system, such as that using the National Institute of Health Stroke Scale (NIHSS), which is a 15-item neurologic examination stroke scale for use in acute stroke therapy [59]. In addition, an evaluation of the nervous system is conducted using a variety of imaging techniques [2], such as CT or MRI, which comprise a series of cross-sectional images of the head and brain [7]. CT/MRI scans can detect hemorrhage and are therefore useful for differentiating hemorrhagic stroke [60]. The additional existing classification technologies that may be used are: blood tests (glucose, platelet count, prothrombin time (PT), and partial thromboplastin time (PTT)), electrocardiogram, carotid ultrasound, carotid angiography, electrocardiogram, echocardiography and more [2,61]. Additional non-clinical diagnostic tools are needed to help classify stroke patients, and to better identify the patient medical status.

## 2.2.1. Ischemic vs. Hemorrhagic Stroke

Stroke is a heterogeneous disease with more than 150 known causes. Stroke initial classification distinguishes between ischemic and hemorrhagic stroke [62]. Ischemic stroke accounts for 88% of stroke cases and occurs as a result of blood flow interference within a blood vessel which supplies blood to the brain. This usually develops due to clot formation and accumulation of fat in the vessel walls. The blood clot can be formed in the same location as the accumulation of fat, which is then termed cerebral thrombosis, or in another location in the circulatory system, which is then called cerebral embolism. In the case of cerebral embolism, the blood clot is usually formed in the heart and in large arteries of the upper chest/neck. When it breaks down, it can then be released into the bloodstream, which may cause a blockage once it reaches small vessels. A second important cause of embolism is an irregular heartbeat, known as atrial fibrillation (AF). It creates conditions where clots can form in the heart, dislodge and travel to the brain [49]. The remaining 12% of stroke cases are due to Hemorrhagic stroke, which occurs when a weakened blood vessel ruptures and bleeds into the surrounding brain [49]. As a result, the blood accumulates and compresses the surrounding brain tissue. There are two types of weakened blood vessels which usually cause Hemorrhagic stroke: aneurysms and arteriovenous malformations (AVM), with the most common cause of hemorrhagic stroke being uncontrolled hypertension (high blood pressure) [2]. An aneurysm is a ballooning of a weakened region of a blood vessel. If left untreated, the aneurysm continues to weaken until it ruptures and bleeds into the brain. An AVM is a cluster of abnormally formed blood vessels. Any one of these vessels can rupture, also causing bleeding into the brain [49]. Of the hemorrhagic strokes, 9% are due to an intra-cerebral hemorrhage (ICH), and 3% are due to a subarachnoid hemorrhage (SAH) [63]. Victims of hemorrhagic strokes are often younger and the fatality rate is higher than for ischemic

Biosensors 2017, 7, 30 8 of 39

stroke. Overall prognosis is also poorer for those who have hemorrhagic strokes. The symptoms of a hemorrhagic stroke usually appear suddenly and often include: very severe headaches, nausea and vomiting, partial or total loss of consciousness [2]. This distinction between hemorrhagic and ischemic stroke is thus critical for stroke management and treatment decisions. It is usually completed using various imaging technologies that may be available albeit often limited by their availability, high cost and the need for professional personnel.

# 2.2.2. Ischemic Stroke Subtypes

The most common stroke is AIS [49], and therefore ischemic stroke etiologies (subtypes) classification is the second most important classification in stroke prognostic clinical care [64]. The 4 main etiologies of ischemic stroke are: (1) 20% atherothrombotic (large artery atherosclerosis, LAA), (2) 20% cardioembolic (CEI), (3) 25% small vessel disease (lacunar, LAC), and (4) 5% other causes. In addition, a fifth subtype is termed 'cryptogenic strokes', which refers to the cases of stroke from unknown causes. 'Cryptogenic strokes' accounts for 30% of stroke cases [65]. The main value of subtyping ischemic stroke is in classifying patients for a better targeted therapeutic decision-making process in clinical practice, to minimize time-to-thrombosis with treatment of IV-tPA admission. The average time-to-thrombosis is 3hr from stroke onset. In addition, stroke classification serves other purposes, such as describing patients' characteristics and grouping and phenotyping patients in a clinical study [65]. There are 3 common ischemic stroke subtypes classification schemes: (1) Trial of Org 10172 in Acute Stroke Treatment (TOAST) classification, (2) National Institute of Neurological Disorders and Stroke (NINDS) classification and (3) The Oxford Community Stroke Project (OCSP) classification, see Table 2.

Biosensors **2017**, 7, 30

 Table 2. Ischemic Stroke Subtype Classification Methods.

Description	Stroke Subtypes	Strengths	Weaknesses	Reference
	Trial of Org 10172 in Acute S	Stroke Treatment (TOAST) classificati	on	
Since 1993, the most clinically used method for ischemic stroke subtyping is TOAST, which is mainly based on clinical symptoms.	<ol> <li>LAA - Thrombosis or embolism from atherosclerosis of a large artery.</li> <li>CEI - Embolism from a cardiac origin.</li> <li>LAC - Occlusion of a small blood vessel.</li> <li>Other determined cause.</li> <li>Undetermined cause - includes either of the following cases:         <ol> <li>more than one possible cause;</li> <li>no cause is identified;</li> <li>incomplete investigation.</li> </ol> </li> </ol>	Reliability has been improved by the use of a computerized algorithm	Stroke from undetermined cause is the most heterogeneous group in the TOAST system, as well as in the Stroke Data Bank. Once a patient matches more than one possible cause, he is equally grouped as a patient with a no cause identified or an incomplete investigation. This weakness could flaw the medical decision-making process.	[38,66–69]
	National Institute of Neurological	Disorders and Stroke (NINDS) Class	sification	
Derived from the Harvard Stroke Registry classification.	<ol> <li>Brain tHemorrhages</li> <li>Brain Infarctions, which include atherothrombotic and tandem arterial abnormalities (LAA)</li> <li>CEI stroke</li> <li>LAC</li> <li>Stroke from rare causes or undetermined etiology</li> </ol>	The best option in the search of new causes of stroke amongst patients with no known causes or with another disease not causally related to the stroke event.	Most of the currently used diagnostic tools were not available at that time, such as modern MRI with diffusion-weighted imaging, transesophageal echocardiography, TEE, magnetic resonance angiography, MRA, duplex ultrasound examination, and transcranial Doppler	[70]
	Oxford Community Stroke Pro	ject (OCSP, Bamford/Oxford) classific	ation	
Relies on the initial symptoms and based on their extent.	<ol> <li>Total anterior circulation stroke (TAC)</li> <li>Partial anterior circulation stroke (PAC)</li> <li>LAC stroke</li> <li>Posterior circulation stroke (POC)</li> <li>The type of stroke is then coded by adding a final letter to the above:</li> <li>I—for infarct (e.g., TACI)</li> <li>H—for hemorrhage (e.g., TACH)</li> <li>S—for syndrome (e.g., TACS)</li> </ol>	Patients are easily classified into groups based on clinical grounds and CT scanning, which are usually done in all stroke patients. The outcome of the stroke event is driven strongly by the severity of the stroke, which is well reflected in this classification, without addressing the cause of the stroke.	The extent and site of the brain infarct is unlikely to be specific to a particular stroke etiology. Patients classified as having a LAC infarct may have a missed cardiac source of embolism. In addition, this classification should no longer be used to investigate potential risk factors or causes of stroke.	[71,72]

Biosensors 2017, 7, 30 10 of 39

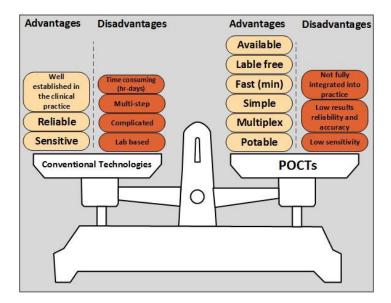
## 2.3. Post-Hospital: Recovery and Prevention of Stroke Reoccurrence

After therapeutic admission and patient stabilization, the patient continues to the recovery process which usually takes place in long-term care (LTC) facilities. The recovery procedure is customized to the patient's state and can last between a few days to up to a few months. The main focus for the stroke patient recovery is outcome improvement and second stroke prevention [73]. There are approximately 30 million stroke survivors globally and they are approximately one-quarter of the residents in LTC facilities. More than half of all global stroke survivors are left dependent on others to complete daily tasks. According to the U.S. National Stroke Association, some 10% of stroke survivors recover completely, 25% recover with minor impairments, 40% recover with moderate to severe impairments and require special care. In addition, 10% require care in a nursing home or other LTC facility, and 15% die shortly after the stroke [2]. The prediction of motor recovery assists in patient's rehabilitation planning. Neuroimaging and neurophysiological assessments are used to measure the extent of stroke damage to the motor system and predict subsequent recovery of function. Voluntary finger extension and shoulder abduction within five days of stroke predicted subsequent recovery of upper-limb function. Diffusion-weighted imaging within seven days detected the effects of stroke on caudal motor pathways and was predictive of lasting motor impairment. Moreover fMRI activation pattern at the acute phase might also be of great interest, both in motor recovery and in language recovery. MRI DTI sequence is also promising for prediction of motor outcome [39]. As stroke patient's recovery process focuses mainly on outcome improvement and second stroke prevention, there are many non-clinical values that can add to a better planning of the patient's recovery.

## 3. POCTs Expedite Stroke Prognostics

Stroke diagnostic tools consist of a wide range of available technologies, from simple to daunting in size and complicity, see Table 1 and Figure 1. A POCT technology is defined as an on-site test. Moreover, it is usually characterized as an easy to use, robust and mobile test which targets a specific need in the clinical practice, see Figure 2. Conventional technologies are usually found in an analytical lab due to their usually more complicated set-up and operation. POCT value proposition includes mobility, affordability, selectivity, being user-friendly, robust, and allows rapid result processing. Current conventional technologies do provide sufficient sensitivity and reliable results; however, they are not always considered practical. An important factor of a POCT development is that the test will be considered effective if action is taken based on its result. For example, POCT results could reduce hospital stay, improve adherence to treatment, and reduce complications [74]. The current POCT development and design research focus on minimizing the device size, while still obtaining highly sensitive and accurate results [75]. POCT successes include the glucose biosensor strips [76] and lateral flow immunoassay strips with the most well-known being the pregnancy test. The conventional clinical technology is ELISA (enzyme-linked immunosorbent assay) [77], reaching higher sensitivities then most POCTs available, but they require multiple steps and a complicated testing procedure. It is important to note that sensitivity is not always the most critical factor, especially in cases where there needs to be faster and more accessible treatments. POCT using biosensor related technologies may result in sufficient sensitivity and accuracy standards being obtained. This section presents different POCTs tested for their use in stroke prognostic management, see Table 3.

Biosensors 2017, 7, 30 11 of 39



**Figure 2.** POCT vs. Conventional Technologies. The advantage of POCT vs. conventional technologies are: portability, a simple structure, easy to use, allows multiplex detection, gives results within min, and also does not require labelling. However, a POCT demonstrates lower sensitivity and gives less reliable and accurate results. Conventional technologies are usually characterized with a more sensitive and reliable detection, but the limitations deny its usage as an on-site diagnostic tool. The disadvantages of conventional technologies are: lab facility requirement, results take hours or even days and complicated usage that requires professional personnel which results in a time-consuming process.

## 3.1. POCTs in Pre-Hospital Setting: Telemedicine and Mobile Stroke Unit

Stroke is a time-dependent medical emergency in which early presentation to specialist care reduces death and improves outcome. Identifying 'true stroke' in an EMS call is challenging, with over 50% of strokes being misclassified [54]. Recent innovations have opened up new perspectives for stroke diagnosis and treatment before the patient arrives at the hospital. These include improved stroke recognition by dispatchers and paramedics, mobile telemedicine for remote clinical examination and imaging, and integration of CT scanners and POCTs in ambulances. Several clinical trials were performed in the prehospital setting, aimed at testing prehospital delivery of neuroprotective, antihypertensive, and thrombolytic therapy. These new approaches shorten time to treatment and improve outcome [17]. The use of telemedicine improves the pre-hospital diagnosis of stroke and enhances the supervision of delivery of IV-tPA in AIS. If available, integrating stroke specialists in pre-hospital stroke response teams significantly reduces time to treatment [78], however this is not possible in a large proportion of locations. Remote access to a stroke specialist is now possible, and recent studies comparing in-person consultation with remote consultation suggest that telemedicine is a promising solution. NIHSS assessment of stroke patients using telemedicine is as reliable as face-to-face assessments [79]. And radiological review of brain CT in stroke management is both feasible and reliable, with the use of, for example, the 'ResolutionMD' mobile application which runs on a Smartphone and affords vascular neurologists access to radiological images of patients with stroke from remote sites in the context of a telemedicine evaluation [80]. There are as yet few definitive studies that have demonstrated whether it has an effect on clinical outcomes [81-84]. The integration of mobile CT scanner and POCTs in ambulances was first clinically tested in the mobile stroke unit (MSU) project of the University of Homburg, Saarland, Germany, where, an IV-tPA treatment can be started at the scene after exclusion of intracranial hemorrhage and coagulopathies, enabling patients to be then transported to a hospital in a normal ambulance. The results of the controlled study showed a remarkable reduction of time from alarm to therapy decision (median 35 min compared to 76 min in regular care) [85]. Time to treatment in those 12 patients who received IV-tPA was approximately

Biosensors 2017, 7, 30 12 of 39

halved and onset-to-treatment time was only 72 min (median) [86]. However, scanning failures (mainly technical) were reported in a number of patients (12 of 53). The MSU concept was introduced in Germany, demonstrating prehospital treatment of more patients within the first hour of symptom onset [87]. Additionally, The Stroke Emergency Mobile (STEMO) project of the Charité in Berlin has added new features [88,89] such as a scanner and POCTs in a fully equipped ambulance, enabling hyperacute treatment and transport in the same vehicle. This pilot study showed encouraging results for treatment safety and number of prehospital IV-tPA applications (23 treatments within 52 days) with a mean call-to-needle time of 62 min compared to 98 min. The data suggest that prehospital stroke care in STEMO is feasible, and no safety concerns have been raised so far. A third similar clinical study uses the mobile stroke treatment unit (MSTU) in the 'Cleveland Pre-Hospital Acute Stroke Treatment Study Group' [90], using a registered nurse, paramedic, emergency medical technician, and a CT technologist. Then, a cerebrovascular specialist evaluates the patient via telemedicine, whereas a neuroradiologist remotely evaluates images obtained by a portable CT scanner. In addition, a variety of POCT was performed, such as coagulation profile, complete blood count, and blood chemistry.

Biosensors **2017**, 7, 30

 Table 3. Stroke Prognostic POCT Devices Summary.

POCT Device	Description	Application	Clinical Value	Reference
Mobile Stroke Unit (MSU)	Imaging and a variety of Blood-tests. Integration of CT scanners and POCTs in ambulances, IV-tPA treatment can be started on-site	Consists of a registered nurse, paramedic, emergency medical technician, and a CT technologist, in addition, POCT are used, which includes coagulation profile, complete blood count, and blood chemistry	Pre-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[79,86–91]
CoaguChek®(Roche)	Test strips with electro-chemical detection. Based on amperometric (electrochemical) determination of the PT time after activation of the coagulation with human recombinant thromboplastin, results are obtained within 1 min		Pre/In-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[91–95]
Hemochron®Junior (ITC)	Optical detection. Micro-coagulation system, results within minutes	POCTs monitoring of: (1) ACT-LR, (2) ACT, (3) PT, (4) Citrate PT, (5) APTT and (6) Citrate APTT	Pre/In-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[96]
PocH-100i hematology analyzer (Sysmex)	Micro-fluidics. WBCs, RBCs and PLTs are counted using the direct current detection method with hydrodynamic focusing technology. Hemoglobin analysis is conducted using a non-cyanide method	Provides a full blood count and a 3-part differential leukocyte count	Pre/In-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[91,96,97]
i-STAT (Abbott)	Micro-fluidics. Based on advanced microfluidic and deliver fast, reliable lab accurate results within 2 min	Bedside care tests such as blood gases, electrolytes, metabolites and coagulation	Pre/In-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[91,98–100]
Reflotron®plus analyzer (Roche, Cobas series)	Test strips with optical detection (Reflectance photometry). Single-test clinical chemistry system which is able to measure whole blood, plasma or serum for: liver and pancreas enzymes, metabolites and blood lipids. Results within 2–3 mins	Used for blood clinical-chemistry parameters measurement, such as c-glutamyltransferase, p-amylase, and glucose	Pre/In-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[96]
Abbott AxSYM®BNP/Alere Triage®BNP/i-STAT BNP	Optical Detection (AxSYM)/Fluorescent detection (Triage)/Electro-chemical sensor on a silicon chip (i-STAT). BNP POCT.	BNP elevated serum levels in stroke patients show (1) correlation with CEI stroke, (2) increased mortality and (3) indication on second stroke recurrence	In/Post-hospital: Improves stroke prognostic (correlation to CEI) and stroke recovery (indication on second stroke reoccurrence)	[88,101–126]
Cornell University, State University of New York and the New York Presbyterian Hospital	Luminescent detection. NSE POCT based on enzymes tethered to nanoparticles	NSE elevated serum levels in stroke patients assist in distinguishing stroke from mimics, an important first step in expediting the diagnostic process	In-hospital: Improves stroke diagnosis and reduces time-to-IV-tPA admission	[127–133]

Biosensors **2017**, 7, 30

Table 3. Cont.

POCT Device	Description	Application	Clinical Value	Reference
Prediction Sciences LLC	Lateral flow POCT for the proteomic marker cellular fibronectin (c-Fn)	Fibronectin (c-Fn) elevated serum levels in stroke patients at IV-tPA admission can identify if the patient is at high or low risk for a subsequent hemorrhage	In-hospital: Reduces time-to-IV-tPA admission	[124]
ReST™ (Valtari Bio™ Inc.)	Rapid evaluation stroke triage POCT for the measurement of blood brain-specific biomarkers associated with immune responses, results within 10 min	Following stroke, the immune system is activated. The degree and direction of the immune system activation allow the accurate identification of acute stroke from non-stroke	In-hospital: Initial stroke versus no stroke diagnosis	[134–136]
SMARTChip (Sarissa Biomedical)	Micro-electrode POCT device for stroke diagnosis that measures purines from a drop of whole blood and give the reading within minutes	Can be used by paramedics, which will allow faster identification of stoke victims at the point of injury	In-hospital: Stroke diagnosis	[137]
PFA-100®, Platelet Function Analyzer (Dade Behring)	High-shear force dynamic flow system POCT that assesses platelet aggregation under high shear, mimicking platelet-rich thrombus formation after injury to a small vessel wall under flow conditions	Rapid and reliable identification of aspirin non-responsive patients, without the requirement of a specialized laboratory	Post-hospital: Prevention of second stroke recurrence	[138–155]
Ultegra-RPFA VerifyNow Aspirin®test (Accumetrics)	Optical detection. POCT based on turbidimetric optical detection of platelet aggregation in whole blood. As aggregation occurs, the system converts luminosity transmittance results into 'Aspirin Reaction Units'	Rapid and reliable identification of aspirin non-responsive patients, without the requirement of a specialized laboratory	Post-hospital: Prevention of second stroke recurrence	[138–154]

Biosensors 2017, 7, 30 15 of 39

#### 3.2. POCTs in In-Hospital Setting: Coagumeters, Blood-Count, Blood-Chemistry and Biomarkers

Prior to the admission of IV-tPA, laboratory results such as coagulation profile (international normalized ratio (INR) and activated partial thromboplastin time (APTT)), blood count (platelet, leukocyte and erythrocyte count) and blood chemistry (hemoglobin, glucose, c-glutamyltransferase and p-amylase test) are required. However, in conventional clinical practices, these valuable tests are not completed due to the time consuming diagnostic procedure [13,95]. Most common prognostic guidelines of stroke recommend that thrombolytic therapy should not be delayed while waiting for these test results unless there is clinical suspicion of a bleeding abnormality or thrombocytopenia, or in cases where either the patient has received anticoagulants (heparin or warfarin) or if the use of anticoagulants is not known [13]. This might lead to an increased risk of overlooking stroke mimics or patients with contraindications for thrombolysis treatment admission [95]. In ED, 50% of non-strokes are misdiagnosed as stroke [134]. The current clinical standard is based solely on a clinician's assessment of symptoms and rudimentary stroke scale tools. There is no simple, immediate, and unbiased way to diagnose stroke [135]. For example, studies have showed that serum glucose could be safely obtained by paramedics, and that INR POCT can reduce door-to-needle times [24,155]. Part of these tests can be completed even before arriving to the hospital when integrated to a mobile stroke unit. However, in some cases, the use of such POCT as a bed-side tool in the emergence department has enabled faster IV-tPA admission and stroke patient's improved outcome. In a recent study [95], they compared the results obtained from POCT to those obtained from the central hospital laboratory. They showed that when using POCT instead of using the central hospital laboratory, the time-to-therapy was reduced from  $84 \pm 26$  to  $40 \pm 24$  min (p < 0.001) and the results of most laboratory tests (except APTT and INR) revealed close agreement with the results from a standard centralized hospital laboratory. However, the accuracy and effectiveness of POCT in emergency management of AIS has not been fully clinically tested [91].

#### 3.2.1. Coagumeters

By using coagumeters POCT, INR values can be measured immediately at the bedside [91]. Roche Company commercialized CoaguChek®, a convenient, portable and user-friendly instrument for monitoring oral anticoagulation therapy and determining the INR value from a drop of capillary whole blood. This POCT was used for both pre-hospital stroke care [90,92] and in-hospital stroke care [91,93]. CoaguChek® test principle is based on amperometric (electrochemical) detection of the coagulation response of the plateletafter activation with the human recombinant thromboplastin. The test's user interface is a simple one with icon-based liquid crystal display (LCD). After placing a blood drop on the test strip, results are obtained within 1 min. INR POCT results correlates well with laboratory values and can be used to shorten door-to-needle time [92,94]. As mentioned previously, an APTT POCT, like the Hemochron® Junior (ITC) which was tested as an improved strategy for in-hospital stroke care, is also required [95]. Being a micro-coagulation system, it offers point-of-care monitoring of: (1) ACT-LR: Low Range Activated Clotting Time, (2) ACT+: Activated Clotting Time Plus, (3) PT, (4) Citrate PT, (5) APTT and (6) Citrate APTT. The test results are received within minutes.

## 3.2.2. Blood-Count POCT

Blood-count tests are also needed in stroke patient prognosis. A POCT specific for blood-count is the PocH-100i hematology analyzer (Sysmex), which is a compact device designed specifically for a POCT environment. The analyzer provides a full blood count and a 3-part differential leukocyte count [96]. This POCT is used for both pre-hospital stroke care [90] and in-hospital stroke care [95]. This POCT is designed for laboratories testing of up to 25 samples per day, and detects red blood cells and platelets count. White blood cells (WBCs), red blood cells (RBCs) and platelets (PLTs) are counted using the direct current detection method with hydrodynamic focusing technology to minimize coincidence or recirculation. Hemoglobin analysis is conducted using a non-cyanide

Biosensors 2017, 7, 30 16 of 39

method. Hematocrit is directly determined based on the red cell count and volume detection of each individual RBC.

#### 3.2.3. Blood-Chemistry POCTs

There are two commercial blood-chemistry POCT; the i-STAT (Abbott) and the Reflotron<sup>®</sup>. The first one is a portable clinical analyzer designed to be used at the patient's bedside for critical care tests for blood gases, electrolytes, metabolites and coagulation, and was used for both pre-hospital stroke care [90] and in-hospital stroke care [97,98]. In a recent stroke clinical study [97], they tested a 3-tiered system together with a POCT INR in determining use of tissue-type plasminogen activator. This portable POCT uses advanced microfluidics and delivers fast, reliable, lab accurate results within 2 min. The Wide test menu includes: blood gases, electrolytes, blood chemistries, coagulation, cardiac markers (cTnI) and hematology. A clinical study was supervised by Abbott and conducted in an anonymous hospital [99], which aimed to revitalize the ED's systems by including new emergent protocols that integrated bedside POCTs. As part of this new protocol, the i-STAT® System CHEM8+ (basic metabolic panel), cTnI (troponin I), CG4+ (blood gas with lactate), and PT/INR were implemented in an effort to improve diagnostic efficiency and patient flow. Incorporating i-STAT CHEM8+, cTnI, CG4+, and PT/INR into nurse-driven emergent protocols made measurable advancements in the diagnostic-process efficiency. The ED staff was empowered to accelerate diagnosis, treatment, and disposition of patients. Another blood-chemistry POCT is the Reflotron® plus analyzer (Roche, Cobas series), which is used for the measurement of c-glutamyltransferase, p-amylase, and glucose. It was tested as an improved strategy for in-hospital stroke care [95]. This is a single-test clinical chemistry system which is able to measure whole blood, plasma or serum for: liver and pancreas enzymes, metabolites and blood lipids. The full 17 parameters include: bilirubin, cholesterol, creatinine, glucose, hemoglobin, , potassium, triglycerides, uric acid, urea, alkaline phosphates, amylase, pancreatic, creatine kinase (CK), gamma glutamyl transpeptidase (GGT), glutamic oxaloacetic transaminase (GOT), aspartate aminotransferase (AST), glutamic-pyruvic transaminase (GPT) and alanine aminotransferase (ALT). The test gives on-site and reliable test results within 2–3 min.

#### 3.2.4. POCTs for Biomarkers Measurement

There are also a few POCT that are directed against a specific stroke related biomarkers measurement. A recent clinical study [109] demonstrated the usefulness of using the B-Type Natriuretic Peptide (BNP) POCT platform on suspected ischemic stroke patients in the ED in order to complete stroke subtype classification. BNP is a well-known biomarker for heart failure (HF), and its measurement is integrated into the established clinical practice of cardiologists. BNP is now identified as potentially useful for stroke patient prognostic and recovery management. BNP elevated serum levels in stroke patients show correlation with CEI stroke [87,100–108,112–114,120]. It is recommended to add a plasma BNP test at the bedside and integrate it into stroke guidelines in the ED, so that suspected stroke patients can obtain their laboratory assessments within 10 min of arrival at the ED, while the plasma BNP concentration can be measured immediately at the bedside [125]. In the case of a high plasma BNP level, emergency physicians and neurologists should strongly consider CEI stroke subtype. There are more than 20 registered devices for HF BNP measurement in the FDA, which include: Abbott AxSYM® BNP, Alere Triage® BNP, i-STAT BNP test and more.

Another POCT directed against stroke biomarker measurement, was developed by researchers from Cornell University, State University of New York and the New York Presbyterian Hospital. This POCT is based on enzymes tethered to nanoparticles [126] for the detection of neuron-specific enolase (NSE). Immobilization of pyruvate kinase (PK) and luciferase on silica NPs was used to achieve rapid and sensitive detection of NSE, a well-known clinically relevant biomarker for stroke. The researchers show that their data match well (r = 0.815) with the current gold standard for biomarker detection, ELISA. Moreover, they have a great advantage over ELISA as they can achieve detection in 10 min as opposed to the several hours required for traditional ELISA. Although no single biomarker

Biosensors 2017, 7, 30 17 of 39

will likely provide a definitive diagnosis of any disease, the glycolytic enzyme, NSE, is released from damaged neurons and has been suggested to be valuable for the diagnosis of various brain injuries. NSE has been suggested to be useful in distinguishing stroke from mimics, an important first step in expediting the diagnostic process [127–129]. As an alternative to antibody capture, POCTs based on fluid phase enzymatic activities [130,131] or semi-solid phase bioluminescence [132] are used for plasma NSE monitoring.

A third POCT directed against a stroke biomarker was developed by Prediction Sciences LLC (California, USA). This POCT is directed for the measurement of proteomic marker cellular fibronectin (c-Fn) [123], which has been shown in recent studies to predict hemorrhagic transformation in tPA-treated patients with a sensitivity of 100%. As mentioned previously, admission of IV-tPA is limited to a critical time window of 3 h and the amount of c-Fn in the blood of stroke patients at admission can identify if the patient is at high or low risk for a subsequent hemorrhage. This POCT platform is based on lateral flow technology detection of c-Fn, with the ability to obtain results within 10 min.

Valtari Bio<sup>TM</sup> Inc., a company from West-Virginia (USA), is also developing a POCT for the detection of stroke related biomarkers. ReST<sup>TM</sup>, is a rapid evaluation stroke triage test [133], which is aimed at improving the initial stroke versus no stroke determination in ten minutes or less. This approach is based on measurement of blood brain-specific biomarkers associated with immune responses, for better stroke identification. The degree and direction of the immune system activation, following stroke and brain injury, allow the accurate identification of acute stroke from non-stroke. In addition, they employ machine learning and pattern recognition tools in order to identify different immune response patterns in the peripheral blood following various types of neuro-related brain injury. Their method is optimized due to their use of pattern recognition and ratios of biomarkers, rather than the absolute measurement of specific biomarkers. The company conducted clinical studies on over 500 real-world patients, and their preliminary clinical trial data suggests that the sensitivity and specificity for diagnosing stroke, using a pattern of expression of associated immune related biomarkers, are much higher than current clinical practice.

Furthermore, Sarissa Biomedical, a spin-off company from Coventry (UK), is developing POCT for stroke related biomarkers. SMARTChip [136] is a POCT device for stroke diagnosis that can be used by a paramedic, which will allow faster identification of stroke victims at the point of injury, and facilitate rapid coordination of the clinical treatment pathway to maximize the chances of the best possible patient outcomes. The SMARTChip can measure purines from just a drop of whole unprocessed blood and give the reading within minutes. Purines (e.g., adenosine, inosine and hypoxanthine) are neurochemicals that influence the function of the nervous system and can be used as stroke biomarkers as they are released after stroke onset. This SMARTChip POCT consist of a 3-layer structure, that converts a "dumb" microelectrode into a "smart" device capable of measuring specific analytes in real-time. The analyte sensitivity depends upon the nature of the enzymes included in the sensing layer. The upper sensing layer is termed 'biolayer' and contains the bioreporter (enzyme) and placed sample, with/without the target analyte. The layer underneath the 'biolayer' is termed 'mediator' layer, and the bottom layer is the electrode which transmits the signal. The current SMARTChip platforms available are for the detection of the following analytes: adenosine triphosphate (ATP), adenosine, inosine, hypoxanthine, acetylcholine, choline, glutamate, glucose, lactate and D-serine.

## 3.3. POCTs in Post-Hospital Setting: Aspirin Resistance and Biomarkers

## 3.3.1. POCT for Biomarkers Measurement

Recent clinical studies clearly show potential significance for adapting clinical bio markers in a stroke patient's rehabilitation procedures, which would help to ensure appropriate individual care of the patient [39]. As mentioned previously, BNP POCT were presented in the correlation with CEI stroke subtype. However, BNP measurement can also be valuable for stroke patient's recovery. BNP elevated

Biosensors 2017, 7, 30 18 of 39

serum levels in stroke patients also show a correlation with increased mortality [115–119,121,122,124] and indication of second stroke recurrence [110,111,156]. By monitoring BNP levels in stroke patients in their recovery process, the clinical staff will be able to better understand the stroke patient's status by predicting mortality and preventing second stroke recurrence. As previously mentioned, there are more than 20 registered devices for HF BNP measurement in the FDA.

### 3.3.2. Aspirin Resistance POCTs

Another potentially useful POCT for stroke recovery is to screen for aspirin responsiveness after transient ischemic attack (TIA) and stroke [137]. Aspirin (ASA), the most commonly used antiplatelet agent, reduces the relative risk of major vascular events and vascular death by 20% after ischemic stroke [138]. However, the antiplatelet properties of ASA are not uniform between individuals and recurrent events, which may be caused by 'ASA resistance' or ASA non-responsiveness [139-146]. There is evidence that ASA nonresponsive individuals may be at increased risk of ischemic vascular events [147,148]. The response to ASA should be monitored in post-hospital care of stroke patients for the prevention of second stroke recurrence [150,153]. However, the platelet function tests for ASA monitoring are time-consuming and difficult to follow as a routine practice. There are simpler platelet function POCT, such as the PFA-100®(Platelet Function Analyzer, Dade) and the Ultegra-RPFA Verify Now Aspirin®test (RPFA) [149,151,154] available to screen stroke patients for 'ASA resistance'. These POCT offer the possibility of a rapid and reliable identification of ASA non-responsive patients, without the requirement of a specialized laboratory [152]. Harrison et al compared the use of both the PFA-100® and the RPFA in 100 patients with transient ischemic attack or stroke receiving daily low-dose ASA treatment. Aspirin non-responsiveness is highly test-specific and large prospective studies should determine the prognostic value for each POCT used.

#### 4. Key Elements in Novel Stroke-POCTs

#### 4.1. What is an Ideal Brain Biomarker

Novel POCT devices should show the use of stroke related biomarkers. Ideal brain biomarkers are usually proteins which can be measured frequently from bio-fluids using safe methods, in order to provide non-clinical data on specific organs, mainly the brain and spinal cord. Those biomarkers demonstrate: (1) specificity—uniquely present in the central nervous system (CNS) and reflect the extent of brain damage; (2) sensitivity—abundant and easily detected; (3) selectivity-for example, reflect therapeutic efficacy [157]. Also, there is a need for resemblance to injury biomarker characteristics, for example, the biomarker needs to be resistant to cytoplasmic and extracellular proteolytic activity and not be dependent on renal excretion [158]. Biomarkers can be classified into three main classes: (1) susceptibility—reveal subjects with genetically mediated predisposition to a specific condition; (2) effect—measure early biological effect (structural or functional changes in affected cells or tissues); (3) exposure-measure chemicals or their metabolites to determine a patient's exposure to them [159]. The identification and use of such ideal brain biomarkers would be useful for identification of patients at risk for stroke and also to detect and monitor their treatment [158].

#### 4.2. Specific Stroke Related Biomarkers

Stroke is associated with a variety of pathophysiological changes, which leads to triggering different bio-chemical processes [14]. This results in a big variety of stroke related biomarkers (Table 4) for which their clinical practice values are yet to be fully determined [160]. A convenient way to categorize stroke biomarkers is by their origin, which includes the following groups: glial cells origin, neuronal cells origin, heart muscle cells (cardiomyocytes) origin, blood vessels cells (myocytes) origin, general inflammatory cytokines, cytoskeleton proteins, hemostatic proteins, lipids, metabolic proteins and others. Glial cells' origin stroke biomarkers include Protein S100-Beta (S100B) [157, 158,161–165], Glial Fibrillary Acidic Protein (GFAP) [157,161,162,166–171] and Myelin Basic Protein

Biosensors 2017, 7, 30 19 of 39

(MBP) [172–175], whereas that for Neuronal cells include, Neuron-Specific Enolase (NSE) [129,157,158, 161-163,166,176,177], Ubiquitin Carboxyl-terminal Hydrolase L1 (UCH-L1) [157,158,178-181] and Creatine Kinase-BB (CK-BB) [182,183]. On the other hand, Heart muscle cells (cardiomyocytes)  $include \ B-Type \ Natriuretic \ Peptide \ (BNP) \ [87,100-108,110-122,124,156,184-186] \ and \ Blood \ vessels \ cells$ (myocytes) exhibit Matrix Metallo-Proteinase 9 (MMP-9) [187–191]. General inflammatory cytokines and proteins stroke biomarkers include, interleukin-6 (IL-6), interleukin-1b (IL-1b), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) [166,192–199] and inflammatory protein Neutrophil Lymphocyte Ratios (NLR) [199–206], while Cytoskeleton proteins include, neurofilaments (NFs) [207-209], cleaved-tau (C-tau) [157,158, 172,210–213], microtubule-associated protein 2 (MAP2) [214–217] and alpha-II spectrin break-down products (SBDPs) [157,218–221], Finally, Hemostatic stroke biomarkers include, D-dimer [11,222–240], C-reactive protein (CRP) [11,241–245], Fibrin monomer complex (FMC) [246], soluble fibrin (SF) [246], fibrinogen [246,247], fibrin/fibrinogen degradation products (FDPs) [246] and von willebrand factor (vWF) [247], and those of Lipid origin include, Triglycerides [248-250], Low density lipoprotein (LDL)/High density lipoprotein (HDL) [248–250], heart fatty acid binding protein (H-FABP) [251,252], free fatty acid (FFA) [253], ApoA [254,255] and ApoE4 [249,256], Whereas Metabolic proteins will have: Lactate dehydrogenase (LD) [182,257] and Albumin [258]. Other origin stroke biomarker is Decorin [259].

Biosensors **2017**, 7, 30

 Table 4. Stroke Related Biomarkers Summary.

Family	Biomarker	Expression Association	Stroke Clinical Value	Reference
	S100-Beta	Associated with blood-brain barrier (BBB) dysfunction	Diagnosis	[157,158,161–165]
Glial cells origin	Glial Fibrillary Acidic Protein (GFAP)	Associated with the size of brain lesions, the neurological status and short-term functional outcome	Prognosis: outcome prediction	[157,161,162,166–171]
	Myelin Basic Protein (MBP)	Associated with worsened outcome	Prognosis: outcome prediction	[172–175]
	Neuron-Specific Enolase (NSE)	Associated with neurological outcomes and infarct volume	Prognosis: outcome prediction	[129,157,158,161–163,166,176,177]
Neuronal cells origin	Ubiquitin Carboxyl-terminal Hydrolase L1 (UCH-L1)	Associated with the extent of the neuronal injury	Prognosis: outcome prediction	[157,158,178–181]
	Creatinine Kinase-BB (CKBB)	Associated with the extent and severity of the brain damage and the recovery potential	Prognosis and recovery	[182,183]
Heart muscle cells (cardio-myocytes) origin	B-Type Natriuretic Peptide (BNP)	Associated with CEI ischemic stroke, increased mortality and second stroke indication	Prognosis: subtype classification and recovery: second stroke prevention	[87,100–108,110–122,124,156,184–186]
Blood vessels cells (myocytes) origin	Matrix Metallo-Proteinase 9 (MMP-9)	Associated with blood–brain barrier (BBB) dysfunction and second stroke indication	Diagnosis and recovery: second stroke prevention	[187–191]
General inflammatory cytokines and proteins	Interleukin-6 (IL-6), interleukin-1b (IL-1b), tumor necrosis factor- $\alpha$ (TNF- $\alpha$ )	Associated with CEI ischemic stroke subtype	Prognosis: subtype classification	[166,192–199]
	Neutrophil Lymphocyte Ratios (NLR)	Associated with increased mortality and LAA ischemic stroke subtype	Prognosis: subtype classification and recovery	[199–206]
	Neurofilaments (NF)	Associated with disconnected axons	Prognosis	[207–209]
Cytoskeleton proteins	Cleaved-tau (C-tau)	Associated with neuronal degeneration and disease progression	Prognosis	[157,158,172,210–213]
Cytosicion proteins	Microtubule-associated protein 2 (MAP2)	Associated with dendritic injury	Prognosis	[214–217]
	Alpha-II spectrin break-down products (SBDPs)	Associated with apoptosis and necrotic neuronal death	Prognosis	[157,218–221]

Biosensors **2017**, 7, 30

 Table 4. Cont.

Family	Biomarker	Expression Association	Stroke Clinical Value	Reference
	D-dimer (DD)	Associated with CEI ischemic stroke subtype and recurrence strokes	Prognosis: subtype classification	[11,222–240]
	C-reactive protein (CRP)	Associated with AIS diagnosis, stroke severity and LAA ischemic stroke subtype	Prognosis: subtype classification	[11,241-245]
	Fibrin monomer complex (FMC)	Associated with stroke early recognition and CEI ischemic stroke subtype	Diagnosis and prognosis: subtype classification	[246]
Hemostatic proteins	Soluble fibrin (SF)	Associated with stroke early recognition and CEI ischemic stroke subtype	Diagnosis and prognosis: subtype classification	[246]
	Fibrinogen	Associated with stroke early recognition	Diagnosis	[246,247]
	Fibrin/fibrinogen degradation products (FDPs)	Associated with stroke early recognition	Diagnosis	[246]
	Von willebrand factor (vWF)	Associated with stroke early recognition and LAC ischemic stroke subtype	Diagnosis and prognosis: subtype classification	[247]
	Triglycerides	Associated with LAA ischemic stroke subtype	Prognosis: subtype classification	[248–250]
	Low density lipoprotein (LDL)/High density lipoprotein (HDL)	Associated with LAA ischemic stroke subtype	Prognosis: subtype classification	[248–250]
	Heart fatty acid binding protein (H-FABP)	Associated with early diagnosis of stroke	Diagnosis	[251,252]
Lipids origin	Free fatty acid (FFA)	Associated with CEI ischemic stroke subtype	Prognosis: subtype classification	[253]
	ApoA	Associated with LAA ischemic stroke subtype, and stroke severity	Prognosis: subtype classification and outcome prediction	[254,255]
	ApoE4	Associated with ischemic stroke diagnosis (vs. hemorrhagic stroke) and with LAC and LAA ischemic stroke subtypes	Diagnosis and prognosis: subtype classification	[249,256]
Metabolic proteins	lactate dehydrogenase (LD)	Associated with the extent and severity of the brain damage and the recovery potential	Prognosis and recovery	[182,257]
	Albumin	Associated with CEI ischemic stroke subtype	Prognosis: subtype classification	[258]
Others	Decorin	Associated with LAA ischemic stroke subtype	Prognosis: subtype classification	[259]

Biosensors 2017, 7, 30 22 of 39

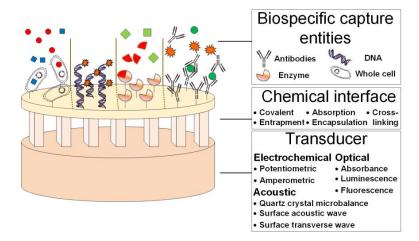
#### 4.3. Multiplex and Quantitative Detection

The use of a multi biomarker panel strategy, instead of the measurement of a single biomarker, is probably the more useful approach. However, it is still mostly research based and of unproven value. While the results for certain biomarkers fulfil certain clinical requirements, there are currently no available biomarkers that can be recommended for immediate use in clinical settings. Multi biomarker studies will make effective panels in clinical settings [166], increasing diagnostic accuracy by minimizing the cross-effect of any individual biomarker [157]. The majority of the brain biomarkers tested so far for stroke also show association to other medical conditions. Hence, a multi-biomarker study would be especially significant [11,189]. In addition, there is a need for a quantitative biomarker detection for a useful clinical value. The use of biomarker measurement as a diagnostic value also requires a specific cut-off value which will allow an improved decision-making process in clinical practice. In order to develop such a multiplex and quantitative POCT for stroke biomarker measurement panel, there is a need to engineer novel POCT platforms.

#### 4.4. POCT-based Sensors

POCT represents an on-site tool that expedites and targets specific diagnostic delays. However, not all POCT devices demonstrate the full features required for a diagnostic device. A POCT is not defined by any particular technology or method of use, for example, it does not require reagent-free operation, battery-powered operation, or a specific degree of operator training [260]. However a biosensor as a POCT will be a more accurate approach for a novel-stroke POCT. There is a variety of POCT-based sensor analytical formats, such as microfluidics [261], microarrays [262], paper-based immunoassays [263-265] and optical-based sensors [266-268]. Biosensors represent a rapidly expanding field and have been widely used in drug discovery, diagnosis, biomedicine, food safety and processing, environmental monitoring, defense, and security applications. A biosensor is usually described as a self-containeddevice, capable of providing selective quantitative or semi-quantitative analytical information and which uses a biological recognition element and a transducer placed in intimate contact via some form of chemical immobilization [268–270]. A typical biosensor is based on three different parts: the biospecific capture entity-biological detection of the target molecule; the chemical interface-controls the main function of the system; and transducer – signal detection and measurement, see Figure 3. The biospecific capture entity (e.g., whole cells, enzymes, DNA or RNA strands, antibodies, antigens or biomimetic molecules) is chosen according to the target analyte, while the interfacial chemistry ensures that the biospecific capture entity molecule is immobilized upon the relevant transducer. A successful POCT sensor must possess at least three critical conditions after the immobilization steps: (1) maintaining stability and activity of the biological part during and after functionalization; (2) maximize proximity of the biological layer to the transducer; (3) maintaining sensitivity and specificity of the biological components to a target analyte [271]. Adsorption, cross-linking, covalent binding, entrapment, and less useful Langmuir-Blodgett (LMB) deposition, self-assembled monolayers, bulk modifications are the most reported methods used in biosensor functionalization applications [271,272]. The transducer, which converts the molecular recognition event to a measurable signal, may be electrochemical, acoustic or optical origin etc. [273]. An ongoing trend in the area of biosensor devices is the development of healthcare diagnostics tools. While recent advances in these technologies relate to the integration of microfluidics and optics, the miniaturization of devices and communication, and the advent of simplified fabrication technologies will allow the creation of commercial applications in the future [274].

Biosensors 2017, 7, 30 23 of 39



**Figure 3.** Biosensor Structure. A Biosensor consists of three components: biospecific capture entity, chemical interface and transducer. A variety of biospecific capture entities are used for the bio-detection of the target analyte in biosensors, such as antibodies, enzymes, DNA and whole cells. The biosensor chemical interface can be based on either a covalent, entrapment, absorption, encapsulation or cross-linking binding. In addition, the transducer, which is used for the signal transmission and measurement, can be based on electrochemical signal (potentiometric or amperometric), optical signal (absorbance, luminescence or fluorescence) or acoustic signal (quartz crystal microbalance, surface acoustic wave or surface transverse wave).

#### 4.5. Novel POCT-based Sensors Platforms

Two novel POCT platforms that originated from the present author's work are the electro-lateral-flow-immunoassay (ELFI) [263,275,276], which enables quantitative detection, and the stack-pad [264,265], which allows multiplex detection, see Figure 4. The ELFI platform consists of a convenient flow test strip, combined with screen-printed gold electrode (SPGE), enabling quantification and a robust set-up. The ELFI bio-recognition is based on novel immune-electroactive nanobeads that become part of a highly sensitive sandwich recognition complex. Lastly, the test results are recorded electrochemically, with a quantitative signal measurement. The electrochemical signal is contributed by a redox label present on the immobilized bead. The second novel POCT platform is the Stack-pad, which is based on stacked membranes, each layer with a specific function. The sample is added onto the bottom-most layer, and as each layer is wetted, the analyte is pushed through to the next layer of the membrane. Through the analyte migration, attached with corresponding antibody conjugated with HRP, a measurable signal is produced. In order to prevent a false positive result, a proprietary blocking layer membrane is added to stop unbounded antibodies from reaching to the top membrane. Thus, only the analyte/antibody-HRP complex will generate a signal. These two novel POCT platforms present a robust set-up which also allows multiplex and quantitative stroke biomarkers detection.

#### 4.6. Use of Data-Mining for Efficient POCT Clinical Integration

As modern research is developing, the use of big-data studies became increasingly common. Incorporating data engineering tools in scientific research helps to achieve better results' analysis and conclusions, and has become mandatory in clinical research, with a powerful impact on the research directions and choices. Big data analytics in healthcare have evolved significantly as an innovative approach [277]. Data mining and knowledge discovery as an approach to examining medical data can limit some of the inherent bias in the hypothesis assumptions that can be found in traditional clinical data analysis [278]. Big data has four characteristics—Volume, Variety, Velocity and Value (the 4 Vs)—which traditional systems are incapable of processing. Medical data sets are continuously becoming larger, thus making it increasingly difficult for traditional systems to process them [279]. Data mining is one of the technologies called to improve the quality of service in clinical medicine

Biosensors 2017, 7, 30 24 of 39

through the intelligent analysis of biomedical information. From the enunciation of evidence-based medicine in early 1990s [280], the need for creating evidence that could be quickly transferred to physician daily practice is one of the most important challenges in medicine. The use of statistics to prove the validity of the treatment over discrete populations; the creation of predictive models for diagnosis, prognosis, and treatment; and the inference of clinical guidelines as decision trees or workflows from instances of healthcare protocols are examples of how data mining can help in the application of evidence based medicine. For example, in a recent study [235], N. Dougu et al developed an algorithm combining AF and D-dimer level in order to classify the diagnosis of cerebral infarction (CI). CI patients were classified into LAA, CEI, LAC infarction or others. If the D-dimer level exceeds a certain cutoff point, the patient would be suspected of having cardioembolic disease. However, despite the great penetration of these techniques in literature, their application in real clinical practice is far from complete [281]. In order to make use of the strong clinical potential of a POCT for stroke biomarkers measurement, there is a need to develop a multi biomarker panel, combined with a strong analytical tool, to create a customized useful POCT for improved stroke prognosis care.

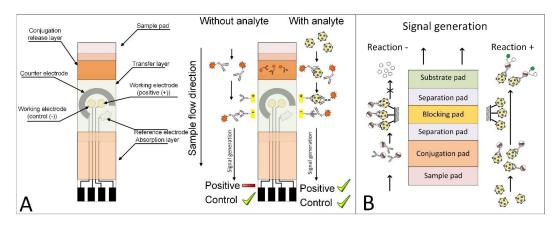


Figure 4. Novel POCT Platforms. (A) Left: Schematic illustration of electro-lateral-flow-immunoassay (ELFI) prototype. This POCT consists of a sample pad, conjugation layer immobilized with specific immune-nanobeads, a transfer layer and a SPGE immobilized with second specific Antibody for the formation of sandwich bio-recognition. Right: ELFI test concept. In case the tested sample contains the target analyte, the formulated immune-nanobeads (AuNPs-Ab-Fc) bind to it and there is a sandwich immune-complex formation on the SPGE. The signal is then measured by electrochemical reaction. In case the tested sample does not contain the target analyte, there will not be a sandwich immune-complex formation on the screen-printed gold electrode (SPGE), which will change the electrochemical signal. (B) Schematic illustration of Stack-Pad prototype. This POCT consists of a sample pad, conjugation pad immobilized with specific HRP-conjugated antibodies, a separation pad, blocking pad immobilized with the target analyte and a substrate pad immobilized with HRP substrate, which later produce a measurable signal. In case the tested sample contains the target analyte, the HRP-conjugated antibodies will bind to it, and continue to flow through the stack, until reaching the last membrane and producing a measurable quantitative signal. In case the tested sample doesn't contain the target analyte, the HRP-conjugated antibodies will bind to the analyte immobilized on the blocking pad, will not continue to flow through the stack and therefore will not produce a signal.

#### 5. Future Trends

The use of stroke prognosis POCTs is only now starting to be integrated into the established clinical practice. However, the most commonly used POCTs are mainly well-known devices for the management of the general ED's patients. However, these POCTs, such as coagulation system parameters and blood chemistry measurements, are not specifically customized to a stroke patient's care. There are only a few innovative POCTs that are still in the research phase and have not been tested in a clinical study, and which show a direction for the development of tailored-made stroke POCT

Biosensors 2017, 7, 30 25 of 39

device. As stroke diagnosis and outcome improvement mostly rely on time, POCT incorporation in stroke prognosis will improve patient's care, at their most critical state. There is a clear need for new stroke POCT devices, specifically aimed at resolving gaps in stroke clinical practice. It is within the means of current scientific knowledge to develop such POCT devices aimed at improving stroke prognosis, with the use of novel on-site immunoassays to detect stroke related biomarkers. Stroke related biomarkers research is a growing field, with new biomarkers being discovered regularly. In order to fully make use of biomarkers measurement for the integration of POCT devices in stroke care, there is a need to make use of modern analytical tools.

**Acknowledgments:** This publication is supported by the National Research Foundation (NRF) of Singapore under the Campus for Research Excellence and Technological Enterprise (CREATE) and the Singapore-MIT Alliance for Research and Technology (SMART), The Institute for Sport Research (ISR) and the Singapore International Graduate Award (SINGA). The authors thank Yeo Loo Pin for correcting the manuscript.

Conflicts of Interest: The authors declare no conflict of interest

#### References

- 1. Strong, K.; Mathers, C.; Bonita, R. Preventing stroke: Saving lives around the world. *Lancet Neurolol.* **2007**, *6*, 182–187. [CrossRef]
- 2. Bhavna, J. Stroke Diagnostics and Therapeutics: Global Markets; BCC research: Wellesley, MA, USA, 2015.
- 3. Goldstein, L.B.; Adams, R.; Alberts, M.J.; Appel, L.J.; Brass, L.M.; Bushnell, C.D.; Culebras, A.; Degraba, T.J.; Gorelick, P.B.; Guyton, J.R.; et al. Primary prevention of ischemic stroke: A guideline from the american heart association/american stroke association stroke council: Cosponsored by the atherosclerotic peripheral vascular disease interdisciplinary working group; cardiovascular nursing council; clinical cardiology council; nutrition, physical activity, and metabolism council; and the quality of care and outcomes research interdisciplinary working group: The American academy of neurology affirms the value of this guideline. *Stroke* 2006, 37, 1583–1633. [PubMed]
- 4. Allender, S.; Scarborough, P.; Peto, V.; Rayner, M.; Leal, J.; Luengo-Fernandez, R.; Gray, A. *European Cardiovascular Disease Statistics*; European Heart Network: Brussels, UK, 2008.
- 5. Charles, P.W.; Jan, V.G.; Martin, S.D.; Joanna, M.W.; John, M.B.; Graeme, J.H.; Peter, A.G.S.; Gabriel, R.; Peter, L.; Cathie, S.; et al. *Stroke: Practical Management*, 3rd ed.; Wiley-Blackwell: Tokyo, Japan, 2008.
- 6. Great Britain: National Audit Office. *Reducing brain damage: Faster access to better stroke care;* National Audit Office: London, UK, 2005.
- 7. Kidwell, C.S.; Chalela, J.A.; Saver, J.L.; Starkman, S.; Hill, M.D.; Demchuk, A.M.; Butman, J.A.; Patronas, N.; Alger, J.R.; Latour, L.L.; et al. Comparison of mri and ct for detection of acute intracerebral hemorrhage. *Jama* **2004**, 292, 1823–1830. [CrossRef] [PubMed]
- 8. Singh, T.P.; Weinstein, J.R.; Murphy, S.P. Stroke: Basic and clinical. *Adv. Neurobiol.* **2017**, *15*, 281–293. [PubMed]
- 9. Patel, R.A.G.; White, C.J. Stroke treatment and prevention. *Prog. Cardiovasc. Dis.* **2017**, 59, 525–526. [CrossRef] [PubMed]
- 10. Tsivgoulis, G.; Kargiotis, O.; Alexandrov, A.V. Intravenous thrombolysis for acute ischemic stroke: A bridge between two centuries. *Expert Rev. Neurother.* **2017**, *17*, 819–837. [CrossRef] [PubMed]
- 11. Glickman, S.W.; Phillips, S.; Anstrom, K.J.; Laskowitz, D.T.; Cairns, C.B. Discriminative capacity of biomarkers for acute stroke in the emergency department. *J. Emerg. Med.* **2011**, *41*, 333–339. [CrossRef] [PubMed]
- Caplan, L.R. Tissue plasminogen activator for acute ischemic stroke. N. Engl. J. Med. 1995, 333, 1581–1587.
- 13. Adams, H.P., Jr.; del Zoppo, G.; Alberts, M.J.; Bhatt, D.L.; Brass, L.; Furlan, A.; Grubb, R.L.; Higashida, R.T.; Jauch, E.C.; Kidwell, C.; et al. Guidelines for the early management of adults with ischemic stroke: A guideline from the american heart association/american stroke association stroke council, clinical cardiology council, cardiovascular radiology and intervention council, and the atherosclerotic peripheral vascular disease and quality of care outcomes in research interdisciplinary working groups: The american academy of neurology affirms the value of this guideline as an educational tool for neurologists. *Circulation* 2007, 115, e478–e534. [PubMed]

Biosensors 2017, 7, 30 26 of 39

14. Ng, G.J.; Quek, A.M.; Cheung, C.; Arumugam, T.V.; Seet, R.C. Stroke biomarkers in clinical practice: A critical appraisal. *Neurochem. Int.* **2017**, 107, 11–22. [CrossRef] [PubMed]

- 15. Siegel, J.; Pizzi, M.A.; Brent Peel, J.; Alejos, D.; Mbabuike, N.; Brown, B.L.; Hodge, D.; David Freeman, W. Update on neurocritical care of stroke. *Curr. Cardiol. Reports* **2017**, *19*, 67. [CrossRef] [PubMed]
- 16. Khaku, A.D. Stroke; Statpearls: Treasure Island, FL, USA, 2017.
- 17. Audebert, H.J.; Saver, J.L.; Starkman, S.; Lees, K.R.; Endres, M. Prehospital stroke care: New prospects for treatment and clinical research. *Neurology* **2013**, *81*, 501–508. [CrossRef] [PubMed]
- 18. Lees, K.R.; Bluhmki, E.; von Kummer, R.; Brott, T.G.; Toni, D.; Grotta, J.C.; Albers, G.W.; Kaste, M.; Marler, J.R.; Hamilton, S.A.; et al. Time to treatment with intravenous alteplase and outcome in stroke: An updated pooled analysis of ecass, atlantis, ninds, and epithet trials. *Lancet* **2010**, *375*, 1695–1703. [CrossRef]
- 19. Hacke, W.; Kaste, M.; Bluhmki, E.; Brozman, M.; Davalos, A.; Guidetti, D.; Larrue, V.; Lees, K.R.; Medeghri, Z.; Machnig, T.; et al. Thrombolysis with alteplase 3 to 4.5 hours after acute ischemic stroke. *N. Engl. J. Med.* **2008**, 359, 1317–1329. [CrossRef] [PubMed]
- Sandercock, P.; Wardlaw, J.M.; Lindley, R.I.; Dennis, M.; Cohen, G.; Murray, G.; Innes, K.; Venables, G.; Czlonkowska, A.; Kobayashi, A.; et al. The benefits and harms of intravenous thrombolysis with recombinant tissue plasminogen activator within 6 h of acute ischaemic stroke (the third international stroke trial [ist-3]): A randomised controlled trial. *Lancet* 2012, 379, 2352–2363. [PubMed]
- 21. Leys, D.; Ringelstein, E.B.; Kaste, M.; Hacke, W. Facilities available in european hospitals treating stroke patients. *Stroke*; *J. Cereb. Circ.* **2007**, *38*, 2985–2991. [CrossRef] [PubMed]
- 22. Nolte, C.H.; Audebert, H.J. [management of acute ischemic stroke]. *Dtsch. Med. Wochenschr.* **2015**, 140, 1583–1586. [PubMed]
- 23. Evenson, K.R.; Rosamond, W.D.; Morris, D.L. Prehospital and in-hospital delays in acute stroke care. *Neuroepidemiology* **2001**, *20*, 65–76. [CrossRef] [PubMed]
- 24. Tai, Y.J.; Yan, B. Minimising time to treatment: Targeted strategies to minimise time to thrombolysis for acute ischaemic stroke. *Intern. Med. J.* **2013**, *43*, 1176–1182. [CrossRef] [PubMed]
- 25. Bustamante, A.; Garcia-Berrocoso, T.; Rodriguez, N.; Llombart, V.; Ribo, M.; Molina, C.; Montaner, J. Ischemic stroke outcome: A review of the influence of post-stroke complications within the different scenarios of stroke care. *Eur. J. Intern. Med.* **2016**, *29*, 9–21. [CrossRef] [PubMed]
- 26. McMullan, J.T.; Knight, W.A.; Clark, J.F.; Beyette, F.R.; Pancioli, A. Time-critical neurological emergencies: The unfulfilled role for point-of-care testing. *Intern. J. Emerg. Med.* **2010**, *3*, 127–131. [CrossRef] [PubMed]
- 27. Rooney, K.D.; Schilling, U.M. Point-of-care testing in the overcrowded emergency department—Can it make a difference? *Crit. Care* **2014**, *18*, 692. [CrossRef] [PubMed]
- 28. Cummins, B.M.; Ligler, F.S.; Walker, G.M. Point-of-care diagnostics for niche applications. *Biotech. Adv.* **2016**, 34, 161–176. [CrossRef] [PubMed]
- 29. Vasan, A.S.; Mahadeo, D.M.; Doraiswami, R.; Huang, Y.; Pecht, M. Point-of-care biosensor system. *Front. Biosci.* **2013**, *5*, 39–71. [CrossRef]
- 30. Chalela, J.A.; Kidwell, C.S.; Nentwich, L.M.; Luby, M.; Butman, J.A.; Demchuk, A.M.; Hill, M.D.; Patronas, N.; Latour, L.; Warach, S. Magnetic resonance imaging and computed tomography in emergency assessment of patients with suspected acute stroke: A prospective comparison. *Lancet* **2007**, *369*, 293–298. [CrossRef]
- 31. Dzialowski, I.; Hill, M.D.; Coutts, S.B.; Demchuk, A.M.; Kent, D.M.; Wunderlich, O.; von Kummer, R. Extent of early ischemic changes on computed tomography (CT) before thrombolysis. Prognostic Value of the Alberta Stroke Program Early CT Score in ECASS II. *Stroke* 2006, *37*, 973–978. [CrossRef] [PubMed]
- 32. Dunbabin, D.W.; Sandercock, P.A. Investigation of acute stroke: What is the most effective strategy? *Postgrad. Med. J.* **1991**, *67*, 259–270. [CrossRef] [PubMed]
- 33. Cramer, S.C. Functional imaging in stroke recovery. Stroke 2004, 35, 2695–2698. [CrossRef] [PubMed]
- 34. Schellinger, P.D.; Chalela, J.A.; Kang, D.W.; Latour, L.L.; Warach, S. Diagnostic and prognostic value of early mr imaging vessel signs in hyperacute stroke patients imaged <3 hours and treated with recombinant tissue plasminogen activator. *Am. J. Neuroradiol.* **2005**, *26*, 618–624. [PubMed]
- 35. Schellinger, P.D.; Jansen, O.; Fiebach, J.B.; Hacke, W.; Sartor, K. A standardized mri stroke protocol. Comparison with CT in Hyperacute Intracerebral Hemorrhage. *Stroke* 1999, 30, 765–768. [CrossRef] [PubMed]
- 36. Caplan, L.R. Stroke: A Clinical Approach, 4th ed.; Saunders Elsevier: Philadelphia, PA, USA, 2009.
- 37. Jiang, Q.; Zhang, Z.G.; Chopp, M. Mri of stroke recovery. Stroke 2010, 41, 410–414. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 27 of 39

38. Lee, L.J.; Kidwell, C.S.; Alger, J.; Starkman, S.; Saver, J.L. Impact on stroke subtype diagnosis of early diffusion-weighted magnetic resonance imaging and magnetic resonance angiography. *Stroke* **2000**, *31*, 1081–1089. [CrossRef] [PubMed]

- 39. Chollet, F.; Albucher, J.F. Strategies to augment recovery after stroke. *Curr. Treat. Options Neurol.* **2012**, *14*, 531–540. [CrossRef] [PubMed]
- 40. Heller, S.L.; Heier, L.A.; Watts, R.; Schwartz, T.H.; Zelenko, N.; Doyle, W.; Devinsky, O. Evidence of cerebral reorganization following perinatal stroke demonstrated with fmri and dti tractography. *Clin. Imag.* **2005**, 29, 283–287. [CrossRef] [PubMed]
- 41. Kim, Y.H.; You, S.H.; Kwon, Y.H.; Hallett, M.; Kim, J.H.; Jang, S.H. Longitudinal fmri study for locomotor recovery in patients with stroke. *Neurology* **2006**, *67*, 330–333. [CrossRef] [PubMed]
- 42. Kim, Y.-H.; Jang, S.H.; Chang, Y.; Byun, W.M.; Son, S.; Ahn, S.H. Bilateral primary sensori-motor cortex activation of post-stroke mirror movements: An fMRI study. *Neuroreport* **2003**, *14*, 1329–1332. [CrossRef] [PubMed]
- 43. Eaton, K.P.; Szaflarski, J.P.; Altaye, M.; Ball, A.L.; Kissela, B.M.; Banks, C.; Holland, S.K. Reliability of fMRI for studies of language in post-stroke aphasia subjects. *Neuroimage* **2008**, *41*, 311–322. [CrossRef] [PubMed]
- 44. O'Sullivan, M.; Morris, R.G.; Huckstep, B.; Jones, D.K.; Williams, S.C.; Markus, H.S. Diffusion tensor MRI correlates with executive dysfunction in patients with ischaemic leukoaraiosis. *J. Neurol. Neurosurg. Psychiatry* 2004, 75, 441–447. [CrossRef] [PubMed]
- 45. Jiang, Q.; Zhang, Z.G.; Chopp, M. Mri evaluation of white matter recovery after brain injury. *Stroke* **2010**, *41*, S112–S113. [CrossRef] [PubMed]
- Kunimatsu, A.; Aoki, S.; Masutani, Y.; Abe, O.; Mori, H.; Ohtomo, K. Three-dimensional white matter tractography by diffusion tensor imaging in ischaemic stroke involving the corticospinal tract. *Neuroradiology* 2003, 45, 532–535. [CrossRef] [PubMed]
- 47. Moore, W.S.; Barnett, H.J.; Beebe, H.G.; Bernstein, E.F.; Brener, B.J.; Brott, T.; Caplan, L.R.; Day, A.; Goldstone, J.; Hobson, R.W.; et al. Guidelines for carotid endarterectomy. A multidisciplinary consensus statement from the ad hoc committee, american heart association. *Stroke* 1995, 26, 188–201. [CrossRef] [PubMed]
- 48. Barnett, H.J.M. *Stroke Pathophysiology, Diagnosis, and Management*, 2nd ed.; Churchill Livingstone: New York, NY, USA, 1992.
- 49. American Heart & Stroke Association. Available online: https://www.heart.org/HEARTORG/General/Heart-and-Stroke-AssociationStatistics\_UCM\_319064\_SubHomePage.jsp (accessed on 1 April 2017).
- 50. Govindarajan, P.; Ghilarducci, D.; McCulloch, C.; Pierog, J.; Bloom, E.; Johnston, C. Comparative evaluation of stroke triage algorithms for emergency medical dispatchers (meds): Prospective cohort study protocol. *BMC Neurol.* **2011**, *11*, 14. [CrossRef] [PubMed]
- 51. Wojner-Alexandrov, A.W.; Alexandrov, A.V.; Rodriguez, D.; Persse, D.; Grotta, J.C. Houston paramedic and emergency stroke treatment and outcomes study (hopsto). *Stroke* **2005**, *36*, 1512–1518. [CrossRef] [PubMed]
- 52. Zhai, S.; Gardiner, F.; Neeman, T.; Jones, B.; Gawarikar, Y. The cost-effectiveness of a stroke unit in providing enhanced patient outcomes in an australian teaching hospital. Available online: http://www.sciencedirect.com/science/article/pii/S1052305717302550 (accessed on 3 August 2017).
- 53. Miller, E.C.; Blum, C.; Rostanski, S.K. Developing a stroke center. *Stroke* **2017**, *48*, e155–e156. [CrossRef] [PubMed]
- 54. Watkins, C.L.; Leathley, M.J.; Jones, S.P.; Ford, G.A.; Quinn, T.; Sutton, C.J. Training emergency services' dispatchers to recognise stroke: An interrupted time-series analysis. *BMC Health. Serv. Res.* **2013**, *13*, 318. [CrossRef] [PubMed]
- 55. Deng, Y.Z.; Reeves, M.J.; Jacobs, B.S.; Birbeck, G.L.; Kothari, R.U.; Hickenbottom, S.L.; Mullard, A.J.; Wehner, S.; Maddox, K.; Majid, A. Iv tissue plasminogen activator use in acute stroke: Experience from a statewide registry. *Neurology* **2006**, *66*, 306–312. [CrossRef] [PubMed]
- 56. Lebrun, L.; Rusterholtz, T.; Fery-Lemonnier, E.; Woimant, F.; Leroyer, J.; Hommel, M. Improving stroke care: a French health-care organiser's perspective. *Intern. J. Stroke.* **2011**, *6*, 123–124. [CrossRef] [PubMed]
- 57. Katz, B.S.; Adeoye, O.; Sucharew, H.; Broderick, J.P.; McMullan, J.; Khatri, P.; Widener, M.; Alwell, K.S.; Moomaw, C.J.; Kissela, B.M.; et al. Estimated impact of emergency medical service triage of stroke patients on comprehensive stroke centers: An urban population-based study. *Stroke* **2017**, *48*, 2164–2170. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 28 of 39

58. Waqar Faiz, K.; Sundseth, A.; Thommessen, B.; Ronning, O.M. Prehospital path in acute stroke. *Tidsskr. Nor.* **2017**, 137, 798–802. [CrossRef] [PubMed]

- 59. Brott, T.; Adams, H.P., Jr.; Olinger, C.P.; Marler, J.R.; Barsan, W.G.; Biller, J.; Spilker, J.; Holleran, R.; Eberle, R.; Hertzberg, V.; et al. Measurements of acute cerebral infarction: A clinical examination scale. *Stroke* **1989**, 20, 864–870. [CrossRef] [PubMed]
- 60. Assis, Z.A.; Menon, B.K.; Goyal, M. Imaging department organization in a stroke center and workflow processes in acute stroke. Available online: http://www.sciencedirect.com/science/article/pii/S0720048X17302607 (accessed on 3 August 2017).
- 61. Ferro, J.M.; Massaro, A.R.; Mas, J.L. Aetiological diagnosis of ischaemic stroke in young adults. *Lancet. Neurol.* **2010**, *9*, 1085–1096. [CrossRef]
- 62. Thomas, S.M.; Thomas, S.R. Stroke and Transient Ischemic Attack. Available online: https://scholar.google.com/scholar?q=Thomas%2C+S.%3B+Maldonado%2C+M.D.%3B+Thomas%2C+S.%3B+Riles%2C+M.D.%3B+F.A.C.S.+Stroke+and+transient+ischemic+attack-1+1+stroke+and+transient+ischemic+attack.+Vasc.+Syst.+2007&btnG=&hl=en&as\_sdt=0%2C5 (accessed on 3 August 2017).
- 63. Go, A.S.; Mozaffarian, D.; Roger, V.L.; Benjamin, E.J.; Berry, J.D.; Borden, W.B.; Bravata, D.M.; Dai, S.; Ford, E.S.; Fox, C.S.; et al. Heart disease and stroke statistics–2013 update: A report from the american heart association. *Circulation* **2013**, 127, e6–e245. [CrossRef] [PubMed]
- 64. Radu, R.A.; Terecoasa, E.O.; Bajenaru, O.A.; Tiu, C. Etiologic classification of ischemic stroke: Where do we stand? *Clin. Neurol. Neurosurg.* **2017**, *159*, 93–106. [CrossRef] [PubMed]
- 65. Amarenco, P.; Bogousslavsky, J.; Caplan, L.R.; Donnan, G.A.; Hennerici, M.G. Classification of stroke subtypes. *Cerebrovasc. Dis.* **2009**, 27, 493–501. [CrossRef] [PubMed]
- 66. Adams, H.P., Jr.; Bendixen, B.H.; Kappelle, L.J.; Biller, J.; Love, B.B.; Gordon, D.L.; Marsh, E.E. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial. Toast. Trial of org 10172 in acute stroke treatment. *Stroke* 1993, 24, 35–41. [CrossRef] [PubMed]
- 67. Landau, W.M.; Nassief, A. Editorial comment-time to burn the toast. Stroke 2005, 36, 902-904. [PubMed]
- 68. Amarenco, P. Patent foramen ovale and the risk of stroke: Smoking gun guilty by association? *Heart* **2005**, *91*, 441–443. [CrossRef] [PubMed]
- 69. Gokcal, E.; Niftaliyev, E.; Asil, T. Etiological classification of ischemic stroke in young patients: A comparative study of TOAST, CCS, and ASCO. *Acta. Neurologica. Belgica.* **2017**, 1–6.
- 70. Sacco, R.L.; Ellenberg, J.H.; Mohr, J.P.; Tatemichi, T.K.; Hier, D.B.; Price, T.R.; Wolf, P.A. Infarcts of undetermined cause: The nincds stroke data bank. *Ann. Neurol.* **1989**, 25, 382–390. [CrossRef] [PubMed]
- 71. Bamford, J.; Sandercock, P.; Dennis, M.; Burn, J.; Warlow, C. Classification and natural history of clinically identifiable subtypes of cerebral infarction. *Lancet* **1991**, *337*, 1521–1526. [CrossRef]
- 72. Lindley, R.I.; Warlow, C.P.; Wardlaw, J.M.; Dennis, M.S.; Slattery, J.; Sandercock, P.A. Interobserver reliability of a clinical classification of acute cerebral infarction. *Stroke* **1993**, 24, 1801–1804. [CrossRef] [PubMed]
- 73. Teo, K.; Slark, J. A systematic review of studies investigating the care of stroke survivors in long-term care facilities. *Disabil. Rehabil.* **2016**, *38*, 715–723. [CrossRef] [PubMed]
- 74. Price, C.P. Point of care testing. BMJ 2001, 322, 1285–1288. [CrossRef] [PubMed]
- 75. St John, A.; Price, C.P. Existing and emerging technologies for point-of-care testing. *Clin. Biochem. Rev.* **2014**, 35, 155–167. [PubMed]
- 76. Yoo, E.H.; Lee, S.Y. Glucose biosensors: An overview of use in clinical practice. *Sensors* **2010**, *10*, 4558–4576. [CrossRef] [PubMed]
- 77. Shah, D.; Maghsoudlou, D. Enzyme-linked immunosorbent assay (elisa). *Basics Br. J. Hospital Med.* **2016**, 77, C98–C101. [CrossRef] [PubMed]
- 78. Weber, J.E.; Ebinger, M.; Rozanski, M.; Waldschmidt, C.; Wendt, M.; Winter, B.; Kellner, P.; Baumann, A.; Fiebach, J.B.; Villringer, K.; et al. Prehospital thrombolysis in acute stroke: Results of the phantom-s pilot study. *Neurology* **2013**, *80*, 163–168. [CrossRef] [PubMed]
- 79. Handschu, R.; Littmann, R.; Reulbach, U.; Gaul, C.; Heckmann, J.G.; Neundorfer, B.; Scibor, M. Telemedicine in emergency evaluation of acute stroke: Interrater agreement in remote video examination with a novel multimedia system. *Stroke* **2003**, *34*, 2842–2846. [CrossRef] [PubMed]
- 80. Demaerschalk, B.M.; Vargas, J.E.; Channer, D.D.; Noble, B.N.; Kiernan, T.E.; Gleason, E.A.; Vargas, B.B.; Ingall, T.J.; Aguilar, M.I.; Dodick, D.W.; et al. Smartphone teleradiology application is successfully incorporated into a telestroke network environment. *Stroke* **2012**, *43*, 3098–3101. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 29 of 39

81. Amadi-Obi, A.; Gilligan, P.; Owens, N.; O'Donnell, C. Telemedicine in pre-hospital care: A review of telemedicine applications in the pre-hospital environment. *Intern. J. Emerg. Med.* **2014**, *7*, 29. [CrossRef] [PubMed]

- 82. Agrawal, K.; Raman, R.; Ernstrom, K.; Claycomb, R.J.; Meyer, D.M.; Hemmen, T.M.; Modir, R.F.; Kachhi, P.; Meyer, B.C. Accuracy of stroke diagnosis in telestroke-guided tissue plasminogen activator patients. *J. Stroke Cerebrovasc. Dis.* **2016**, *25*, 2942–2946. [CrossRef] [PubMed]
- 83. Jeon, S.B.; Ryoo, S.M.; Lee, D.H.; Kwon, S.U.; Jang, S.; Lee, E.J.; Lee, S.H.; Han, J.H.; Yoon, M.J.; Jeong, S.; et al. Multidisciplinary approach to decrease in-hospital delay for stroke thrombolysis. *J. Stroke* **2017**, *19*, 196–204. [CrossRef] [PubMed]
- 84. Vecchiato, A. Can we imagine a survival chain also for ischemic stroke? Available online: http://www.sciencedirect.com/science/article/pii/S0735675717304370 (accessed on 3 August 2017).
- 85. Walter, S.; Kostopoulos, P.; Haass, A.; Keller, I.; Lesmeister, M.; Schlechtriemen, T.; Roth, C.; Papanagiotou, P.; Grunwald, I.; Schumacher, H.; et al. Diagnosis and treatment of patients with stroke in a mobile stroke unit versus in hospital: A randomised controlled trial. *Lancet. Neurol.* **2012**, *11*, 397–404. [CrossRef]
- 86. Fassbender, K.; Walter, S.; Liu, Y.; Muehlhauser, F.; Ragoschke, A.; Kuehl, S.; Mielke, O. "Mobile stroke unit" for hyperacute stroke treatment. *Stroke* **2003**, *34*, e44. [CrossRef] [PubMed]
- 87. Parker, S.A.; Bowry, R.; Wu, T.C.; Noser, E.A.; Jackson, K.; Richardson, L.; Persse, D.; Grotta, J.C. Establishing the first mobile stroke unit in the united states. *Stroke* **2015**, *46*, 1384–1391. [CrossRef] [PubMed]
- 88. Ebinger, M.; Rozanski, M.; Waldschmidt, C.; Weber, J.; Wendt, M.; Winter, B.; Kellner, P.; Baumann, A.M.; Malzahn, U.; Heuschmann, P.U.; et al. Phantom-s: The prehospital acute neurological therapy and optimization of medical care in stroke patients-study. *Int. J. Stroke* 2012, 7, 348–353. [CrossRef] [PubMed]
- 89. Wendt, M.; Ebinger, M.; Kunz, A.; Rozanski, M.; Waldschmidt, C.; Weber, J.E.; Winter, B.; Koch, P.M.; Freitag, E.; Reich, J.; et al. Improved prehospital triage of patients with stroke in a specialized stroke ambulance: Results of the pre-hospital acute neurological therapy and optimization of medical care in stroke study. *Stroke* 2015, 46, 740–745. [CrossRef] [PubMed]
- 90. Gomes, J.A.; Ahrens, C.L.; Hussain, M.S.; Winners, S.; Rasmussen, P.A.; Uchino, K. Prehospital reversal of warfarin-related coagulopathy in intracerebral hemorrhage in a mobile stroke treatment unit. *Stroke* **2015**, *46*, e118–e120. [CrossRef] [PubMed]
- 91. Rizos, T.; Herweh, C.; Jenetzky, E.; Lichy, C.; Ringleb, P.A.; Hacke, W.; Veltkamp, R. Point-of-care international normalized ratio testing accelerates thrombolysis in patients with acute ischemic stroke using oral anticoagulants. *Stroke* **2009**, *40*, 3547–3551. [CrossRef] [PubMed]
- 92. Nusa, D.; Harvey, I.; Almansouri, A.Y.; Wright, S.; Neeman, T.; Ahmad, O.; Hughes, A.R.; Lueck, C.J. Assessment of point-of-care measurement of international normalised ratio using the coaguchek xs plus system in the setting of acute ischaemic stroke. *Intern. Med. J.* 2013, 43, 1205–1209. [CrossRef] [PubMed]
- 93. Green, T.L.; Mansoor, A.; Newcommon, N.; Stephenson, C.; Stewart, E.; Hill, M.D. Reliability of point-of-care testing of inr in acute stroke. *Can. J. Neurol. Sci.* **2008**, *35*, 348–351. [CrossRef] [PubMed]
- 94. Thorne, K.; McNaughton, H.; Weatherall, M. An audit of coagulation screening in patients presenting to the emergency department for potential stroke thrombolysis. *Intern. Med. J.* **2017**, 47, 189–193. [CrossRef] [PubMed]
- 95. Walter, S.; Kostopoulos, P.; Haass, A.; Lesmeister, M.; Grasu, M.; Grunwald, I.; Keller, I.; Helwig, S.; Becker, C.; Geisel, J.; et al. Point-of-care laboratory halves door-to-therapy-decision time in acute stroke. *Ann. Neurol.* **2011**, *69*, 581–586. [CrossRef] [PubMed]
- 96. Briggs, C.; Kunka, S.; Pennaneach, C.; Forbes, L.; Machin, S.J. Performance evaluation of a new compact hematology analyzer, the sysmex poch-100i. *Lab. Hematol.* **2003**, *9*, 225–233. [PubMed]
- 97. Drescher, M.J.; Spence, A.; Rockwell, D.; Staff, I.; Smally, A.J. Point-of-care testing for coagulation studies in a stroke protocol: A time-saving innovation. *Am. J. Emerg. Med.* **2011**, 29, 82–85. [CrossRef] [PubMed]
- 98. Nanduri, S.; Tayal, A.H.; Hegde, G.G.; Shang, J.; Venkat, A. An analysis of discrepancy between point-of-care and central laboratory international normalized ratio testing in ED patients with cerebrovascular disease. *Am. J. Emerg. Med.* **2012**, *30*, 2025–2029. [CrossRef] [PubMed]
- 99. Abbott, Inc. A single, Integrated Point-of-care Testing Solution. Available online: https://www.pointofcare.abbott/int/en/offerings/istat/istat-handheld (accessed on 2 August 2017).

Biosensors 2017, 7, 30 30 of 39

100. Chaudhuri, J.R.; Sharma, V.K.; Mridula, K.R.; Balaraju, B.; Bandaru, V.C. Association of plasma brain natriuretic peptide levels in acute ischemic stroke subtypes and outcome. *J. Stroke Cerebrovasc. Dis.* **2015**, 24, 485–491. [CrossRef] [PubMed]

- 101. Clerico, A.; Franzini, M.; Masotti, S.; Prontera, C.; Passino, C. State of the art of immunoassay methods for b-type natriuretic peptides: An update. *Crit. Rev. Clin. Lab. Sci.* **2015**, 52, 56–69. [CrossRef] [PubMed]
- 102. Kawase, S.; Kowa, H.; Suto, Y.; Fukuda, H.; Kusumi, M.; Nakayasu, H.; Nakashima, K. Plasma brain natriuretic peptide is a marker of prognostic functional outcome in non-cardioembolic infarction. *J. Stroke Cerebrovasc. Dis.* **2015**, 24, 2285–2290. [CrossRef] [PubMed]
- 103. Llombart, V.; Antolin-Fontes, A.; Bustamante, A.; Giralt, D.; Rost, N.S.; Furie, K.; Shibazaki, K.; Biteker, M.; Castillo, J.; Rodriguez-Yanez, M.; et al. B-type natriuretic peptides help in cardioembolic stroke diagnosis: Pooled data meta-analysis. *Stroke* 2015, 46, 1187–1195. [CrossRef] [PubMed]
- 104. Balion, C.; McKelvie, R.; Don-Wauchope, A.C.; Santaguida, P.L.; Oremus, M.; Keshavarz, H.; Hill, S.A.; Booth, R.A.; Ali, U.; Brown, J.A.; et al. B-type natriuretic peptide-guided therapy: A systematic review. *Heart Fail. Rev.* **2014**, *19*, 553–564. [CrossRef] [PubMed]
- 105. Kara, K.; Gronewold, J.; Neumann, T.; Mahabadi, A.A.; Weimar, C.; Lehmann, N.; Berger, K.; Kalsch, H.I.; Bauer, M.; Broecker-Preuss, M.; et al. B-type natriuretic peptide predicts stroke of presumable cardioembolic origin in addition to coronary artery calcification. *Eur. J. Neurol.* **2014**, *21*, 914–921. [CrossRef] [PubMed]
- 106. Maruyama, K.; Shiga, T.; Iijima, M.; Moriya, S.; Mizuno, S.; Toi, S.; Arai, K.; Ashihara, K.; Abe, K.; Uchiyama, S. Brain natriuretic peptide in acute ischemic stroke. *J. Stroke Cerebrovasc. Dis.* **2014**, 23, 967–972. [CrossRef] [PubMed]
- 107. Qihong, G.; Zhixin, W.; Mingfeng, H.; Lianhong, Y.; Wenchong, X. Experiences and the use of bnp poct platform on suspected stroke patients by a chinese emergency department. *Annals Indian Acad. Neurol.* **2014**, 17, 243–244.
- 108. Yang, H.L.; Lin, Y.P.; Long, Y.; Ma, Q.L.; Zhou, C. Predicting cardioembolic stroke with the b-type natriuretic peptide test: A systematic review and meta-analysis. *J. Stroke Cerebrovasc. Dis.* **2014**, 23, 1882–1889. [CrossRef] [PubMed]
- 109. Wu, Z.; Yang, L.; Guo, Q.; He, M.; Li, K. Experiences and the use of bnp poct platform on suspected ischemic stroke patients in the emergency department setting. *Clin. Neurol. Neurosurg.* **2014**, *123*, 199–200. [CrossRef] [PubMed]
- 110. Shibazaki, K.; Kimura, K.; Aoki, J.; Sakai, K.; Saji, N.; Uemura, J. Brain natriuretic peptide level on admission predicts recurrent stroke after discharge in stroke survivors with atrial fibrillation. *Clin. Neurol. Neurosurg.* **2014**, 127, 25–29. [CrossRef] [PubMed]
- 111. Shibazaki, K.; Kimura, K.; Aoki, J.; Sakai, K.; Saji, N.; Uemura, J. Plasma brain natriuretic peptide as a predictive marker of early recurrent stroke in cardioembolic stroke patients. *J. Stroke Cerebrovasc. Dis.* **2014**, 23, 2635–2640. [CrossRef] [PubMed]
- 112. Cojocaru, I.M.; Cojocaru, M.; Sapira, V.; Ionescu, A.; Barlan, S.; Tacu, N. Could pro-bnp, uric acid, bilirubin, albumin and transferrin be used in making the distinction between stroke subtypes? *Rom. J. Intern. Med.* **2013**, *51*, 188–195. [CrossRef] [PubMed]
- 113. Hajsadeghi, S.; Kashani Amin, L.; Bakhshandeh, H.; Rohani, M.; Azizian, A.R.; Jafarian Kerman, S.R. The diagnostic value of n-terminal pro-brain natriuretic peptide in differentiating cardioembolic ischemic stroke. *J. Stroke Cerebrovasc. Dis.* **2013**, 22, 554–560. [CrossRef] [PubMed]
- 114. Sakai, K.; Shibazaki, K.; Kimura, K.; Aoki, J.; Kobayashi, K.; Fujii, S.; Okada, Y. Brain natriuretic peptide as a predictor of cardioembolism in acute ischemic stroke patients: Brain natriuretic peptide stroke prospective study. *Eur. Neurol.* **2013**, *69*, 246–251. [CrossRef] [PubMed]
- 115. Garcia-Berrocoso, T.; Giralt, D.; Bustamante, A.; Etgen, T.; Jensen, J.K.; Sharma, J.C.; Shibazaki, K.; Saritas, A.; Chen, X.; Whiteley, W.N.; et al. B-type natriuretic peptides and mortality after stroke: A systematic review and meta-analysis. *Neurology* **2013**, *81*, 1976–1985. [CrossRef] [PubMed]
- 116. Jickling, G.C.; Foerch, C. Predicting stroke mortality: Bnp could it be? *Neurology* **2013**, *81*, 1970–1971. [CrossRef] [PubMed]
- 117. Shibazaki, K.; Kimura, K.; Sakai, K.; Fujii, S.; Aoki, J.; Saji, N. Brain natriuretic peptide on admission as a biological marker of long-term mortality in ischemic stroke survivors. *Eur. Neurol.* **2013**, 70, 218–224. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 31 of 39

118. Chen, X.; Zhan, X.; Chen, M.; Lei, H.; Wang, Y.; Wei, D.; Jiang, X. The prognostic value of combined nt-pro-bnp levels and nihss scores in patients with acute ischemic stroke. *Intern. Med.* **2012**, *51*, 2887–2892. [CrossRef] [PubMed]

- 119. Montaner, J.; Garcia-Berrocoso, T.; Mendioroz, M.; Palacios, M.; Perea-Gainza, M.; Delgado, P.; Rosell, A.; Slevin, M.; Ribo, M.; Molina, C.A.; et al. Brain natriuretic peptide is associated with worsening and mortality in acute stroke patients but adds no prognostic value to clinical predictors of outcome. *Cerebrovasc. Dis.* **2012**, 34, 240–245. [CrossRef] [PubMed]
- 120. Shibazaki, K.; Kimura, K.; Iguchi, Y.; Okada, Y.; Inoue, T. Plasma brain natriuretic peptide can be a biological marker to distinguish cardioembolic stroke from other stroke types in acute ischemic stroke. *Intern. Med.* **2009**, *48*, 259–264. [CrossRef] [PubMed]
- 121. Shibazaki, K.; Kimura, K.; Okada, Y.; Iguchi, Y.; Uemura, J.; Terasawa, Y.; Aoki, J. Plasma brain natriuretic peptide as an independent predictor of in-hospital mortality after acute ischemic stroke. *Intern. Med.* **2009**, 48, 1601–1606. [CrossRef] [PubMed]
- 122. Jensen, J.K.; Atar, D.; Kristensen, S.R.; Mickley, H.; Januzzi, J.L., Jr. Usefulness of natriuretic peptide testing for long-term risk assessment following acute ischemic stroke. *Am. J. Cardiol.* **2009**, *104*, 287–291. [CrossRef] [PubMed]
- 123. United States Government; Development of stroke point of care immunoassay for cellular fibronectin. Available online: https://www.sbir.gov/sbirsearch/detail/280110 (accessed on 3 August 2017).
- 124. Mäkikallio, A.M.; Mäkikallio, T.H.; Korpelainen, J.T.; Vuolteenaho, O.; Tapanainen, J.M.; Ylitalo, K.; Sotaniemi, K.A.; Huikuri, H.V.; Myllylä, V.V. Natriuretic peptides and mortality after stroke. *Stroke* **2005**, *36*, 1016–1020. [CrossRef] [PubMed]
- 125. He, M.; Wu, Z.; Li, Y.; Lei, J. A new algorithm of suspected stroke patient management with brain natriuretic peptide/n-terminal pro-brain natriuretic peptide point of care testing platform in the emergency department. *Annals Indian Acad. Neurol.* **2017**, *20*, 81–82.
- 126. Cohen, R.; Lata, J.P.; Lee, Y.; Hernández, J.C.C.; Nishimura, N.; Schaffer, C.B.; Mukai, C.; Nelson, J.L.; Brangman, S.A.; Agrawal, Y.; et al. Use of tethered enzymes as a platform technology for rapid analyte detection. Available online: http://journals.plos.org/plosone/article?id=10.1371/journal.pone.0142326. (accessed on 3 August 2017).
- 127. Dash, P.K.; Zhao, J.; Hergenroeder, G.; Moore, A.N. Biomarkers for the diagnosis, prognosis, and evaluation of treatment efficacy for traumatic brain injury. *Neurotherapeutics* **2010**, 7, 100–114. [CrossRef] [PubMed]
- 128. Ahmad, O.; Wardlaw, J.; Whiteley, W.N. Correlation of levels of neuronal and glial markers with radiological measures of infarct volume in ischaemic stroke: A systematic review. *Cerebrovasc. Dis.* **2012**, *33*, 47–54. [CrossRef] [PubMed]
- 129. Wunderlich, M.T.; Lins, H.; Skalej, M.; Wallesch, C.W.; Goertler, M. Neuron-specific enolase and tau protein as neurobiochemical markers of neuronal damage are related to early clinical course and long-term outcome in acute ischemic stroke. *Clin. Neurol. Neurosurg.* **2006**, *108*, 558–563. [CrossRef] [PubMed]
- 130. Wevers, R.A.; Jacobs, A.A.; Hommes, O.R. A bioluminescent assay for enolase (ec 4.2.1.11) activity in human serum and cerebrospinal fluid. *Clin. Chim. Acta.* **1983**, *135*, 159–168. [CrossRef]
- 131. Viallard, J.L.; Murthy, M.R.; Dastugue, B. An ultramicro bioluminescence assay of enolase: Application to human cerebrospinal fluid. *Neurochem. Res.* **1985**, *10*, 1555–1566. [CrossRef] [PubMed]
- 132. Wevers, R.A.; Theunisse, A.W.; Rijksen, G. An immunobioluminescence assay for gamma-gamma enolase activity in human serum and cerebrospinal fluid. *Clin. Chim. Acta.* **1988**, *178*, 141–150. [CrossRef]
- 133. Valtari Bio™ Inc. Overview. Available online: http://valtaribio.com/executive-overview/ (accessed on 1 August 2017).
- 134. Karlinski, M.; Gluszkiewicz, M.; Czlonkowska, A. The accuracy of prehospital diagnosis of acute cerebrovascular accidents: An observational study. *AMS* **2015**, *11*, 530–535. [CrossRef] [PubMed]
- 135. Brandler, E.S.; Sharma, M.; Sinert, R.H.; Levine, S.R. Prehospital stroke scales in urban environments: A systematic review. *Neurology* **2014**, *82*, 2241–2249. [CrossRef] [PubMed]
- 136. Sarissa Biomedical Ltd. Sarissa's stroke diagnostic: SMARTChip, wins SBRI grant award. Available online: http://www.sarissa-biomedical.com/news/sarissa%E2%80%99s-stroke-diagnostic-smartchip, -wins-sbri-grant-award.aspx (accessed on 1 August 2017).

Biosensors 2017, 7, 30 32 of 39

137. Harrison, P.; Segal, H.; Blasbery, K.; Furtado, C.; Silver, L.; Rothwell, P.M. Screening for aspirin responsiveness after transient ischemic attack and stroke. Comparison of 2 Point-of-Care Platelet Function Tests With Optical. *Aggregometry* **2005**, *36*, 1001–1005. [CrossRef] [PubMed]

- 138. Antithrombotic Trialists' Collaboration Collaborative meta-analysis of randomised trials of antiplatelet therapy for prevention of death, myocardial infarction, and stroke in high risk patients. *BMJ* **2002**, 324, 71–86.
- 139. Howard, P.A. Aspirin resistance. Ann. Pharmacother. 2002, 36, 1620–1624. [CrossRef] [PubMed]
- 140. McKee, S.A.; Sane, D.C.; Deliargyris, E.N. Aspirin resistance in cardiovascular disease: A review of prevalence, mechanisms, and clinical significance. *Thromb. Haemost.* **2002**, *88*, 711–715. [PubMed]
- 141. Altman, R.; Luciardi, H.L.; Muntaner, J.; Herrera, R.N. The antithrombotic profile of aspirin. Aspirin resistance, or simply failure? *Thromb. J.* **2004**, *2*, 1. [CrossRef] [PubMed]
- 142. Hankey, G.J.; Eikelboom, J.W. Aspirin resistance. BMJ 2004, 328, 477-479. [CrossRef] [PubMed]
- 143. Eikelboom, J.W.; Hankey, G.J. Failure of aspirin to prevent atherothrombosis: Potential mechanisms and implications for clinical practice. *Am. J. Cardiovasc. Drugs* **2004**, *4*, 57–67. [CrossRef] [PubMed]
- 144. Gum, P.A.; Kottke-Marchant, K.; Poggio, E.D.; Gurm, H.; Welsh, P.A.; Brooks, L.; Sapp, S.K.; Topol, E.J. Profile and prevalence of aspirin resistance in patients with cardiovascular disease. *Am. J. Cardiol.* **2001**, *88*, 230–235. [CrossRef]
- 145. Helgason, C.M.; Tortorice, K.L.; Winkler, S.R.; Penney, D.W.; Schuler, J.J.; McClelland, T.J.; Brace, L.D. Aspirin response and failure in cerebral infarction. *Stroke* **1993**, 24, 345–350. [CrossRef] [PubMed]
- 146. Patrono, C. Aspirin resistance: Definition, mechanisms and clinical read-outs. *JTH* **2003**, *1*, 1710–1713. [CrossRef] [PubMed]
- 147. Gum, P.A.; Kottke-Marchant, K.; Welsh, P.A.; White, J.; Topol, E.J. A prospective, blinded determination of the natural history of aspirin resistance among stable patients with cardiovascular disease. *J. Am. Coll. Cardiol.* **2003**, *41*, 961–965. [CrossRef]
- 148. Eikelboom, J.W.; Hankey, G.J. Aspirin resistance: A new independent predictor of vascular events? *J. Am. Coll. Cardiol.* **2003**, *41*, 966–968. [CrossRef]
- 149. Rand, M.L.; Leung, R.; Packham, M.A. Platelet function assays. *Transfus. Apheresis Sci.* **2003**, 28, 307–317. [CrossRef]
- 150. Baigent, C.; Blackwell, L.; Collins, R.; Emberson, J.; Godwin, J.; Peto, R.; Buring, J.; Hennekens, C.; Kearney, P.; Meade, T.; et al. Aspirin in the primary and secondary prevention of vascular disease: Collaborative meta-analysis of individual participant data from randomised trials. *Lancet* 2009, 373, 1849–1860. [PubMed]
- 151. Harrison, P. Progress in the assessment of platelet function. Br. J. Haematol. 2000, 111, 733–744. [PubMed]
- 152. Lordkipanidzé, M.; Pharand, C.; Schampaert, E.; Turgeon, J.; Palisaitis, D.A.; Diodati, J.G. A comparison of six major platelet function tests to determine the prevalence of aspirin resistance in patients with stable coronary artery disease. *Eur. Heart J.* **2007**, *28*, 1702–1708. [CrossRef] [PubMed]
- 153. Rothwell, P.M.; Algra, A.; Chen, Z.; Diener, H.C.; Norrving, B.; Mehta, Z. Effects of aspirin on risk and severity of early recurrent stroke after transient ischaemic attack and ischaemic stroke: Time-course analysis of randomised trials. *Lancet* **2016**, *388*, 365–375. [CrossRef]
- 154. Pearson, C.; Przyklenk, K.; Mika, V.H.; Ayaz, S.I.; Ellis, M.; Varade, P.; Tolomello, R.; Welch, R.D. Utility of point of care assessment of platelet reactivity (using the PFA-100(r)) to aid in diagnosis of stroke. *Am. J. Emerg. Med.* 2017, 35, 802.e1–802.e5. [CrossRef] [PubMed]
- 155. Drescher, M.J.; Spence, A.; Rockwell, D.; Smally, A.J. Point-of-care testing for coagulation studies in an emergency department stroke protocol: A time-saving innovation. *Annals Emerg. Med.* **2007**, *50*, S25. [CrossRef]
- 156. Mortezabeigi, H.R.; Taghizadeh, A.; Talebi, M.; Amini, K.; Goldust, M. ABCD2 score and BNP level in patients with TIA and cerebral stroke. *Pak. J. Biolog. Sciences* **2013**, *16*, 1393–1397.
- 157. Yokobori, S.; Hosein, K.; Burks, S.; Sharma, I.; Gajavelli, S.; Bullock, R. Biomarkers for the clinical differential diagnosis in traumatic brain injury—a systematic review. *CNS. Neurosci. Ther.* **2013**, *19*, 556–565. [CrossRef] [PubMed]
- 158. Cata, J.P.; Abdelmalak, B.; Farag, E. Neurological biomarkers in the perioperative period. *Br. J. Anaesth.* **2011**, 107, 844–858. [CrossRef] [PubMed]
- 159. AP., D. Biomarkers: Coming of age for environmental health and risk assessment. *Environ. Sci. Technol.* **1997**, 31, 1837–1848.

Biosensors 2017, 7, 30 33 of 39

160. Monbailliu, T.; Goossens, J.; Hachimi-Idrissi, S. Blood protein biomarkers as diagnostic tool for ischemic stroke: A systematic review. *Biomark. Med.* **2017**, *11*, 503–512. [CrossRef] [PubMed]

- 161. Herrmann, M.; Ehrenreich, H. Brain derived proteins as markers of acute stroke: Their relation to pathophysiology, outcome prediction and neuroprotective drug monitoring. *Restorative Neurol. Neurosci.* **2003**, *21*, 177–190.
- 162. Herrmann, M.; Vos, P.; Wunderlich, M.T.; de Bruijn, C.H.; Lamers, K.J. Release of glial tissue-specific proteins after acute stroke: A comparative analysis of serum concentrations of protein s-100b and glial fibrillary acidic protein. *Stroke* **2000**, *31*, 2670–2677. [CrossRef] [PubMed]
- 163. Wunderlich, M.T.; Ebert, A.D.; Kratz, T.; Goertler, M.; Jost, S.; Herrmann, M. Early neurobehavioral outcome after stroke is related to release of neurobiochemical markers of brain damage. *Stroke* **1999**, *30*, 1190–1195. [CrossRef] [PubMed]
- 164. Kapural, M.; Krizanac-Bengez, L.; Barnett, G.; Perl, J.; Masaryk, T.; Apollo, D.; Rasmussen, P.; Mayberg, M.R.; Janigro, D. Serum s-100beta as a possible marker of blood-brain barrier disruption. *Brain Res.* **2002**, *940*, 102–104. [CrossRef]
- 165. Gazzolo, D.; Abella, R.; Frigiola, A.; Giamberti, A.; Tina, G.; Nigro, F.; Florio, P.; Colivicchi, M.; Temporini, F.; Ricotti, A.; et al. Neuromarkers and unconventional biological fluids. *J. Maternal-fetal Neonatal Med.* **2010**, 23, 66–69. [CrossRef] [PubMed]
- 166. Mir, I.N.; Chalak, L.F. Serum biomarkers to evaluate the integrity of the neurovascular unit. *Early. Hum. Dev.* **2014**, *90*, 707–711. [CrossRef] [PubMed]
- 167. Brouns, R.; De Vil, B.; Cras, P.; De Surgeloose, D.; Marien, P.; De Deyn, P.P. Neurobiochemical markers of brain damage in cerebrospinal fluid of acute ischemic stroke patients. *Clin. Chem.* **2010**, *56*, 451–458. [CrossRef] [PubMed]
- 168. Lian, T.; Qu, D.; Zhao, X.; Yu, L.; Gao, B. Identification of site-specific stroke biomarker candidates by laser capture microdissection and labeled reference peptide. *Int. J. Mol. Sci.* **2015**, *16*, 13427–13441. [CrossRef] [PubMed]
- 169. Pelinka, L.E.; Kroepfl, A.; Leixnering, M.; Buchinger, W.; Raabe, A.; Redl, H. Gfap versus s100b in serum after traumatic brain injury: Relationship to brain damage and outcome. *J. Neurotrauma* **2004**, *21*, 1553–1561. [CrossRef] [PubMed]
- 170. Dvorak, F.; Haberer, I.; Sitzer, M.; Foerch, C. Characterisation of the diagnostic window of serum glial fibrillary acidic protein for the differentiation of intracerebral haemorrhage and ischaemic stroke. *Cerebrovasc. Dis.* 2009, 27, 37–41. [CrossRef] [PubMed]
- 171. Yang, Z.; Wang, K.K. Glial fibrillary acidic protein: From intermediate filament assembly and gliosis to neurobiomarker. *Trends Neurosci.* **2015**, *38*, 364–374. [CrossRef] [PubMed]
- 172. Teunissen, C.E.; Dijkstra, C.; Polman, C. Biological markers in csf and blood for axonal degeneration in multiple sclerosis. *Lancet. Neurol.* **2005**, *4*, 32–41. [CrossRef]
- 173. Shibata, D.; Cain, K.; Tanzi, P.; Zierath, D.; Becker, K. Myelin basic protein autoantibodies, white matter disease and stroke outcome. *J. Neuroimmunol.* **2012**, 252, 106–112. [CrossRef] [PubMed]
- 174. Zierath, D.; Kunze, A.; Fecteau, L.; Becker, K. Promiscuity of autoimmune responses to mbp after stroke. *J. Neuroimmunol.* **2015**, 285, 101–105. [CrossRef] [PubMed]
- 175. Becker, K.J.; Tanzi, P.; Zierath, D.; Buckwalter, M.S. Antibodies to myelin basic protein are associated with cognitive decline after stroke. *J. Neuroimmunol.* **2016**, 295, 9–11. [CrossRef] [PubMed]
- 176. Basile, A.M.; Fusi, C.; Conti, A.A.; Paniccia, R.; Trefoloni, G.; Pracucci, G.; Di Carlo, A.; Noferi, D.; Carbonetto, F.; Pretelli, P.; et al. S-100 protein and neuron-specific enolase as markers of subclinical cerebral damage after cardiac surgery: Preliminary observation of a 6-month follow-up study. *Eur. Neurol.* **2001**, *45*, 151–159. [CrossRef] [PubMed]
- 177. Karkela, J.; Bock, E.; Kaukinen, S. Csf and serum brain-specific creatine kinase isoenzyme (ck-bb), neuron-specific enolase (nse) and neural cell adhesion molecule (ncam) as prognostic markers for hypoxic brain injury after cardiac arrest in man. *J. Neurol. Sci.* **1993**, *116*, 100–109. [CrossRef]
- 178. Liu, M.C.; Akinyi, L.; Scharf, D.; Mo, J.; Larner, S.F.; Muller, U.; Oli, M.W.; Zheng, W.; Kobeissy, F.; Papa, L.; et al. Ubiquitin c-terminal hydrolase-l1 as a biomarker for ischemic and traumatic brain injury in rats. *Eur. J. Neurosci.* **2010**, *31*, 722–732. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 34 of 39

179. Papa, L.; Akinyi, L.; Liu, M.C.; Pineda, J.A.; Tepas, J.J.; Oli, M.W.; Zheng, W.; Robinson, G.; Robicsek, S.A.; Gabrielli, A.; et al. Ubiquitin c-terminal hydrolase is a novel biomarker in humans for severe traumatic brain injury. *Crit. Care Med.* **2010**, *38*, 138–144. [CrossRef] [PubMed]

- 180. Ren, C.; Zoltewicz, S.; Guingab-Cagmat, J.; Anagli, J.; Gao, M.; Hafeez, A.; Li, N.; Cao, J.; Geng, X.; Kobeissy, F.; et al. Different expression of ubiquitin c-terminal hydrolase-l1 and alphaii-spectrin in ischemic and hemorrhagic stroke: Potential biomarkers in diagnosis. *Brain Res.* **2013**, *1540*, 84–91. [CrossRef] [PubMed]
- 181. Tongaonkar, P.; Chen, L.; Lambertson, D.; Ko, B.; Madura, K. Evidence for an interaction between ubiquitin-conjugating enzymes and the 26s proteasome. *Mol. Cell. Biol.* **2000**, 20, 4691–4698. [CrossRef] [PubMed]
- 182. Vaagenes, P.; Urdal, P.; Melvoll, R.; Valnes, K. Enzyme level changes in the cerebrospinal fluid of patients with acute stroke. *Arch. Neurol.* **1986**, *43*, 357–362. [CrossRef] [PubMed]
- 183. Ingebrigtsen, T.; Romner, B. Biochemical serum markers for brain damage: A short review with emphasis on clinical utility in mild head injury. *Restor. Neurol. Neurosci.* **2003**, *21*, 171–176. [PubMed]
- 184. Atisha, D.; Bhalla, M.A.; Morrison, L.K.; Felicio, L.; Clopton, P.; Gardetto, N.; Kazanegra, R.; Chiu, A.; Maisel, A.S. A prospective study in search of an optimal b-natriuretic peptide level to screen patients for cardiac dysfunction. *Am. Heart J.* **2004**, *148*, 518–523. [CrossRef] [PubMed]
- 185. Ioannou, A.; Papageorgiou, N.; Falconer, D.; Rehal, O.; Sewart, E.; Zacharia, E.; Toutouzas, K.; Vlachopoulos, C.; Siasos, G.; Tsioufis, C.; et al. Biomarkers associated with stroke risk in atrial fibrillation. *Curr. Med. Chem.* 2017. [CrossRef] [PubMed]
- 186. Bustamante, A.; Lopez-Cancio, E.; Pich, S.; Penalba, A.; Giralt, D.; Garcia-Berrocoso, T.; Ferrer-Costa, C.; Gasull, T.; Hernandez-Perez, M.; Millan, M.; et al. Blood biomarkers for the early diagnosis of stroke: The stroke-chip study. *Stroke* **2017**. [CrossRef] [PubMed]
- 187. Castellanos, M.; Leira, R.; Serena, J.; Pumar, J.M.; Lizasoain, I.; Castillo, J.; Davalos, A. Plasma metalloproteinase-9 concentration predicts hemorrhagic transformation in acute ischemic stroke. *Stroke* **2003**, 34, 40–46. [CrossRef] [PubMed]
- 188. Sood, R.; Yang, Y.; Taheri, S.; Candelario-Jalil, E.; Estrada, E.Y.; Walker, E.J.; Thompson, J.; Rosenberg, G.A. Increased apparent diffusion coefficients on mri linked with matrix metalloproteinases and edema in white matter after bilateral carotid artery occlusion in rats. *J. Cereb. Blood Flow MeTable* 2009, 29, 308–316. [CrossRef] [PubMed]
- 189. Laskowitz, D.T.; Kasner, S.E.; Saver, J.; Remmel, K.S.; Jauch, E.C. Clinical usefulness of a biomarker-based diagnostic test for acute stroke: The biomarker rapid assessment in ischemic injury (brain) study. *Stroke* 2009, 40, 77–85. [CrossRef] [PubMed]
- 190. Zhang, X.Q.; Zhang, Z.M.; Yin, X.L.; Zhang, K.; Cai, H.; Ling, F. Exploring the optimal operation time for patients with hypertensive intracerebral hemorrhage: Tracking the expression and progress of cell apoptosis of prehematomal brain tissues. *Chin. Med. J.* **2010**, *123*, 1246–1250. [PubMed]
- 191. Taurino, M.; Raffa, S.; Mastroddi, M.; Visco, V.; Rizzo, L.; Torrisi, M.R.; Faraglia, V. Metalloproteinase expression in carotid plaque and its correlation with plasma levels before and after carotid endarterectomy. *Vascular Endovasc. Surg.* **2007**, *41*, 516–521. [CrossRef] [PubMed]
- 192. Tayal, V.; Kalra, B.S. Cytokines and anti-cytokines as therapeutics—an update. *Eur. J. Pharmacol.* **2008**, 579, 1–12. [CrossRef] [PubMed]
- 193. Rostene, W.; Dansereau, M.A.; Godefroy, D.; Van Steenwinckel, J.; Reaux-Le Goazigo, A.; Melik-Parsadaniantz, S.; Apartis, E.; Hunot, S.; Beaudet, N.; Sarret, P. Neurochemokines: A menage a trois providing new insights on the functions of chemokines in the central nervous system. *J. Neurochem.* **2011**, *118*, 680–694. [CrossRef] [PubMed]
- 194. Tuttolomondo, A.; Di Raimondo, D.; Pecoraro, R.; Arnao, V.; Pinto, A.; Licata, G. Inflammation in ischemic stroke subtypes. *Curr. Pharm. Des.* **2012**, *18*, 4289–4310. [CrossRef] [PubMed]
- 195. Licata, G.; Tuttolomondo, A.; Di Raimondo, D.; Corrao, S.; Di Sciacca, R.; Pinto, A. Immuno-inflammatory activation in acute cardio-embolic strokes in comparison with other subtypes of ischaemic stroke. *Thromb. Haemostasis* **2009**, *101*, 929–937. [CrossRef]
- 196. Abbott, N.J.; Ronnback, L.; Hansson, E. Astrocyte-endothelial interactions at the blood-brain barrier. *Nat. Rev. Neurosci.* **2006**, *7*, 41–53. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 35 of 39

197. Bailey, S.L.; Carpentier, P.A.; McMahon, E.J.; Begolka, W.S.; Miller, S.D. Innate and adaptive immune responses of the central nervous system. *Crit. Rev. Immunol.* **2006**, *26*, 149–188. [CrossRef] [PubMed]

- 198. Vela, J.M.; Molina-Holgado, E.; Arevalo-Martin, A.; Almazan, G.; Guaza, C. Interleukin-1 regulates proliferation and differentiation of oligodendrocyte progenitor cells. *Mol. Cell. Neurosci.* **2002**, *20*, 489–502. [CrossRef] [PubMed]
- 199. Rodriguez-Yanez, M.; Castillo, J. Role of inflammatory markers in brain ischemia. *Current Opin. Neurol.* **2008**, 21, 353–357. [CrossRef] [PubMed]
- 200. Gokhan, S.; Ozhasenekler, A.; Mansur Durgun, H.; Akil, E.; Ustundag, M.; Orak, M. Neutrophil lymphocyte ratios in stroke subtypes and transient ischemic attack. *Eur. Rev. Med. Pharmacol. Sci.* **2013**, 17, 653–657. [PubMed]
- 201. Huang, G.; Zhong, X.N.; Zhong, B.; Chen, Y.Q.; Liu, Z.Z.; Su, L.; Ling, Z.Y.; Cao, H.; Yin, Y.H. Significance of white blood cell count and its subtypes in patients with acute coronary syndrome. *Eur. J. Clin. Investig.* **2009**, 39, 348–358. [CrossRef] [PubMed]
- 202. Papa, A.; Emdin, M.; Passino, C.; Michelassi, C.; Battaglia, D.; Cocci, F. Predictive value of elevated neutrophil-lymphocyte ratio on cardiac mortality in patients with stable coronary artery disease. *Clin. Chim. Acta.* 2008, 395, 27–31. [CrossRef] [PubMed]
- 203. Cook, E.J.; Walsh, S.R.; Farooq, N.; Alberts, J.C.; Justin, T.A.; Keeling, N.J. Post-operative neutrophil-lymphocyte ratio predicts complications following colorectal surgery. *Intern. J. Surg.* **2007**, 5, 27–30. [CrossRef] [PubMed]
- 204. Karabinos, I.; Koulouris, S.; Kranidis, A.; Pastromas, S.; Exadaktylos, N.; Kalofoutis, A. Neutrophil count on admission predicts major in-hospital events in patients with a non-st-segment elevation acute coronary syndrome. *Clin. Cardiol.* 2009, 32, 561–568. [CrossRef] [PubMed]
- 205. Buck, B.H.; Liebeskind, D.S.; Saver, J.L.; Bang, O.Y.; Yun, S.W.; Starkman, S.; Ali, L.K.; Kim, D.; Villablanca, J.P.; Salamon, N.; et al. Early neutrophilia is associated with volume of ischemic tissue in acute stroke. *Stroke* **2008**, *39*, 355–360. [CrossRef] [PubMed]
- 206. Elkind, M.S.; Sciacca, R.R.; Boden-Albala, B.; Rundek, T.; Paik, M.C.; Sacco, R.L. Relative elevation in baseline leukocyte count predicts first cerebral infarction. *Neurology* **2005**, *64*, 2121–2125. [CrossRef] [PubMed]
- 207. Petzold, A. Neurofilament phosphoforms: Surrogate markers for axonal injury, degeneration and loss. *J. Neurol. Sci.* **2005**, 233, 183–198. [CrossRef] [PubMed]
- 208. Chen, X.H.; Meaney, D.F.; Xu, B.N.; Nonaka, M.; McIntosh, T.K.; Wolf, J.A.; Saatman, K.E.; Smith, D.H. Evolution of neurofilament subtype accumulation in axons following diffuse brain injury in the pig. *J. Neuropathol. Experim. Neurol.* **1999**, *58*, 588–596. [CrossRef]
- 209. Jafari, S.S.; Maxwell, W.L.; Neilson, M.; Graham, D.I. Axonal cytoskeletal changes after non-disruptive axonal injury. *J. Neurocytol.* **1997**, *26*, 207–221. [CrossRef] [PubMed]
- 210. Ost, M.; Nylen, K.; Csajbok, L.; Ohrfelt, A.O.; Tullberg, M.; Wikkelso, C.; Nellgard, P.; Rosengren, L.; Blennow, K.; Nellgard, B. Initial csf total tau correlates with 1-year outcome in patients with traumatic brain injury. *Neurology* **2006**, *67*, 1600–1604. [CrossRef] [PubMed]
- 211. Folkerts, M.M.; Berman, R.F.; Muizelaar, J.P.; Rafols, J.A. Disruption of map-2 immunostaining in rat hippocampus after traumatic brain injury. *J. Neurotrauma* **1998**, *15*, 349–363. [CrossRef] [PubMed]
- 212. Zemlan, F.P.; Jauch, E.C.; Mulchahey, J.J.; Gabbita, S.P.; Rosenberg, W.S.; Speciale, S.G.; Zuccarello, M. C-tau biomarker of neuronal damage in severe brain injured patients: Association with elevated intracranial pressure and clinical outcome. *Brain Res.* **2002**, *947*, 131–139. [CrossRef]
- 213. Zemlan, F.P.; Mulchahey, J.J.; Gudelsky, G.A. Quantification and localization of kainic acid-induced neurotoxicity employing a new biomarker of cell death: Cleaved microtubule-associated protein-tau (c-tau). *Neuroscience* 2003, 121, 399–409. [CrossRef]
- 214. Olmsted, J.B. Microtubule-associated proteins. Ann. Rev. Cell Biol. 1986, 2, 421–457. [CrossRef] [PubMed]
- 215. Matus, A. Neurofilament protein phosphorylation–where, when and why. *Trends Neurosci.* **1988**, *11*, 291–292. [CrossRef]
- 216. Garner, C.C.; Tucker, R.P.; Matus, A. Selective localization of messenger RNA for cytoskeletal protein MAP2 in dendrites. *Nature* **1988**, 336, 674–677. [CrossRef] [PubMed]
- 217. Kobeissy, F.H.; Sadasivan, S.; Oli, M.W.; Robinson, G.; Larner, S.F.; Zhang, Z.; Hayes, R.L.; Wang, K.K. Neuroproteomics and systems biology-based discovery of protein biomarkers for traumatic brain injury and clinical validation. *Proteomics. Clin. Appl.* **2008**, *2*, 1467–1483. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 36 of 39

218. Buki, A.; Siman, R.; Trojanowski, J.Q.; Povlishock, J.T. The role of calpain-mediated spectrin proteolysis in traumatically induced axonal injury. *J. Neuropathol. Experim. Neurol.* **1999**, *58*, 365–375. [CrossRef]

- 219. Reeves, T.M.; Greer, J.E.; Vanderveer, A.S.; Phillips, L.L. Proteolysis of submembrane cytoskeletal proteins ankyrin-g and alphaii-spectrin following diffuse brain injury: A role in white matter vulnerability at nodes of ranvier. *Brain Pathol.* **2010**, *20*, 1055–1068. [CrossRef] [PubMed]
- 220. Wang, K.K.; Posmantur, R.; Nath, R.; McGinnis, K.; Whitton, M.; Talanian, R.V.; Glantz, S.B.; Morrow, J.S. Simultaneous degradation of alphaii- and betaii-spectrin by caspase 3 (cpp32) in apoptotic cells. *J. Biol. Chem.* 1998, 273, 22490–22497. [CrossRef] [PubMed]
- 221. Cox, C.D.; West, E.J.; Liu, M.C.; Wang, K.K.; Hayes, R.L.; Lyeth, B.G. Dicyclomine, an m1 muscarinic antagonist, reduces biomarker levels, but not neuronal degeneration, in fluid percussion brain injury. *J. Neurotrauma* 2008, 25, 1355–1365. [CrossRef] [PubMed]
- 222. Adam, S.S.; Key, N.S.; Greenberg, C.S. D-dimer antigen: Current concepts and future prospects. *Blood* **2009**, 113, 2878–2887. [CrossRef] [PubMed]
- 223. Mai, H.; Xia, J.; Wu, Y.; Ke, J.; Li, J.; Pan, J.; Chen, W.; Shao, Y.; Yang, Z.; Luo, S.; et al. Clinical presentation and imaging characteristics of occult lung cancer associated ischemic stroke. *J. Clin. Neurosci.* 2015, 22, 296–302. [CrossRef] [PubMed]
- 224. Kim, K.; Lee, J.H. Risk factors and biomarkers of ischemic stroke in cancer patients. *J. Stroke* **2014**, *16*, 91–96. [CrossRef] [PubMed]
- 225. Liu, L.B.; Li, M.; Zhuo, W.Y.; Zhang, Y.S.; Xu, A.D. The role of hs-crp, d-dimer and fibrinogen in differentiating etiological subtypes of ischemic stroke. *PLoS. One* **2015**, *10*, e0118301. [CrossRef] [PubMed]
- 226. Zi, W.J.; Shuai, J. Plasma d-dimer levels are associated with stroke subtypes and infarction volume in patients with acute ischemic stroke. *PLoS One* **2014**, *9*, e86465. [CrossRef] [PubMed]
- 227. Zecca, B.; Mandelli, C.; Maino, A.; Casiraghi, C.; Bolla, G.; Consonni, D.; Santalucia, P.; Torgano, G. A bioclinical pattern for the early diagnosis of cardioembolic stroke. *Emerg. Med. Intern.* **2014**, 2014, 242171. [CrossRef] [PubMed]
- 228. Yuan, W.; Shi, Z.H. The relationship between plasma d-dimer levels and outcome of chinese acute ischemic stroke patients in different stroke subtypes. *J. Neur. Transm.* **2014**, 121, 409–413. [CrossRef] [PubMed]
- 229. Wiseman, S.; Marlborough, F.; Doubal, F.; Webb, D.J.; Wardlaw, J. Blood markers of coagulation, fibrinolysis, endothelial dysfunction and inflammation in lacunar stroke versus non-lacunar stroke and non-stroke: Systematic review and meta-analysis. *Cerebrovasc. Dis.* 2014, 37, 64–75. [CrossRef] [PubMed]
- 230. Okazaki, T.; Yamamoto, Y.; Yoda, K.; Nagahiro, S. The ratio of d-dimer to brain natriuretic peptide may help to differentiate between cerebral infarction with and without acute aortic dissection. *J. Neurol. Sci.* **2014**, *340*, 133–138. [CrossRef] [PubMed]
- 231. Isenegger, J.; Meier, N.; Lammle, B.; Alberio, L.; Fischer, U.; Nedeltchev, K.; Gralla, J.; Kohler, H.P.; Mattle, H.P.; Arnold, M. D-dimers predict stroke subtype when assessed early. *Cerebrovasc. Dis.* **2010**, 29, 82–86. [CrossRef] [PubMed]
- 232. Ilhan, D.; Ozbabalik, D.; Gulcan, E.; Ozdemir, O.; Gulbacs, Z. Evaluation of platelet activation, coagulation, and fibrinolytic activation in patients with symptomatic lacunar stroke. *Neurologist* **2010**, *16*, 188–191. [CrossRef] [PubMed]
- 233. Haapaniemi, E.; Tatlisumak, T. Is d-dimer helpful in evaluating stroke patients? A systematic review. *Acta Neurol. Scand.* **2009**, *119*, 141–150. [CrossRef] [PubMed]
- 234. Montaner, J.; Perea-Gainza, M.; Delgado, P.; Ribo, M.; Chacon, P.; Rosell, A.; Quintana, M.; Palacios, M.E.; Molina, C.A.; Alvarez-Sabin, J. Etiologic diagnosis of ischemic stroke subtypes with plasma biomarkers. Stroke 2008, 39, 2280–2287. [CrossRef] [PubMed]
- 235. Dougu, N.; Takashima, S.; Sasahara, E.; Taguchi, Y.; Toyoda, S.; Hirai, T.; Nozawa, T.; Tanaka, K.; Inoue, H. Differential diagnosis of cerebral infarction using an algorithm combining atrial fibrillation and d-dimer level. *Eur. J. Neurol.* **2008**, *15*, 295–300. [CrossRef] [PubMed]
- 236. Squizzato, A.; Ageno, W.; Finazzi, S.; Mera, V.; Romualdi, E.; Bossi, A.; Venco, A. D-dimer is not a long-term prognostic marker following acute cerebral ischemia. *Blood Coagul. Fibrinol.* 2006, 17, 303–306. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 37 of 39

237. Squizzato, A.; Ageno, W. D-dimer testing in ischemic stroke and cerebral sinus and venous thrombosis. *Semin. Vasc. Med.* **2005**, *5*, 379–386. [CrossRef] [PubMed]

- 238. Koch, H.J.; Horn, M.; Bogdahn, U.; Ickenstein, G.W. The relationship between plasma d-dimer concentrations and acute ischemic stroke subtypes. *J. Stroke Cerebrovasc. Dis.* **2005**, *14*, 75–79. [CrossRef] [PubMed]
- 239. Ageno, W.; Finazzi, S.; Steidl, L.; Biotti, M.G.; Mera, V.; Melzi D'Eril, G.; Venco, A. Plasma measurement of d-dimer levels for the early diagnosis of ischemic stroke subtypes. *Arch. Intern. Med.* **2002**, *162*, 2589–2593. [CrossRef]
- 240. Takano, K.; Yamaguchi, T.; Uchida, K. Markers of a hypercoagulable state following acute ischemic stroke. *Stroke* **1992**, 23, 194–198. [CrossRef] [PubMed]
- 241. Elkind, M.S.; Tai, W.; Coates, K.; Paik, M.C.; Sacco, R.L. High-sensitivity c-reactive protein, lipoprotein-associated phospholipase a2, and outcome after ischemic stroke. *Arch. Intern. Med.* **2006**, *166*, 2073–2080. [CrossRef] [PubMed]
- 242. Muir, K.W.; Weir, C.J.; Alwan, W.; Squire, I.B.; Lees, K.R. C-reactive protein and outcome after ischemic stroke. *Stroke* **1999**, *30*, 981–985. [CrossRef] [PubMed]
- 243. Audebert, H.J.; Rott, M.M.; Eck, T.; Haberl, R.L. Systemic inflammatory response depends on initial stroke severity but is attenuated by successful thrombolysis. *Stroke* **2004**, *35*, 2128–2133. [CrossRef] [PubMed]
- 244. Suwanwela, N.C.; Chutinet, A.; Phanthumchinda, K. Inflammatory markers and conventional atherosclerotic risk factors in acute ischemic stroke: Comparative study between vascular disease subtypes. *J. Med. Assoc. Thail.* **2006**, *89*, 2021–2027.
- 245. Masotti, L.; Ceccarelli, E.; Forconi, S.; Cappelli, R. Prognostic role of c-reactive protein in very old patients with acute ischaemic stroke. *J. Intern. Med.* **2005**, 258, 145–152. [CrossRef] [PubMed]
- 246. Hirano, K.; Takashima, S.; Dougu, N.; Taguchi, Y.; Nukui, T.; Konishi, H.; Toyoda, S.; Kitajima, I.; Tanaka, K. Study of hemostatic biomarkers in acute ischemic stroke by clinical subtype. *J. Stroke Cerebrovasc. Dis.* **2012**, 21, 404–410. [CrossRef] [PubMed]
- 247. Iskra, T.; Turaj, W.; Slowik, A.; Zwolinska, G.; Strojny, J.; Szczudlik, A. [hemostatic markers of endothelial injury in ischaemic stroke caused by large or small vessel disease]. *Pol. Merkur. Lekarski.* **2006**, 21, 429–433. [PubMed]
- 248. Bang, O.Y.; Saver, J.L.; Liebeskind, D.S.; Pineda, S.; Ovbiagele, B. Association of serum lipid indices with large artery atherosclerotic stroke. *Neurology* **2008**, *70*, 841–847. [CrossRef] [PubMed]
- 249. Slowik, A.; Iskra, T.; Turaj, W.; Hartwich, J.; Dembinska-Kiec, A.; Szczudlik, A. Ldl phenotype b and other lipid abnormalities in patients with large vessel disease and small vessel disease. *J. Neurol. Sci.* 2003, 214, 11–16. [CrossRef]
- 250. Iskra, T.; Turaj, W.; Hartwich, J.; Slowik, A.; Pankiewicz, J.; Dembinska-Kiec, A.; Szczudlik, A. [ldl phenotype a and b in ischemic stroke]. *Prz. Lekarski* **2002**, *59*, 7–10.
- 251. Zimmermann-Ivol, C.G.; Burkhard, P.R.; Le Floch-Rohr, J.; Allard, L.; Hochstrasser, D.F.; Sanchez, J.C. Fatty acid binding protein as a serum marker for the early diagnosis of stroke: A pilot study. *MCP.* **2004**, *3*, 66–72. [CrossRef] [PubMed]
- 252. Watson, M.A.; Scott, M.G. Clinical utility of biochemical analysis of cerebrospinal fluid. *Clin. Chem.* **1995**, *41*, 343–360. [PubMed]
- 253. Sun, G.J.; Ding, S.C.; Ling, W.Y.; Wang, F.; Yang, X.P. Cerebrospinal fluid free fatty acid levels are associated with stroke subtypes and severity in chinese patients with acute ischemic stroke. *World Neurosurg.* **2015**, *84*, 1299–1304. [CrossRef] [PubMed]
- 254. Zambrelli, E.; Emanuele, E.; Marcheselli, S.; Montagna, L.; Geroldi, D.; Micieli, G. Apo(a) size in ischemic stroke: Relation with subtype and severity on hospital admission. *Neurology* **2005**, *64*, 1366–1370. [CrossRef] [PubMed]
- 255. Iskra, T.; Turaj, W.; Slowik, A.; Szczudlik, A.; Dembinska-Kiec, A. [lipoprotein (a) in stroke patients with large and small vessel disease]. *Prz. Lekarski* 2002, *59*, 877–880.
- 256. Saidi, S.; Slamia, L.B.; Ammou, S.B.; Mahjoub, T.; Almawi, W.Y. Association of apolipoprotein E gene polymorphism with ischemic stroke involving large-vessel disease and its relation to serum lipid levels. *J. Stroke Cerebrovasc. Dis.* **2007**, *16*, 160–166. [CrossRef] [PubMed]

Biosensors 2017, 7, 30 38 of 39

257. Lampl, Y.; Paniri, Y.; Eshel, Y.; Sarova-Pinhas, I. Cerebrospinal fluid lactate dehydrogenase levels in early stroke and transient ischemic attacks. *Stroke* **1990**, *21*, 854–857. [CrossRef] [PubMed]

- 258. Alvarez-Perez, F.J.; Castelo-Branco, M.; Alvarez-Sabin, J. Albumin level and stroke. Potential association between lower albumin level and cardioembolic aetiology. *Int. J. Neurosci.* **2011**, *121*, 25–32. [CrossRef] [PubMed]
- 259. Xu, Y.Z.; Yang, Z.G.; Zhang, Y.H.; Zhang, Y.W.; Hong, B.; Liu, J.M. Dynamic reduction of plasma decorin following ischemic stroke: A pilot study. *Neurochem. Res.* **2012**, *37*, 1843–1848. [CrossRef] [PubMed]
- 260. Drain, P.K.; Hyle, E.P.; Noubary, F.; Freedberg, K.A.; Wilson, D.; Bishai, W.R.; Rodriguez, W.; Bassett, I.V. Diagnostic point-of-care tests in resource-limited settings. *Lancet. Infect. Dis.* **2014**, *14*, 239–249. [CrossRef]
- 261. Kumar, S.; Kumar, S.; Ali, M.A.; Anand, P.; Agrawal, V.V.; John, R.; Maji, S.; Malhotra, B.D. Microfluidic-integrated biosensors: Prospects for point-of-care diagnostics. *Biotechnol. J.* **2013**, *8*, 1267–1279. [CrossRef] [PubMed]
- 262. Eltzov, E.; Cohen, A.; Marks, R.S. Bioluminescent liquid light guide pad biosensor for indoor air toxicity monitoring. *Anal. Chem.* **2015**, *87*, 3655–3661. [CrossRef] [PubMed]
- 263. Eltzov, E.; Guttel, S.; Low Yuen Kei, A.; Sinawang, P.D.; Ionescu, R.E.; Marks, R.S. Lateral flow immunoassays from paper strip to smartphone technology. *Electroanalysis* **2015**, *27*, 2116–2130. [CrossRef]
- 264. Eltzov, E.; Marks, R.S. Colorimetric stack pad immunoassay for bacterial identification. *Biosens. Bioelectron.* **2016**, *87*, 572–578. [CrossRef] [PubMed]
- 265. Eltzov, E.; Marks, R.S. Miniaturized flow stacked immunoassay for detecting escherichia coli in a single step. *Anal. Chem.* **2016**, *88*, 6441–6449. [CrossRef] [PubMed]
- 266. Algaar, F.; Eltzov, E.; Vdovenko, M.M.; Sakharov, I.Y.; Fajs, L.; Weidmann, M.; Mirazimi, A.; Marks, R.S. Fiber-optic immunosensor for detection of crimean-congo hemorrhagic fever igg antibodies in patients. *Anal. Chem.* 2015, 87, 8394–8398. [CrossRef] [PubMed]
- 267. Eltzov, E.; Cosnier, S.; Marks, R.S. Biosensors based on combined optical and electrochemical transduction for molecular diagnostics. *Exp. Rev.Molec. Diagn.* **2011**, *11*, 533–546. [CrossRef] [PubMed]
- 268. Eltzov, E.; Marks, R.S. Fiber-optic based cell sensors. In *Whole Cell Sensing Systems I*; Springer: Berlin/Heidelberg, Germany, 2010; pp. 131–154.
- 269. Thevenot, D.R.; Toth, K.; Durst, R.A.; Wilson, G.S. Electrochemical biosensors: Recommended definitions and classification. *Biosens. Bioelectron.* **2001**, *16*, 121–131. [CrossRef] [PubMed]
- 270. Eltzov, E.; Marks, R.S. Whole-cell aquatic biosensors. *Anal. Bioanal. Chem.* **2011**, *400*, 895–913. [CrossRef] [PubMed]
- 271. Collings, A.F.; Caruso, F. Biosensors: Recent advances. Rep. Prog. Phys. 1997, 60, 1397–1445. [CrossRef]
- 272. Eltzov, E.; Kushmaro, A.; Marks, R.S. Biosensors and related techniques for endocrine disruptors. In *Endocrine Disrupting Chemicals in Food, Snow, I., Ed.*; Woodhead: Cambridge, UK, 2009.
- 273. Marks, R.S.; Cullen, D.C.; Karube, I.; Lowe, C.R.; Weetall, H.H. *Handbook of Biosensors and Biochips*; Wiley-Blackwell: Tokyo, Japan, 2007.
- 274. Chin, C.D.; Linder, V.; Sia, S.K. Commercialization of microfluidic point-of-care diagnostic devices. *Lab Chip* **2012**, *12*, 2118–2134. [CrossRef] [PubMed]
- 275. Sinawang, P.D.; Rai, V.; Ionescu, R.E.; Marks, R.S. Electrochemical lateral flow immunosensor for detection and quantification of dengue ns1 protein. *Biosens. Bioelectron.* **2016**, 77, 400–408. [CrossRef] [PubMed]
- Sinawang, P.D.; Harpaz, D.; Fajs, L.; Seet, R.C.S.; Tok, A.I.Y.; Marks, R.S. Electrochemical impedimetric detection of stroke biomarker nt-probnp using disposable screen-printed gold electrodes. *EuroBiotech J.* 2017, 5, 165–176. [CrossRef]
- 277. Stephenson, D.; Hu, M.T.; Romero, K.; Breen, K.; Burn, D.; Ben-Shlomo, Y.; Bhattaram, A.; Isaac, M.; Venuto, C.; Kubota, K.; et al. Precompetitive data sharing as a catalyst to address unmet needs in parkinson's disease1. *J. Parkinsons. Dis.* 2015, *5*, 581–594. [CrossRef] [PubMed]
- 278. Easton, J.F.; Stephens, C.R.; Angelova, M. Risk factors and prediction of very short term versus short/intermediate term post-stroke mortality: A data mining approach. *Comput. Biol. Med.* **2014**, *54*, 199–210. [CrossRef] [PubMed]
- 279. Yao, Q.; Tian, Y.; Li, P.F.; Tian, L.L.; Qian, Y.M.; Li, J.S. Design and development of a medical big data processing system based on hadoop. *J. Med. Syst.* 2015, 39, 23. [CrossRef] [PubMed]

Biosensors **2017**, 7, 30 39 of 39

280. Davidoff, F.; Haynes, B.; Sackett, D.; Smith, R. Evidence based medicine. *BMJ* 1995, 310, 1085–1086. [CrossRef] [PubMed]

281. Fernández-Llatas, C.; García-Gómez, J.M. (Eds.) *Data Mining in Clinical Medicine*; Humana Press: New York, NY, USA, 2015.



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).