

## **Supporting Information**

### **Evanescent Wave Optical-Fiber Aptasensor for Rapid Detection of Zearalenone in Corn with Unprecedented Sensitivity**

Haixu Zhao<sup>†</sup>, Shang Ren<sup>†</sup>, Zhenzhe Wei, and Xinhui Lou\*

Department of Chemistry, Capital Normal University, Xisanhuan North Road. 105,  
Beijing 100048, China

<sup>†</sup> The authors contributed equally to this work.

\*To whom correspondence should be addressed. Tel: +86-10-68902491 ext. 808;  
E-mail: xinhuiou@cnu.edu.cn

**Table S1.** ZEN-binding aptamers used in the aptasensors

Aptamer name	Sequence (5'-3')	Length (nt)	Kd (nM)	Ref.
Z31N	ATGGTACATTACTATCTGTAATGTGATAT	29	11.77 ± 1.44	1
8Z <sub>31</sub>	TCATCTATCTATGGTACATTACTATCTGTAATGTGATATG	40	41±5	2
ZEN-62	CTACCAGCTTTGAGGCTCGATCCAGCTTATTCAATTATACCAGCTTATTCAA TTATACCAGC	62	15.2±3.4	3
ZEN-38	GATGGGGAAAGGGTCCCCCTGGGTTGGAGCATCGGACA	38	2200±400	4
Number46	GGAATTCTTGATGTTGCCTGGGATTGTTTGGGCCTTGTGTTTTCTTCCGTTC CAACTTAGTAGGATCCCGAA	72	50.5 ± 5.4	5
ZEN-80	AGCAGCACAGAGGTCAGATGTCATCTATCTATGGTACATTACTATCTGTAAT GTGATATGCCTATGCGTGCTACCGTGAA	80	-	6
Z100	ATACCAGCTTATTCAATTCTACCAGCTTTGAGGCTCGATCCAGCTTATTCAA TTATACCAGCTTATTCAATTATACCAGCACAATCGTAATCAGTTAG	98	15.2 ± 3.4	7

Note: The aptamer names shown in the Table are either the names used in the reported literatures or named according to the length of the aptamer.

**Table S2.** Literature reported aptasensors for the detection ZEN

No	Aptasensor	Aptamer probe	LOD (ng/mL)	Dynamic range (ng/mL)	Ref.
1	Colorimetric competitive ELAA	Biotin-8Z <sub>31</sub>	0.7	1-10000	<sup>8</sup>
2	Colorimetric lateral flow assay	HS-8Z <sub>31</sub> -T <sub>29</sub>	20	5–200	<sup>9</sup>
3	Colorimetric	ZEN-98	4	4–128	<sup>7</sup>
4	Enhanced ELAA	HS–GAT-8Z <sub>31</sub> -Biotin	0.08	0.1–160	<sup>10</sup>
5	Colorimetric-AuNP	ZEN-38	10	10–250	<sup>11</sup>
6	Colorimetric-aptamer walker, Exo III, and AuNPs	HS-ZEN-80	0.01	0.02–80	<sup>12</sup>
7	Fluorescence quenching	ZEN-80-FAM	0.5	0.5- 64	<sup>6</sup>
8	AIR-SE combined with SPR	HS–T <sub>10</sub> -8Z <sub>31</sub>	0.08	0.01–1000	<sup>13</sup>
9	Green ELAA	Biotin-8Z <sub>31</sub>	0.377	-	<sup>14</sup>
10	Colorimetric-porous platinum nanotubes, p-PtNTs/AuNPs and thionine (Thi) labelled graphene oxide (GO)	HS-ZEN-98	$1.7 \times 10^{-4}$	$5 \times 10^{-4}$ –500	<sup>15</sup>
11	Fluorescent sensing-Genefinder	8Z <sub>31</sub>	0.1	0.1-200	<sup>16</sup>
12	Fluorescent sensing-mesoporous silica nanoparticle	8Z <sub>31</sub> -FAM	0.012	0.005–150	<sup>17</sup>
13	SERS	HS-8Z <sub>31</sub>	0.001	0.005 -500	<sup>18</sup>
14	Electrochemical	8Z <sub>31</sub>	0.017	0.01-1000	<sup>19</sup>
15	Competitive electrochemical	ZEN-29	$1.5 \times 10^{-3}$	0.001-100	<sup>1</sup>
16	Ratiometric fluorescent-CdTe quantum dots (QDs)	8Z <sub>31</sub> and NH <sub>2</sub> -ZEN-80	2.4	10 -200	<sup>20</sup>
17	FRET- dual-color AuNCs	NH <sub>2</sub> -8Z <sub>31</sub>	$5.3 \times 10^{-4}$	0.005–100	<sup>21</sup>
18	Optical fiber- localized SPR	HS-ZEN-80	0.102	1- 480	<sup>22</sup>
19	Nanofibers for dispersive solid phase extraction (dSPE)	HOOC-8Z <sub>31</sub>	$1.8 \times 10^{-5}$	$6 \times 10^{-5}$ –0.01	<sup>23</sup>

No	Aptasensor	Aptamer probe	LOD (ng/mL)	Dynamic range (ng/mL)	Ref ·
20	Magnetic separation and time-resolved fluorescent $\text{Ln}^{3+}$ inorganic nanoparticles.	Biotin-8Z <sub>31</sub>	$2.1 \times 10^{-4}$	0.001–10	24
21	Ratiometric fluorescence	8Z <sub>31</sub> -NH <sub>2</sub>	$3.2 \times 10^{-4}$	$3.2 \times 10^{-4}$ -0.32	25
22	Electrochemical-DNA assembly	AAA-8Z <sub>31</sub> -SH	$5.1 \times 10^{-7}$	$5.0 \times 10^{-6}$ -50	26
23	Fluorescence-upconverting nanoparticles	8Z <sub>31</sub>	0.007	0.05–100	27
24	AuNP-affinity monolithic column	8Z <sub>31</sub>	0.05	0.16-60	28
25	Photoelectrochemical-ZnO-NGQDs	ZEN-72-NH <sub>2</sub>	$3.3 \times 10^{-5}$	$1.0 \times 10^{-4}$ –100	29
26	Fluorometric-lighting-up silver nanocluster	GGTCTATTAACGA AGGTCATTAAC-8Z <sub>3</sub> 1	$3.2 \times 10^{-4}$	$1.3 \times 10^{-3}$ -100	30
27	Competitive ELAA	ZEN-72	0.2	0.03–2.5	5
28	Fluorometric-gold nanorods and upconversion nanoparticles	8Z <sub>31</sub> -AAATCATGTC	0.01	0.05–100	31
29	Dual-colored persistent luminescence	HS-8Z <sub>31</sub>	$2.2 \times 10^{-4}$	0.001–50	32
30	Electrochemical-CS@AB-MWCNTs	8Z <sub>31</sub> -NH <sub>2</sub>	$3.64 \times 10^{-6}$	$10^{-5}$ –1	33
31	Fluorometric-DNA tweezer nanomachine	8Z <sub>31</sub>	0.037	0.05- 50	34
32	Fluorometric	8Z <sub>31</sub>	0.25	0.1-1000	2
33	Fluorometric	Biotin-8Z <sub>31</sub>	$5.1 \times 10^{-4}$	0.001 - 100	35
34	Electrochemical-3D sakura-shaped Cu@L-Glu combined with palladium-platinum nanoparticle (Pd-PtNPs) to obtain Cu@L-Glu/Pd-PtNPs	NH <sub>2</sub> -ZEN-80	$4.5 \times 10^{-7}$	$10^{-6}$ - 100	36
35	Fluorometric-dual-enzyme	Biotin-ZEN-62	$2.13 \times 10^{-4}$	$10^{-3}$ –1	3
36	Electrochemical-PEI-MoS <sub>2</sub> -MWCNTs	8Z <sub>31</sub> -SH	$1.7 \times 10^{-4}$	$5 \times 10^{-4}$ -50	37

No	Aptasensor	Aptamer probe	LOD (ng/mL)	Dynamic range (ng/mL)	Ref ·
37	Colorimetric-metal-organic framework nano-container and trivalent DNAzyme	8Z <sub>31</sub>	$3.6 \times 10^{-4}$	0.01–100	<sup>38</sup>
38	SERS-Au@Ag core-shell nanoparticles and gold nanorods	HS-8Z <sub>31</sub>	0.054	0.05 - 500	<sup>39</sup>
39	Electrochemical-AuNPs (rMoS <sub>2</sub> -Au)	HS-8Z <sub>31</sub>	$5 \times 10^{-4}$	$1 \times 10^{-3}$ –10	<sup>40</sup>
40	Electrochemical-CoSe <sub>2</sub> /AuN Rs and 3D structured DNA-PtNi@Co-MOF	CCTCAGCCATATCA CATTAGC TGA-8Z <sub>31</sub>	$1.37 \times 10^{-6}$	$10^{-5}$ - 10	<sup>41</sup>
41	Pregnancy test strip-catalytic hairpin assembly	HS-8Z <sub>31</sub> -TTGTTCTTC ACATTACAGATAGT AATGTA	0.05	0.05–5,000	<sup>42</sup>

Note: ELAA: enzyme-linked aptamer assay; AIR-SE: attenuated internal reflection spectroscopic ellipsometry; SPR: surface plasmon resonance; SERS: surface-enhanced Raman scattering;

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