

*Supplementary Materials*

# Factors Influencing Properties of Spider Silk Coatings and Their Interactions within a Biological Environment

Vanessa T. Trossmann <sup>1</sup>, Sarah Lentz <sup>1</sup> and Thomas Scheibel <sup>1,2,3,4,5,6,\*</sup>

- <sup>1</sup> Chair of Biomaterials, Faculty of Engineering Science, University of Bayreuth, Prof.-Rüdiger-Bormann-Straße 1, 95447 Bayreuth, Germany; vanessa.trossmann@uni-bayreuth.de (V.T.T.); sarah1.lentz@uni-bayreuth.de (S.L.)  
<sup>2</sup> Bayreuth Center for Colloids and Interfaces (BZKG), University of Bayreuth, Universitätsstraße 30, 95440 Bayreuth, Germany  
<sup>3</sup> Bavarian Polymer Institute (BPI), University of Bayreuth, Universitätsstraße 30, 95440 Bayreuth, Germany  
<sup>4</sup> Bayreuth Center for molecular Biosciences (BZMB), University of Bayreuth, Universitätsstraße 30, 95440 Bayreuth, Germany  
<sup>5</sup> Bayreuth Materials Center (BayMAT), University of Bayreuth, Universitätsstraße 30, 95440 Bayreuth, Germany  
<sup>6</sup> Faculty of Medicine, University of Würzburg, Pleicherwall 2, 97070 Würzburg, Germany  
\* Correspondence: thomas.scheibel@uni-bayreuth.de

**Table S1.** Summary of recombinant spider silk proteins, spider species, production host and respective amino acid sequence.

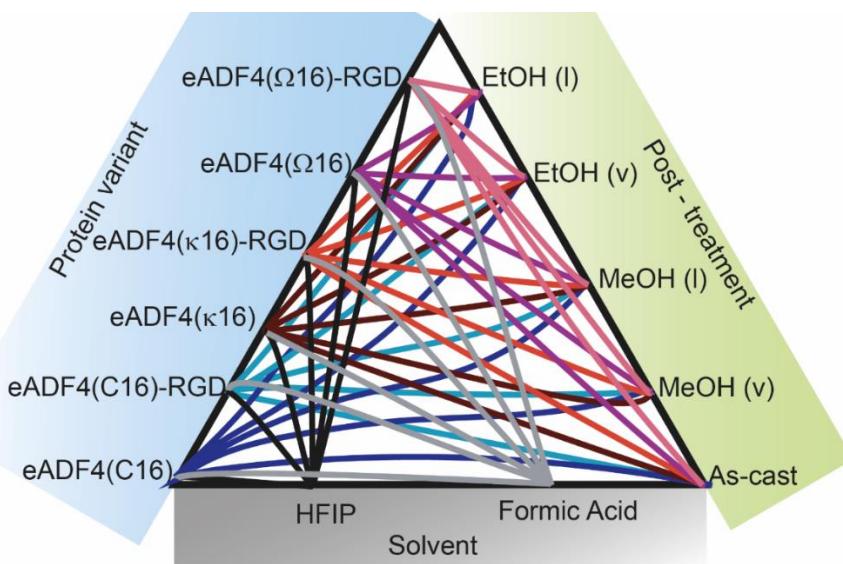
Protein	Spider species	Production host	Amino acid sequence	Reference
eADF4(C16)	<i>Araneus diadematus</i>	<i>E. coli</i>	C- module repeated 16 or 32 times GSSAAAAAAAASGPGGYGPENQGPSPGPGGYGPGGP + cell adhesion peptides (e.g. RGD, IKVAV) + biomineralization peptides + cystein-containing peptide (e.g. ntagCys)	[1-7]
eADF4(C32)	<i>Araneus diadematus</i>	<i>E. coli</i>	$\kappa$ -module repeated 16 times GSSAAAAAAAASGPGGYGPKNQGPSPGPGGYGPGGP + cell adhesion peptides (e.g. RGD, IKVAV) + cystein-containing peptide (e.g. ntagCys)	[1,3-6]
eADF4( $\Omega$ 16)	<i>Araneus diadematus</i>	<i>E. coli</i>	$\Omega$ -module repeated 16 times GSSAAAAAAAASGPGGYGPQNQGPSPGPGGYGPGGP + cell adhesion peptides (e.g. RGD, IKVAV)	[1-4]
eADF3(AQ) <sub>12</sub>	<i>Araneus diadematus</i>	<i>E. coli</i>	A and Q module repeated 12 or 24 times (A) GPYGPGASAAAAAGGYGPGSGQQ (Q) GPGQQGPGQQGPGQQGPGQQ	[1,2]
eADF3(AQ) <sub>24</sub>				
eADF3 with functional domains			Domains: cellulose-binding domains (CBM), peptide interacting domains (SPY_C), gamma-crystalline D domain (Crys), fibronectin III domain (FN)	[8-10]

---

4RepCT	<i>Euprosthenops australis</i>	<i>E. coli</i>	GGSGNSGIQGQGGYGGLGQGGYGGAGSSAAAAAAAAAA AGGQGGQGQGGYGGQGSGSAAAAAAGGGGGQGQGQGQGQG GYGQGSGNAAAAAAAGGGGGQGQGQGQGQGQGQGQGQGQG GSAAAAAAAAAGSGQGGYGGQGQGQGQGQGQGQGQGQGQG AASAASTVANSVSRLLSPSAVSRVSSAVSSLVSNGQVNMAALP NIISNISSVSASAPGASGCEVIVQALLEVITALVQIVSSSVGYIN PSAVNQITNVVANAMAQVMG (contains C-terminal domain) + cell adhesion peptides (e.g. RGD, IKVAV) + antimicrobial peptides (e.g. Lac and Mag) + specific ligand-interaction peptides (e.g. IgG and albumin) + entire proteins (e.g. FGF)	[11-36]
2RepCT			GNSGRGQGGYGGSGGNAAAAAAAAAAAAGQGGQGG YGRQSQGAGSAAAAAAAAAAAGSGQGGYGGQGQGGY GQSGNSVTSGGYGYGTSAAAGAGVAAGSYAGAVNRLSSAEA SRVSSNIAAIASGGASALPSVISNIYSCVVASGVSSNEALIQALLE LLSALVHVLSSASIGNVSSVGVDSTLNVQDSVGQYVG + N-terminal domain	[37-41]
rMaSp1	<i>Trichonephila clavipes</i>	Transgenic goats	QGAGAAAAAAGGAGQGGYGLGGQAGQGGYGLGGQGA GQGAGAAAAAAGGAGQGGYGLGSQGAGRGGLGGQGAGAAA AAAGGAGQGGYGLGSQGAGRGGLGGQGAGAAAAAAGG AGQGGYGLGNQGAGRGGQGAAAAAAGGAGQGGYGLGS QGAGRGGLGGQGAGAAAAAAGGAGQGGYGLGGQGAGQQ GYGLGSQGAGRGGLGGQGAGAAAAAAGGAGQGGLGGQ GAGQGAGASAAAAGGAGQGGYGLGSQGAGRGGEGAGAA AAAAGGAGQGGYGLGGQGAGQGGYGLGSQGAGRGGL GQGAGAAAAGGAGQGGLGGQGAGQQGAGAAAAAAGGAGQ GGYGLGSQGAGRGGLGGQGAGAVAAAAAGGAGQGGYGG LGSQGAGRGGQGAGAAAAAAGGAGQRGYGGLGNQGAGR GLGGQGAGAAAAAAGGAGQGGYGLGNQGAGRGGQGAA AAAGGAGQGGYGLGSQGAGRGGQGAGAAAAAVGAGQE GIRGQGAGQGGYGLGSQGAGRGGQGAGAAAAAAGGAG GQGGLGGQGAGQGAGAAAAAAGGVRQGGYGLGSQGAGR GGQGAGAAAAAAGGAGQGGYGLGGQGAGVGRGGLGGQGAG AAAAGGAGQGGYGGVGVGSGASAASAAAASRLSSPQASSR NLVATGPTNSAALSSTISNVVSQIGASNPGLSGCDVLIQAL VSALIQILGSSSIGQVNYGSAGQATQIVQSVYQALG	[42-48]
rMaSp2			PGGYGPGQQGPQGGYGPQQGPSPGPGSAAAAAAAAAGPG GPGQQGPQGGYGPQQGPGRYGPQQGPSPGPGSAAAAAAGSG QQGPQGGYGPQQGPQGGYGPQQGPSPGPGSAAAASAAASA GQQGPQGGYGPQQGPQGGYGPQQGPSPGPGSAAAASAA AAAAAAASGPQQGPQGGYGPQQGPQGGYGPQQGPSPG AAAAAAASGPQQGPQGGYGPQQGPQGGYGPQQGPSPG AAAAAAAGPGQQGPQGGYGPQQGPSPGGSAAAAAAAAAGPG	

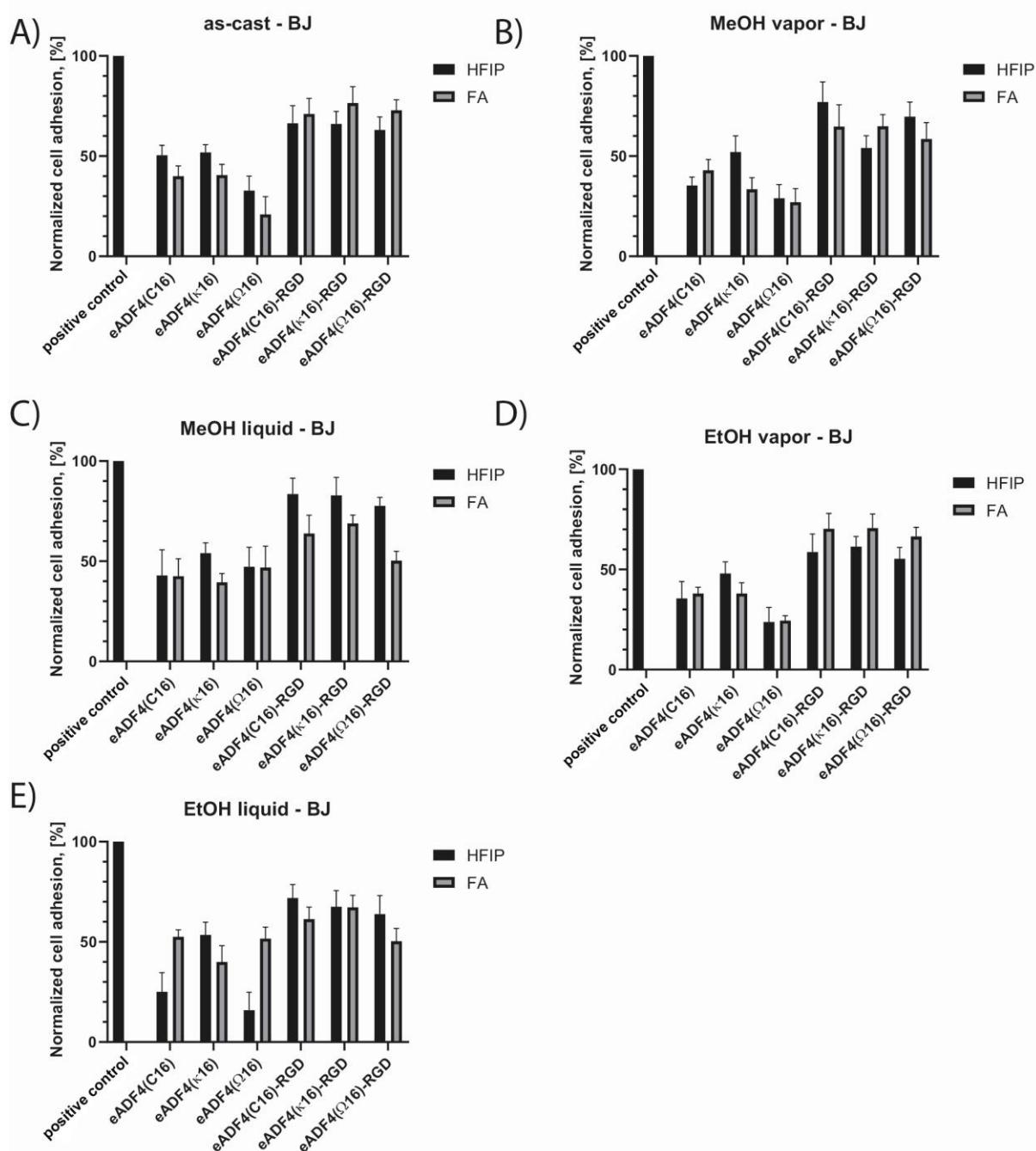
---

			GYGPGQQGPQGGYGPQQGPSAGSAAAAAAAGPGQQGLGG YGPQQGPQGGYGPQQGPSAGSAAAAAAAGPGQQGPQGP GYGPGQQGPSPGSASAAAAAAAGPGGYGPQQGPQGGYAP QQQGPSPGSASAAAAAAAGPGGYGPQGPQGPQGGYAPGQQ GPSPGSASAAAAAAAGPGGYGPQGPQGPQGGYAPGQQ GGYGPQGPAGYGPQSAVAASAGAGSAGYGPQSQASAAAS RLASPDGARVASAVSNLVSSGPTSSAALSSVISNAVSQIGASNP GLSGCDVLIQALLEIVSACVTILSSSIGQVNYGAASQFAQVVVGQ SVLSAF	
FLYS, FLYS <sub>3</sub> FLYS <sub>4</sub>	<i>Trichonephila</i> <i>clavipes</i>	<i>E. coli</i>	Module repeated up to 4 times	[44,47]
1-mer until 192-mer	<i>Trichonephila</i> <i>clavipes</i>	<i>E. coli</i>	1 mer module repeated up to 96mer SGRGGLGGQGAGAAAAGGAGQGGYGGGLGSQGT + cell adhesion peptides (e.g. RGD) + antimicrobial peptides + biomineralization peptides + cancer treating peptides	[49-72]



**Figure S1.** Scheme of the sample matrix of experimental settings used to analyze how the spider silk protein sequence, the casting solvent and the post-treatment influence cell adhesion. The recombinant ADF4 variants (eADF4(C16), eADF4(κ16), eADF4(Ω16), eADF4(C16)-RGD, eADF4(κ16)-RGD and eADF4(Ω16)-RGD) were either solved in FA or HFIP and cast on ozone-pre-treated PS surfaces. Furthermore, coatings were differently post-treated (MeOH vapor or liquid, EtOH vapor or liquid) and compared to non-post-treated films regarding adhesion of human BJ skin fibroblasts and human MG63 bone fibroblasts. The results thereof are summarized in Figure 6, S2 and S3.





**Figure S3.** Cell adhesion on films made of BJ of eADF4 variants (A) as cast or post-treated with (B) MeOH vapor, (C) MeOH liquid, (D) EtOH vapor, (E) EtOH liquid.

## References

- Lentz, S.; Trossmann, V.T.; Borkner, C.B.; Beyersdorfer, V.; Rottmar, M.; Scheibel, T. Structure–Property Relationship Based on the Amino Acid Composition of Recombinant Spider Silk Proteins for Potential Biomedical Applications. *ACS Applied Materials & Interfaces* **2022**, *14*, 31751–31766, doi:10.1021/acsami.2c09590.
- Kumari, S.; Lang, G.; DeSimone, E.; Spengler, C.; Trossmann, V.T.; Lücker, S.; Hudel, M.; Jacobs, K.; Krämer, N.; Scheibel, T. Engineered spider silk-based 2D and 3D materials prevent microbial infestation. *Materials Today* **2020**, *41*, 21–33, doi:<https://doi.org/10.1016/j.mattod.2020.06.009>.
- Esser, T.U.; Trossmann, V.T.; Lentz, S.; Engel, F.B.; Scheibel, T. Designing of spider silk proteins for human induced pluripotent stem cell-based cardiac tissue engineering. *Materials Today Bio* **2021**, *11*, 100114, doi:<https://doi.org/10.1016/j.mtbiobio.2021.100114>.
- Trossmann, V.T.; Scheibel, T. Design of Recombinant Spider Silk Proteins for Cell Type Specific Binding. *Advanced Healthcare Materials* **2023**, *12*, 2202660, doi:<https://doi.org/10.1002/adhm.202202660>.







65. Gomes, S.; Gallego-Llamas, J.; Leonor, I.B.; Mano, J.F.; Reis, R.L.; Kaplan, D.L. In Vivo Biological Responses to Silk Proteins Functionalized with Bone Sialoprotein. *Macromolecular Bioscience* **2013**, *13*, 444-454, doi:<https://doi.org/10.1002/mabi.201200372>.
66. Gomes, S.; Leonor, I.B.; Mano, J.F.; Reis, R.L.; Kaplan, D.L. Spider silk–bone sialoprotein fusion proteins for bone tissue engineering. *Soft Matter* **2011**, *7*, 4964-4973, doi:10.1039/C1SM05024A.
67. Numata, K.; Kaplan, D.L. Silk-Based Gene Carriers with Cell Membrane Destabilizing Peptides. *Biomacromolecules* **2010**, *11*, 3189-3195, doi:10.1021/bm101055m.
68. Plowright, R.; Dinjaski, N.; Zhou, S.; Belton, D.J.; Kaplan, D.L.; Perry, C.C. Influence of silk–silica fusion protein design on silica condensation in vitro and cellular calcification. *RSC Advances* **2016**, *6*, 21776-21788, doi:10.1039/C6RA03706B.
69. Belton, D.J.; Mieszawska, A.J.; Currie, H.A.; Kaplan, D.L.; Perry, C.C. Silk-silica composites from genetically engineered chimeric proteins: materials properties correlate with silica condensation rate and colloidal stability of the proteins in aqueous solution. *Langmuir* **2012**, *28*, 4373-4381, doi:10.1021/la205084z.
70. Zhou, S.; Huang, W.; Belton, D.J.; Simmons, L.O.; Perry, C.C.; Wang, X.; Kaplan, D.L. Control of silicification by genetically engineered fusion proteins: silk-silica binding peptides. *Acta Biomater* **2015**, *15*, 173-180, doi:10.1016/j.actbio.2014.10.040.
71. Numata, K.; Subramanian, B.; Currie, H.A.; Kaplan, D.L. Bioengineered silk protein-based gene delivery systems. *Biomaterials* **2009**, *30*, 5775-5784, doi:10.1016/j.biomaterials.2009.06.028.
72. Mieszawska, A.J.; Nadkarni, L.D.; Perry, C.C.; Kaplan, D.L. Nanoscale control of silica particle formation via silk-silica fusion proteins for bone regeneration. *Chem Mater* **2010**, *22*, 5780-5785, doi:10.1021/cm101940u.