

Supplementary material

Antibacterial electrodeposited copper-doped calcium phosphate coatings for dental implants

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1 – CaP coating on dental implant by electrodeposition

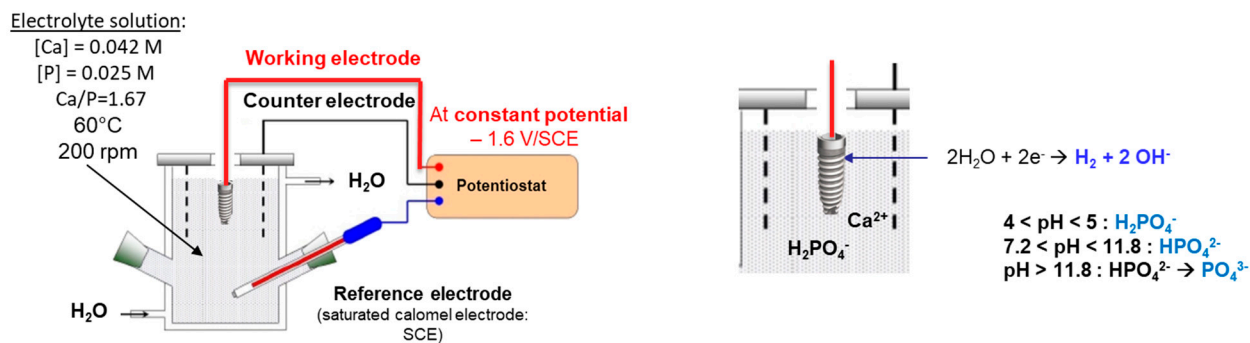


Figure S1: Experimental set for CaP electrodeposition on dental implant: a) General overview and, b) Focus on the dental implant placed at the cathode and the reaction of reduction of water occurring at the surface.

2 - FTIR-ATR analysis of CaP reference compounds and CaP coated samples

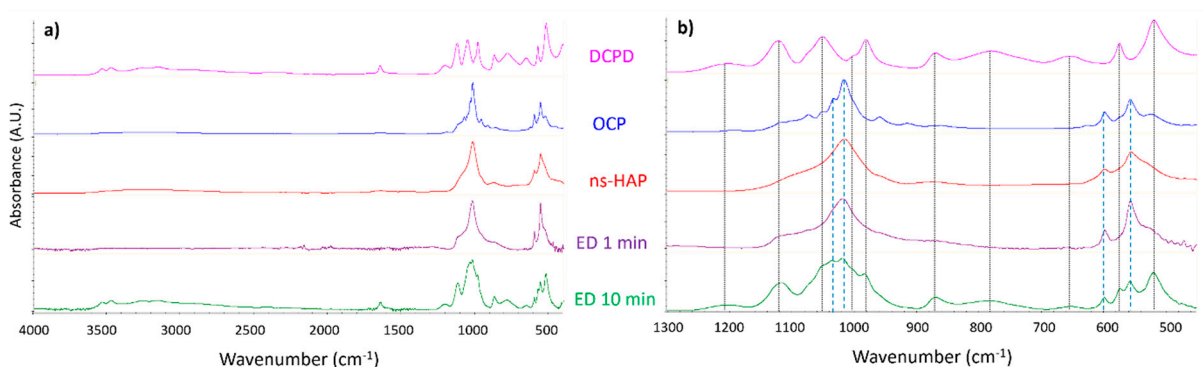


Figure S2: FTIR-ATR spectra of DCPD, OCP, ns-HAP reference compounds and CaP coated samples after 1 and 10 minutes of electrodeposition : a) Full spectrum ($4000\text{--}400\text{ cm}^{-1}$) and b) $1300\text{--}450\text{ cm}^{-1}$ domain.

3 – SEM-EDX analysis of Cu-doped CaP coated titanium

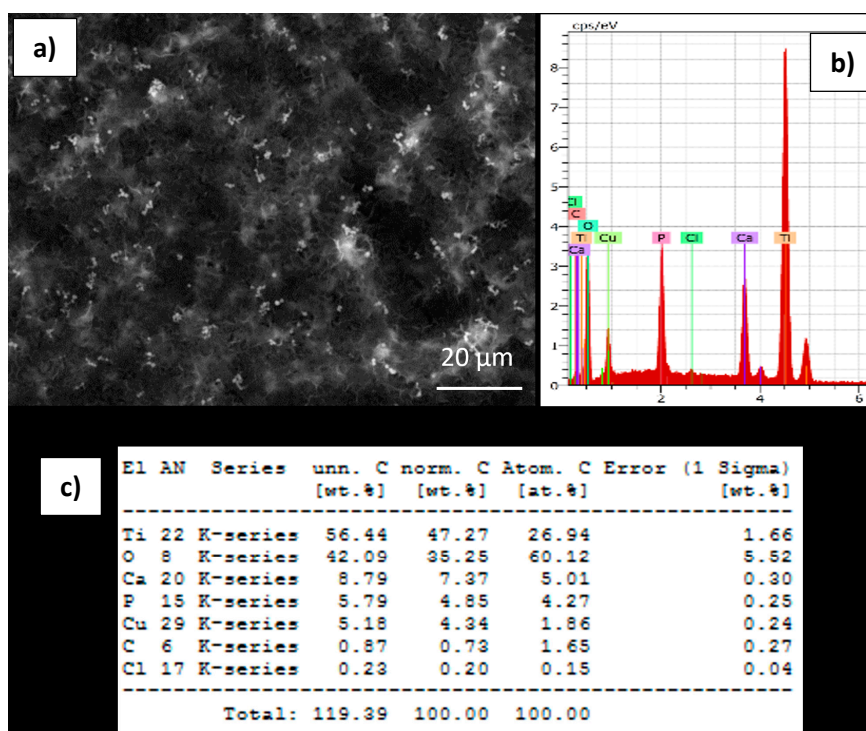


Figure S3: SEM micrograph (a) of a 1 min electrodeposited CaP coating doped with an ion exchange post-treatment using a copper ion solution of 0.001 M, its corresponding EDX spectrum (b) and elemental analysis report (c).

4 - Cytotoxicity on mammalian cells

Evaluation of CaP coated titanium coupons cytotoxicity was performed using the MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) colorimetric assay based on Mosmann *et al.* procedure [1] , according to the EN/ISO 10993-5 standard (tests for *in vitro* cytotoxicity) for direct contact test. The potential cytotoxic activity of the copper doped CaP-coated coupons (after ionic exchange in 0.005, 0.001 and 0.01 M copper nitrate solution) versus non-doped CaP-coated coupons was assessed against Vero cells (ATCC® CCL-81) purchased from ATCC® (Manassas, United-States) and cultivated in RPMI medium (RPMI 1640 – PAN-Biotech). 96-wells microtiter plates were first filled with 100 μL of a cell suspension prepared in RPMI medium (2×10^4 cells/100 μL). After overnight incubation at 37 °C in a humidified 5-6.5% CO₂ incubator, the cell confluence was checked by microscopic observation. Then, the medium was replaced by fresh medium and the coupons were placed gently on the cell layer in the centre of the well. Wells corresponding to the negative control were supplemented

with 100 μ L of fresh medium. Tween 40 (5%, Sigma, St. Quentin Fallavier, France) was used as positive control for cytotoxicity. The microplate was then incubated for 24 h at 37 °C in a humidified 5-6.5% CO₂ incubator. The supernatant was then discarded followed by a rinsing with 100 μ L of phosphate buffer solution (PBS). Then 100 μ L of a MTT solution prepared in PBS at a concentration of 0.5 mg/mL were added in all wells. After 60 min of incubation at 37 °C and in order to solubilize the formed formazan, indicator of cell viability, 100 μ L of DMSO were added. After agitation, the optical density (OD) was measured at 570 nm using a CLARIOstar Plus plate reader (BMG Labtech). Finally, viability percentages were determined by measuring the OD at 570 nm for each Cu-doped CaP-coated coupons, in comparison to the non-doped ones. Assays were performed in triplicate during a single test. The results are presented in Figure S3.

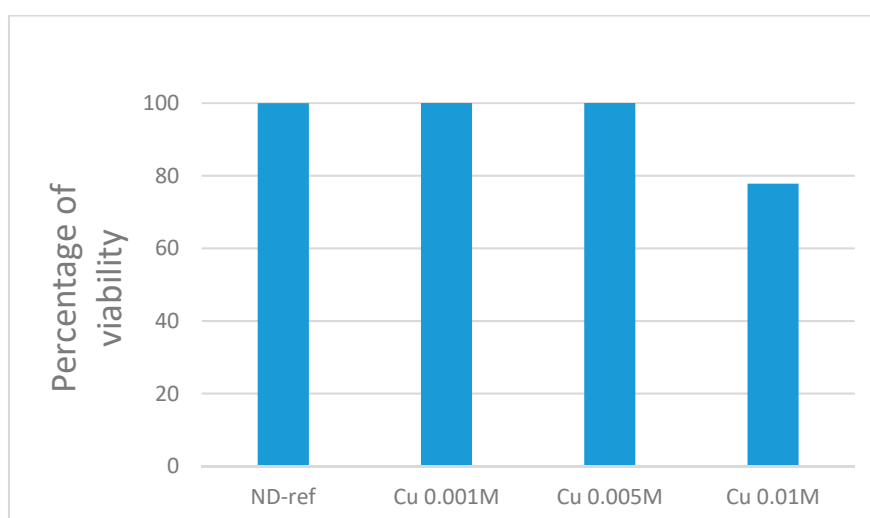


Figure S4: Percentage of viability of Vero cells after direct contact with non-doped CaP coated samples (ND ref) and Cu-doped CaP coated samples after exchange in 0.001, 0.005 or 0.01 M copper ions solutions.

Reference:

[1] Mosmann, T. Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays. *Journal of Immunological Methods* **1983**, 65, 55-63, D.O.I.: [https://doi.org/10.1016/0022-1759\(83\)90303-4](https://doi.org/10.1016/0022-1759(83)90303-4).