

Article

Depth Profiles of Microplastics in Sediment Cores from Two Mangrove Forests in Northern Vietnam

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Abstract: Plastics are essential materials that play critical roles in people's lives in the modern world. However, plastic pollution in the ocean has become a critical environmental problem due to the fact of its durability and long life span. In the present study, we analyze the contamination of microplastics in sediment cores from mangrove forests of the Red River Delta and Tien Yen Bay in Northern Vietnam. The results showed that the concentration of microplastics in sediment cores ranged from 0 to 4941 particles/kg, with four types of microplastics: microfiber, microfragment, microfoam, and microfilm. Microplastics were not observed in sediment samples lower than 65 and 70 cm core depth in sediment cores from the Tien Yen Bay and Red River Delta. The microplastic concentration in sediment cores from the Red River Delta was significantly higher than those of Tien Yen Bay, reflecting the influence of anthropogenic activities on microplastics pollution. The present results highlight that the accumulation of microplastics may have occurred a long time ago and tends to increase in the future. Further investigation on microplastic accumulation rates, sources of microplastics, and reducing plastic pollution is necessary to minimize the impacts of microplastic pollution on aquatic life and the environment.

Keywords: microplastics; mangrove sediment; Red River Delta; Tien Yen Bay



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1. Introduction

Plastic products have been produced and widely used since the 1940s, playing important roles in many economic sectors and people's lives [1]. However, with the increasing demand for plastic products (estimated to be up to 12 billion tons by 2025 [2]), plastic wastes are increasingly affecting the environment, especially coastal ecosystems [2]. Every year, the ocean receives approximately 4.8–12.7 million tons of plastic waste from fishing activities, tourism, and especially the increasing amount of domestic and industrial wastes [3]. Most plastic components are difficult to biodegrade, and they are broken into smaller pieces by physical factors, such as UV rays and waves, and deposited in coral reefs and sediments. Pieces of plastic < 5 mm in size are called microplastics [4,5]. In recent years, large amounts of plastic waste discharged into the environment, causing environmental quality degradation and negatively affecting the coastal ecosystems [6]. Microplastics are classified into primary and secondary microplastics [7]. Primary microplastics are produced for the plastics industry with specific shapes and sizes and are usually spherical. Secondary microplastics are products of plastic waste during the decomposition process in the environment [7]. Microplastics are widespread in coastal and ocean environments, and they can float on the ocean's surface or settle in sediments throughout the world's oceans. Microplastics are widely distributed from coastal to offshore environments, even in

Arctic Sea regions [8–10]. Studies on microplastics accumulation in sediments from coastal zones of China showed that large estuaries in this country are highly contaminated by microplastic wastes [11–15]. Recent studies have recorded the appearance of microplastics in the bodies of many fish, turtles, seabirds [16], plankton [17], and the digestive system of bivalves [18]. Microplastics accumulated in the organism can be enriched in the food chain, affecting many other species including humans. Moreover, microplastics can adsorb organic pollutants, heavy metals, and PCBs, causing harmful effects on various animals and ecosystems [19–21].

Research on microplastics pollution has expanded worldwide, focusing on the distribution and characteristics of microplastics in coastal marine environments. Unfortunately, there are few studies on microplastics pollution in developing countries, the largest source of untreated plastic waste discharges into the ocean. Vietnam is considered a significant source of marine plastic wastes (0.28–0.73 MMT/year) dumped into the ocean [3]. However, microplastics accumulation in sediment cores is rarely observed, particularly in Vietnam and Southeast Asia. A lack of information on plastic pollution makes government efforts to reduce plastic waste hard to implement in the future. In the present study, we analyzed microplastics in five mangrove sediment cores to understand the current status and temporal variations of microplastics pollution in Northern Vietnam. The research results will play essential roles in reducing microplastics pollution in coastal areas, particularly in Vietnam and Asia–Pacific regions.

2. Materials and Methods

2.1. Study Area

The present study was conducted in two mangrove forests of Northern Vietnam including the Red River Delta and Tien Yen Bay (Figure 1). The first sampling location in the Red River Delta belongs in the Tien Hai District, Thai Binh Province, Vietnam, with the dominant plantation mangrove species being *Kandelia obovata* and *Sonneratia caseolaris*. In Tien Yen Bay, the dominated mangrove species are *Bruguiera gymnorrhiza*, *Avicennia marina*, *Kandelia obovata*, and *Rhizophora stylosa* (Table 1). The true mangrove species, *Aegiceras corniculatum*, is widely distributed in both Tien Yen Bay and the Red River Delta and grows near tidal creeks and estuaries. The mangrove forests in the Red River Delta have been replanted since the 1990s with the dominant planted species being *K. obovata* and *S. caseolaris*, which play essential roles in coastal protection, pollutant filtration, and providing habitats for aquatic organisms. Mangrove forests and surrounding areas in the Red River Delta are vital stopovers and breeding sites for migratory birds and waterfowls. Mangrove forests in Tien Yen Bay are natural forests in northeastern Vietnam that play essential roles in biodiversity conservation and provide habitats for animals. All sampling sites are located in the monsoon climate zone, with the dry season from November to April and the rainy season from May to October. The Red River Estuary receives large amounts of pollutants from large cities and industrial areas in Northern Vietnam [22]. Mangroves in this area play an important role as a “natural filter” for pollutants, creating favorable conditions for microplastics accumulation. Tien Yen Bay is a semi-closed Bay with weak urbanization and industrial activities [23]. However, the fishing activities in Tien Yen Bay are highly concentrated in this area, with the main seafood products being peanut worms, clam, shrimps, and sea groupers.

2.2. Field Sampling

The sediment cores were collected by a peat sampler (Eijkelkamp Soil & Water, The Netherlands) up to 1 m in depth at five sampling sites from the Red River Delta and Tien Yen Bay, Vietnam (Figure 1). The samples were immediately transferred in a cleaned PVC tube, wrapped with aluminum foils, and stored in a cool box until further analysis. The sediment samples were processed within 24 h of collection in a field laboratory near sampling sites. In the field laboratory, the outer layers of sediment cores (~0.5 cm in thickness) were removed and then sliced with a stainless-steel knife for reducing any

contamination of plastic particles. The slicing intervals were 5 and 10 cm for the depths of 0–50 cm and 50–100 cm in sediment cores from the Red River Delta, respectively. In Tien Yen Bay, due to the high density of mangrove roots, the slicing intervals were modified to 10 and 15 cm for the depths of 0–50 cm and 50–100 cm, respectively. All sliced samples were stored in washed polyethylene bags (three times washed with distilled water) on a cool box and then transported to the laboratory for further processing. The subsamples of mangrove sediment (approximately 5 g in weight) were also collected to analyze sediment grain size.

Table 1. The sampling sites’ description in two mangrove forests in Northern Vietnam.

Sample Site	Location	Type of Mangrove Forest	Dominant Mangrove Species	Sedimentation Rate (cm/Year)
TH-01	Red River Delta	Re-plantation mangrove forest	<i>Aegiceras corniculatum</i> , <i>Kandeliaobovata</i> , and <i>Sonneratia caseolaris</i>	1.2 cm [22]
TH-02	Red River Delta	Re-plantation mangrove forest	<i>Aegiceras corniculatum</i> and <i>Kandeliaobovata</i>	1.2 cm [22]
TH-03	Red River Delta	Re-plantation mangrove forest	<i>Aegiceras corniculatum</i> , <i>Kandeliaobovata</i> , and <i>Sonneratia caseolaris</i>	1.2 cm [22]
TY-01	Tien Yen Bay	Natural mangrove forest	<i>Bruguiera gymnorrhiza</i> , <i>Kandelia obovata</i> , <i>Avicenia marina</i> , and <i>Rhizophora stylosa</i>	0.82 cm [24]
TY-02	Tien Yen Bay	Natural mangrove forest	<i>Avicenia marina</i> , <i>Kandelia obovata</i> , and <i>Rhizophora stylosa</i>	0.82 cm [24]

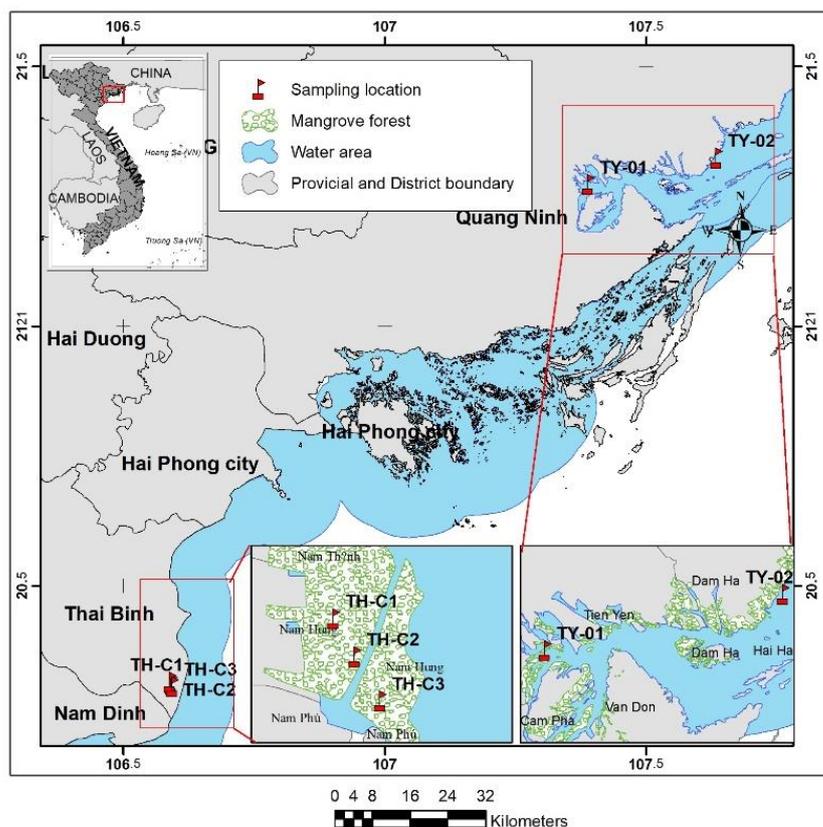


Figure 1. Map of the sampling locations in Northern Vietnam.

2.3. Sample Analysis

2.3.1. Sediment Grain Size Analysis

The sediment grain size was analyzed by laser diffraction methods using the Horiba Partica LA-950 system. Before the analysis, all visible organic fragments in the samples were removed by hand using stainless-steel tweezers. Approximately 2 g of fresh sediment sample was placed in a ceramic cup, and then we added 10 mL of hydroperoxide 10% solution for organic matter oxidation in 24 h. The organic matter removal process was repeated two or three times until all organic materials were decomposed. Due to the high clay and silt particle concentration in mangrove sediment samples, we added 2 mL of sodium hexametaphosphate 5% solution to each ceramic cup to reduce sediment flocculation [25,26]. All sediment samples in the ceramic cup were transferred in an ultrasonic bath for sonication 10 min before analysis. The sediment grain size was reported as the median size and grain size fractions (i.e., sand, silt, and clay contents). The number of samples was 45 and 18 for the Red River Delta and Tien Yen Bay, respectively.

2.3.2. Microplastics Analysis

The microplastics analysis method in the sediment cores followed US National Oceanic and Atmospheric Administration (NOAA)'s practices with a slightly modified procedure [21]. Briefly, approximately 50 g of fresh sediment samples were dried at 40 °C for 48 h or until constant weight. The dried sample was placed in a beaker, and we slowly added 20–50 mL of a 30% peroxide solution. Then, approximately 10 mL of catalyst solution (0.05M FeSO₄ solution) was added to each beaker to enhance the oxidation rates of organic matter. The nearshore environment is the habitat of many benthic species with a high density of shell debris in the sediment. For samples with a lot of shell debris, approximately 10–20 mL of 1 M HCl solution was added to remove the carbonate component within 12 h (or until bubbles stopped appearing). The sample was sieved using a 0.3 mm test sieve (20 cm in diameter) to remove small sediment particles, and the residue on the sieve was washed into a beaker before being transferred to an electric oven to remove water at 40 °C in 48 h.

After removing organic matter, carbonate, and water, the samples were placed in 50 mL centrifuge tubes and filled with 1.6 g/L ZnCl₂ solutions. The mixture was centrifuged at high speed (3000 RCF for 5 min) 3 times to separate the microplastics (having low-density and floating on top of each tube). The microplastic particles floating on top were filtered through a filter membrane (47 mm diameter grid filter, pore size of 0.7 µm) using a Nagalene vacuum filter holder. During filtration, Milli-Q water was added to ensure no residual ZnCl₂ on the filter membrane. After filtration, all filter membranes were stored in aluminum foil bags, then dried at 45 °C to a constant weight. A stereomicroscope coupled with a digital camera (Euromex DC.5000) was used to determine microplastic particles in the sediments. The types of microplastics were identified according to the guidelines of the NOAA [4,19].

All reagents and solutions were firstly filtered through 47 mm Whatman GF/F filters and then stored in a glass beaker before analysis to reduce the contaminants in the experiment. Aluminum foils covered all beakers for microplastics analysis during the experimental processes. For each sediment core, blank samples were performed during analysis to measure background contamination. If the contaminants were detected in a blank sample, the total microplastic in a batch of samples was substrated by the number of contaminated plastic particles. We found that two blank procedural samples were contaminated with 2 particles of microfibers during the experiment processes.

2.3.3. Chemical Characteristics of Microplastic by FTIR Analysis

Due to the small size and super lightweight of microplastics in sediment samples, the selected particles of microplastics were collected from filter paper to determine the chemical composition. We merged selected plastic particles in each sediment core before microplastics chemical identification by the FTIR method. Microplastic particles were finely

ground with the KBr substrate using an agate mortar and pestle, then compressed by a compressor before being analyzed by the Jasco FTIR 4600 system at the University of Science, Vietnam National University, Hanoi. The FTIR analysis spectrum of the microplastics sample was compared and classified according to the chemical composition published by Gerrit Renner [27].

2.3.4. Statistical Analysis

Principal component analysis (PCA) was used to explain the relationship between the concentration of microplastics and sediment characteristics (Md, sand, silt, and clay contents). Pearson correlation was also applied to interpret the connection among the number of microplastics and sediment characteristics. The statistical difference of microplastics distribution in depth and the location was identified by non-parametric Kruskal–Wallis test. Statistical difference between groups was recorded when $p < 0.05$. All statistical tests were performed by IBM SPSS v20.0.

3. Results

3.1. Sediment Grain Size Distribution in Mangrove Forests

The sediment samples from Tien yen Bay and the Red River Delta had high concentrations of silt and clay and varied along with the depth of the sediment cores. The Md of the sediment particles ranged from 11.55 to 45.66 μm and 7.06 to 13.1 μm in Tien Yen Bay and the Red River Delta, respectively. The Md of mangrove sediment in Tien yen Bay was significantly higher than those of the Red River Delta, and a similar trend was observed for sand content. However, there was no statistical difference between silt concentrations in sediment among sampling locations. Md value tends to be stable along with sediment depth, except core TY-01 showed an increasing trend from top to bottom of the core. The sediment grain size distribution indicated that the main sediment types in sampling locations were silty and sandy silt sediment for Tien Yen Bay and the Red River Delta, respectively.

3.2. Microplastic Abundance in Sediment Core

3.2.1. Microplastic Types in Mangrove Sediment Cores

In the present study, we observed four types of microplastic in mangrove sediments including microfiber, microfoam, microfragment, and microfilm. All types of microplastics in two sampling locations are secondary plastics directly related to plastic wastes from inland and coastal areas. We did not focus on determining the color and size of each microplastic particle in the samples. However, the main colors of microplastics were mainly transparent, white, clear, blue, and green. We also recorded that most microplastic particles were smaller than 1 mm in all sediment cores. Microfiber was the most popular microplastic type in all sediment cores, ranging from 0 to 4253 particles/kg in two sampling locations. The microfiber concentration from 0 to 50 cm in depth in sediment cores from Tien Yen Bay was statistically lower than those of the Red River Delta (Kruskal–Wallis, $p < 0.05$). A similar trend was detected in microfragment concentration, with the variation in microfragment ranging from 0 to 79 and 0 to 439 particles/kg for Tien Yen Bay and the Red River Delta, respectively. However, a location difference for microplastic types was not observed in sediment cores from 50 to 10 cm in depth (Kruskal–Wallis, $p > 0.05$, Table 2). The correlation between microplastic types and sediment characteristics was also not observed in the present study. Based on previous studies on sedimentation rates, we estimated that the microplastics existed in the sediment stratum from 1935 to 1959 and from 1953 to 1962 for Tien Yen Bay and the Red River Delta, respectively (Table 1, Figures 2 and 3).

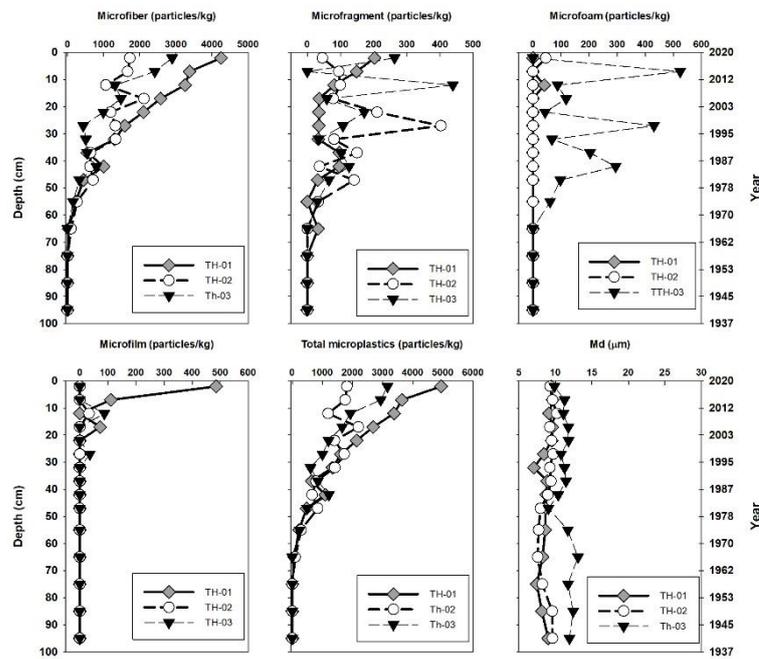


Figure 2. The characteristics of microplastics and sediment grain size in the Red River Delta. Data for the year axis were calculated from [22].

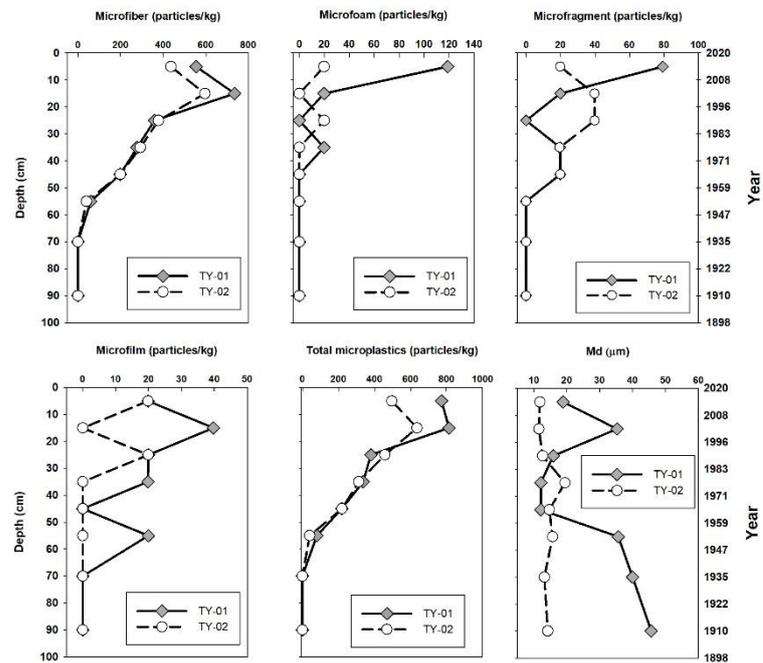


Figure 3. The characteristics of microplastics and sediment grain size in Tien Yen Bay. Data for the year axis were calculated from [24].

3.2.2. Total Microplastics in Mangrove Sediment Cores

The total microplastics in sediment cores ranged from 0 to 4941 and from 0 to 815 particles/kg for the Red River Delta and Tien Yen Bay, respectively. In all sampling locations, the total microplastics decreased from the top to the bottom of each core (Figures 2 and 3). The total microplastics in 0–50 cm deep sediment cores from Tien Yen Bay was significantly lower than those of the Red River Delta (Kruskal–Wallis, $p < 0.05$). However, a similar trend was not observed for sediment samples from 50 to 100 cm in depth (Table 2). The microplastics in sediment samples from Tien Yen Bay were detected in the sediment layer from 50

to 65 cm, corresponding to the 1930–1950s [24]. In the Red River Delta, the microplastics were not detected in the sediment layer from 70 to 80 cm in depth, corresponding to the 1940–1960s [22]. These results suggested that microplastics have been present in mangrove sediments from Northern Vietnam since the 1950s.

Table 2. The Kruskal–Wallis test results between two sampling locations in Northern Vietnam.

Depth Zone		Md	Sand	Silt	Clay	MFB	MFA	MFO	MFI	MPs
0–50 cm	Chi-Square	21.370	19.142	3.750	21.370	15.244	13.016	0.000	2.874	17.518
	<i>p</i>	<0.001	<0.001	0.053	<0.001	<0.001	<0.001	0.986	0.090	<0.001
50–100 cm	Chi-Square	12.273	12.353	12.273	12.273	0.077	1.324	0.400	2.500	0.138
	<i>p</i>	<0.001	<0.001	<0.001	0.000	0.781	0.250	0.527	0.114	0.711

MFB, MFA, MFO, MFI, and MPs denote microfiber, microfragment, microfoam, microfilm, and total microplastics, respectively.

The PCA analysis was applied to determine the relationship between microplastics concentration and sediment characteristics (Md, sand, silt, and clay contents). The first factor accounted for 72.94% and 61.64% of the variance in Tien Yen Bay and the Red River Delta, respectively. The high loading of the first factors was observed in MFB, MFA, MFO, MFI, and MPs from both Tien Yen Bay and the Red River Delta, which indicated that all microplastic types might originate from the same sources (Figure 4). The second factor accounted for 32.15% and 26.26% of the variance in Tien Yen Bay and the Red River Delta, respectively. High loading factors of Md and sand content may relate to the grain size distribution of each sediment sample. Overall, the relationship between sediment characteristics and microplastics was not clearly observed in the present study.

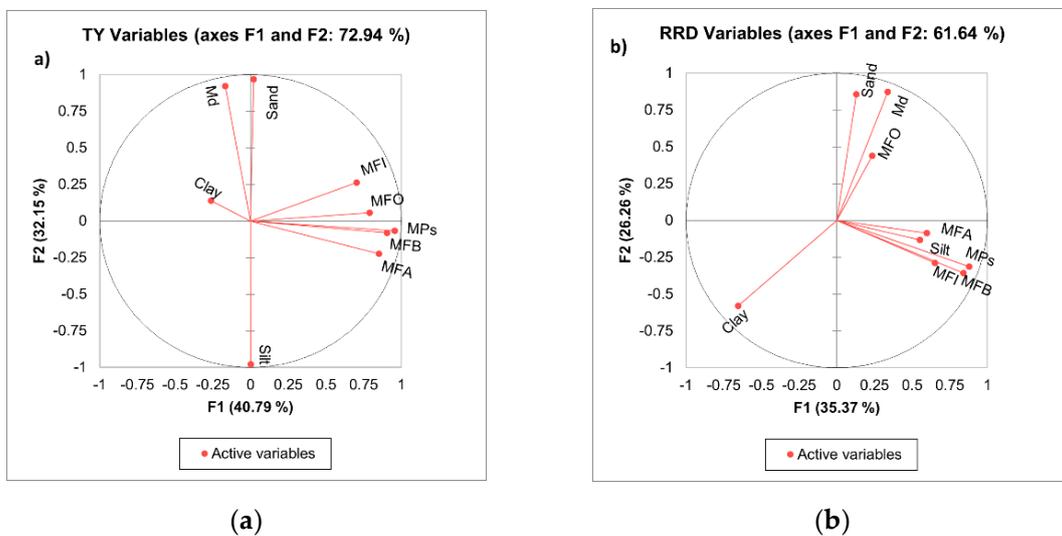


Figure 4. The PCA analysis among microplastics composition and sediment grain size: (a) all samples in Tien Yen Bay (*n* = 18); (b) all samples in the Red River Delta (*n* = 45). MFB, MFA, MFO, MFI, and MPs denote microfiber, microfragment, microfoam, microfilm, and total microplastics.

3.3. Microplastic Polymer Type in Sediment Core

The results from the FTIR analysis of polymer type in microplastics samples in the Red River Delta and Tien Yen Bay were polyethylene (PE), polypropylene (PP), polystyrene (PS), polyethylene terephthalates (PET), polyamide (PA), and polyester (PLE). Popular plastics, such as polyvinyl chloride (PVC), acrylic, and polyurethane (PU), were not observed in all sediment cores from this area. However, the small microfibers or weathered particles in the samples may not be identifiable using the FTIR method. Microplastics’ chemical characteristics indicated that the most popular plastics were found in sediment cores from mangrove forests in Northern Vietnam.

4. Discussion

The present study showed that the concentration of microplastics in sediment from Northern Vietnam was significantly higher than those of other wetland types in Southeast Asia and China [28–30] and equivalent to the concentration of microplastics in estuaries in China [11,31] (Table 3). The decreasing trend in microplastics abundance in sediment cores was similar to previous studies in Asia and Africa [12,29]. The vertical distribution of microplastics may relate to the accumulation of plastic wastes in Northern Vietnam since the 1950s. Most microplastics were observed from a 0 to 30 cm depth, and microfiber was the dominant plastic type in all sampling areas. Microplastic size in the study areas was mainly smaller than 1 mm, similar to previous reports in the Australian estuary and Asia–Africa regions [29,32]. The small-sized microplastics in the study areas resulted from the breakdown of plastics in the sea environment [15] and the low dynamic environment in mangrove forests. A study in mangrove forests of Singapore reported that finer sediments have a high concentration of microplastics due to the microplastics retention in flocculation processes [28]. Overall, a clear relationship between sediment grain size and microplastics abundance was not observed in mangrove forests in Northern Vietnam.

The lower concentration of microplastics in sediment from Tien Yen Bay compared to the Red River Delta may be related to anthropogenic activities in Northern Vietnam. The sampling site in the Red River Delta was located in the largest estuary in Northern Vietnam, which receives a large amount of untreated plastic wastes from the mainland. The increasing trend of microplastics in sediment cores from the Red River Delta from the 1990s to the present may relate to the economic development and growth of plastics consumption in Vietnam. A similar trend was also observed in Tien Yen Bay, with the concentration of microplastics increasing from the 1980s to the present (Figure 3). Recent research in the East China Sea indicated that microplastics concentration in sediment cores was directly related to plastic consumption and production [12], and a similar trend was observed in Northern Vietnam. The other microplastic sources may relate to cross-border transportation of plastic wastes in the ocean environment, which brings floating plastic wastes from developed countries in the Asia–Pacific region to the study areas [33]. Therefore, an international framework in the Asia–Pacific region for reducing marine debris and microplastics pollution is necessary to be implemented for enhancing environmental protection and conservation [34].

Table 3. The microplastics distribution in mangrove and wetland sediments from this study and selected regions.

Location	Type of Wetlands	Plastic Particle Size (mm)	Total Microplastics (Particles/kg)	Data Sources
Tien Yen Bay, Vietnam	Mangrove forest	0.3–5	0–815	This study
Red River Delta, Vietnam	Mangrove forest	0.3–5	0–4941	This study
Thanh Hoa, Vietnam	Tidal flats	0.25–5	2921–5365	[35]
Singapore	Mangrove forest	0.02–5	12–62.8	[28]
Gulf of Thailand	Bay	0.3–5	83–165	[29]
Andong salt marsh,	Saltmarsh	0.1–5	40–480	[12]
Hangzhou Bay, China				
Hong Kong	Bay	0.1–5	44–458	[36]
Changjiang Estuary, China	Estuary and tidal flats	<5	20–340	[37]
Sishili Bay,	Estuary and bay	<5	140–1873	[30]
North Yellow Sea, China				
Sanggou Bay, China	Bay and estuary	<5	699–2824	[14]
Pearl River Estuary, South China	Mangrove forest	<5	100–7900	[31]
Brisbane River, Australia	River	<5	10–520	[38]
Southeast of China	Mangrove forest	0.05–5	8.3–5738.3	[13]
Tampabay Bay, Florida	Bay	<5	30–790	[39]

Table 3. Cont.

Location	Type of Wetlands	Plastic Particle Size (mm)	Total Microplastics (Particles/kg)	Data Sources
Guanabara Bay, Brazilian	Bay	0.1–5	160–1000	[40]
Jinjang Enstuary, China	Estuary	<5	980–2340	[11]
Jagir Estuary, Indonesida	Estuary	0.3–5	92–590	[41]
Liaohe Estuary, china	Estuary	<5	80–220	[42]
Xiangshan Bay, China	Bay	<5	1739–2153	[43]
South Yellow Sea, China	Sea	<5	560–4205	[15]
Pearl River, China	Estuary	1–5	258	[44]

Both natural conditions and anthropogenic activities have also influenced the spatial distribution of microplastics sediment cores. Tien Yen Bay is a closed bay with a water current from the North and exchanging water with the Gulf of Tonkin through two small channels [45], indicating that sources of microplastics may relate to local anthropogenic activities. Additionally, microplastic types in sediment cores are secondary plastics, mainly originating from untreated plastics wastes discharged to the coastal marine environment. The chemical composition of microplastics in sediment (i.e., PE, PP, PS, PET, PA, and polyester) suggested that the sources of microplastics come from households and aquaculture products such as fishnet, clothes, styrofoam, bottles, and others. The spatial variation of microplastics in the Red River Delta also supported this assumption, with a high concentration of microplastics observed in sediment cores near dikes and aquaculture zones. Therefore, the concentration of microplastics has a positive relationship with anthropogenic activities.

The vertical abundance of microplastics in sediment core is influenced by sedimentation rates, anthropogenic activities, and other natural processes. The sedimentation rates are influenced by mangrove standings and the tidal dynamic in mangrove forests [22,46,47]. These processes may be affecting the accumulation of microplastics in sediment cores. In mangrove forests, the bioturbation of mangrove crabs has changed sediment properties [48], and microplastics in surface sediment can be transferred into deeper layers through crab burrows on the forest floor. Furthermore, the anthropogenic activities of local people, such as aquatic organism harvesting in mangrove forests and fishing, may disturb the sediment stratum, leading to microplastic contamination in deeper sediment layers. Based on previous sediment dating data, the microplastics contamination in sediment cores from Northern Vietnam may have occurred from the 1950s to the 1970s. Further analysis, such as fine scales and associated radionuclides methods, such as Pb-210 and Cs-137, is necessary to clarify this process.

5. Conclusions

The present study showed that the mangrove sediments in Northern Vietnam had a high concentration of microplastics, mainly originating from anthropogenic wastes. Microplastics contamination in sediments from Northern Vietnam was similar to microplastics pollution reports in developing countries in the Asia–Pacific region. The depth variation of microplastics indicated that the microplastics might exist in sediment from 40 to 50 years ago. Sedimentation rate analysis using radionuclides methods, such as Pb-210 and Cs-137, is necessary to confirm the temporal changes in microplastics in the environment. The present study is the first observation of microplastics contamination in sediment cores from Vietnam’s mangrove forests and coastal areas. Further investigation toward the identification of the sources of microplastics is necessary to reduce microplastics pollution in Vietnam and Asia-Pacific regions.

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