



Review

Effect of Obesity on Lung Function in the Pediatric and Adult Populations with Asthma: A Review

Nayely Reyes Noriega ¹, Blanca E. Del-Río-Navarro ¹, *, Arturo Berber ¹, Sergio de Jesús Romero Tapia ² and Darío Jorge Mario Molina Díaz ³

- Allergy and Immunology Pediatric Department, Hospital Infantil de México Federico Gómez, Ciudad de México 06720, Mexico; naye.rey.nor@gmail.com (N.R.N.); arturoberber@aol.com (A.B.)
- División Académica de Ciencias de la Salud, Universidad Juárez Autónoma de Tabasco, Villahermosa 86280, Mexico; sergio.romero@ujat.mx
- ³ Pediatric Endocrinology Department, Hospital Infantil de México Federico Gómez, Ciudad de México 06720, Mexico; drjorgemariomolina@gmail.com
- * Correspondence: blancadelrionavarro@gmail.com

Abstract: Obesity and asthma are major global health concerns, particularly in industrialized nations. Obesity has been shown to have detrimental effects on the respiratory system and lung function owing to metabolic issues and immunological consequences. Research has indicated that obese patients with asthma (atopic or T2-high and non-atopic or T2-low) have diminished lung function in terms of functional residual capacity (FRC), residual volume (RV), expiratory reserve volume (ERV), the FEV1/FVC ratio, and FEF 25–75% due to mechanical fat loading on the diaphragm and central adiposity when compared to non-obese asthmatic patients. Therefore, it is plausible that changes in lung function are the result of a combination of mechanical (fat loading on the diaphragm, central adiposity, bronchial hyper-reactivity, and an increase in cholinergic tone), environmental (diet and exercise), and inflammatory factors (local and systemic), which can lead to the obesity-related asthma phenotype characterized by severe asthma symptoms, poor response to corticosteroid treatment, loss of lung function, and poor quality of life from an early age.

Keywords: lung function; obesity; asthma; pediatric population



Citation: Reyes Noriega, N.; Del-Río-Navarro, B.E.; Berber, A.; de Jesús Romero Tapia, S.; Molina Díaz, D.J.M. Effect of Obesity on Lung Function in the Pediatric and Adult Populations with Asthma: A Review. J. Clin. Med. 2023, 12, 5385. https://doi.org/10.3390/jcm12165385

Academic Editor: Ildiko Horvath

Received: 20 July 2023 Revised: 16 August 2023 Accepted: 17 August 2023 Published: 19 August 2023



Copyright: © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).

1. Introduction

Asthma and obesity are chronic, non-transmittable diseases that are widespread in industrialized countries [1]. According to the National Health and Nutrition Examination Survey 2017–March 2020 Prepandemic Data Files Development of Files and Prevalence Estimates for Selected Health Outcomes, between 2017 and 2020, the prevalence of obesity was higher than 20% among children and adolescents, and 40% among adults [2]. This condition is characterized by excessive fat accumulation caused by a person's genetic inclination towards overeating from an early age, along with the distribution of fatty tissue (which varies depending on the reference population) and the individual's interaction with environmental factors such as the availability of food, healthcare services, and family income [1]. Obesity leads to changes in multiple organs and systems, including the respiratory-system-originating alterations in lung volume, augmented resistance, and gas exchange at the alveolar level [3].

On the other hand, asthma is a heterogeneous disorder characterized by persistent airway inflammation and symptoms such as breathlessness, coughing, chest tightness, and wheezing, which can range in severity and expiratory flow limitation [4]. The Global Asthma Network Phase 1 study reported that the worldwide prevalence of asthma in 6–7-year-old schoolchildren and 13–14-year-old adolescents was 9.0% and 11%, respectively, with the highest prevalence of asthma symptoms reported in low- to middle-income

countries [5]. Similarly, in 2019, the estimated prevalence of asthma in adults was 9.8%, with the African region being most affected [6].

As the severity of this disease is related to the age of symptom onset, patient phenotype, and the presence of metabolic comorbidities, the purpose of this review was to update and describe the clinical effects of obesity and its repercussions on lung function in asthma patients, providing a clinical and practical overview for physicians.

2. The Obesity-Related Asthma Phenotypes

It is generally accepted that obesity can increase the risk of asthma through a variety of mechanisms and is connected to both extrinsic and intrinsic factors that can be linked to the onset of this condition. For example, studies have revealed that maternal obesity and weight gain during pregnancy are linked to an increased probability of asthma symptoms during school age (OR = 1.31: 95% confidence interval [CI] 1.16–1.49) as well as a heightened likelihood of developing obesity during childhood and school age [7,8]. Also, during adolescence, it has been suggested that the effect of obesity on the likelihood of developing asthma is affected by sex hormones, particularly in females with obesity and early menarche [9]. Asthma symptoms in these patients are more intense during the perimenstrual period (also known as perimenstrual asthma) and can result in severe or difficult-to-manage asthma [10] due to decreased progesterone and testosterone levels, as well as fluctuations in estradiol, luteinizing hormone (LH), and follicle-stimulating hormone (FSH) levels, which can trigger mast cell degranulation and result in a heightened respiratory response to aeroallergens [11–13].

Chronic inflammation associated with obesity and the early accumulation of central adipose tissue can result in alterations in respiratory mechanics [14,15] and may be further exacerbated by other metabolic conditions such as hyperlipidemia, hyperinsulinemia, and hypercholesterolemia from an early age [16,17]. This persistent inflammation causes changes in the large and small airways, leading to epithelial cell apoptosis, fibroblast activation, increased collagen deposition, thickening of the basement membrane, and increased hypertrophy of bronchial smooth muscle cells, resulting in airway remodeling [18,19]. These histological changes are influenced by different inflammatory pathways based on the patient's asthma phenotype, which can be identified based on a record of atopy, pulmonary symptoms, age and time of symptom onset, body mass index (BMI), Immunoglobin E serum levels (IgE), cellularity in sputum, the degree of sensitivity to aeroallergens through skin tests, the severity of airway obstruction, and the level of fractional exhaled nitric oxide (FeNO) [20,21]. As a result, asthma patients with obesity can be classified into two subclusters: early-onset (atopic or T2-high) and late-onset (non-atopic or T2-low) phenotypes [22–24].

The early-onset or atopic phenotype is characterized by an eosinophilic inflammatory response, with high allergen sensitization since childhood being the most frequent. In addition to the inflammatory markers of the TH2 response, such as interleukin-4 (IL-4), IL-5, and IL-13, high levels of IL-6 and adipokines (such as leptin) due to obesity may have a synergistic effect on airway inflammation and remodeling by stimulating the accumulation of eosinophils, mast cells, and CD4+ T lymphocytes in the airway [25,26]. In contrast, the late-onset phenotype (which is strongly linked to the low-grade systemic inflammation state of obesity also called "meta-inflammation") is seen in 10-20% of cases and mainly in female patients [27,28]. It is characterized by reduced TH2 markers, neutrophilic airway inflammation induced by type 3 innate lymphoid cells, and airway hyper-responsiveness due to premature closure caused by adipose tissue mediators and TH17 inflammatory cytokines such as IL-17, IL-6, IL-21, IL-22, and Interferon-gamma (IFN- γ). As a result, Tidal volume (TV) is mechanically restricted, and weight gain can exacerbate asthma symptoms and decrease the response to inhaled corticosteroids (ICS) and long-acting beta agonist (LABA) combination therapy [29,30]. For this reason, other treatment alternatives, such as leukotriene modifiers, nonsurgical weight loss strategies, and bariatric surgery, have been investigated with contrasting results [29–34]. For example, adult patients with eosinophilic

asthma, high levels of IgE, and obesity who underwent bariatric surgery have reported improvements in their lung function (forced expiratory volume in 1 (FEV1) and forced vital capacity (FVC)) one year after the surgical intervention [35,36], control of asthma symptoms or remission [35], and decreased use of inhaled medications compared to patients with late-onset asthma or with normal serum IgE levels [35–38]. As for children, some studies have reported that a low-calorie diet, exercise, and counseling can reduce exercise-induced bronchospasm with a significant reduction in post-exercise FEV1 drop [39] and can improve FEV1/FVC, reduce asthma symptoms, and increase adiponectin levels [40], suggesting that the asthma phenotype and obesity onset are key factors for the clinical improvement of patients.

Interestingly, these phenotypes are not mutually exclusive, and some studies have indicated that the expression of TH17 and TH2 inflammatory mediators is possible, likely because IL17A can be stimulated by IL-13 during allergic inflammation, which then leads to increased expression of TH2 mediators by IL-17, resulting in airway hyper-responsiveness mediated by complement component C5 [41] (See Figure 1).

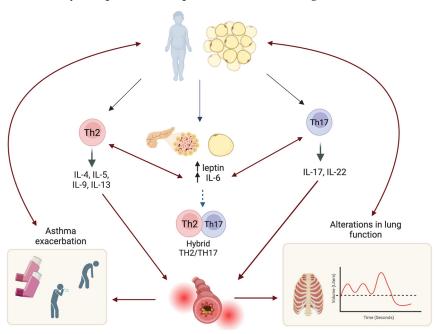


Figure 1. Inflammatory pathways related to asthma phenotypes (Th2, Th17, and Th2/Th17). These pathways can be activated by metabolic alterations, such as hyperinsulinemia and high levels of leptin, due to the meta-inflammation caused by obesity manifesting as severe asthma symptoms and alterations in lung volumes and resistance from an early age. Created with BioRender.com (https://www.biorender.com/, accessed on 17 May 2023).

Another potential modifier of the asthma phenotype is the gut/lung microbiome axis. The microbiome involves the microbiota, metagenome, and surrounding environment that allows microorganisms to interact with different cells, including the immune system [42]. It has already been described that the airway is not sterile, and the presence of *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Moraxella catarrhalis*, and *Haemophilus* spp. since infancy has been associated with the eosinophilic asthma phenotype [43,44]. In contrast, the presence of enriched Proteobacteria increases the expression of TH17 inflammatory pathways, reflecting severe asthma symptoms and suboptimal control [44]. Interestingly, gut dysbiosis has also been associated with asthma since childhood [42]. Recent studies have shown that children with a lower relative abundance of *Faecalibacterium*, *Lachnospira*, *Rothia*, and *Viollenella* (FLVR), slower diversification of the gut microbiome [45,46], and colonization of Clostridium difficile since infancy express higher serum levels of IL-17A, IL-6, and TNF- α , thereby increasing the risk of severe asthma symptoms [45,47]. Consequently, dysbiosis in both the airway and intestinal microbiota could have a synergistic effect on the

J. Clin. Med. 2023, 12, 5385 4 of 13

clinical expression of asthma, as they have the potential to stimulate metabolites (such as short-chain free fatty acids) and pro- or anti-inflammatory cytokines at local and systemic levels, which directly affects the regulation and maturation of mucosal-associated immune tissues (MALT), B-cell differentiation, and dendritic cell (DC) activation, among other immune-critical cells related to protection against pathogens [48]. In addition, there are other external factors that can alter gut and lung microbiota, such as diet, route of birth, breastfeeding, use of antibiotics and antipyretics during the first year of life, the frequency of viral infections, and a history of family atopy, which have been widely described not only as risk factors for asthma but also for obesity [49,50].

3. Lung Obstructive Pattern vs. Dysanapsis in Children with Asthma and Obesity

Understanding the impact of inflammation on lung function in patients with asthma is essential for successful treatment and prognosis. In this regard, international guidelines for diagnosing and treating asthma recommend spirometry as the preferred pulmonary study for measuring lung volumes and evaluating the bronchial muscle response to short-acting β2 adrenergic agonists, with FEV1 serving as an indicator of the severity of bronchial obstruction. [4,51]. However, this value relies on the patient's cooperation and does not always achieve a 12% or 200 milliliter increase after bronchodilator use [4]. Moreover, children with asthma and obesity have a lower FEV1/FVC ratio (a decrease of at least 1% in the FEV1/FVC ratio for every 5 unit increase in body mass index (BMI)) but normal or even increasing FEV1 and FVC, which is in line with either an obstructive pattern or airway dysanapsis [12,52-55]. Dysanapsis is defined as an imbalance in the growth of the lung parenchyma and airway calibration and is more prevalent in males and in those with obesity from a young age [56]. Even though the mechanisms underlying this phenomenon are still being studied, factors such as weight gain during pregnancy [57], in utero exposure to tobacco [58], and vitamin D deficiency have been associated with it [59,60]. Furthermore, the widespread presence of M1 macrophages and leptin polymorphisms due to excess fat tissue during childhood may impede lung growth and development by stimulating proinflammatory cytokines such as IL-6, IL-12, tumor necrosis factor-alpha (TNF-α), and monocyte chemoattractant protein 1 (MCP-1), which are associated with chronic inflammation and other comorbidities such as insulin resistance or metabolic syndrome, as mentioned later in this review [61–63].

A meta-analysis conducted by Forno et al. found that overweight or obese children and adults with asthma had significantly reduced maximum mid-expiratory flow (MMEF of FEF25–75) and a slight decrease in total lung capacity (TLC), residual volume (RV), and functional residual capacity (FRC) [64]. In comparison, children with dysanapsis had a greater FVC, vital capacity, RV, and TLC but a normal RV/TLC ratio, indicating that their lungs were larger with no significant air entrapment [15] (for lung parameter differences, see Table 1). As a clinical resource, it has been observed that the dysanaptic index (FEF50/FVC×Pst(l)50% or FEF25–75/FVC ratio) can be used to predict expiratory flow limitation and response to methacholine, suggesting that dysanapsis may be involved in airway obstruction and hyper-reactivity [65,66]. In addition, patients with dysanapsis are more likely to take three or more asthma medications (odds ratio (OR) 1.72; 95% confidence interval (CI) 1.02–2.90; p = 0.04), use short-acting beta 2 agonists daily (OR, 8.3; 95% CI, 1.1–64.0; p = 0.04), or experience severe asthma attacks that necessitate hospitalization or systemic steroid treatment (hazard ratio (HR) of 1.40; 95% confidence interval of 1.07 to 1.75; p-value of 0.014) [15,55].

Table 1. Differences between obstructive and dysanapsis patterns in children with obesity.

Lung Parameters	Obstructive Pattern	Dysanapsis Pattern
FEV1/FVC	↓	↓
FEV1	NL or ↑	NL or ↑
FVC	NL or ↑	NL or ↑↑
FEF 25–75%	+	↓

J. Clin. Med. 2023, 12, 5385 5 of 13

Table 1. Cont.

Lung Parameters	Obstructive Pattern	Dysanapsis Pattern
MEF50		
MEF75%	NI	↓
RV/TLC	↓	NL
TLC	<u> </u>	†

Forced expiratory volume in 1 s/forced vital capacity index (FEV1/FVC); forced expiratory volume in 1 s (FEV1); forced vital capacity (FVC); maximum mid-expiratory flow (MMEF or FEF 25–75%); maximal expiratory flow at 50% of the forced vital capacity (MEF50); maximal expiratory flow at 75% of the forced vital capacity (MEF75%); residual volume (RV); total lung capacity (TLC); NL: normal; NI: no information; ↑: increased; ↓: reduced.

4. Complementary Studies to Evaluate Lung Function in Patients with Asthma and Obesity

Currently, there are other techniques that evaluate small airway mechanics and monitor pulmonary inflammation, such as the impulse oscillometry system (IOS) and fractional exhaled nitric oxide (FeNO). Both techniques require minimal effort from the patient and are non-invasive [67,68]. While the usefulness of these resources is still being studied for different lung diseases, including chronic obstructive pulmonary disease and bronchiectasis, and in different populations to determine cut-off points due to variability between ethnic groups, age, and comorbidities, various studies have demonstrated their effectiveness in identifying and diagnosing obstructive patterns in the airway and phenotyping patients with asthma [67].

4.1. Impulse Oscillometry

The impulse oscillometry system (IOS) is a method that allows the measurement of impedance (Z) and its components, resistance (R) and reactance (X), which are the forces that must be overcome for air to enter and exit the respiratory system to detect alterations in the distal airway by utilizing multiple oscillatory frequencies. In addition, the IOS provides information on the resonant frequency (Fres) and reactance area (AX) [69]. This is useful in patients with alterations in thoracoabdominal compliance, obstructive pulmonary symptoms with normal spirometry, or in obese patients for whom it is not clear whether they have an obstructive or restrictive pulmonary pattern [69,70]. Previous studies have shown a strong correlation between an increase in resistance at 5 Hertz (Hz) minus 20 Hz (R5 - R20) in patients with small airway dysfunction and uncontrolled asthma [71,72]. In addition, the distal obstruction pattern can be identified by the R5 Hz value, which is higher than the upper limit of normality or greater than 1.64 standard deviations (SD) of the predicted value with normal R20 and AX values. According to the Z-value, the degree of obstruction can be classified as mild (1.64 to 2 standard deviations), moderate (≥ 2 standard deviations), or severe (≥ 4 standard deviations) [72]. As for IOS bronchodilator responsiveness, it has been suggested that there should be a change of 20–25% to 40% in R5, depending on the cut-off points of each population [72–74].

However, it has been observed that in children with obesity, the R5, Fres, and AX parameters are higher than those in healthy individuals, and this is reflected in the reference values [75]. The effect of R5 (suggestive of affected small- and large-caliber airways) and the changes in Fres and AX suggest the presence of airway obstruction [75–77]. In adults, studies have found that when severe obesity (body mass index (BMI) $\geq 40~{\rm kg/m^2}$) is present, there is a notable rise in peripheral system resistance (R) and a decrease in reactance (X) [69], and have also established a correlation between the resistance at 5 Hz (R5) and the FEV1 spirometry value (2.5% change in %R5 for every unit in FEV1) [78]. Nevertheless, this association is not always present in children with an obstructive pattern, particularly in those with a history of bronchiolitis or pneumonia during childhood [77]. Consequently, it is recommended that both spirometry and impulse oscillometry be conducted in patients with asthma to define the type and severity of obstruction [79–81] (see Figure 2).

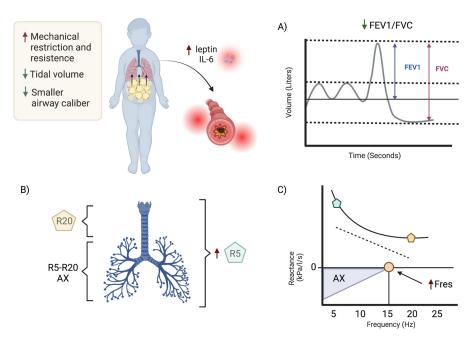


Figure 2. Obesity and proinflammatory metabolic alterations cause changes in respiratory mechanics, promoting a smaller volume and caliber of the airway in patients with asthma. This is reflected in a decrease in the FEV1/FVC ratio (**A**) and an increase in the resistances of both the proximal (R20) and distal airways (R5) (**B**), leading to changes in the reactance area (AX) and resonant frequency (Fres), which is the point where the magnitudes of the capacitive and inertial reactance are equal and have a value of zero at different frequencies (**C**). Created with BioRender.com (https://www.biorender.com/, accessed on 17 May 2023).

4.2. Exhaled Fraction of Nitric Oxide

As part of the medical approach for asthma patients, the exhaled fraction of nitric oxide (FeNO) allows for the estimation and monitoring of airway inflammation, which has been relevant in recent years [82]. The inducible or type II isoform of nitric oxide (iNOS) is responsible for the extravasation of macrophages, neutrophils, eosinophils, and T lymphocytes through chemotaxis as well as the expression of proinflammatory cytokines such as TNF- α , IL-1, and IFN- γ , with the consequent overproduction of mucus through the indirect stimulation of TH2 cells in the lungs [83,84]. In relation to the above, an increase in fractional exhaled nitric oxide (FeNO) has been linked to the allergic or eosinophilic asthma phenotype [82,85]. However, patients with asthma and obesity have been found to have low or normal FeNO levels [86,87]. This may have been caused by neutrophils and a low T2-type inflammatory response, which are characteristic of an obesity-related asthma phenotype [88]. Interestingly, some studies in adults have revealed an inverse correlation between FeNO levels and IBM regardless of the presence of asthma [89–92], and this may be due to the fact that obese patients have narrower airways, which increases the speed of airflow and decreases the time for pulmonary diffusion. Therefore, the amount of nitric oxide in the exhaled air is minimal [93]. Still, further research is needed to determine the usefulness of this measurement in distinguishing between allergic and non-allergic asthma in obese patients.

5. Metabolic Alterations Related to Lung Function in Obesity-Related Asthma Phenotype

It is common for individuals with obesity since childhood to display signs of metabolic alterations, such as insulin resistance, elevated levels of leptin, and vitamin D deficiency. In addition to the reduced functional residual capacity due to obesity, these factors can be linked to increased respiratory resistance and changes in lung structure, thereby increasing the risk of bronchial hyper-reactivity and asthma [94].

Research in children and adolescents with metabolic syndrome has shown that both asthmatics and non-asthmatics have decreased FEV1 and FVC values (up to 90 mL) and a reduced FEV1/FVC ratio when insulin resistance, as measured by the Homeostatic Model Assessment Insulin Resistance (HOMA-IR), is elevated, and these changes in lung volume are more pronounced in individuals with obesity and insulin resistance [95,96]. In addition, children with hyperinsulinemia are susceptible to airway dysfunction and heightened airway responsiveness due to epithelial damage and the stimulation of airway smooth muscle growth through insulin growth factor 1 [97,98], which also inhibits neuronal M2 muscarinic receptors and increases the release of acetylcholine from airway parasympathetic nerves, leading to obstructive pulmonary symptoms from an early age [99,100]. These pulmonary changes may also be due to the interaction of other inflammatory agents such as leptin, a hormone that is structurally and functionally similar to IL-6 and directly related to fat tissue [101]. Leptin can activate different signaling pathways that can induce asthma by promoting the proliferation of hybrid TH2/TH17 and TH17-like cells and decreasing T regulatory function by leptin-producing monocytes, resulting in a severe asthma phenotype [101–106]. From a clinical standpoint, leptin levels have been observed to be inversely correlated with FEV1 and the FEV1/FVC ratio [107], with a significant decrease during exercise [108] and a poor response to inhaled steroid treatment, which mainly affects adolescent females (see Figure 1) [32,109].

Given the close relationship between obesity, leptin, and insulin resistance, there are various recommendations for the treatment of these conditions with the aim of reducing cardiovascular risk and diabetes mellitus, as well as improving asthma symptoms [110]. Dietary interventions, physical activity, and lifestyle changes have been proposed as the first-choice treatment to promote weight loss and improve insulin sensitivity, respiratory symptoms, and lung function in pediatric patients with asthma. In addition, various studies have reported the beneficial effect of the adjuvant use of pharmacological treatments like metformin (approved by the US Food and Drug Administration in patients older than 10 years) [111], glucagon-like peptide-1 (GLP-1) [112] and dipeptidylpeptidase-4 (DPP4) inhibitors in order to obtain beneficial changes in body weight, insulin, and glycated hemoglobin (HbA1C) [113]. As per a recent study by Foer et al., adult patients with moderate to severe asthma and diabetes mellitus who were treated with glucagon-like peptide-1 receptor agonists (GLP1-RAs) experienced fewer asthma exacerbations over a 6-month period compared to those receiving treatment with insulin, DPP4 inhibitors, or sulfonylureas. The study found that weight loss was a significant contributor to this clinical outcome [114]. Therefore, personalized treatment that takes into account the patient's age, the severity of obesity, and asthma symptoms is crucial in choosing the best therapeutic option for metabolic alterations.

Vitamin D deficiency has been linked to lung disease due to its role in regulating the immune system by stimulating the expression of anti-inflammatory cytokines such as IL-4, IL-5, IL-10, and IL-13 and inhibiting the expression and transcription of proinflammatory cytokines, particularly IL-17 [115,116]. Additionally, vitamin D plays a critical role in the development of the lung during pregnancy by stimulating type II pneumocytes with receptors for 1,25-(OH)2D3 to increase surfactant production [117,118]. It also suppresses smooth muscle growth and is essential for epithelial–mesenchymal interactions during lung development [119].

Since childhood, low vitamin D levels have been linked to decreased lung function, including lower forced vital capacity (FVC) and forced expiratory volume (FEV1), as well as an increased risk of wheezing, asthma hospitalizations, and an inadequate response to glucocorticoids [120–123]. These associations may be due to the fact that low vitamin D levels can increase inflammation by elevating the expression of TNF- α and IL-17, while reducing the expression of IL-10, leading to a more intense TH2/TH17 immune response [124]. As a result, studies have been conducted to evaluate the effects of vitamin D supplementation in patients with asthma, and evidence suggests that vitamin D supplementation may improve

the response to inhaled corticosteroids in patients with asthma and reduce the frequency of asthma exacerbations [125,126].

Although there are recommendations regarding the supplementation dose in pediatric and adult patients (400–2000 IU/day) [127], there is still no consensus on the appropriate dose to supplement patients with asthma and obesity using high doses of inhaled or oral corticosteroids. This has been the subject of multiple clinical trials (mostly in adults) that have sought to improve the control of asthma symptoms and lung function (specifically FEV1). However, the results have been contrasting because the routes of administration, doses (standard and loading doses), and time of administration are variable, depending on the population and study criteria. A meta-analysis conducted by Lui et al., which analyzed the results of clinical trials of vitamin D supplementation in asthma patients, reported great heterogeneity in the dosage and time of administration of vitamin D, differences in the asthma severity of the included patients, the treatment of asthma at the time of supplementation, and the measurement of clinical outcomes (such as asthma exacerbations, asthma attacks, and control of symptoms using the ACT questionnaire) [128]. Therefore, it is difficult to determine the actual effect of vitamin D supplementation in asthmatic patients with obesity, and more clinical trials with this specific phenotype are required.

6. Conclusions

The increasing number of patients with asthma and obesity suggests that the interaction between these two diseases is complex and is influenced by intrinsic and extrinsic factors, which have an impact at the pulmonary level and on the clinical expression of asthma since childhood. Obesity increases the expression of proinflammatory cytokines at the respiratory and systemic levels, which affects the hyper-reactivity of the respiratory tract and conditions the response to drug treatment, which is a challenge for the prevention and treatment of asthmatic patients. Although there are currently diagnostic tools for the evaluation of lung function and asthma phenotyping as diagnostic support, obesity is a comorbidity that must be addressed to improve the quality of life and evolution of asthma. Therefore, patients must be evaluated in an interdisciplinary way to choose the best therapeutic strategy that can modify the progression of both diseases and improve lung function.

Author Contributions: Conceptualization, N.R.N., B.E.D.-R.-N., A.B., S.d.J.R.T. and D.J.M.M.D.; methodology, N.R.N. and B.E.D.-R.-N.; investigation, N.R.N., B.E.D.-R.-N., A.B., S.d.J.R.T. and D.J.M.M.D.; writing—original draft preparation, N.R.N. and B.E.D.-R.-N.; writing—review and editing, B.E.D.-R.-N., A.B., S.d.J.R.T. and D.J.M.M.D.; supervision, B.E.D.-R.-N. and S.d.J.R.T. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Purnell, J.Q. Definitions, Classification, and Epidemiology of Obesity. In *Endotext*; Feingold, K.R., Anawalt, B., Blackman, M.R., Boyce, A., Chrousos, G., Corpas, E., de Herder, W.W., Dhatariya, K., Dungan, K., Hofland, J., et al., Eds.; MDText.com, Inc.: South Dartmouth, MA, USA, 2023.

- 2. Stierman, B.; Afful, J.; Carroll, M.D.; Chen, T.C.; Davy, O.; Fink, S.; Fryar, C.D.; Gu, Q.; Hales, C.; Hughes, J.P.; et al. *National Health and Nutrition Examination Survey* 2017–March 2020 Prepandemic Data Files Development of Files and Prevalence Estimates for Selected Health Outcomes; National Health Statistics Reports; National Center for Health Statistics (U.S.): Hyattsville, MD, USA, 2021; p. 158.
- 3. Parameswaran, K.; Todd, D.C.; Soth, M. Altered respiratory physiology in obesity. Can. Respir. J. 2006, 13, 203–210. [CrossRef]
- 4. Reddel, H.K.; Bacharier, L.B.; Bateman, E.D.; Brightling, C.E.; Brusselle, G.G.; Buhl, R.; Cruz, A.A.; Duijts, L.; Drazen, J.M.; FitzGerald, J.M.; et al. Global Initiative for Asthma (GINA) Strategy 2021—Executive summary and rationale for key changes. *Eur. Respir. J.* 2021, 59, 2102730. [CrossRef]

J. Clin. Med. 2023, 12, 5385 9 of 13

5. Asher, M.I.; Rutter, C.E.; Bissell, K.; Chiang, C.Y.; El Sony, A.; Ellwood, E.; Ellwood, P.; García-Marcos, L.; Marks, G.B.; Morales, E.; et al. Worldwide trends in the burden of asthma symptoms in school-aged children: Global Asthma Network Phase I cross-sectional study. *Lancet* 2021, 398, 1569–1580. [CrossRef]

- Song, P.; Adeloye, D.; Salim, H.; Dos Santos, J.P.; Campbell, H.; Sheikh, A.; Rudan, I. Global, regional, and national prevalence of asthma in 2019: A systematic analysis and modelling study. J. Glob. Health 2022, 12, 4052. [CrossRef]
- Forno, E.; Young, O.M.; Kumar, R.; Simhan, H.; Celedón, J.C. Maternal obesity in pregnancy, gestational weight gain, and risk of childhood asthma. *Pediatrics* 2014, 134, e535–e546. [CrossRef]
- 8. Harskamp-van Ginkel, M.W.; London, S.J.; Magnus, M.C.; Gademan, M.G.; Vrijkotte, T.G. A Study on Mediation by Offspring BMI in the Association between Maternal Obesity and Child Respiratory Outcomes in the Amsterdam Born and Their Development Study Cohort. *PLoS ONE* **2015**, *10*, e0140641. [CrossRef]
- 9. Zein, J.G.; Erzurum, S.C. Asthma is Different in Women. Curr. Allergy Asthma Rep. 2015, 15, 28. [CrossRef]
- 10. Graziottin, A.; Serafini, A. Perimenstrual asthma: From pathophysiology to treatment strategies. *Multidiscip. Respir. Med.* **2016**, 11, 30. [CrossRef] [PubMed]
- 11. Owen, J.A., Jr. Physiology of the menstrual cycle. Am. J. Clin. Nutr. 1975, 28, 333–338. [CrossRef] [PubMed]
- 12. Pereira-Vega, A.; Sánchez, J.L.; Maldonado, J.A.; Borrero, F.; Rico, I.V.; Vázquez, R.; Alvarez, F. Premenstrual asthma and atopy markers. *Ann. Allergy Asthma Immunol.* **2010**, 105, 218–222. [CrossRef] [PubMed]
- 13. Kalogeromitros, D.; Katsarou, A.; Armenaka, M.; Rigopoulos, D.; Zapanti, M.; Stratigos, I. Influence of the menstrual cycle on skin-prick test reactions to histamine, morphine and allergen. *Clin. Exp. Allergy* **1995**, 25, 461–466. [CrossRef] [PubMed]
- 14. Jones, R.L.; Nzekwu, M.M. The effects of body mass index on lung volumes. Chest 2006, 130, 827–833. [CrossRef] [PubMed]
- 15. Forno, E.; Weiner, D.J.; Mullen, J.; Sawicki, G.; Kurland, G.; Han, Y.Y.; Cloutier, M.M.; Canino, G.; Weiss, S.T.; Litonjua, A.A.; et al. Obesity and Airway Dysanapsis in Children with and without Asthma. *Am. J. Respir. Crit. Care Med.* **2017**, 195, 314–323. [CrossRef] [PubMed]
- 16. Robertson, A.K.; Zhou, X.; Strandvik, B.; Hansson, G.K. Severe hypercholesterolaemia leads to strong Th2 responses to an exogenous antigen. *Scand. J. Immunol.* **2004**, *59*, 285–293. [CrossRef] [PubMed]
- 17. Li, Y.; Schwabe, R.F.; DeVries-Seimon, T.; Yao, P.M.; Gerbod-Giannone, M.C.; Tall, A.R.; Davis, R.J.; Flavell, R.; Brenner, D.A.; Tabas, I. Free cholesterol-loaded macrophages are an abundant source of tumor necrosis factor-alpha and interleukin-6: Model of NF-kappaB- and map kinase-dependent inflammation in advanced atherosclerosis. *J. Biol. Chem.* **2005**, *280*, 21763–21772. [CrossRef]
- 18. Lloyd, C.M.; Hessel, E.M. Functions of T cells in asthma: More than just T(H)2 cells. *Nat. Rev. Immunol.* **2010**, 10, 838–848. [CrossRef]
- 19. Grainge, C.L.; Lau, L.C.; Ward, J.A.; Dulay, V.; Lahiff, G.; Wilson, S.; Holgate, S.; Davies, D.E.; Howarth, P.H. Effect of bronchoconstriction on airway remodeling in asthma. *N. Engl. J. Med.* **2011**, *364*, 2006–2015, Correction in *N. Engl. J. Med.* **2018**, *378*, 2450. https://doi.org/10.1056/NEJMx180023. [CrossRef]
- 20. Ray, A.; Oriss, T.B.; Wenzel, S.E. Emerging molecular phenotypes of asthma. *Am. J. Physiol. Lung Cell Mol. Physiol.* **2015**, 308, L130–L140. [CrossRef]
- 21. Schleich, F.N.; Manise, M.; Sele, J.; Henket, M.; Seidel, L.; Louis, R. Distribution of sputum cellular phenotype in a large asthma cohort: Predicting factors for eosinophilic vs. neutrophilic inflammation. *BMC Pulm. Med.* **2013**, *13*, 11. [CrossRef]
- 22. Moore, W.C.; Hastie, A.T.; Li, X.; Li, H.; Busse, W.W.; Jarjour, N.N.; Wenzel, S.E.; Peters, S.P.; Meyers, D.A.; Bleecker, E.R.; et al. Sputum neutrophil counts are associated with more severe asthma phenotypes using cluster analysis. *J. Allergy Clin. Immunol.* **2014**, 133, 1557–1563.e5. [CrossRef]
- 23. Loza, M.J.; Djukanovic, R.; Chung, K.F.; Horowitz, D.; Ma, K.; Branigan, P.; Barnathan, E.S.; Susulic, V.S.; Silkoff, P.E.; Sterk, P.J.; et al. Validated and longitudinally stable asthma phenotypes based on cluster analysis of the ADEPT study. *Respir. Res.* **2016**, *17*, 165. [CrossRef] [PubMed]
- Lefaudeux, D.; De Meulder, B.; Loza, M.J.; Peffer, N.; Rowe, A.; Baribaud, F.; Bansal, A.T.; Lutter, R.; Sousa, A.R.; Corfield, J.; et al. U-BIOPRED clinical adult asthma clusters linked to a subset of sputum omics. *J. Allergy Clin. Immunol.* 2017, 139, 1797–1807. [CrossRef] [PubMed]
- 25. Sideleva, O.; Dixon, A.E. The many faces of asthma in obesity. J. Cell Biochem. 2014, 115, 421–426. [CrossRef] [PubMed]
- 26. Sideleva, O.; Suratt, B.T.; Black, K.E.; Tharp, W.G.; Pratley, R.E.; Forgione, P.; Dienz, O.; Irvin, C.G.; Dixon, A.E. Obesity and asthma: An inflammatory disease of adipose tissue not the airway. *Am. J. Respir. Crit. Care Med.* **2012**, *186*, 598–605. [CrossRef]
- 27. Unamuno, X.; Gómez-Ambrosi, J.; Rodríguez, A.; Becerril, S.; Frühbeck, G.; Catalán, V. Adipokine dysregulation and adipose tissue inflammation in human obesity. *Eur. J. Clin. Investig.* **2018**, *48*, 12997. [CrossRef] [PubMed]
- 28. Kuruvilla, M.E.; Lee, F.E.; Lee, G.B. Understanding Asthma Phenotypes, Endotypes, and Mechanisms of Disease. *Clin. Rev. Allergy Immunol.* **2019**, *56*, 219–233. [CrossRef]
- 29. Jensen, M.E.; Wood, L.G.; Gibson, P.G. Obesity and childhood asthma—Mechanisms and manifestations. *Curr. Opin. Allergy Clin. Immunol.* **2012**, 12, 186–192. [CrossRef]
- 30. Boulet, L.P.; Franssen, E. Influence of obesity on response to fluticasone with or without salmeterol in moderate asthma. *Respir. Med.* **2007**, *101*, 2240–2247. [CrossRef]
- 31. Dixon, A.E.; Holguin, F.; Sood, A.; Salome, C.M.; Pratley, R.E.; Beuther, D.A.; Celedón, J.C.; Shore, S.A. An official American Thoracic Society Workshop report: Obesity and asthma. *Proc. Am. Thorac. Soc.* **2010**, *7*, 325–335. [CrossRef]

32. Forno, E.; Lescher, R.; Strunk, R.; Weiss, S.; Fuhlbrigge, A.; Celedón, J.C.; Childhood Asthma Management Program Research Group. Decreased response to inhaled steroids in overweight and obese asthmatic children. *J. Allergy Clin. Immunol.* 2011, 127, 741–749. [CrossRef]

- 33. Peters-Golden, M.; Swern, A.; Bird, S.S.; Hustad, C.M.; Grant, E.; Edelman, J.M. Influence of body mass index on the response to asthma controller agents. *Eur. Respir. J.* **2006**, *27*, 495–503. [CrossRef] [PubMed]
- 34. Murr, M.M.; Siadati, M.R.; Sarr, M.G. Results of Bariatric Surgery for Morbid Obesity in Patients Older than 50 Years. *Obes. Surg.* **1995**, *5*, 399–402. [CrossRef] [PubMed]
- 35. Xie, L.; Chandrasekhar, A.; DeSantis, S.M.; Almandoz, J.P.; de la Cruz-Muñoz, N.; Messiah, S.E. Discontinuation and reduction of asthma medications after metabolic and bariatric surgery: A systematic review and meta-analysis. *Obes. Rev.* 2023, 24, 13527. [CrossRef] [PubMed]
- 36. Dixon, A.E.; Pratley, R.E.; Forgione, P.M.; Kaminsky, D.A.; Whittaker-Leclair, L.A.; Griffes, L.A.; Garudathri, J.; Raymond, D.; Poynter, M.E.; Bunn, J.Y.; et al. Effects of obesity and bariatric surgery on airway hyperresponsiveness, asthma control, and inflammation. *J. Allergy Clin. Immunol.* **2011**, 128, 508–515.e152. [CrossRef]
- 37. Chaaban, T.A. Bariatric surgery: A potential cure for asthma? Eur. Respir. Rev. 2019, 28, 190003. [CrossRef] [PubMed]
- 38. Van Huisstede, A.; Rudolphus, A.; Cabezas, M.C.; Biter, L.U.; van de Geijn, G.J.; Taube, C.; Hiemstra, P.S.; Braunstahl, G.J. Effect of bariatric surgery on asthma control, lung function and bronchial and systemic inflammation in morbidly obese subjects with asthma. *Thorax* **2015**, *70*, 659–667. [CrossRef] [PubMed]
- 39. Da Silva, P.L.; de Mello, M.T.; Cheik, N.C.; Sanches, P.L.; de Piano, A.; Corgosinho, F.C.; da Silveira Campos, R.M.; Carnier, J.; Inoue, D.; do Nascimento, C.M.; et al. The role of pro-inflammatory and anti-inflammatory adipokines on exercise-induced bronchospasm in obese adolescents undergoing treatment. *Respir. Care* 2012, 57, 572–582. [CrossRef] [PubMed]
- 40. Da Silva, P.L.; de Mello, M.T.; Cheik, N.C.; Sanches, P.L.; Correia, F.A.; de Piano, A.; Corgosinho, F.C.; da Silveira Campos, R.M.; do Nascimento, C.M.; Oyama, L.M.; et al. Interdisciplinary therapy improves biomarkers profile and lung function in asthmatic obese adolescents. *Pediatr. Pulmonol.* **2012**, *47*, 8–17. [CrossRef]
- 41. Lajoie, S.; Lewkowich, I.P.; Suzuki, Y.; Clark, J.R.; Sproles, A.A.; Dienger, K.; Budelsky, A.L.; Wills-Karp, M. Complement-mediated regulation of the IL-17A axis is a central genetic determinant of the severity of experimental allergic asthma. *Nat. Immunol.* **2010**, 11, 928–935. [CrossRef]
- 42. Belkaid, Y.; Hand, T.W. Role of the microbiota in immunity and inflammation. Cell 2014, 157, 121–141. [CrossRef]
- 43. Van Nimwegen, F.A.; Penders, J.; Stobberingh, E.E.; Postma, D.S.; Koppelman, G.H.; Kerkhof, M.; Reijmerink, N.E.; Dompeling, E.; Van Den Brandt, P.A.; Ferreira, I.; et al. Mode and place of delivery, gastrointestinal microbiota, and their influence on asthma and atopy. *J. Allergy Clin. Immunol.* **2011**, *128*, 948–955.e553. [CrossRef] [PubMed]
- 44. Green, B.J.; Wiriyachaiporn, S.; Grainge, C.; Rogers, G.B.; Kehagia, V.; Lau, L.; Carroll, M.P.; Bruce, K.D.; Howarth, P.H. Potentially pathogenic airway bacteria and neutrophilic inflammation in treatment resistant severe asthma. *PLoS ONE* **2014**, *9*, 100645. [CrossRef] [PubMed]
- 45. Stokholm, J.; Blaser, M.J.; Thorsen, J.; Rasmussen, M.A.; Waage, J.; Vinding, R.K.; Schoos, A.M.M.; Kunøe, A.; Fink, N.R.; Chawes, B.L.; et al. Maturation of the gut microbiome and risk of asthma in childhood. *Nat. Commun.* **2018**, *9*, 704. [CrossRef] [PubMed]
- 46. Arrieta, M.C.; Stiemsma, L.T.; Dimitriu, P.A.; Thorson, L.; Russell, S.; Yurist-Doutsch, S.; Kuzeljevic, B.; Gold, M.J.; Britton, H.M.; Lefebvre, D.L.; et al. Early infancy microbial and metabolic alterations affect risk of childhood asthma. *Sci. Transl. Med.* **2015**, 7, 307ra152. [CrossRef]
- 47. Durack, J.; Kimes, N.E.; Lin, D.L.; Rauch, M.; McKean, M.; McCauley, K.; Panzer, A.R.; Mar, J.S.; Cabana, M.D.; Lynch, S.V. Delayed gut microbiota development in high-risk for asthma infants is temporarily modifiable by Lactobacillus supplementation. *Nat. Commun.* 2018, *9*, 707. [CrossRef] [PubMed]
- Bouskra, D.; Brézillon, C.; Bérard, M.; Werts, C.; Varona, R.; Boneca, I.G.; Eberl, G. Lymphoid tissue genesis induced by commensals through NOD1 regulates intestinal homeostasis. *Nature* 2008, 456, 507–510. [CrossRef]
- 49. Hilty, M.; Burke, C.; Pedro, H.; Cardenas, P.; Bush, A.; Bossley, C.; Davies, J.; Ervine, A.; Poulter, L.; Pachter, L.; et al. Disordered microbial communities in asthmatic airways. *PLoS ONE* **2010**, *5*, 8578. [CrossRef]
- 50. Huang, Y.J.; Nariya, S.; Harris, J.M.; Lynch, S.V.; Choy, D.F.; Arron, J.R.; Boushey, H. The airway microbiome in patients with severe asthma: Associations with disease features and severity. *J. Allergy Clin. Immunol.* **2015**, *136*, 874–884. [CrossRef]
- 51. Expert Panel Working Group of the National Heart, Lung, and Blood Institute (NHLBI) administered and coordinated National Asthma Education and Prevention Program Coordinating Committee (NAEPPCC); Cloutier, M.M.; Baptist, A.P.; Blake, K.V.; Brooks, E.G.; Bryant-Stephens, T.; DiMango, E.; Dixon, A.E.; Elward, K.S.; Hartert, T.; et al. 2020 Focused Updates to the Asthma Management Guidelines: A Report from the National Asthma Education and Prevention Program Coordinating Committee Expert Panel Working Group. J. Allergy Clin. Immunol. 2020, 146, 1217–1270. [CrossRef]
- 52. Kolsum, U.; Borrill, Z.; Roy, K.; Starkey, C.; Vestbo, J.; Houghton, C.; Singh, D. Impulse oscillometry in COPD: Identification of measurements related to airway obstruction, airway conductance and lung volumes. *Respir. Med.* 2009, 103, 136–143. [CrossRef]
- 53. Chu, Y.T.; Chen, W.Y.; Wang, T.N.; Tseng, H.I.; Wu, J.R.; Ko, Y.C. Extreme BMI predicts higher asthma prevalence and is associated with lung function impairment in school-aged children. *Pediatr. Pulmonol.* **2009**, 44, 472–479. [CrossRef] [PubMed]
- 54. Starr, S.; Wysocki, M.; DeLeon, J.D.; Silverstein, G.; Arcoleo, K.; Rastogi, D.; Feldman, J.M. Obesity-related pediatric asthma: Relationships between pulmonary function and clinical outcomes. *J. Asthma* **2023**, *60*, 1418–1427. [CrossRef] [PubMed]

55. Tantisira, K.G.; A Litonjua, A.; Weiss, S.T.; Fuhlbrigge, A.L.; Childhood Asthma Management Program Research Group. Association of body mass with pulmonary function in the Childhood Asthma Management Program (CAMP). *Thorax* **2003**, 58, 1036–1041. [CrossRef] [PubMed]

- 56. Taussig, L.M.; Cota, K.; Kaltenborn, W. Different mechanical properties of the lung in boys and girls. *Am. Rev. Respir. Dis.* **1981**, 123, 640–643.
- 57. Dumas, O.; Varraso, R.; Gillman, M.W.; Field, A.E.; Camargo, C.A., Jr. Longitudinal study of maternal body mass index, gestational weight gain, and offspring asthma. *Allergy* **2016**, *71*, 1295–1304. [CrossRef]
- 58. Lieberman, E.; Torday, J.; Barbieri, R.; Cohen, A.; Van Vunakis, H.; Weiss, S.T. Association of intrauterine cigarette smoke exposure with indices of fetal lung maturation. *Obstet. Gynecol.* **1992**, *79*, 564–570.
- 59. Zosky, G.R.; Hart, P.H.; Whitehouse, A.J.O.; Kusel, M.M.; Ang, W.; Foong, R.E.; Chen, L.; Holt, P.G.; Sly, P.D.; Hall, G.L. Vitamin D deficiency at 16 to 20 weeks' gestation is associated with impaired lung function and asthma at 6 years of age. *Ann. Am. Thorac. Soc.* **2014**, *11*, 571–577. [CrossRef]
- 60. Loddo, F.; Nauleau, S.; Lapalus, D.; Tardieu, S.; Bernard, O.; Boubred, F. Association of Maternal Gestational Vitamin D Supplementation with Respiratory Health of Young Children. *Nutrients* **2023**, *15*, 2380. [CrossRef]
- 61. Chylikova, J.; Dvorackova, J.; Tauber, Z.; Kamarad, V. M1/M2 macrophage polarization in human obese adipose tissue. *Biomed. Pap.* **2018**, *162*, 79–82. [CrossRef]
- 62. Van den Borst, B.; Souren, N.Y.; Loos, R.J.; Paulussen, A.D.; Derom, C.; Schols, A.M.; Zeegers, M.P. Genetics of maximally attained lung function: A role for leptin? *Respir. Med.* **2012**, *106*, 235–242. [CrossRef]
- 63. Peters, M.C.; McGrath, K.W.; A Hawkins, G.; Hastie, A.T.; Levy, B.D.; Israel, E.; Phillips, B.R.; Mauger, D.T.; A Comhair, S.; Erzurum, S.C.; et al. Plasma interleukin-6 concentrations, metabolic dysfunction, and asthma severity: A cross-sectional analysis of two cohorts. *Lancet Respir. Med.* **2016**, *4*, 574–584. [CrossRef] [PubMed]
- 64. Forno, E.; Han, Y.Y.; Mullen, J.; Celedón, J.C. Overweight, Obesity, and Lung Function in Children and Adults-A Meta-analysis. *J. Allergy Clin. Immunol. Pract.* **2018**, *6*, 570–581.e10. [CrossRef] [PubMed]
- 65. Mead, J. Dysanapsis in normal lungs assessed by the relationship between maximal flow, static recoil, and vital capacity. *Am. Rev. Respir. Dis.* **1980**, 121, 339–342. [PubMed]
- 66. Smith, J.R.; Rosenkranz, S.K.; Harms, C.A. Dysanapsis ratio as a predictor for expiratory flow limitation. *Respir. Physiol. Neurobiol.* **2014**, *198*, 25–31. [CrossRef]
- 67. Houghton, C.M.; Woodcock, A.A.; Singh, D. A comparison of lung function methods for assessing dose-response effects of salbutamol. *Br. J. Clin. Pharmacol.* **2004**, *58*, 134–141. [CrossRef]
- 68. Gemicioglu, B.; Musellim, B.; Dogan, I.; Guven, K. Fractional exhaled nitric oxide (FeNo) in different asthma phenotypes. *Allergy Rhinol.* **2014**, *5*, 157–161. [CrossRef]
- 69. De Albuquerque, C.G.; de Andrade, F.M.D.; Rocha, M.A.d.A.; de Oliveira, A.F.F.; Ladosky, W.; Victor, E.G.; Rizzo, J. Determining respiratory system resistance and reactance by impulse oscillometry in obese individuals. *J. Bras. Pneumol.* **2015**, *41*, 422–426. [CrossRef]
- 70. De Oliveira Jorge, P.P.; de Lima, J.H.P.; Chong ESilva, D.C.; Medeiros, D.; Solé, D.; Wandalsen, G.F. Impulse oscillometry in the assessment of children's lung function. *Allergol. Immunopathol.* **2019**, *47*, 295–302. [CrossRef]
- 71. Lin, L.-M.; Chang, Y.-J.; Yang, K.D.; Lin, C.-H.; Chien, J.-W.; Kao, J.-K.; Lee, M.-S.; Chiang, T.-I.; Tsai, Y.-G. Small Airway Dysfunction Measured by Impulse Oscillometry and Fractional Exhaled Nitric Oxide Is Associated with Asthma Control in Children. *Front. Pediatr.* **2022**, *10*, 877681. [CrossRef]
- Gochicoa-Rangel, L.; Cantú-González, G.; Miguel-Reyes, J.L.; Rodríguez-Moreno, L.; Torre-Bouscoulet, L. Oscilometría de Impulso: Recomendaciones y Procedimiento. Neumol. Cir. Torax 2014, 78, 124. Available online: http://www.scielo.org.mx/scielo.php?script=sci_arttext&pid=S0028-37462014000200006&lng=es (accessed on 17 May 2023). [CrossRef]
- 73. Marotta, A.; Klinnert, M.D.; Price, M.R.; Larsen, G.L.; Liu, A.H. Impulse oscillometry provides an effective measure of lung dysfunction in 4-year-old children at risk for persistent asthma. *J. Allergy Clin. Immunol.* **2003**, 112, 317–322. [CrossRef] [PubMed]
- 74. Hellinckx, J.; De Boeck, K.; Bande-Knops, J.; van der Poel, M.; Demedts, M. Bronchodilator response in 3–6.5 years old healthy and stable asthmatic children. *Eur. Respir. J.* 1998, 12, 438–443. [CrossRef]
- 75. Assumpção, M.S.; Ribeiro, J.D.; Wamosy, R.M.G.; Figueiredo, F.C.X.S.; Parazzi, P.L.F.; Schivinski, C.I.S. Impulse oscillometry and obesity in children. *J. Pediatr.* **2018**, *94*, 419–424. [CrossRef]
- 76. Van de Kant, K.D.; Paredi, P.; Meah, S.; Kalsi, H.S.; Barnes, P.J.; Usmani, O.S. The effect of body weight on distal airway function and airway inflammation. *Obes. Res. Clin. Pract.* **2016**, *10*, 564–573. [CrossRef] [PubMed]
- 77. Lauhkonen, E.; Koponen, P.; Nuolivirta, K.; Paassilta, M.; Toikka, J.; Saari, A.; Korppi, M. Obesity and bronchial obstruction in impulse oscillometry at age 5-7 years in a prospective post-bronchiolitis cohort. *Pediatr. Pulmonol.* **2015**, *50*, 908–914. [CrossRef]
- 78. Nair, A.; Ward, J.; Lipworth, B.J. Comparison of bronchodilator response in patients with asthma and healthy subjects using spirometry and oscillometry. *Ann. Allergy Asthma Immunol.* **2011**, 107, 317–322. [CrossRef] [PubMed]
- 79. Dencker, M.; Malmberg, L.P.; Valind, S.; Thorsson, O.; Karlsson, M.K.; Pelkonen, A.; Pohjanpalo, A.; Haahtela, T.; Turpeinen, M.; Wollmer, P. Reference values for respiratory system impedance by using impulse oscillometry in children aged 2–11 years. *Clin. Physiol. Funct. Imaging* **2006**, *26*, 247–250. [CrossRef]
- 80. Frei, J.; Jutla, J.; Kramer, G.; Hatzakis, G.E.; Ducharme, F.M.; Davis, G.M. Impulse oscillometry: Reference values in children 100 to 150 cm in height and 3 to 10 years of age. *Chest* 2005, 128, 1266–1273. [CrossRef]

81. Lee, J.-Y.; Seo, J.-H.; Kim, H.Y.; Jung, Y.H.; Kwon, J.-W.; Kim, B.-J.; Bin Kim, H.; Lee, S.-Y.; Jang, G.C.; Song, D.J.; et al. Reference values of impulse oscillometry and its utility in the diagnosis of asthma in young Korean children. *J. Asthma* **2012**, *49*, 811–816. [CrossRef]

- 82. Karrasch, S.; Linde, K.; Rücker, G.; Sommer, H.; Karsch-Völk, M.; Kleijnen, J.; A Jörres, R.; Schneider, A. Accuracy of FENO for diagnosing asthma: A systematic review. *Thorax* **2017**, 72, 109–116. [CrossRef]
- 83. Meng, Q.; Polak, J.M.; Edgar, A.J.; Chacon, M.R.; Evans, T.J.; Gruenert, D.C.; Bishop, A.E. Neutrophils enhance expression of inducible nitric oxide synthase in human normal but not cystic fibrosis bronchial epithelial cells. *J. Pathol.* **2000**, 190, 126–132. [CrossRef]
- 84. Moncada, S.; Palmer, R.M.; Higgs, E.A. Nitric oxide: Physiology, pathophysiology, and pharmacology. *Pharmacol. Rev.* **1991**, 43, 109–142.
- 85. Schneider, A.; Schwarzbach, J.; Faderl, B.; Welker, L.; Karsch-Völk, M.; Jörres, R.A. FENO measurement and sputum analysis for diagnosing asthma in clinical practice. *Respir. Med.* **2013**, *107*, 209–216. [CrossRef]
- 86. Han, Y.Y.; Forno, E.; Celedón, J.C. Adiposity, fractional exhaled nitric oxide, and asthma in U.S. children. *Am. J. Respir. Crit. Care Med.* **2014**, *190*, 32–39. [CrossRef]
- 87. Santamaria, F.; Montella, S.; De Stefano, S.; Sperlì, F.; Barbarano, F.; Spadaro, R.; Franzese, A. Asthma, atopy, and airway inflammation in obese children. *J. Allergy Clin. Immunol.* **2007**, *120*, 965–967. [CrossRef] [PubMed]
- 88. Rastogi, D.; Holguin, F. Metabolic Dysregulation, Systemic Inflammation, and Pediatric Obesity-related Asthma. *Ann. Am. Thorac. Soc.* **2017**, *14*, S363–S367. [CrossRef] [PubMed]
- 89. Yao, T.-C.; Ou, L.-S.; Lee, W.-I.; Yeh, K.-W.; Chen, L.-C.; Huang, J.-L.; for the PATCH Study Group. Exhaled nitric oxide discriminates children with and without allergic sensitization in a population-based study. *Clin. Exp. Allergy* **2011**, *41*, 556–564. [CrossRef] [PubMed]
- 90. De Winter-de Groot, K.M.; Van der Ent, C.K.; Prins, I.; Tersmette, J.M.; Uiterwaal, C.S. Exhaled nitric oxide: The missing link between asthma and obesity? *J. Allergy Clin. Immunol.* **2005**, *115*, 419–420. [CrossRef] [PubMed]
- 91. Kazaks, A.; Uriu-Adams, J.Y.; Stern, J.S.; Albertson, T.E. No significant relationship between exhaled nitric oxide and body mass index in people with asthma. *J. Allergy Clin. Immunol.* **2005**, *116*, 929–930. [CrossRef] [PubMed]
- 92. Flashner, B.M.; Rifas-Shiman, S.L.; Oken, E.; Camargo, C.A.; Platts-Mills, T.J.; Workman, L.; Litonjua, A.A.; Gold, D.R.; Rice, M.B. Obesity, sedentary lifestyle, and exhaled nitric oxide in an early adolescent cohort. *Pediatr. Pulmonol.* **2020**, *55*, 503–509. [CrossRef] [PubMed]
- 93. Ho, L.P.; Wood, F.T.; Robson, A.; Innes, J.A.; Greening, A.P. The current single exhalation method of measuring exhales nitric oxide is affected by airway calibre. *Eur. Respir. J.* **2000**, *15*, 1009–1013. [CrossRef]
- 94. Kongkiattikul, L.; Sritippayawan, S.; Chomtho, S.; Deerojanawong, J.; Prapphal, N. Relationship between Obesity Indices and Pulmonary Function Parameters in Obese Thai Children and Adolescents. *Indian J. Pediatr.* **2015**, *82*, 1112–1116. [CrossRef] [PubMed]
- 95. Del Río-Camacho, G.; Domínguez-Garrido, M.N.; Pita, J.; Aragón, I.; Collado, R.; Soriano-Guillén, L. Masa ventricular izquierda, espirometría basal forzada y perfil de adipocitocinas en niños obesos con y sin síndrome metabólico [Left ventricular mass, forced baseline spirometry and adipocytokine profiles in obese children with and without metabolic syndrome]. *An. Pediatr.* **2013**, 78, 27–34. [CrossRef]
- 96. Forno, E.; Han, Y.Y.; Muzumdar, R.H.; Celedón, J.C. Insulin resistance, metabolic syndrome, and lung function in US adolescents with and without asthma. *J. Allergy Clin. Immunol.* **2015**, *136*, 304–311.e8. [CrossRef]
- 97. Zosky, G.R.; Berry, L.J.; Elliot, J.G.; James, A.L.; Gorman, S.; Hart, P.H. Vitamin D deficiency causes deficits in lung function and alters lung structure. *Am. J. Respir. Crit. Care Med.* **2011**, *183*, 1336–1343. [CrossRef]
- 98. Noveral, J.P.; Bhala, A.; Hintz, R.L.; Grunstein, M.M.; Cohen, P. Insulin-like growth factor axis in airway smooth muscle cells. *Am. J. Physiol.* **1994**, 267, L761–L765. [CrossRef] [PubMed]
- 99. Agrawal, A.; Mabalirajan, U.; Ahmad, T.; Ghosh, B. Emerging interface between metabolic syndrome and asthma. *Am. J. Respir. Cell Mol. Biol.* **2011**, *44*, 270–275. [CrossRef]
- 100. Nie, Z.; Jacoby, D.B.; Fryer, A.D. Hyperinsulinemia potentiates airway responsiveness to parasympathetic nerve stimulation in obese rats. *Am. J. Respir. Cell Mol. Biol.* **2014**, *51*, 251–261. [CrossRef]
- 101. Zheng, H.; Zhang, X.; Castillo, E.F.; Luo, Y.; Liu, M.; Yang, X.O. Leptin Enhances TH2 and ILC2 Responses in Allergic Airway Disease. *J. Biol. Chem.* **2016**, 291, 22043–22052. [CrossRef]
- 102. Atamas, S.P.; Chapoval, S.P.; Keegan, A.D. Cytokines in chronic respiratory diseases. F1000 Biol. Rep. 2013, 5, 3. [CrossRef]
- 103. Lessard, A.; St-Laurent, J.; Turcotte, H.; Boulet, L.P. Leptin and adiponectin in obese and non-obese subjects with asthma. *Biomarkers* 2011, 16, 271–273. [CrossRef]
- 104. Watanabe, K.; Suzukawa, M.; Kawauchi-Watanabe, S.; Igarashi, S.; Asari, I.; Imoto, S.; Tashimo, H.; Fukami, T.; Hebisawa, A.; Tohma, S.; et al. Leptin-producing monocytes in the airway submucosa may contribute to asthma pathogenesis. *Respir. Investig.* **2023**, *61*, 5–15. [CrossRef] [PubMed]
- 105. Vollmer, C.M.; Dias, A.S.O.; Lopes, L.M.; Kasahara, T.M.; Delphim, L.; Silva, J.C.C.; Lourenço, L.P.; Gonçalves, H.C.; Linhares, U.C.; Gupta, S.; et al. Leptin favors Th17/Treg cell subsets imbalance associated with allergic asthma severity. *Clin. Transl. Allergy* **2022**, *12*, e12153. [CrossRef] [PubMed]

106. Sánchez-Ortega, H.; Jiménez-Cortegana, C.; Novalbos-Ruiz, J.P.; Gómez-Bastero, A.; Soto-Campos, J.G.; Sánchez-Margalet, V. Role of Leptin as a Link between Asthma and Obesity: A Systematic Review and Meta-Analysis. *Int. J. Mol. Sci.* 2022, 24, 546. [CrossRef] [PubMed]

- 107. Huang, F.; Del-Río-Navarro, B.E.; Torres-Alcántara, S.; Pérez-Ontiveros, J.A.; Ruiz-Bedolla, E.; Saucedo-Ramírez, O.J.; Villafaña, S.; Muñoz, F.S.; Bravo, G.; Hong, E. Adipokines, asymmetrical dimethylarginine, and pulmonary function in adolescents with asthma and obesity. *J. Asthma* **2017**, *54*, 153–161. [CrossRef] [PubMed]
- 108. Baek, H.S.; Kim, Y.D.; Shin, J.H.; Kim, J.H.; Oh, J.W.; Lee, H.B. Serum leptin and adiponectin levels correlate with exercise-induced bronchoconstriction in children with asthma. *Ann. Allergy Asthma Immunol.* **2011**, *107*, 14–21. [CrossRef] [PubMed]
- 109. Kilic, H.; Oguzulgen, I.K.; Bakir, F.; Turktas, H. Asthma in obese women: Outcomes and factors involved. *J. Investig. Allergol. Clin. Immunol.* **2011**, 21, 290–296. [PubMed]
- 110. Tagi, V.M.; Giannini, C.; Chiarelli, F. Insulin Resistance in Children. Front. Endocrinol. 2019, 10, 342. [CrossRef]
- 111. Khokhar, A.; Umpaichitra, V.; Chin, V.L.; Perez-Colon, S. Metformin Use in Children and Adolescents with Prediabetes. *Pediatr. Clin. N. Am.* **2017**, *64*, 1341–1353. [CrossRef]
- 112. Guo, C.; Huang, T.; Chen, A.; Chen, X.; Wang, L.; Shen, F.; Gu, X. Glucagon-like peptide 1 improves insulin resistance in vitro through anti-inflammation of macrophages. *Braz. J. Med. Biol. Res.* **2016**, *49*, e5826. [CrossRef]
- 113. Guo, H.; Fang, C.; Huang, Y.; Pei, Y.; Chen, L.; Hu, J. The efficacy and safety of DPP4 inhibitors in patients with type 1 diabetes: A systematic review and meta-analysis. *Diabetes Res. Clin. Pract.* **2016**, 121, 184–191. [CrossRef] [PubMed]
- 114. Foer, D.; Beeler, P.E.; Cui, J.; Karlson, E.W.; Bates, D.W.; Cahill, K.N. Asthma Exacerbations in Patients with Type 2 Diabetes and Asthma on Glucagon-like Peptide-1 Receptor Agonists. *Am. J. Respir. Crit. Care Med.* **2021**, 203, 831–840. [CrossRef] [PubMed]
- 115. Lang, P.O.; Aspinall, R. Vitamin D Status and the Host Resistance to Infections: What It Is Currently (Not) Understood. *Clin. Ther.* **2017**, 39, 930–945. [CrossRef] [PubMed]
- 116. Joshi, S.; Pantalena, L.C.; Liu, X.K.; Gaffen, S.L.; Liu, H.; Rohowsky-Kochan, C.; Ichiyama, K.; Yoshimura, A.; Steinman, L.; Christakos, S.; et al. 1,25-dihydroxyvitamin D(3) ameliorates Th17 autoimmunity via transcriptional modulation of interleukin-17A. *Mol. Cell Biol.* 2011, 31, 3653–3669. [CrossRef] [PubMed]
- 117. Dogan, M.; Erol, M.; Cesur, Y.; Yuca, S.A.; Doğan, Z. The effect of 25-hydroxyvitamin D3 on the immune system. *J. Pediatr. Endocrinol. Metab.* **2009**, 22, 929–935. [CrossRef] [PubMed]
- 118. Nguyen, M.; Trubert, C.; Rizk-Rabin, M.; Rehan, V.; Besançon, F.; Cayre, Y.; Garabédian, M. 1,25-Dihydroxyvitamin D3 and fetal lung maturation: Immunogold detection of VDR expression in pneumocytes type II cells and effect on fructose 1,6 bisphosphatase. *J. Steroid Biochem. Mol. Biol.* 2004, 89–90, 93–97. [CrossRef] [PubMed]
- 119. Damera, G.; Fogle, H.W.; Lim, P.; Goncharova, E.A.; Zhao, H.; Banerjee, A.; Tliba, O.; Krymskaya, V.P.; Panettieri, R.A., Jr. Vitamin D inhibits growth of human airway smooth muscle cells through growth factor-induced phosphorylation of retinoblastoma protein and checkpoint kinase 1. *Br. J. Pharmacol.* **2009**, *158*, 1429–1441. [CrossRef]
- 120. Zhu, Y.; Jing, D.; Liang, H.; Li, D.; Chang, Q.; Shen, M.; Pan, P.; Liu, H.; Zhang, Y. Vitamin D status and asthma, lung function, and hospitalization among British adults. *Front. Nutr.* **2022**, *9*, 954768. [CrossRef]
- 121. Brehm, J.M.; Celedón, J.C.; Soto-Quiros, M.E.; Avila, L.; Hunninghake, G.M.; Forno, E.; Laskey, D.; Sylvia, J.S.; Hollis, B.W.; Weiss, S.T.; et al. Serum vitamin D levels and markers of severity of childhood asthma in Costa Rica. *Am. J. Respir. Crit. Care Med.* **2009**, 179, 765–771. [CrossRef]
- 122. Pfeffer, P.E.; Mann, E.H.; Hornsby, E.; Chambers, E.S.; Chen, Y.-H.; Rice, L.; Hawrylowicz, C.M. Vitamin D influences asthmatic pathology through its action on diverse immunological pathways. *Ann. Am. Thorac. Soc.* **2014**, *11*, S314–S321. [CrossRef]
- 123. Wang, M.; Liu, M.; Wang, C.; Xiao, Y.; An, T.; Zou, M.; Cheng, G. Association between vitamin D status and asthma control: A meta-analysis of randomized trials. *Respir. Med.* **2019**, *150*, 85–94. [CrossRef]
- 124. Berry, M.A.; Hargadon, B.; Shelley, M.; Parker, D.; Shaw, D.E.; Green, R.H.; Bradding, P.; Brightling, C.E.; Wardlaw, A.J.; Pavord, I.D. Evidence of a role of tumor necrosis factor alpha in refractory asthma. *N. Engl. J. Med.* 2006, 354, 697–708. [CrossRef] [PubMed]
- 125. Sutherland, E.R.; Goleva, E.; Jackson, L.P.; Stevens, A.D.; Leung, D.Y. Vitamin D levels, lung function, and steroid response in adult asthma. *Am. J. Respir. Crit. Care Med.* **2010**, *181*, 699–704. [CrossRef] [PubMed]
- 126. Li, Q.; Zhou, Q.; Zhang, G.; Tian, X.; Chen, Y.; Cun, Y.; Xu, X.; Luo, Z. Long-term effects of vitamin D on exacerbation rate, health care utilization and lung function in children with asthma. *Ann. Transl. Med.* **2022**, *10*, 1094. [CrossRef] [PubMed]
- 127. Pludowski, P.; Holick, M.F.; Grant, W.B.; Konstantynowicz, J.; Mascarenhas, M.R.; Haq, A.; Povoroznyuk, V.; Balatska, N.; Barbosa, A.P.; Karonova, T.; et al. Vitamin D supplementation guidelines. *J. Steroid Biochem. Mol. Biol.* **2018**, *175*, 125–135. [CrossRef] [PubMed]
- 128. Liu, M.; Wang, J.; Sun, X. A Meta-Analysis on Vitamin D Supplementation and Asthma Treatment. *Front. Nutr.* **2022**, *9*, 860628. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.