



Review

Proprotein Convertase Subtilisin/Kexin Type 9 Inhibitors Use for Atherogenic Dyslipidemia in Solid Organ Transplant Patients

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Abstract: Dyslipidemia is a widespread risk factor in solid organ transplant patients, due to many reasons, such as the use of immunosuppressive drugs, with a consequent increase in cardiovascular diseases in this population. PCSK9 is an enzyme mainly known for its role in altering LDL levels, consequently increasing cardiovascular risk. Monoclonal antibody PCSK9 inhibitors demonstrated remarkable efficacy in the general population in reducing LDL cholesterol levels and preventing cardiovascular disease. In transplant patients, these drugs are still poorly used, despite having comparable efficacy to the general population and giving fewer drug interactions with immunosuppressants. Furthermore, there is enough evidence that PCSK9 also plays a role in other pathways, such as inflammation, which is particularly dangerous for graft survival. In this review, the current evidence on the function of PCSK9 and the use of its inhibitors will be discussed, particularly in transplant patients, in which they may provide additional benefits.

Keywords: dyslipidemia; immunosuppressants; transplant; PCSK9; PCSK9 inhibitors; alirocumab; evolocumab

1. Introduction

Dyslipidemia is defined as the occurrence of abnormal plasma levels of any lipid and/or lipoprotein fraction. This condition is globally widespread, but there is higher prevalence (about 50%) in developed countries, such as in Europe and in North America, whereas in developing countries, the prevalence is about 25% [1]. Dyslipidemia, particularly high levels of serum low-density lipoprotein (LDL), is the main risk factor for the development of atherosclerosis and cardiovascular disease (CVD) [2], which caused about 523 million cases and 18.5 million deaths in 2019: Moreover, it is a relevant cause of disability, with about 194 million disability-adjusted life years (DALYs) estimated in the same year [3].

Transplant recipients are part of a particular population suffering from atherogenic dyslipidemia. In fact, lipid alterations have been found in 80% of kidney transplant recipients [4] and heart transplant recipients [5], and 50% of liver transplant recipients [6]. Consequently, this lipid elevation is associated with an increase in CVDs, which become the first cause of mortality in kidney and heart transplant recipients [7,8]. In addition, dyslipidemia is also associated with other complications occurring in these patients, such

as cardiac allograft vasculopathy (CAV) in heart transplant recipients [9], and chronic allograft nephropathy (CAN) in kidney transplant recipients [10].

However, in these patients, the management of atherogenic dyslipidemia is challenging, since lipid elevation is caused by immunosuppressants (that cannot be withdrawn) [11,12] and due to the interactions between immunosuppressants and statins or other lipid-lowering drugs [13]. For this reason, often, statins and ezetimibe are used at a lower dosage, complicating LDL level reduction and CVD prevention [14].

In recent years, a therapy with Proprotein Convertase Subtilisin/Kexin type 9 (PCSK9) inhibitors is available for patients suffering from familial hypercholesterolemia or who are defined as very-high risk for the occurrence of cardiovascular events by the European Society of Cardiology guidelines on dyslipidemia [15,16]. They are human monoclonal antibodies which have proven to be safe and effective in reducing the incidence of major cardiac adverse events (MACE) in the general population [17–19].

This clinical success could be too simplistically attributed to the reduction in total cholesterol, LDL, and lipoprotein (a) (Lp(a)) levels alone: in fact, these drugs can also counteract the atherosclerotic process through other mechanisms, such as anti-inflammatory effects, plaque stabilization, anti-aggregation, and anticoagulant effects [20,21].

In transplant patients, due to the restrictive criteria for the prescription of PCSK9 inhibitors (PCSK9i), few cases have been reported with efficacy comparable to the general population [22]. Moreover, these drugs undergo a metabolism pattern different than immunosuppressants; therefore, there are no pharmacokinetic interferences [13,14].

In addition, elevation in PCSK9 levels due to the use of immunosuppressants has been shown [23,24], and this could be one more reason to use PCSK9i in this population.

This review aims at describing the role of the PCSK9 pathway in atherogenic dyslipidemia in transplant recipients, comparing the effectiveness of PCSK9i use in both the general population and in transplant patients. New emerging categories of patients that could benefit from the use of PCSK9i have been proposed.

2. Role of PCSK9 in Dyslipidemia and Atherosclerosis

PCSK9 is a proteolytic enzyme first discovered in neurons and involved in the apoptosis mechanism [25].

This protein has a ubiquitous distribution, but is predominantly found in the liver, small intestine, kidney, and muscular tissue [26]. In recent years, it has received much attention for its role in lipid metabolism.

The hepatocytes, where PCSK9 is primarily expressed, are the main drivers for LDL particle removal from blood, through endocytosis mediated by the LDL receptor (LDLR) expressed on these cells' membrane [27]. PCSK9 target the epidermal growth factor-like repeat homology domain A (EGFA-like) located on the LDLR, so LDLR and LDL particles are enabled to separate, and the entire complex is directed to lysosomes for degradation, avoiding the receptor recycling on the cell surface, and reducing, in this way, the clearance of LDL particles from plasma [17,26].

Gain-of-function mutations of the PCSK9 gene have been shown to cause familial hypercholesterolemia (FH), enhancing the risk for atherosclerosis and CVDs [28,29]. Missense mutations' gain-of-function in the same gene is linked to hypercholesterolemia, due to reduction in LDLR expression on the cell surface; in reverse, loss-of-function mutations cause hypocholesterolemia and lower LDL plasma levels until 40% [30].

Furthermore, PCSK9 is also related to the modulation of total cholesterol and VLDL levels, through overproduction of apolipoprotein B (ApoB) [31]. PCSK9 overexpression in hepatocytes increases VLDL production and circulating triglycerides levels [32,33], and mice without PCSK9 present reduced post-prandial triglycerides levels [34]. PCSK9 implication in VLDL regulation is also proven by the finding that mice with homozygous deletion for the PCSK9 gene accumulate more visceral adipose tissue and have higher levels of VLDL receptor [35].

In addition to these effects on metabolism, PCSK9 also directly promotes the atherosclerotic process. In fact, lipid accumulation in the intima layer of the vessel is not the only factor responsible for atheroma formation, as other factors promote the initiation and progression of atherosclerosis. In particular, inflammatory cells play a main role: macrophages start the process of phagocytosing oxidized LDL particles and transforming into foam cells, which begins a local inflammatory reaction, with consequent release of cytokines that attract other monocytes [36]; in later stages, T lymphocytes recruited into the lesion release other cytokines, causing the accumulation of collagen and glycosaminoglycans, which enlarge the plaque [37]. PCSK9 has been shown to have a role in the inflammatory response: on the one hand, promoting monocytes recruitment in the atherosclerotic lesion [38], and, on the other hand, activating leukocytes in a pro-inflammatory state, with consequent release of cytokines, in particular IL-17, which is associated with the atherosclerotic process [39]. Furthermore, PCSK9 is also produced by endothelial cells, and as a consequence of reactive oxygen species (ROS) production, the expression of this enzyme is enhanced, causing the hyperexpression of adhesion molecules which promote monocytes adhesion, favoring plaque progression [40].

Moreover, PCSK9 has also been shown to play a role in hemostasis, and, therefore, in clot formation that causes CVD: on the one hand, it can promote CVD, also increasing platelet aggregation and, consequently, thrombus formation, binding the CD36 receptor present on the thrombocyte membrane [41]; on the other hand, it increases the concentration of coagulation factors both directly, as on fibrinogen [42], and indirectly through the action on LDLR, which influences the factor VIII levels [43,44].

CD36, as well as lecithin-like oxidized low-density lipoprotein receptor-1 (LOX-1), also act as receptors for oxidized LDL particles in macrophages, provoking their transformation in foam cells, and both are enhanced by PCSK9 in inflammatory status [42].

In addition, PCSK9 levels are associated with a higher plasma concentration of fibrinogen ($r = 0.211$, $p = 0.002$) and C reactive protein (CRP) ($r = 0.153$, $p = 0.023$) in patients with coronary artery disease (CAD) [45].

Therefore, PCSK9 is involved in several pathways promoting atherosclerosis, and for this reason, it has become a target for atherosclerotic dyslipidemia therapies, particularly in patients who have a very-high risk for CVD.

Figure 1 shows the different mechanisms by which PCSK9 promotes atherosclerosis.

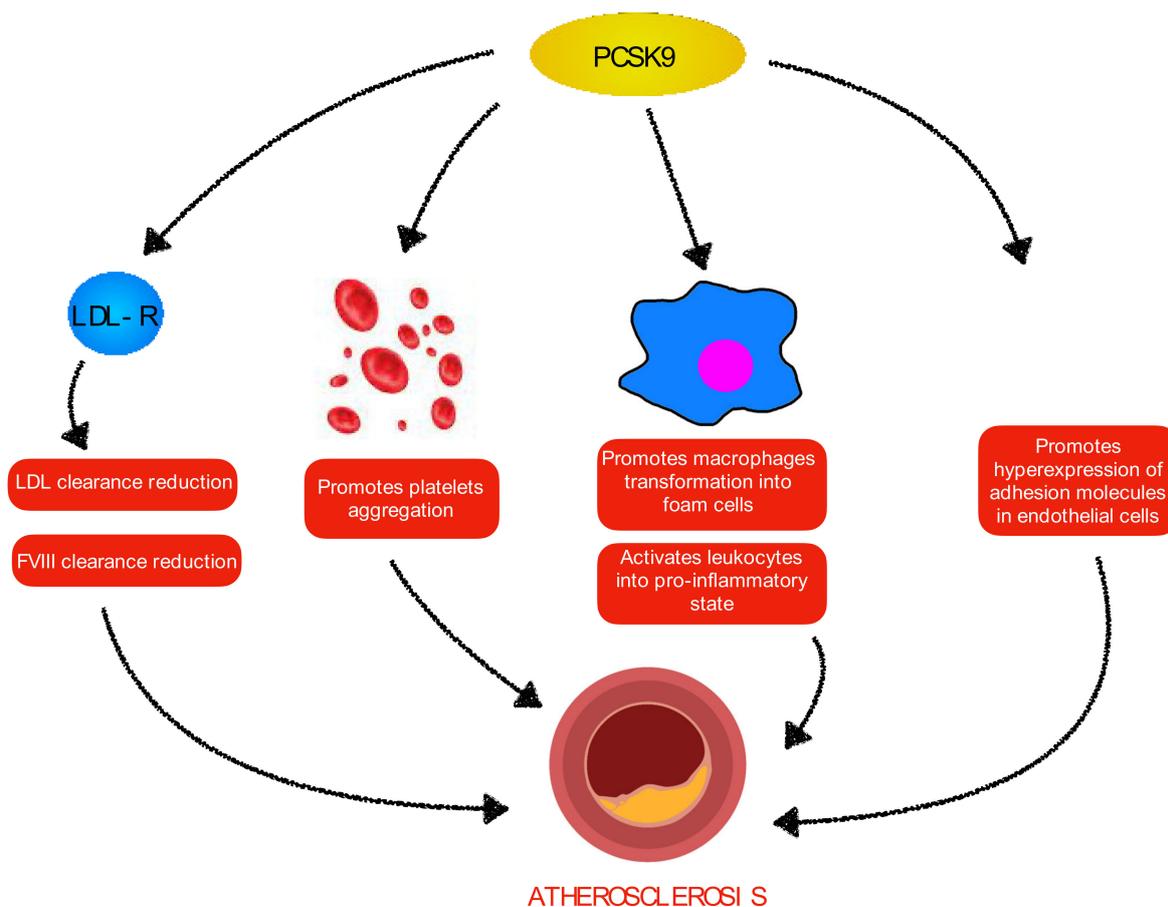


Figure 1. Mechanisms by which PCSK9 leads to atherosclerosis. FVIII: coagulation factor VIII; LDL: low-density lipoprotein; LDL-R: low-density lipoprotein receptor; PCSK9: proprotein convertase subtilisin/kexin type 9.

3. PCSK9 Inhibitors Efficacy in Cardiovascular Diseases

The two PCSK9i used in clinical practice for optimizing LDL-lowering therapy are the human monoclonal antibodies, evolocumab and alirocumab. They are administered through a 2- or 4-week subcutaneous or intramuscular injection, and have been shown to decrease LDL plasma concentration and reduce ischemic cardiovascular events. Another PCSK9i, the humanized monoclonal antibody called bococizumab, has not been placed on the market because of its higher rate of side reactions and increased production of neutralizing antibodies [46].

Alirocumab and evolocumab have been shown in clinical trials to reduce LDL serum levels, on average, by 60%, so the latest European Society of Cardiology guidelines on dyslipidemia recommend their use (class IA) in combination with the maximum-tolerated dose of a statin and ezetimibe in secondary prevention patients, and in FH patients with another major risk factor for CVD who do not achieve the LDL levels target [15].

The Further Cardiovascular Outcomes Research with PCSK9 Inhibition in Subjects with Elevated Risk (FOURIER) trial investigated the efficacy of evolocumab in addition to statin therapy in reducing the incidence of MACE in 27,564 patients (75% were males; average age, 63 years) with CVD and LDL levels ≥ 70 mg/dL (median LDL levels, 92 mg/dL), who were randomized in an evolocumab group ($n = 13,784$) and a placebo group ($n = 13,780$). A mean reduction in LDL levels of 59% (from 92 mg/dL to 30 mg/dL, $p < 0.001$) and a reduction of primary MACE endpoints (HR 0.85; 95% CI: 0.79–0.92; $p < 0.001$) were observed in the treatment group [18]. In addition, prespecified analysis showed a doubling of MACE prevented with evolocumab over a longer observation time, suggesting the benefit of continuing aggressive lipid-lowering therapy [47].

In the ODYSSEY LONG TERM trial, alirocumab at 150 mg every 2 weeks showed a statistically significant reduction in LDL levels (-62% , $p < 0.001$) compared to placebo, and in a post hoc analysis, the reduction in MACE was also lower (HR 0.52; 95% CI: 0.31–0.90; $p = 0.02$) [48].

Moreover, in the ODYSSEY OUTCOMES trial, 18,924 patients who experienced an acute coronary syndrome in the previous 1–12 months and had LDL levels ≥ 70 mg/dL or ApoB level ≥ 80 mg/dL despite statin therapy were randomly assigned to alirocumab at a 75 mg subcutaneous dose every 2 week group ($n = 9462$) or a placebo group ($n = 9462$): alirocumab showed great results in terms of LDL levels reduction (about 60% lower than the placebo group), and a reduction in composite primary endpoints (HR 0.85; 95% CI: 0.78–0.93; $p < 0.001$), which were death from CAD, nonfatal myocardial infarction, ischemic stroke, or unstable angina requiring hospitalization [19].

Furthermore, for both evolocumab and alirocumab, there were no differences in the rate of serious adverse events, allergic reactions, and diabetes, but injection-site reactions were more common than with placebo (3.8% for alirocumab, and 2.1% for evolocumab) [18,19].

In addition, PCSK9i showed other effects. Unlike statins, these monoclonal antibodies are effective in reducing, by about 25–30%, the circulating levels of lipoprotein (a) (Lp(a)) [49], which is associated with a higher risk of CVD [50,51].

Moreover, PCSK9i can reduce the atherosclerotic plaque: the GLAGOV trial showed that the addition of evolocumab to statin therapy in patients with CAD decreases the total atheroma volume by -4.9 mm³, and stimulates a strong plaque regression compared to placebo (64.3% vs. 47.3%) [52].

Finally, PCSK9i efficacy in preventing CVD is also plausibly due to the pleiotropic effects on hemostasis [20]: in 21 patients with isolated hypercholesterolemia, these drugs showed a significant reduction in plasma levels of fibrinogen ($p = 0.01$), factor VII ($p = 0.01$), and plasminogen activator inhibitor-1 (PAI) ($p = 0.001$).

4. Atherogenic Dyslipidemia and Cardiovascular Diseases in Transplant Patients

Solid organ transplantation is a therapy which allows patients with end-stage organ disease to survive. Due to recent innovations in the field of transplantation, there has been a reduction in early complications, such as rejection and infections, which have long-been the major cause of mortality in these patients, and, consequently, the life expectancy of these patients has increased [53].

Nevertheless, other diseases now threaten the survival of these patients, such as CVDs, which have become the first causes of death in heart transplant patients [8] and kidney transplant patients [7], and the second causes of death in liver transplant patients [54,55]. It should be considered that kidney transplant recipients are particularly at risk for CVD because, due to end-stage chronic kidney disease, they are classified as having a very-high risk for CVD [56].

However, this increase is not solely due to the longer life expectancy, which entails the possibility of the appearance of diseases common to the general population. In fact, transplant recipients appear to be more exposed to risk factors for CVD. In particular, atherogenic dyslipidemia is present in about 80% of kidney transplant recipients [4], 80% of heart transplant recipients [5], and 70% of liver transplant recipients [57], compared to about 30–40% of the general population [58,59]. These variations are due to multiple factors, such as genetic [60–62] and lifestyle [63,64] factors; however, the main role is probably played by immunosuppressants, which exert pharmacodynamic and pharmacokinetic interferences on lipid and glycemic metabolism [11,65]: in fact, in the first months after transplantation, during which immunosuppressants are administered at a higher dose, higher total cholesterol serum levels were found [66].

Among immunosuppressants, the mammalian target of rapamycin (mTOR) inhibitors, sirolimus and everolimus, are strongly associated with an increase in triglycerides, VLDL, and LDL, because they inhibit LPL function; reduce catabolism of the apolipoproteins, ApoB100 and apoCIII; alter insulin secretion; and induce pancreatic β -cells apoptosis [67–69]; these alterations are probably responsible for the significantly higher incidence of cardiovascular disease shown in patients treated with mTOR inhibitors (ROR 1.95, 95% CI: 1.70–2.23) [70].

The calcineurin inhibitors, cyclosporine (CsA) and tacrolimus, reduce the activity of lipoproteinlipase (LPL) and hepatic lipase with an increase in hepatic lipogenesis and apolipoprotein CIII (apoCIII), increasing serum levels of total cholesterol, LDL, and triglycerides [71–73]. CsA also causes drug interactions with statins, making the achievement of the LDL target by lipid-lowering therapies difficult [13,14].

In addition to atherogenic dyslipidemia, with increased levels of LDL and triglycerides, these drugs are responsible for other risk factors promoting atherosclerosis, such as arterial hypertension and hyperglycemia [11,65,74]. In fact, 20–40% of patients who have undergone solid organ transplant develop post-transplant diabetes mellitus (PTDM) [75], which involves an increase in CVD and reduced survival [76]. Several predisposing factors have been investigated for PTDM [77–80], but this complication is mostly associated with the use of immunosuppressants, particularly glucocorticoids [81,82], calcineurin inhibitors [83,84], and mTOR inhibitors [85,86].

Moreover, some immunosuppressants cause arterial hypertension, which occurs in more than 50% of transplant recipients, and is a well-known risk factor for CVD [87]. Calcineurin inhibitors, and particularly CsA, are the principal cause of arterial hypertension in transplant patients, both by activating production of vasoconstrictor molecules and reducing the release of nitric oxide (NO) [88,89]: proving this, the discontinuation of these drugs often resolves hypertension [65].

A main role in initiation and progression of atherosclerotic plaque in transplant recipients is also played by inflammation, which is increased due to the immune reaction against donor organs [90,91]. For this reason, in heart transplant patients, there is a particular form of CAD, named CAV, which is caused by antibodies against donor antigens that activate T lymphocytes, and the consequent inflammatory reaction leads to endothelial proliferation and vessel occlusion [9]. CAV is responsible for about 10% of deaths in these patients, and is distinguished from the usual CAD for its pathognomonic lesions, which affect the intramuscular arteries and microvascular bed [92–94].

In kidney transplant recipients, the increased inflammatory state causes the formation of oxidized LDL particles [95], which are associated with CAN, which can lead to transplant failure [10,96,97].

In conclusion, CVDs result more frequently in transplant patients due to a greater exposure to risk factors, caused both by the inflammatory state of the transplant, and by the side effects of immunosuppressants.

In Figure 2, the mechanisms of CVD in transplant recipients are summarized.

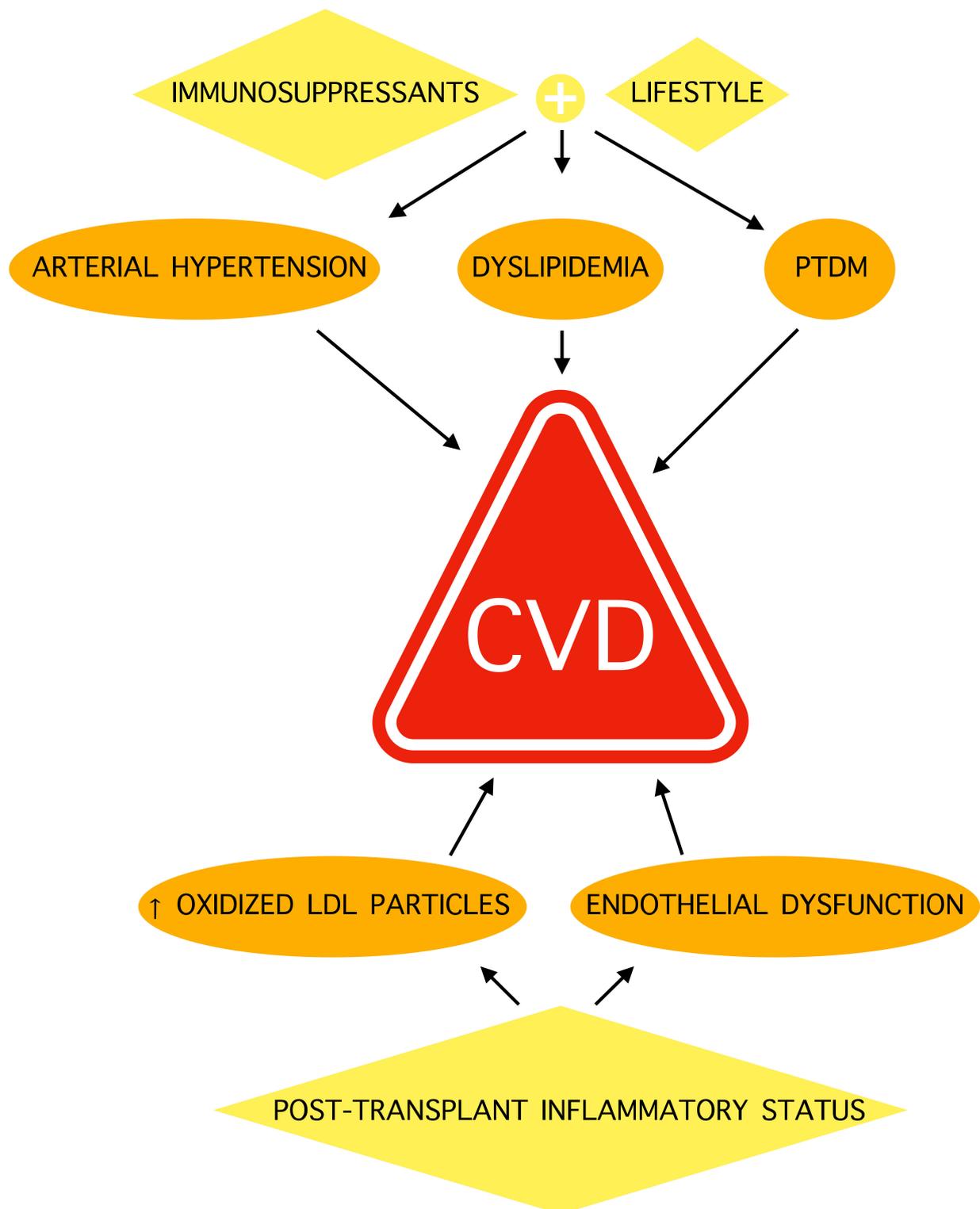


Figure 2. Cardiovascular disease mechanisms in transplant patients. CVD: cardiovascular disease; PTDM: post-transplant diabetes mellitus. LDL: low-density lipoprotein.

5. PCSK9 Inhibitors Use in Transplant Patients

The 2019 European Society of Cardiology guidelines on dyslipidemia consider transplant patients a special population that need attention [15].

However, as there is no risk category for transplant recipients, each patient should be assessed individually based on their clinical history. Kidney transplant patients are an

exception, as end-stage chronic kidney disease places them in the very-high risk class [98]. Moreover, heart transplant patients should keep LDL serum levels < 100 mg/dL to avoid the occurrence of CAV [99].

Anyway, lipid alterations in these patients cause not only CVD, but are also associated with transplant failure, as in CAN [10].

In these patients, statin use is often used at a lower dosage due to drug interactions with immunosuppressants, and as a result, it is not sufficient to achieve LDL target levels [13,14].

PCSK9i have been shown to be effective in reducing LDL levels in the general population by about 60% if used alone, 75% if used with high-intensity statins, and 85% if ezetimibe is added [16,100].

Based on their pharmacodynamic and pharmacokinetic properties, these monoclonal antibodies should not create drug interactions with immunosuppressants, and do not act on the cytochrome system or other enzymes involved in their metabolism [13].

To date, less evidence is available on the use of these drugs in transplant patients, since the indications for their use are limited to patients in the secondary prevention for CVD, or with familial hypercholesterolemia [15,16].

The first case reports were carried out in heart transplant recipients, and showed encouraging data on LDL level reduction, calling for larger trials or the creation of registries to also evaluate the safety of these drugs in solid organ transplant patients [101,102].

More recently, Sammour et al. [103] investigated efficacy and safety of PCSK9i in 65 patients who underwent a heart transplant in last 10 years: a mean reduction of 58% in LDL circulating levels was found 3 months after therapy initiation (from 130 mg/dL to 55 mg/dL; $p < 0.001$), and this significant reduction was confirmed at last follow-up (median follow-up time, 1.6 years) [103]. A total of 72% of patients achieved an LDL level target of <70 mg/dL at first follow-up, and significant reductions in total cholesterol and triglycerides, and significant increases in HDL were shown [103]. In addition, in 33 patients, through coronary angiography with intravascular ultrasound (IVUS), no progression of atherosclerotic plaque was observed [103].

In regard to this action on coronary disease, in the next years, the EVOLVD trial will try to prove the reduction in CAV incidence in heart transplant patients treated with evolocumab [104].

In kidney transplant recipients, there is scarce evidence about PCSK9i use. A recent case report described the experience of a 54-year-old female patient who underwent kidney transplantation in 2011 with atherogenic dyslipidemia arising during everolimus and tacrolimus treatment [105]: one year after initiation of alirocumab, a significant decrease of 83.1 mg/dL in LDL circulating levels ($p = 0.04$) and of 94.9 mg/dL in total cholesterol ($p = 0.03$) were observed, while no side effects or drug interactions with immunosuppressants occurred, and kidney function remained stable [105].

Another report described the case of a male kidney transplant patient who experienced two episodes of respiratory infections, the first of which required hospitalization in the intensive care unit for 48 h, after the initiation of therapy with alirocumab [106]; however, these episodes did not recur after everolimus was replaced with azathioprine.

Warden et al. described the use of PCSK9i in a more heterogeneous population, including nine heart transplant patients, one kidney transplant patient, one liver transplant patient, and one lung transplant patient [22]: 6 months after treatment, a median decrease of 60% in LDL circulating levels was observed, and all patients achieved the target of <70 mg/dL. Moreover, there were no problems with immunosuppressant therapeutic ranges, and no transplant rejection occurred [22].

The possibility that PCSK9i may cause an immune-interfering effect in transplant patients should be taken into account, especially given the reasons that led to bococizumab withdrawal [46]. However, it must be considered that other types of monoclonal antibodies with immunosuppressive effects are already successfully used in transplant recipients,

and currently, no immune-interfering effect has been found with the human monoclonal antibodies, alirocumab and evolocumab.

In reverse, there are some findings about PCSK9 in transplant patients, suggesting potential additional benefits that PCSK9i could have in this population.

For example, PCSK9 serum levels have been shown to increase 6 months after kidney transplantation [107]: although it can be hypothesized that this is due to the role of PCSK9 in inflammation [108], this does not seem plausible given that an inverse trend has been observed in inflammatory markers, such as leukocytes, IL-6, and CRP [107]. Authors hypothesized that this increase should be responsible for the higher incidence of atherogenic dyslipidemia in kidney transplant recipients and graft dysfunction, hoping for new research on the possibility that PCSK9i could promote longer renal graft survival [107].

Moreover, the administration of everolimus in renal and heart transplant patients demonstrated a statistically significant increase in PCSK9 levels [23,24], especially in patients with the mTORC1 rs2295080G polymorphism ($p = 0.006$) [24]: although it was not correlated with an increase in LDL levels, as the authors suggested, PCSK9 level elevation should represent an independent risk factor for the onset of CVD [109] or graft vasculopathy [23].

Finally, recent evidence showed that higher PCSK9 serum levels are associated with the development of PTDM, and this association is independent of statin use [110]. This association could be due to the regulatory role that PCSK9 has on the LDLR, which, in turn, appears to influence each other with the insulin receptor [111].

In addition, insulin resistance has been observed to cause an increase in hepatocyte PCSK9 transcription [112], so the increase in PCSK9 levels in patients with PTDM may be just a sign of insulin resistance from other causes.

However, the role of PCSK9 in diabetes, especially with this type in transplant patients, is still unclear.

So, the alterations in PCSK9 levels in transplant recipients are due to several factors. The use of PCSK9i, in addition to the well-known effect on lipid metabolism and cardiovascular risk, may underline, in future studies, more benefit in transplant patients. The assessed and possible benefits of these drugs' use in transplant recipients are summarized in Table 1.

Table 1. Shown and possible PCSK9 inhibitor use benefits in transplant patients, CVD: cardiovascular disease; FVIII: coagulation factor VIII; LDL: low-density lipoprotein.

BENEFITS SHOWN	Reduction in LDL levels in patients already at the highest possible dose of statins, without drug interferences with immunosuppressants.
	Reduction in CVD incidence and atherosclerotic plaque progression.
POSSIBLE BENEFITS	Reduction in cardiac allograft vasculopathy incidence.
	Reduction in post-transplant diabetes mellitus incidence.
	Reduction in chronic allograft nephropathy incidence.
	Graft survival extension.
	Reduction in CVD through platelet aggregation inhibition.
	Reduction in CVD through FVIII clearance.

6. Conclusions

Atherogenic dyslipidemia in solid transplant recipients is often more difficult to treat, due to the role of immunosuppressants, and often, many patients do not reach the LDL level goal, resulting in an increase in cardiovascular disease.

PCSK9i have been shown to be very effective and safe drugs even in these patients; in fact, they can be used together with immunosuppressants without causing drug interactions that could cause serious side effects or reduce graft survival.

Furthermore, post-transplantation status and the use of drugs such as everolimus have been shown not only to favor atherosclerosis and CVD incidence, but also increase

PCSK9 levels to a greater extent, and this can cause well- or less-known consequences: in fact, in addition to causing an increase in plasma levels of LDL and probably favoring post-transplant diabetes mellitus, it remains to be clarified whether elevation in PCSK9 levels can influence graft survival.

The use of PCSK9i in this population should be further investigated, as in addition to improving dyslipidemia and cardiovascular outcomes, they could have additional benefits on transplant complications and graft survival.

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References

1. Pirillo, A.; Casula, M.; Olmastroni, E.; Norata, G.D.; Catapano, A.L. Global epidemiology of dyslipidaemias. *Nat. Rev. Cardiol.* **2021**, *18*, 689–700. [[CrossRef](#)] [[PubMed](#)]
2. Ference, B.A.; Ginsberg, H.N.; Graham, I.; Ray, K.K.; Packard, C.J.; Bruckert, E.; Hegele, R.A.; Krauss, R.M.; Raal, F.J.; Schunkert, H.; et al. Low-density lipoproteins cause atherosclerotic cardiovascular disease. 1. Evidence from genetic, epidemiologic, and clinical studies. A consensus statement from the European Atherosclerosis Society Consensus Panel. *Eur. Heart J.* **2017**, *38*, 2459–2472. [[CrossRef](#)]
3. Roth, G.A.; Mensah, G.A.; Johnson, C.O.; Addolorato, G.; Ammirati, E.; Baddour, L.M.; Barengo, N.C.; Beaton, A.Z.; Benjamin, E.J.; Benziger, C.P.; et al. Global Burden of Cardiovascular Diseases and Risk Factors, 1990–2019: Update from the GBD 2019 Study. *J. Am. Coll. Cardiol.* **2020**, *76*, 2982–3021. [[CrossRef](#)]
4. Markell, M.S.; Armenti, V.; Danovitch, G.; Sumrani, N. Hyperlipidemia and glucose intolerance in the post-renal transplant patient. *J. Am. Soc. Nephrol.* **1994**, *4*, S37–S47. [[CrossRef](#)] [[PubMed](#)]
5. Lund, L.H.; Edwards, L.B.; Dipchand, A.I.; Goldfarb, S.; Kucheryavaya, A.Y.; Levvey, B.J.; Meiser, B.; Rossano, J.W.; Yusen, R.D.; Stehlik, J. The Registry of the International Society for Heart and Lung Transplantation: Thirty-third Adult Heart Transplantation Report—2016; Focus Theme: Primary Diagnostic Indications for Transplant. *J. Heart Lung Transplant.* **2016**, *35*, 1158–1169. [[CrossRef](#)]
6. Parekh, J.; Corley, D.A.; Feng, S. Diabetes, Hypertension and Hyperlipidemia: Prevalence Over Time and Impact on Long-Term Survival After Liver Transplantation. *Am. J. Transplant.* **2012**, *12*, 2181–2187. [[CrossRef](#)] [[PubMed](#)]
7. Israni, A.K.; Snyder, J.J.; Skeans, M.A.; Peng, Y.; Maclean, J.R.; Weinhandl, E.D.; Kasiske, B.L. Predicting Coronary Heart Disease after Kidney Transplantation: Patient Outcomes in Renal Transplantation (PORT) Study. *Am. J. Transplant.* **2010**, *10*, 338–353. [[CrossRef](#)] [[PubMed](#)]
8. Kobashigawa, J.; Starling, R.; Mehra, M.; Kormos, R.; Bhat, G.; Barr, M.; Sigouin, C.; Kolesar, J.; Fitzsimmons, W. Multicenter Retrospective Analysis of Cardiovascular Risk Factors Affecting Long-term Outcome of De Novo Cardiac Transplant Recipients. *J. Heart Lung Transplant.* **2006**, *25*, 1063–1069. [[CrossRef](#)] [[PubMed](#)]
9. Lee, F.; Nair, V.; Chih, S. Cardiac allograft vasculopathy: Insights on pathogenesis and therapy. *Clin. Transplant.* **2020**, *34*, e13794. [[CrossRef](#)]
10. Yates, P.; Nicholson, M. The aetiology and pathogenesis of chronic allograft nephropathy. *Transpl. Immunol.* **2006**, *16*, 148–157. [[CrossRef](#)]
11. Claes, K.; Meier-Kriesche, H.-U.; Schold, J.D.; Vanrenterghem, Y.; Halloran, P.F.; Ekberg, H. Effect of different immunosuppressive regimens on the evolution of distinct metabolic parameters: Evidence from the Symphony study. *Nephrol. Dial. Transplant.* **2011**, *27*, 850–857. [[CrossRef](#)] [[PubMed](#)]
12. Kockx, M.; Kritharides, L. Hyperlipidaemia in immunosuppression. *Curr. Opin. Lipidol.* **2016**, *27*, 631–632. [[CrossRef](#)]
13. Page, R.L.; Miller, G.G.; Lindenfeld, J. Drug therapy in the heart transplant recipient: Part IV: Drug-drug interactions. *Circulation* **2005**, *111*, 230–239. [[CrossRef](#)] [[PubMed](#)]
14. Warden, B.A.; Duell, P.B. Management of dyslipidemia in adult solid organ transplant recipients. *J. Clin. Lipidol.* **2019**, *13*, 231–245. [[CrossRef](#)] [[PubMed](#)]

15. Mach, F.; Baigent, C.; Catapano, A.L.; Koskinas, K.C.; Casula, M.; Badimon, L.; Chapman, M.J.; De Backer, G.G.; Delgado, V.; Ference, B.A.; et al. 2019 ESC/EAS guidelines for the management of dyslipidaemias: Lipid modification to reduce cardiovascular risk. *Atherosclerosis* **2019**, *290*, 140–205. [[CrossRef](#)]
16. Visseren, F.L.J.; Mach, F.; Smulders, Y.M.; Carballo, D.; Koskinas, K.C.; Böck, M.; Benetos, A.; Biffi, A.; Boavida, J.-M.; Capodanno, D.; et al. 2021 ESC Guidelines on cardiovascular disease prevention in clinical practice. *Eur. Heart J.* **2021**, *42*, 3227–3337. [[CrossRef](#)]
17. Iannuzzo, G.; Gentile, M.; Bresciani, A.; Mallardo, V.; Di Lorenzo, A.; Merone, P.; Cuomo, G.; Pacileo, M.; Sarullo, F.; Venturini, E.; et al. Inhibitors of Protein Convertase Subtilisin/Kexin 9 (PCSK9) and Acute Coronary Syndrome (ACS): The State-of-the-Art. *J. Clin. Med.* **2021**, *10*, 1510. [[CrossRef](#)]
18. Sabatine, M.S.; Giugliano, R.P.; Keech, A.C.; Honarpour, N.; Wiviott, S.D.; Murphy, S.A.; Kuder, J.F.; Wang, H.; Liu, T.; Wasserman, S.M.; et al. Evolocumab and Clinical Outcomes in Patients with Cardiovascular Disease. *N. Engl. J. Med.* **2017**, *376*, 1713–1722. [[CrossRef](#)]
19. Schwartz, G.G.; Steg, P.G.; Szarek, M.; Bhatt, D.L.; Bittner, V.A.; Diaz, R.; Edelberg, J.M.; Goodman, S.G.; Hanotin, C.; Harrington, R.A.; et al. Alirocumab and Cardiovascular Outcomes after Acute Coronary Syndrome. *N. Engl. J. Med.* **2018**, *379*, 2097–2107. [[CrossRef](#)]
20. Paciullo, F.; Momi, S.; Gresele, P. PCSK9 in Haemostasis and Thrombosis: Possible Pleiotropic Effects of PCSK9 Inhibitors in Cardiovascular Prevention. *Thromb. Haemost.* **2019**, *119*, 359–367. [[CrossRef](#)]
21. Basiak, M.; Kosowski, M.; Cyrnek, M.; Bułdak, Ł.; Maligłówa, M.; Machnik, G.; Okopień, B. Pleiotropic Effects of PCSK-9 Inhibitors. *Int. J. Mol. Sci.* **2021**, *22*, 3144. [[CrossRef](#)]
22. Warden, B.A.; Kaufman, T.; Minnier, J.; Duell, P.B.; Fazio, S.; Shapiro, M.D. Use of PCSK9 Inhibitors in Solid Organ Transplantation Recipients. *JACC Case Rep.* **2020**, *2*, 396–399. [[CrossRef](#)] [[PubMed](#)]
23. Simha, V.; Qin, S.; Shah, P.; Smith, B.H.; Kremers, W.K.; Kushwaha, S.; Wang, L.; Pereira, N.L. Sirolimus Therapy Is Associated with Elevation in Circulating PCSK9 Levels in Cardiac Transplant Patients. *J. Cardiovasc. Transl. Res.* **2016**, *10*, 9–15. [[CrossRef](#)] [[PubMed](#)]
24. Sato, S.; Akamine, Y.; Kagaya, H.; Saito, M.; Inoue, T.; Numakura, K.; Habuchi, T.; Satoh, S.; Miura, M. Changes in PCSK9 and LDL cholesterol concentrations by everolimus treatment and their effects on polymorphisms in PCSK9 and mTORC1. *Pharmacol. Rep.* **2020**, *72*, 622–630. [[CrossRef](#)]
25. Chiang, L.W.; Grenier, J.M.; Ettwiller, L.; Jenkins, L.P.; Ficenc, D.; Martin, J.; Jin, F.; DiStefano, P.S.; Wood, A. An orchestrated gene expression component of neuronal programmed cell death revealed by cDNA array analysis. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 2814–2819. [[CrossRef](#)]
26. Piper, D.E.; Jackson, S.; Liu, Q.; Romanow, W.G.; Shetterly, S.; Thibault, S.T.; Shan, B.; Walker, N.P. The Crystal Structure of PCSK9: A Regulator of Plasma LDL-Cholesterol. *Structure* **2007**, *15*, 545–552. [[CrossRef](#)] [[PubMed](#)]
27. Dietschy, J.M.; Turley, S.D.; Spady, D.K. Role of liver in the maintenance of cholesterol and low density lipoprotein homeostasis in different animal species, including humans. *J. Lipid Res.* **1993**, *34*, 1637–1659. [[CrossRef](#)]
28. Abifadel, M.; Guerin, M.; Benjannet, S.; Rabès, J.-P.; Le Goff, W.; Julia, Z.; Hamelin, J.; Carreau, V.; Varret, M.; Bruckert, E.; et al. Identification and characterization of new gain-of-function mutations in the PCSK9 gene responsible for autosomal dominant hypercholesterolemia. *Atherosclerosis* **2012**, *223*, 394–400. [[CrossRef](#)]
29. Abboud, S.; Karhunen, P.J.; Lütjohann, D.; Goebeler, S.; Luoto, T.; Friedrichs, S.; Lehtimäki, T.; Pandolfo, M.; Laaksonen, R. Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9) Gene Is a Risk Factor of Large-Vessel Atherosclerosis Stroke. *PLoS ONE* **2007**, *2*, e1043. [[CrossRef](#)]
30. Melendez, Q.M.; Krishnaji, S.T.; Wooten, C.J.; Lopez, D. Hypercholesterolemia: The role of PCSK9. *Arch. Biochem. Biophys.* **2017**, *625–626*, 39–53. [[CrossRef](#)]
31. Ouguerram, K.; Chetiveaux, M.; Zair, Y.; Costet, P.; Abifadel, M.; Varret, M.; Boileau, C.; Magot, T.; Krempf, M. Apolipoprotein B100 Metabolism in Autosomal-Dominant Hypercholesterolemia Related to Mutations in PCSK9. *Arter. Thromb. Vasc. Biol.* **2004**, *24*, 1448–1453. [[CrossRef](#)]
32. Costet, P.; Cariou, B.; Lambert, G.; Lalanne, F.; Lardeux, B.; Jarnoux, A.-L.; Grefhorst, A.; Staels, B.; Krempf, M. Hepatic PCSK9 Expression Is Regulated by Nutritional Status via Insulin and Sterol Regulatory Element-binding Protein 1c. *J. Biol. Chem.* **2006**, *281*, 6211–6218. [[CrossRef](#)] [[PubMed](#)]
33. Tavori, H.; Giunzioni, I.; Predazzi, I.M.; Plubell, D.; Shivinsky, A.; Miles, J.; DeVay, R.M.; Liang, H.; Rashid, S.; Linton, M.F.; et al. Human PCSK9 promotes hepatic lipogenesis and atherosclerosis development via apoE- and LDLR-mediated mechanisms. *Cardiovasc. Res.* **2016**, *110*, 268–278. [[CrossRef](#)] [[PubMed](#)]
34. Le May, C.; Kourimate, S.; Langhi, C.; Chétiveaux, M.; Jarry, A.; Comera, C.; Collet, X.; Kuipers, F.; Krempf, M.; Cariou, B.; et al. Proprotein Convertase Subtilisin Kexin Type 9 Null Mice Are Protected from Postprandial Triglyceridemia. *Arter. Thromb. Vasc. Biol.* **2009**, *29*, 684–690. [[CrossRef](#)] [[PubMed](#)]
35. Roubtsova, A.; Munkonda, M.N.; Awan, Z.; Marcinkiewicz, J.; Chamberland, A.; Lazure, C.; Cianflone, K.; Seidah, N.G.; Prat, A. Circulating Proprotein Convertase Subtilisin/Kexin 9 (PCSK9) Regulates VLDLR Protein and Triglyceride Accumulation in Visceral Adipose Tissue. *Arter. Thromb. Vasc. Biol.* **2011**, *31*, 785–791. [[CrossRef](#)] [[PubMed](#)]

36. Navab, M.; Ananthramaiah, G.M.; Reddy, S.T.; Van Lenten, B.J.; Ansell, B.J.; Fonarow, G.; Vahabzadeh, K.; Hama, S.; Hough, G.; Kamranpour, N.; et al. The oxidation hypothesis of atherogenesis: The role of oxidized phospholipids and HDL. *J. Lipid Res.* **2004**, *45*, 993–1007. [[CrossRef](#)] [[PubMed](#)]
37. Libby, P.; Hansson, G.K. Inflammation and Immunity in Diseases of the Arterial Tree. *Circ. Res.* **2015**, *116*, 307–311. [[CrossRef](#)]
38. Giunzioni, I.; Tavori, H.; Covarrubias, R.; Major, A.S.; Ding, L.; Zhang, Y.; DeVay, R.M.; Hong, L.; Fan, D.; Predazzi, I.M.; et al. Local effects of human PCSK9 on the atherosclerotic lesion. *J. Pathol.* **2015**, *238*, 52–62. [[CrossRef](#)]
39. Kim, Y.U.; Kee, P.; Danila, D.; Teng, B.-B.; Gim, E.; Shim, D.-W.; Hwang, I.; Shin, O.S.; Yu, J.-W. A Critical Role of PCSK9 in Mediating IL-17-Producing T Cell Responses in Hyperlipidemia. *Immune Netw.* **2019**, *19*, e41. [[CrossRef](#)]
40. Ding, Z.; Liu, S.; Wang, X.; Deng, X.; Fan, Y.; Sun, C.; Wang, Y.; Mehta, J.L. Hemodynamic Shear Stress via ROS Modulates PCSK9 Expression in Human Vascular Endothelial and Smooth Muscle Cells and Along the Mouse Aorta. *Antioxid. Redox Signal.* **2015**, *22*, 760–771. [[CrossRef](#)]
41. Qi, Z.; Hu, L.; Zhang, J.; Yang, W.; Liu, X.; Jia, D.; Yao, Z.; Chang, L.; Pan, G.; Zhong, H.; et al. PCSK9 (Proprotein Convertase Subtilisin/Kexin 9) Enhances Platelet Activation, Thrombosis, and Myocardial Infarct Expansion by Binding to Platelet CD36. *Circulation* **2021**, *143*, 45–61. [[CrossRef](#)]
42. Barale, C.; Melchionda, E.; Morotti, A.; Russo, I. PCSK9 Biology and Its Role in Atherothrombosis. *Int. J. Mol. Sci.* **2021**, *22*, 5880. [[CrossRef](#)] [[PubMed](#)]
43. Saenko, E.L.; Yakhyayev, A.V.; Mikhailenko, I.; Strickland, D.K.; Sarafanov, A.G. Role of the Low Density Lipoprotein-related Protein Receptor in Mediation of Factor VIII Catabolism. *J. Biol. Chem.* **1999**, *274*, 37685–37692. [[CrossRef](#)] [[PubMed](#)]
44. Martinelli, N.; Girelli, D.; Lunghi, B.; Pinotti, M.; Marchetti, G.; Malerba, G.; Pignatti, P.F.; Corrocher, R.; Olivieri, O.; Bernardi, F. Polymorphisms at LDLR locus may be associated with coronary artery disease through modulation of coagulation factor VIII activity and independently from lipid profile. *Blood* **2010**, *116*, 5688–5697. [[CrossRef](#)] [[PubMed](#)]
45. Zhang, Y.; Zhu, C.-G.; Xu, R.-X.; Li, S.; Guo, Y.-L.; Sun, J.; Li, J.-J. Relation of circulating PCSK9 concentration to fibrinogen in patients with stable coronary artery disease. *J. Clin. Lipidol.* **2014**, *8*, 494–500. [[CrossRef](#)]
46. Ridker, P.M.; Tardif, J.-C.; Amarenco, P.; Duggan, W.; Glynn, R.J.; Jukema, J.W.; Kastelein, J.J.P.; Kim, A.; Koenig, W.; Nissen, S.; et al. Lipid-Reduction Variability and Antidrug-Antibody Formation with Bococizumab. *N. Engl. J. Med.* **2017**, *376*, 1517–1526. [[CrossRef](#)]
47. Murphy, S.A.; Pedersen, T.R.; Gaciong, Z.A.; Ceska, R.; Ezhov, M.V.; Connolly, D.L.; Jukema, J.W.; Toth, K.; Tikkanen, M.J.; Im, K.; et al. Effect of the PCSK9 Inhibitor Evolocumab on Total Cardiovascular Events in Patients With Cardiovascular Disease: A Prespecified Analysis from the FOURIER Trial. *JAMA Cardiol.* **2019**, *4*, 613–619. [[CrossRef](#)]
48. Robinson, J.G.; Farnier, M.; Krempf, M.; Bergeron, J.; Luc, G.; Averna, M.; Stroes, E.S.; Langslet, G.; Raal, F.J.; El Shahawy, M.; et al. Efficacy and Safety of Alirocumab in Reducing Lipids and Cardiovascular Events. *N. Engl. J. Med.* **2015**, *372*, 1489–1499. [[CrossRef](#)]
49. O'Donoghue, M.L.; Fazio, S.; Giugliano, R.P.; Stroes, E.S.; Kanevsky, E.; Gouni-Berthold, I.; Im, K.; Pineda, A.L.; Wasserman, S.M.; Češka, R.; et al. Lipoprotein(a), PCSK9 Inhibition, and Cardiovascular Risk. *Circulation* **2019**, *139*, 1483–1492. [[CrossRef](#)]
50. Nordestgaard, B.G.; Chapman, M.J.; Ray, K.; Borén, J.; Andreotti, F.; Watts, G.; Ginsberg, H.; Amarenco, P.; Catapano, A.L.; Descamps, O.S.; et al. Lipoprotein(a) as a cardiovascular risk factor: Current status. *Eur. Heart J.* **2010**, *31*, 2844–2853. [[CrossRef](#)]
51. Gentile, M.; Simeon, V.; Iannuzzo, G.; Mattiello, A.; di Taranto, M.D.; Panico, S.; Rubba, P. Lipoprotein (a) is an independent predictor of cardiovascular events in Mediterranean women (Progetto Atena). *Eur. J. Prev. Cardiol.* **2019**, *27*, 2248–2250. [[CrossRef](#)] [[PubMed](#)]
52. Nicholls, S.J.; Puri, R.; Anderson, T.; Ballantyne, C.M.; Cho, L.; Kastelein, J.J.P.; Koenig, W.; Somaratne, R.; Kassahun, H.; Yang, J.; et al. Effect of Evolocumab on Progression of Coronary Disease in Statin-Treated Patients: The GLAGOV Randomized Clinical Trial. *JAMA* **2016**, *316*, 2373–2384. [[CrossRef](#)] [[PubMed](#)]
53. Poggio, E.D.; Augustine, J.J.; Arrigain, S.; Brennan, D.C.; Schold, J.D. Long-term kidney transplant graft survival—Making progress when most needed. *Am. J. Transplant.* **2020**, *21*, 2824–2832. [[CrossRef](#)] [[PubMed](#)]
54. Albeldawi, M.; Aggarwal, A.; Madhwal, S.; Cywinski, J.; Lopez, R.; Eghtesad, B.; Zein, N.N. Cumulative risk of cardiovascular events after orthotopic liver transplantation. *Liver Transplant.* **2011**, *18*, 370–375. [[CrossRef](#)]
55. Tsai, H.-I.; Liu, F.-C.; Lee, C.-W.; Kuo, C.-F.; See, L.-C.; Chung, T.-T.; Yu, H.-P. Cardiovascular disease risk in patients receiving organ transplantation: A national cohort study. *Transpl. Int.* **2017**, *30*, 1161–1171. [[CrossRef](#)]
56. Gansevoort, R.T.; Correa-Rotter, R.; Hemmelgarn, B.R.; Jafar, T.H.; Heerspink, H.J.L.; Mann, J.F.; Matsushita, K.; Wen, C.P. Chronic kidney disease and cardiovascular risk: Epidemiology, mechanisms, and prevention. *Lancet* **2013**, *382*, 339–352. [[CrossRef](#)]
57. Hüsing, A.; Kabar, I.; Schmidt, H. Lipids in liver transplant recipients. *World J. Gastroenterol.* **2016**, *22*, 3315–3324. [[CrossRef](#)] [[PubMed](#)]
58. Tóth, P.P.; Potter, D.; Ming, E.E. Prevalence of lipid abnormalities in the United States: The National Health and Nutrition Examination Survey 2003–2006. *J. Clin. Lipidol.* **2012**, *6*, 325–330. [[CrossRef](#)]
59. Pan, L.; Yang, Z.; Wu, Y.; Yin, R.-X.; Liao, Y.; Wang, J.; Gao, B.; Zhang, L. The prevalence, awareness, treatment and control of dyslipidemia among adults in China. *Atherosclerosis* **2016**, *248*, 2–9. [[CrossRef](#)]
60. González-Amieva, A.; López-Miranda, J.; Marín, C.; Pérez-Martínez, P.; Gómez, P.; Paz-Rojas, E.; Arizón, J.M.; Jiménez-Perepérez, J.A.; Concha, M.; Pérez-Jiménez, F. The apo A-I gene promoter region polymorphism determines the severity of hyperlipidemia after heart transplantation. *Clin. Transplant.* **2003**, *17*, 56–62. [[CrossRef](#)]

61. Taegtmeier, A.B.; Breen, J.B.; Smith, J.; Rogers, P.; Kullak-Ublick, G.A.; Yacoub, M.H.; Banner, N.R.; Barton, P.J.R. Effect of ABCB1 Genotype on Pre- and Post-Cardiac Transplantation Plasma Lipid Concentrations. *J. Cardiovasc. Transl. Res.* **2011**, *4*, 304–312. [[CrossRef](#)] [[PubMed](#)]
62. Numakura, K.; Kagaya, H.; Yamamoto, R.; Komine, N.; Saito, M.; Hiroshi, T.; Akihama, S.; Inoue, T.; Narita, S.; Tsuchiya, N.; et al. Characterization of Clinical and Genetic Risk Factors Associated with Dyslipidemia after Kidney Transplantation. *Dis. Markers* **2015**, *2015*, 179434. [[CrossRef](#)] [[PubMed](#)]
63. Demir, E.; Balal, M.; Paydas, S.; Sertdemir, Y.; Erken, U. Dyslipidemia and Weight Gain Secondary to Lifestyle Changes in Living Renal Transplant Donors. *Transplant. Proc.* **2005**, *37*, 4176–4179. [[CrossRef](#)]
64. Pinto, A.S.; Chedid, M.F.; Guerra, L.T.; Cabeleira, D.D.; Kruel, C.D.P. Dietary management for dyslipidemia in liver transplant recipients. *Arq. Bras. Cir. Dig.* **2016**, *29*, 246–251. [[CrossRef](#)] [[PubMed](#)]
65. Miller, L.W. Cardiovascular Toxicities of Immunosuppressive Agents. *Am. J. Transplant.* **2002**, *2*, 807–818. [[CrossRef](#)] [[PubMed](#)]
66. Miller, L.W.; Schlant, R.C.; Kobashigawa, J.; Kubo, S.; Renlund, D.G. Task force 5: Complications. *J. Am. Coll. Cardiol.* **1993**, *22*, 41–54. [[CrossRef](#)]
67. Ricoult, S.J.H.; Manning, B.D. The multifaceted role of mTORC1 in the control of lipid metabolism. *EMBO Rep.* **2013**, *14*, 242–251. [[CrossRef](#)]
68. Murakami, N.; Riella, L.V.; Funakoshi, T. Risk of Metabolic Complications in Kidney Transplantation After Conversion to mTOR Inhibitor: A Systematic Review and Meta-Analysis. *Am. J. Transplant.* **2014**, *14*, 2317–2327. [[CrossRef](#)]
69. Kurdi, A.; Martinet, W.; De Meyer, G. mTOR Inhibition and Cardiovascular Diseases. *Transplantation* **2018**, *102*, S44–S46. [[CrossRef](#)]
70. Nguyen, V.N.; Abagyan, R.; Tsunoda, S.M. Mtor inhibitors associated with higher cardiovascular adverse events—A large population database analysis. *Clin. Transplant.* **2021**, *35*, e14228. [[CrossRef](#)]
71. Derfler, K.; Hayde, M.; Heinz, G.; Hirschl, M.M.; Steger, G.; Hauser, A.-C.; Balcke, P.; Widhalm, K. Decreased postheparin lipolytic activity in renal transplant recipients with cyclosporin A. *Kidney Int.* **1991**, *40*, 720–727. [[CrossRef](#)]
72. Princen, H.M.; Meijer, P.; Wolthers, B.G.; Vonk, R.J.; Kuipers, F. Cyclosporin A blocks bile acid synthesis in cultured hepatocytes by specific inhibition of chenodeoxycholic acid synthesis. *Biochem. J.* **1991**, *275*, 501–505. [[CrossRef](#)]
73. De Groen, P.C. Cyclosporine, Low-Density Lipoprotein, and Cholesterol. *Mayo Clin. Proc.* **1988**, *63*, 1012–1021. [[CrossRef](#)]
74. Montero, N.; Pascual, J. Immunosuppression and Post-transplant Hyperglycemia. *Curr. Diabetes Rev.* **2015**, *11*, 144–154. [[CrossRef](#)]
75. Jenssen, T.; Hartmann, A. Post-transplant diabetes mellitus in patients with solid organ transplants. *Nat. Rev. Endocrinol.* **2019**, *15*, 172–188. [[CrossRef](#)] [[PubMed](#)]
76. Roccaro, G.A.; Goldberg, D.S.; Hwang, W.-T.; Judy, R.; Thomasson, A.; Kimmel, S.E.; Forde, K.A.; Lewis, J.D.; Yang, Y.-X. Sustained Posttransplantation Diabetes Is Associated with Long-Term Major Cardiovascular Events Following Liver Transplantation. *Am. J. Transplant.* **2017**, *18*, 207–215. [[CrossRef](#)]
77. Cron, D.C.; Noon, K.A.; Cote, D.R.; Terjimanian, M.N.; Augustine, J.J.; Wang, S.C.; Englesbe, M.J.; Woodside, K.J. Using analytic morphomics to describe body composition associated with post-kidney transplantation diabetes mellitus. *Clin. Transplant.* **2017**, *31*, e13040. [[CrossRef](#)] [[PubMed](#)]
78. Sharif, A.; Moore, R.; Baboolal, K. Influence of Lifestyle Modification in Renal Transplant Recipients With Postprandial Hyperglycemia. *Transplantation* **2008**, *85*, 353–358. [[CrossRef](#)]
79. McCaughan, J.A.; McKnight, A.J.; Maxwell, A.P. Genetics of New-Onset Diabetes after Transplantation. *J. Am. Soc. Nephrol.* **2013**, *25*, 1037–1049. [[CrossRef](#)]
80. Gervasini, G.; Luna, E.; García-Cerrada, M.; García-Pino, G.; Cubero, J.J. Risk factors for post-transplant diabetes mellitus in renal transplant: Role of genetic variability in the CYP450-mediated arachidonic acid metabolism. *Mol. Cell. Endocrinol.* **2016**, *419*, 158–164. [[CrossRef](#)] [[PubMed](#)]
81. Midtvedt, K. Insulin Resistance after Renal Transplantation: The Effect of Steroid Dose Reduction and Withdrawal. *J. Am. Soc. Nephrol.* **2004**, *15*, 3233–3239. [[CrossRef](#)]
82. Schäcke, H.; Döcke, W.-D.; Asadullah, K. Mechanisms involved in the side effects of glucocorticoids. *Pharmacol. Ther.* **2002**, *96*, 23–43. [[CrossRef](#)]
83. Vincenti, F.; Friman, S.; Scheuermann, E.; Rostaing, L.; Jenssen, T.; Campistol, J.M.; Uchida, K.; Pescovitz, M.D.; Marchetti, P.; Tuncer, M.; et al. Results of an International, Randomized Trial Comparing Glucose Metabolism Disorders and Outcome with Cyclosporine Versus Tacrolimus. *Am. J. Transplant.* **2007**, *7*, 1506–1514. [[CrossRef](#)] [[PubMed](#)]
84. Chakkerla, H.A.; Mandarino, L.J. Calcineurin Inhibition and New-Onset Diabetes Mellitus After Transplantation. *Transplantation* **2013**, *95*, 647–652. [[CrossRef](#)]
85. Fraenkel, M.; Ketzinel-Gilad, M.; Ariav, Y.; Pappo, O.; Karaca, M.; Castel, J.; Berthault, M.-F.; Magnan, C.; Cerasi, E.; Kaiser, N.; et al. mTOR Inhibition by Rapamycin Prevents β -Cell Adaptation to Hyperglycemia and Exacerbates the Metabolic State in Type 2 Diabetes. *Diabetes* **2008**, *57*, 945–957. [[CrossRef](#)] [[PubMed](#)]
86. Johnston, O.; Rose, C.L.; Webster, A.C.; Gill, J.S. Sirolimus Is Associated with New-Onset Diabetes in Kidney Transplant Recipients. *J. Am. Soc. Nephrol.* **2008**, *19*, 1411–1418. [[CrossRef](#)] [[PubMed](#)]
87. Singer, D.R.; Jenkins, G.H. Hypertension in transplant recipients. *J. Hum. Hypertens.* **1996**, *10*, 395–402.

88. Sudhir, K.; MacGregor, J.S.; DeMarco, T.; De Groot, C.J.; Taylor, R.N.; Chou, T.M.; Yock, P.G.; Chatterjee, K. Cyclosporine impairs release of endothelium-derived relaxing factors in epicardial and resistance coronary arteries. *Circulation* **1994**, *90*, 3018–3023. [[CrossRef](#)]
89. Ventura, H.O.; Mehra, M.; Stapleton, D.D.; Smart, F.W. Cyclosporine-induced hypertension in cardiac transplantation. *Med. Clin. N. Am.* **1997**, *81*, 1347–1357. [[CrossRef](#)]
90. Ruiz, M.; Medina, A.; Moreno, J.; Gómez, I.; Ruiz, N.; Bueno, P.; Asensio, C.; Osuna, A. Relationship Between Oxidative Stress Parameters and Atherosclerotic Signs in the Carotid Artery of Stable Renal Transplant Patients. *Transplant. Proc.* **2005**, *37*, 3796–3798. [[CrossRef](#)]
91. Urbanowicz, T.; Michalak, M.; Gąsecka, A.; Ołasińska-Wiśniewska, A.; Perek, B.; Rodzki, M.; Bociński, M.; Jemielity, M. A Risk Score for Predicting Long-Term Mortality Following Off-Pump Coronary Artery Bypass Grafting. *J. Clin. Med.* **2021**, *10*, 3032. [[CrossRef](#)] [[PubMed](#)]
92. Khush, K.K.; Cherikh, W.S.; Chambers, D.C.; Goldfarb, S.; Hayes, D.; Kucheryavaya, A.Y.; Levvey, B.J.; Meiser, B.; Rossano, J.W.; Stehlik, J. The International Thoracic Organ Transplant Registry of the International Society for Heart and Lung Transplantation: Thirty-fifth Adult Heart Transplantation Report—2018; Focus Theme: Multiorgan Transplantation. *J. Heart Lung Transplant.* **2018**, *37*, 1155–1168. [[CrossRef](#)]
93. Lu, W.-H.; Palatnik, K.; Fishbein, G.A.; Lai, C.; Levi, D.S.; Perens, G.; Alejos, J.; Kobashigawa, J.; Fishbein, M.C. Diverse morphologic manifestations of cardiac allograft vasculopathy: A pathologic study of 64 allograft hearts. *J. Heart Lung Transplant.* **2011**, *30*, 1044–1050. [[CrossRef](#)] [[PubMed](#)]
94. Patel, C.B.; Holley, C. Cardiac Allograft Vasculopathy. *J. Am. Coll. Cardiol.* **2019**, *74*, 52–53. [[CrossRef](#)]
95. Ghanem, H.; Dorpel, M.A.V.D.; Weimar, W.; Veld, A.J.M.I.; El-Kannishy, M.H.; Jansen, H. Increased low density lipoprotein oxidation in stable kidney transplant recipients. *Kidney Int.* **1996**, *49*, 488–493. [[CrossRef](#)]
96. Najafian, B.; Kasiske, B.L. Chronic allograft nephropathy. *Curr. Opin. Nephrol. Hypertens.* **2008**, *17*, 149–155. [[CrossRef](#)] [[PubMed](#)]
97. Bosmans, J.-L.; Holvoet, P.; Dauwe, S.E.; Ysebaert, D.K.; Chapelle, T.; Jürgens, A.; Kovacic, V.; Van Marck, E.A.; De Broe, M.E.; Verpooten, G.A. Oxidative modification of low-density lipoproteins and the outcome of renal allografts at 11/2 years. *Kidney Int.* **2001**, *59*, 2346–2356. [[CrossRef](#)] [[PubMed](#)]
98. Matsushita, K.; Van Der Velde, M.; Astor, B.C.; Woodward, M.; Levey, A.S.; De Jong, P.E.; Coresh, J.; Gansevoort, R.T. Association of estimated glomerular filtration rate and albuminuria with all-cause and cardiovascular mortality in general population cohorts: A collaborative meta-analysis. *Lancet* **2010**, *375*, 2073–2081. [[CrossRef](#)]
99. Harris, J.; Teuteberg, J.; Shullo, M. Optimal low-density lipoprotein concentration for cardiac allograft vasculopathy prevention. *Clin. Transplant.* **2018**, *32*, e13248. [[CrossRef](#)]
100. Schmidt, A.F.; Pearce, L.S.; Wilkins, J.T.; Overington, J.; Hingorani, A.; Casas, J.P. PCSK9 monoclonal antibodies for the primary and secondary prevention of cardiovascular disease. *Cochrane Database Syst. Rev.* **2017**, *4*, CD011748. [[CrossRef](#)]
101. Sandesara, P.B.; Dhindsa, D.; Hirsh, B.; Jokhadar, M.; Cole, R.T.; Sperling, L.S. PCSK9 inhibition in patients with heart transplantation: A case series. *J. Clin. Lipidol.* **2019**, *13*, 721–724. [[CrossRef](#)] [[PubMed](#)]
102. Moayed, Y.; Kozuszko, S.; Knowles, J.W.; Chih, S.; Oro, G.; Lee, R.; Fearon, W.F.; Ross, H.J.; Teuteberg, J.J.; Khush, K.K. Safety and Efficacy of PCSK9 Inhibitors After Heart Transplantation. *Can. J. Cardiol.* **2018**, *35*, 104.e1–104.e3. [[CrossRef](#)] [[PubMed](#)]
103. Sammour, Y.; Dezorzi, C.; Austin, B.A.; Borkon, A.M.; Everley, M.P.; Fendler, T.J.; Khumri, T.M.; Lawhorn, S.L.; Nassif, M.E.; Vodnala, D.; et al. PCSK9 Inhibitors in Heart Transplant Patients: Safety, Efficacy, and Angiographic Correlates. *J. Card. Fail.* **2021**, *27*, 812–815. [[CrossRef](#)] [[PubMed](#)]
104. Broch, K.; Gude, E.; Karason, K.; Dellgren, G.; Rådegran, G.; Gjesdal, G.; Gustafsson, F.; Eiskjaer, H.; Lommi, J.; Pentikäinen, M.; et al. Cholesterol lowering with EVOLocumab to prevent cardiac allograft Vasculopathy in De-novo heart transplant recipients: Design of the randomized controlled EVOLVD trial. *Clin. Transplant.* **2020**, *34*, e13984. [[CrossRef](#)]
105. Papisotiriou, M.; Ntrinas, T.; Savvidaki, E.; Papachristou, E.; Goumenos, D.S. Treatment of Mixed Dyslipidemia with Alirocumab in a Kidney Transplant Recipient: A Case Report. *Transplant. Proc.* **2021**, *53*, 2775–2778. [[CrossRef](#)]
106. Ordóñez-Fernández, L.; Rodríguez-Ferreras, A.; Carriles, C.; Martínez-Torrón, A.; Lázaro-López, E.; Rosado-María, M.C. Pneumonia in a patient with kidney transplant treated with alirocumab and everolimus. *Farm. Hosp.* **2019**, *43*, 74–76. [[CrossRef](#)]
107. Melexopoulou, C.; Marinaki, S.; Oikonomou, E.; Bonios, M.J.; Theofilis, P.; Miliou, A.; Siasos, G.; Tousoulis, D.; Boletis, J.N. PCSK9 and inflammatory biomarkers in the early post kidney transplantation period. *Eur. Rev. Med. Pharmacol. Sci.* **2021**, *25*, 4762–4772. [[CrossRef](#)] [[PubMed](#)]
108. Wu, N.-Q.; Shi, H.-W.; Li, J.-J. Proprotein Convertase Subtilisin/Kexin Type 9 and Inflammation: An Updated Review. *Front. Cardiovasc. Med.* **2022**, *9*, 763516. [[CrossRef](#)]
109. Leander, K.; Mälarstig, A.; Hoof, F.M.V.; Hyde, C.; Hellénus, M.-L.; Troutt, J.S.; Konrad, R.J.; Öhrvik, J.; Hamsten, A.; de Faire, U. Circulating Proprotein Convertase Subtilisin/Kexin Type 9 (PCSK9) Predicts Future Risk of Cardiovascular Events Independently of Established Risk Factors. *Circulation* **2016**, *133*, 1230–1239. [[CrossRef](#)]
110. Eisenga, M.F.; Zelle, D.M.; Sloan, J.H.; Gaillard, C.A.; Bakker, S.J.; Dullaart, R.P. High Serum PCSK9 Is Associated With Increased Risk of New-Onset Diabetes After Transplantation in Renal Transplant Recipients. *Diabetes Care* **2017**, *40*, 894–901. [[CrossRef](#)] [[PubMed](#)]

111. Chandra, N.C. A comprehensive account of insulin and LDL receptor activity over the years: A highlight on their signaling and functional role. *J. Biochem. Mol. Toxicol.* **2021**, *35*, e22840. [[CrossRef](#)] [[PubMed](#)]
112. Ai, D.; Chen, C.; Han, S.; Ganda, A.; Murphy, A.J.; Haeusler, R.; Thorp, E.; Accili, D.; Horton, J.D.; Tall, A.R. Regulation of hepatic LDL receptors by mTORC1 and PCSK9 in mice. *J. Clin. Investig.* **2012**, *122*, 1262–1270. [[CrossRef](#)] [[PubMed](#)]