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Infectious Complications in Autoimmune Hemolytic Anemia

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Abstract: Autoimmune hemolytic anemia (AIHA) may be frequently challenged by infectious complications, mainly as a result of immunosuppressive treatments administered. Furthermore, infectious agents are known triggers of AIHA onset and relapse. Although being risk factors for mortality, infections are an underestimated issue in AIHA. This review will collect the available evidence on the frequency and type of infectious complications in AIHA, detailing the risk related to each treatment (i.e., steroids, rituximab, splenectomy, classic immunosuppressive agents, and new target drugs). Moreover, we will briefly discuss the infectious complications in AIHA secondary to other diseases that harbor an intrinsic infectious risk (e.g., primary immunodeficiencies, systemic autoimmune diseases, lymphoproliferative disorders, solid organ and hematopoietic stem cell transplants). Finally, viral and bacterial reactivations during immune suppressive therapies will be discussed, along with suggested screening and prophylactic strategies.

Keywords: autoimmune hemolytic anemia; steroids; rituximab; immunosuppressants; prophylaxis; viral reactivation



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1. Introduction

Autoimmune hemolytic anemia (AIHA) encompasses a group of heterogeneous conditions mainly characterized by red blood cell (RBC) lysis due to autoantibodies against surface erythrocyte's antigens. Based on the thermal characteristics of the autoantibody, AIHAs can be classified into warm forms, generally caused by IgG antibodies reacting at warm temperatures and able to fix complement in some cases; cold agglutinin disease (CAD), due to IgM antibodies that agglutinate RBCs at low temperatures and lyse them via the complement cascade activation; and mixed forms (coexistence of warm and cold autoantibodies) [1,2]. Infections in AIHA are a known player in the pathogenesis of the autoimmune process. On the other hand, infections can occur also as consequence of the disease and its treatments. There is increasing awareness of infections in AIHA, as they can impact on outcome, including morbidity and fatality. Additionally, AIHA can be secondary to systemic autoimmune diseases and lymphoproliferative disorders, whose treatments may further increase the infectious risk. Likewise, AIHA is frequently observed in primary immunodeficiencies (PIDs) that are characterized by a well-known infectious diathesis. The clinical management of infections and prophylactic measures in primary AIHA remains largely unknown, at variance with secondary forms. The only available data derive mainly from retrospective series and case reports, or from more recent clinical trials with novel drugs.

In this review we will focus on the frequency and type of infectious complications in primary and secondary AIHA. We will discuss the infectious risk related to each treatment, including new target drugs, and the issue of viral and mycobacterial reactivations during immunosuppressive therapies. Screening and prophylactic strategies will be also provided.

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2. Prevalence of Infections in Primary AIHA

In a large multicenter Italian study on 308 primary AIHAs, infectious complications were registered in 26 patients (8.4%), of whom 11 were grade 4 (according to the Common Terminology Criteria for Adverse Events) and five were fatal [3]. They consisted mostly in pneumonia, and the causative agents were *Pneumocystis jirovecii*, Mycoplasma pneumoniae, Staphylococcus epidermidis, Pseudomonas aeruginosa, Varicella Zoster virus (VZV), and Candida albicans. They were not associated with AIHA type and severity nor with the number of medical therapies. Notably, splenectomy was the only factor associated with severe infectious complications (grade ≥ 3). The same authors in a subsequent larger series with an extended follow-up reported a higher proportion of infections (14%), mostly in warm and mixed subtypes [4]. In another study on 33 primary AIHA patients from South India followed for a median period of 50 months, four developed infective episodes (two pneumonias, one popliteal abscess, and one sepsis, with no isolations): one was grade 2, two were grade 4, and one resulted in a fatal sepsis. In detail, one pneumonia occurred on a very low dose of prednisone (<5 mg/day) and azathioprine (AZA), while the other complicated a cerebral vein thrombosis requiring intensive care [5]. More recently, in an Italian single-center analysis of 225 primary AIHAs, a total of 45 infections were recorded in 29 patients (up to four episodes in one patient) over a 3-year follow-up. Two third of infections were >G3 and two were fatal (post-splenectomy sepsis and Pneumocystis jiroveci pneumonia). Of note, 60% occurred during an active phase of the disease and 14% at the time of AIHA diagnosis. Patients with infectious complications also had higher rates of thrombotic events and Evans' syndrome (association with immune thrombocytopenia). Additionally, patients with infections had received more lines of therapy, particularly rituximab, splenectomy, and immunosuppressants (AZA, cyclophosphamide and cyclosporine) [6]. Focusing on fatal outcome, a study of 83 AIHA patients in the period 1980–2000 reported 13 deaths, of which five related to infective episodes [7]. A single-center French experience on 60 warm AIHAs reported two deaths, both occurring in splenectomized patients, and still on immunosuppressive therapy. In detail, the first patient had superimposed pneumococcal-related sepsis complicating a H1N1 influenza pneumonia, and the second a Gram-negative bacilli-related sepsis in a previous history of pneumocystosis and pulmonary aspergillosis [8]. In the Italian study [3], the occurrence of infections was strongly associated with death (hazard ratio 11.47; 95% CI 3.43–38.4, p = 0.0004). Finally, a study of 101 warm AIHA performed in Thailand indicated sepsis as the most common cause of death, supervened in 11% of patients [9]. Altogether these findings indicate that infections in AIHA occur in 6–14% of cases, and may result in fatal outcome.

3. Infectious Risk Associated with AIHA Therapies

The risk and type of infections associated with AIHA treatments differ according to the dose, the time of exposure and the depth of immunosuppression induced by each therapy. Table 1 summarizes the main findings for the different AIHA treatments, detailed separately as follows.

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Table 1. Infectious risk associated with autoimmune hemolytic anemia (AIHA) treatments.

Treatment	Main Warnings	References
Corticosteroids	 Infectious risk is dose-dependent Also prolonged use of low-dose steroids is associated with atypical and opportunistic infections 	[10–12]
Rituximab	 Safe as single agent Risk of hepatitis B virus reactivation, if antiviral prophylaxis not instituted Risk increases in chemotherapy-combined regimens or in the context of severe immunodepression (warning for PML) 	[13–17]
Splenectomy	 Infections in 6–7% of AIHA patients Encapsulated bacteria are the main pathogens isolated in OPSI, which can be fatal Risk decreases with proper patient's education and vaccinations 	[18,19]
Classic immunosuppressive agents	- CTX, MMF, and AZA are associated with increased infectious risk by several pathogens - Cyclosporine seems safer than the abovementioned drugs	
Complement inhibitors	- Increased risk of encapsulated bacterial infections	[23,24]
BCR pathway antagonists	- PI3Kδ inhibitors are associated to PJP - Fostamatinib (used in RA patients) has an increased infectious risk	
Proteasome inhibitors	- Apparently safe in AIHA	[27,28]
FcRn antagonists	- Reported to be safe in ITP patients	[29,30]

PML: progressive multifocal leukoencephalopathy, AIHA: autoimmune hemolytic anemia, OPSI: overwhelming post-splenectomy infection, CTX: cyclophosphamide, MMF: mycophenolate mofetil, AZA: azathioprine, BCR: B-cell receptor, PI3Kδ: phosphoinositide 3-kinase delta, PJP: *Pneumocystis jirovecii* pneumonia, RA: rheumatoid arthritis, FcRn: neonatal Fc receptor, ITP: immune thrombocytopenia.

3.1. Steroids

Steroid-associated infectious risk has been largely reported, together with other side effects (i.e., osteoporosis, diabetes mellitus, and hypertension). The mechanisms by which corticosteroids impair the immune response against pathogens are multiple, including reduced opsonization and phagocytosis of bacteria, impaired T cell function (increasing the risk for mycobacterial, viral, and fungal infection), and enhanced eosinophil apoptosis (favoring parasitic infections). In fact, several opportunistic infections have been reported, i.e., Pneumocystis jirovecii pneumonia (PJP; especially with doses >30 mg/day) [31], aspergillosis, candidiasis, strongyloidiasis, cryptococcosis, and VZV and tuberculosis (TB) reactivations [32]. A recent large study (more than 275,000 adults with various conditions) reported a significantly higher risk of infections in the steroid-exposed group. In detail, hazard risk ranged from 2.01 for cutaneous cellulitis to 5.84 for lower respiratory tract infections, and it correlated positively with steroid dose, independently of the underlying condition [10]. In rheumatoid arthritis (RA), the increased risk of infections was attributable to steroids even at low doses (i.e., 5 mg/day or less) [33]. Moreover, a case-control study conducted on almost 12,000 over-65 RA patients found that continuous treatment with 5 mg prednisolone for the last 3 months, 6 months, or 3 years had a 30%, 46%, or 100% increased risk of serious infections, respectively, halving the risk only many months after discontinuation [11]. Similarly, in systemic lupus erythematosus (SLE) infections are one of the leading causes of morbidity and mortality, and a dose >7.5–10 mg/day of prednisone is a well-recognized risk factor [34]. Although being the backbone therapy, no studies addressed the steroid-related infectious risk in AIHA. Retrospective data showed that primary

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AIHA patients experiencing infections received a mean cumulative dose of corticosteroids of 8.2 kg for a median time of 12 months before the first event [6]. Finally, it is noteworthy the description of six cases of cryptococcal infections in AIHA treated with steroids only. One was an elderly non-HIV non-transplanted patient who developed disseminated cryptococcal disease while receiving high-dose prednisone (100 mg/day) [35]; the other five AIHA patients contracted cryptococcal meningitis while on prednisone >15 mg/day [12]. Altogether, these data suggest that steroids represent a risk factor for infections, particularly at high doses but also at low and prolonged regimens. Moreover, they are also associated with infections caused by uncommon agents, including fungi and parasites.

3.2. Rituximab

Rituximab is a chimeric anti-CD20 monoclonal antibody targeting B cells, used as single agent or combined with chemotherapy. It has proven effective both in warm and cold AIHAs, representing the preferred option at relapse in the former and the first-line treatment in the latter [1,2,13]. Rituximab has been associated with an increased infectious risk, related to its B cell and immunoglobulin-depleting effect. A clear association is established between rituximab and progressive multifocal leukoencephalopathy (PML), mostly in hematologic malignancies and bone marrow transplantations [14]. PML is a devastating demyelinating disease of central nervous system caused by the reactivation of John Cunningham virus (JCV), a polyomavirus that latently infects the kidneys of almost 50% of healthy adults. More uncommon infections related to rituximab are described in retrospective case series, and include PJP, enterovirus encephalitis, parvovirus B19, cytomegalovirus (CMV), West Nile virus, and babesiosis [36]. In follicular lymphoma, a meta-analysis shows that severe infections occurred when the drug is used as maintenance therapy [37]. At variance with lymphoproliferative disorders, data on systemic autoimmune disorders indicate that rituximab is not associated with a significant infectious risk. In fact, PML is only a rare complication, reported to be less than 2/100,000 patients in systemic vasculitides [38]. Moreover, only mild infections are reported in RA clinical trials [39]. Finally, a recent systematic review evaluating rituximab use in autoimmune diseases found no difference in infectious rates between rituximab- vs. non-rituximab-treated patients [15].

As regards AIHA, a meta-analysis including 21 studies reported an incidence of about 5% of severe infections, including one PJP [16]. A similar incidence was found in a French retrospective study of autoimmune cytopenias associated with SLE. In this cohort severe non-opportunistic infections occurred in 4.2% patients, with an estimated incidence of 1.2 severe infections/100 patient-years [40]. In cold AIHA, rituximab showed a good safety profile, with only 3% of G1 infections reported, although one fatal pneumonia 9 months after the end of therapy was recorded [41]. The same good safety profile has been reported in a cohort of elderly AIHA patients, in which only two urinary tract infections were registered over a median 31-month follow-up period [42]. As regards the low-dose rituximab regimen (i.e., 100 mg i.v. weekly for 4 weeks), no infections were registered in a median follow-up of 15 months (range 6–35) [43]. Surprisingly, the only study reporting a higher incidence of infections is an Asian case-control study in which the infectious rate was about 35% with low-dose rituximab, comparable to the cyclophosphamide (CTX)-treated arm [44]. Infection rates rise when rituximab is combined to chemotherapy. In perspective studies, Berentsen et al. reported an infection rate of 11% for bendamustine association and up to 59% when associated to fludarabine, including two fatal pneumonias [45,46]. Finally, no published data exist about PML incidence in AIHA, although this complication should be always considered in immunocompromised subjects. Taken together, these results suggest that rituximab as single agent is safe in AIHA, although associations with chemotherapy deserve higher attention.

3.3. Splenectomy

Splenectomy shows response rates similar to rituximab in warm AIHA, although long-term outcomes are poorly known; it is not effective in CAD, where extravascular

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hemolysis occurs mainly in the liver [1,2]. Splenectomy exposes patients to an increased risk of infections along with thrombotic events. For all these reasons, it is usually deferred after other second-line medical therapies; nonetheless, it still represents an option in multi-refractory warm AIHAs. A literature review for the period 1966–1996, including 6942 splenectomised patients for different reasons, found an infectious crude rate of 3% [47]. More recently, in a Danish nationwide analysis of about 4000 splenectomised patients [48] the overall incidence of infections was 7.7/100 patient-years vs. 2/100 for the general population. Finally, a multicenter analysis of 233 splenectomised immune thrombocytopenic (ITP) patients [49] reported a total number of 159 infections (two of them fatal) in 31% of patients. The main threat in asplenic patients consists of encapsulated bacteria, whose phagocytosis is impaired in the absence of splenic macrophages. In a large study including 349 septic episodes in asplenic patients, Streptococcus pneumonia was responsible for 57% of infections and 59% of deaths; Haemophilus influenzae for 6% of infections, with a mortality rate of 32%; and Neisseria meningitidis caused 3.7% of events [18]. A particularly harmful event is overwhelming post-splenectomy infection (OPSI), i.e., a fulminating sepsis, meningitis, or pneumonia caused by encapsulated bacteria. It occurs more commonly within the first two years (range: 1 week to 20 years) and may rapidly evolve in few hours to death, if not adequately and timely treated. The mortality rate ranges from 10 to 70%, despite adequate treatment [50]. OPSI incidence and mortality greatly depend on age (more common in children <2 years old) and on the underlying disease, being higher in hematological disorders [18]. Finally, malaria and babesiosis may be more severe in asplenic patients, who lack the physiologic filter of the infected erythrocytes [18]. Other microorganisms reported include Ehrlichia, Bacteroides, Enterococcus, Salmonella, and Bartonella [51]. The risk was generally higher in children and hereditary anemias, namely, thalassemia and sickle cell disease, while the lowest risk was observed in ITP patients.

Concerning AIHA, a systematic review including four studies and 48 splenectomised patients, found a post-operative infection rate of 6%, although data of long-term follow-up were missing [52]. Similarly, a more recent study on more than 4500 AIHA patients reported an incidence of 6.7% of sepsis in splenectomised subjects, including the late post-operative period [19]. Finally, some authors indicate splenectomy as a safe option if infections are adequately and promptly treated. In fact, in a series of 255 hematologic patients no cases of splenectomy-related sepsis occurred during a median follow-up of 35 months [53].

3.4. Immunosuppressive Agents

Immunosuppressive drugs are all associated with an intrinsic infectious risk, generally attributable to bone marrow toxicity. A systematic review and network metanalysis in lupus nephritis, including a total of 32 randomized clinical trials with 2611 patients, found that CTX, both low- and high-dose, mycophenolate mofetil (MMF), and AZA were associated with significantly higher risk compared to tacrolimus [20]. For CTX, an incidence of infections (bacterial, fungal, viral, protozoal, and parasitic) ranging from 15 to 34% has been described [9,54]. Notably, AZA and MMF have been associated with atypical pathogens like Listeria monocytogenes and Mycobacterium species [55,56], fungal (Cryptococcus neoformans, Aspergillus, Mucor, and Pneumocystis jirovecii) and parasitic infections (Toxoplasma gondii) [57,58]. Moreover, polyomavirus (BK virus and JCV) infections have been reported with these two drugs [9]. Conversely, the incidence of infections in autoimmune patients treated with cyclosporine A (CSA) is reported as low as 1% in clinical trials, and viral reactivation are rare [21]. Regarding AIHA, CTX toxicity is well described, also at low doses (1-2 mg/kg/day) [22], with bacterial pneumonia being the most common infection. The infectious risk related to CSA, MMF, and AZA in AIHA is less known since their use in this setting is described mostly as case series [59–61]. Taken together, these data indicate that treatment with classic immunosuppressants, especially CTX, is burdened by a relevant infectious risk, often characterized by atypical and opportunistic pathogens. J. Clin. Med. 2021, 10, 164 6 of 19

3.5. New Target Drugs

The progressive availability of new target therapies has involved also AIHA in the last years. Their infectious risk is less clear, and data derive mainly from use in diseases other than AIHA.

Upstream complement inhibitors, targeting C1s, C1q, and C3, are under investigation in cold and warm AIHAs, and the C5-inhibitor eculizumab has been used with some efficacy in CAD [23]. These drugs cause an increased susceptibility to infections, due to the impaired opsonisation and lysis of capsulated microorganisms. Particularly, in eculizumab-treated patients with paroxysmal nocturnal hemoglobinuria there is a warning for Neisseria meningitidis infections. Data from 10-year pharmacovigilance reported 76 cases of meningococcal infections (0.25/100 patient-years), eight of which fatal. With the strict adoption of vaccination policies, the meningococcal infection rate has been decreasing over time, but mortality remains considerable [24]. In addition, a recent study demonstrated that continuous C5 blocking impairs IgG-mediated complement activation, suggesting that even patients receiving adequate vaccinations against Neisseria meningitidis may not be sufficiently protected [62]. Eculizumab has been also rarely associated with pneumonia, cellulitis, bacteremia, and urinary tract infections, due to Staphylococcus, Klebsiella oxytoca, Escherichia hermannii, viruses, and fungi [24,63]. The infectious risk associated with new complement inhibitors appears very low [64], most probably due to the extended vaccination policies required for enrollment. As a general comment, C1s- and C1q-inhibitors block only the classical complement pathway, leaving the alterative and the lectin ones intact [23], while C3 inhibition may impair complement activity more profoundly.

A new treatment option for AIHA is targeting the B cell receptor signaling with drugs successfully used in chronic lymphocytic leukemia (CLL) and other lymphoproliferative disorders, such as Bruton tyrosine kinase (BTK) and phosphatidylinositol 3-kinase delta (PI3K δ) inhibitors [65,66]. Ibrutinib use in lymphoproliferative diseases is associated with increased risk of bacterial and fungal infections, up to 40% in clinical trials and real-life experience [25,67]. Parsaclisib, a next-generation and highly selective PI3K δ inhibitor, has shown to be effective in a phase 1–2 trial in relapsed/refractory B-cell malignancies, and a clinical trial in AIHA is ongoing at the time of writing (NCT03538041). It has the same mechanism of action as idelalisib, which has been associated with severe infectious complications in CLL patients, particularly PJP [25]. However, only three septic episodes in a cohort of 72 lymphoma patients treated with parsaclisib have been registered [68]. Fostamatinib, a spleen tyrosine kinase inhibitor proven effective in RA and ITP, is under study in relapsed AIHA (NCT03764618). In a meta-analysis of patients with RA a 20% increase in infectious risk has been reported [26]. Conversely, studies in ITP patients did not report infectious events [69].

Proteasome inhibitors such as bortezomib have also been used in AIHA with a good safety profile [27,28], at variance with the warnings reported for multiple myeloma [70].

Finally, targeting the neonatal Fc receptor (FcRn) is showing promising results in autoantibody-mediated diseases, including ITP [29,30]. FcRn rescues immunoglobulins (Ig) G from lysosomal degradation, prolonging antibody's (and autoantibody's) half-life. Its inhibition has a therapeutic effect by reducing the pathogenic autoantibodies. However, it causes also the reduction of other protective immunoglobulins, resulting in hypogammaglobulinemia, although not associated with clinically relevant infections [29].

Taken together, data about new target therapies in AIHA show an overall good safety profile, even though each drug carries a specific spectrum of possible related infections.

4. Viral and Mycobacterial Reactivations during AIHA Treatments

4.1. Hepatotropic Viruses

Hepatitis B virus (HBV) reactivation is defined as a sudden significant increase (>100-fold) in HBV-DNA levels in subjects with previously detectable HBV-DNA or reappearance of viral DNA in those who did not have viremia prior to the initiation of an

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immunosuppressive therapy. The increased viral replication can lead to hepatic damage, especially if immunosuppression is abruptly stopped, causing an immune reconstitution inflammatory response. HBV reactivation is reported in 10–40% of patients receiving chemotherapy or immunosuppressants for solid tumors and oncohematologic diseases [17]. In autoimmune disorders the rate is lower, as reported in a recent Italian survey on almost 1000 rheumatologic patients [71]. No cases have been reported in AIHA, although HBV reactivation has been described with steroids, rituximab and immunosuppressants [17]. Reactivation can be related to host (higher in males, elderly, in presence of cirrhosis), viral status (high baseline HBV-DNA, HBeAg positivity, chronic hepatitis B with HbsAg positivity) and type/degree of immunosuppression [72]. More precisely, it can be stratified into high (\geq 10%), moderate (1–10%) and low (<1%) risk according to the type of immunosuppressant and the HBsAg/HBcAb status [17] (Table 2). B-cell depleting agents like rituximab are at high risk of reactivation, irrespective of HbsAg/HBcAb status. Time to reactivation is highly variable and ranged from 0 to 12 months (median 3 months) in a meta-analysis on 183 lymphoma patients [73]. Contrarily, steroids are classified in different risk groups, according to the viral status and dose.

Table 2. Risk of hepatitis B virus (HBV) reactivation for different AIHA treatments according to viral status.

Drug	Viral Status	Risk Category	
High-dose steroids * Moderate-dose steroids **	HBsAg+HBsAg-/antiHBc+ HBsAg+HBsAg-/antiHBc+	high moderate moderate low	
Short-term low-dose steroids ***	irrelevant	low	
Rituximab	irrelevant	high	
Cyclosporine	irrelevant	moderate	
Methotrexate	irrelevant	low	
Azathioprine	irrelevant	low	
Bortezomib	irrelevant	moderate	

HBsAg: hepatitis B surface antigen, antiHBc: hepatitis B core antibodies, high risk: >10% rate of HBV reactivation, moderate risk: 1-10% rate of HBV reactivation, low risk: <1% rate of HBV reactivation, *>20 mg prednisone-equivalent dose/day, *** <10 mg prednisone-equivalent dose/day over 4 weeks.

Hepatitis C virus (HCV) reactivation is rarer than HBV, although its morbidity and mortality rates are not inferior [74]. It has been described anecdotally in rheumatic and lymphoma patients receiving rituximab or chemotherapies [75,76]. In the past, HCV-positive patients displayed a higher rate of severe hepatotoxicity in rituximab-containing regimens for lymphomas [77]. Nowadays, HCV reactivation is becoming less concerning, as treatment of HCV is highly effective (cure rates approaching 100% in adherent patients).

4.2. Herpesviruses

Herpesvirus reactivations are frequent after solid organ or allogeneic stem cell transplants. CMV reactivation occurs in 12–67% of patients undergoing autologous stem cell transplant and 2–39% oncohematologic non-transplant subjects [78]. A clear association with alemtuzumab emerged from prospective studies [79]. Additionally, CMV and Epstein–Barr virus (EBV) reactivations have been largely described in aplastic anemia patients treated with anti-thymocyte globulin and CSA [80,81]. This indicates that an impaired T cell immunity may represent an important risk factor. Other drugs possibly associated with CMV reactivation are high-dose steroids (i.e., prednisone >1 mg/kg/day), rituximab, bortezomib, bendamustine, and fludarabine [82]. Of note, idelalisib and ibrutinib have been related to life-threatening CMV infections [83], recommending laboratory monitoring for CMV-DNA during treatment with the former.

In the context of AIHA, herpesvirus reactivations have been described only sporadically. A fulminant case of multiple organ dysfunction caused by VZV reactivation has been reported in an 80-year-old patient treated with prednisone 1 mg/kg [84]. Moreover, a fatal

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case of viral hepatitis (diagnosis on liver biopsy) in a CLL complicated by AIHA requiring steroids has been described [85]. EBV reactivation has been reported in an 88-year-old man treated with low-dose prednisolone [86]. Finally, a case of CMV reactivation after low-dose steroids in a 21-year-old immunocompetent AIHA has been described [87].

Taken together, these data indicate that herpesvirus reactivations represent a serious issue in older, immunocompromised patients, especially those undergoing T-cell directed therapies. In addition, it is advisable to pay attention to high-dose steroids, rituximab in combination with chemotherapies, and other B cell target therapies.

4.3. Tuberculosis

Host responses against *Mycobacterium tuberculosis* are mediated by a delicate interplay between innate and adaptive immunity, dominated by macrophages and T cells, respectively. An alteration of these regulatory mechanisms may result in active TB infection/reactivation [88]. In RA, glucocorticoids and methotrexate are associated with a slightly increased risk of TB infection, whilst tumor necrosis factor (TNF)-inhibitors carry a 4- to 8-fold risk [89]. A case-control study indicates an adjusted odds ratio for tuberculosis of 2.8 in patients receiving <15 mg/day prednisone-equivalent doses. The risk increases to 7.7-fold for higher doses, particularly for intravenous pulse methylprednisolone [90,91]. B cells appear to play a minor role in controlling TB infection [92]. Consistently, rituximab has not been associated with TB reactivation in patients with RA, Sjogren's syndrome, SLE, mixed cryoglobulinemia, and vasculitides [88]. In AIHA, the risk of TB reactivation seems low, although attention should be paid to high and/or prolonged steroids use.

5. Prevention Strategies

Screening tests and prophylactic strategies before a specific AIHA treatment are lacking, and most indications derive from other diseases (Table 3).

- Haemophilus influenzae type B vaccine

Pathogen **Screening Test Risk Factors Prophylaxis** References - Lamivudine, entecavir, Steroids tenofovir or pre-emptive HBsAg, antiHBs, antiHBc, Rituximab therapy according to EASL **HBV** antiHBe, HBeAg, HBV-DNA [93,94] Immunosuppressors or AGA guidelines when indicated Bortezomib - HBV vaccination of seronegative patients No drugs approved for Anti-HCV (HCV-RNA if Long-term steroids prophylaxis HCV [9,95] - Eradication therapy in Rituximab Ab positive) HCV-RNA+ tuberculin skin test or serum Isoniazid (or rifampicin) in Mycobacterium latent TB, polichemotherapy [96] interferon gamma Long-term steroids tuberculosis release assays +/- chest X-ray in active TB Steroids >10 mg/day + age TMP-SMX (atovaquone, Pneumocystis >65 or pulmonary disease No screening tests available pentamidine, dapsone if not [9,97,98] jirovecii or therapy with tolerated/contraindicated) rituximab/CTX - ACWY and B group meningococcal vaccines Encapsulated Splenectomy - 23-valent and 13-valent [23,99–102] No screening tests available bacteria Complement inhibitors pneumococcal vaccines

Table 3. Prevention strategies detailed per pathogen.

HBV: hepatitis B virus, HCV: hepatitis C virus, HBsAg: hepatits B surface antigen, antiHBs: hepatitis B surface antibodies, antiHBc: hepatitis B e-antibodies, HBeAg: hepatitis B e-antigen, HCV: hepatitis C virus, Ab: antibodies, EASL: European Association for the Study of the Liver, AGA: American Gastroenterological Association, TB: tuberculosis, CTX: cyclophosphamide, TMP-SMX: trimethoprim-sulfamethoxazole.

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5.1. HBV and HCV

HBV screening is indicated before any chemotherapy or immunosuppressive therapy, particularly before rituximab, according to the European Association for the Study of the Liver (EASL) and the Center for Disease Control and Prevention (CDC) guidelines [93,103]. HCV screening is also advisable before immunosuppression, particularly in patients at risk (substance users, tattoos, hemodialysis, transfusions/surgery before 1994, born from HCV-positive mother, HIV-positive, elevated liver enzymes/liver disease) [9,95]. HBV/HCV testing should be preferably performed before or 3 months after immunoglobulin administration, because of possible false positivity derived from passive transmission of anti-HBc antibodies [104]. Concerning HBV prophylaxis (Table 4), the recommended antiviral strategies vary across risk categories (previously detailed) and according different guidelines [93,94] (EASL and American Gastroenterological Association, AGA). The main discrepancy is for the moderate risk category, where AGA suggests active prophylaxis, whilst EASL indicates preemptive therapy (HBsAg and/or HBV-DNA monitoring during and after immunosuppression, starting antiviral therapy in case of positivity) [93,105]. Finally, some authors also consider HBeAg/Ab status in HBsAg-negative patients, suggesting entecavir or tenofovir in case of HBeAg positivity [106]. As a general rule, prophylaxis should continue for at least 18 months after the end of immunosuppressive therapy, monitoring serology/DNA levels for at least 12 months after prophylaxis withdrawal. Of note, EASL guidelines recommend vaccination for HBV seronegative patients who are candidate for immunosuppressive treatments [93].

Table 4. HBV reactivation prophylactic strategies according to risk category.

Risk Category	Preventive Strategy Recommended		
	EASL Guidelines (2017)	AGA Guidelines (2015)	
High (>10%)	Entecavir or tenofovir if HBsAg+ Lamivudine if HBsAg-/antiHBc+	Entecavir or tenofovir	
Moderate (1–10%) Low (<1%)	Pre-emptive therapy Pre-emptive therapy	Entecavir or tenofovir Pre-emptive therapy	

EASL: European Association for the Study of the Liver, AGA: American Gastroenterological Association, HBsAg: hepatitis B surface antigen, antiHBc: hepatitis B core antibodies.

5.2. Tuberculosis

Screening for TB is a well-defined issue in rheumatologic patients undergoing immunosuppressive therapy with anti-TNF agents, with consequent prophylaxis in latent infections. Tuberculin skin test or serum interferon release assays are recommended, with an additional chest radiograph to exclude active TB in positive subjects [107]. Given the described risk of TB reactivation under long-term steroid therapy, the CDC guidelines recommend following the above-mentioned strategies for long-term prednisone users [9,96].

5.3. Pneumocystis jirovecii

PJP is life-threatening in immunocompromised patients. Adequate antimicrobial prophylaxis is mandatory in conditions like allogeneic stem cell transplants, high-dose chemotherapy regimens and patients receiving long-term steroids for hematological malignancies [97]. The mortality of PJP is quite high also in patients with autoimmune diseases, reaching even 50% in some settings [108,109]. In AIHA and other autoimmune disorders, PJP prophylaxis is not defined, and some experts suggest it in patients receiving >10–20 mg/day prednisone-equivalent doses for more than 4 weeks with additional risk factors (age >65, pre-existing pulmonary disease, combination therapy with CTX or rituximab) [9,98]. Additionally, PJP prophylaxis is requested in the clinical trial with the PI3Kd inhibitor parsaclisib, due to the warning of PJP observed with idelalisib (NCT03538041).

5.4. Herpesvirus Reactivations

Clear evidence supports antiviral prophylaxis in transplanted patients or those receiving high-dose chemotherapy [110,111]. The German Society for Hematology and Medical Oncology suggests (val)acyclovir for herpes simplex virus and VZV in patients treated with alemtuzumab, bortezomib, or purine analogs [112]. The European Conference on Infections in Leukaemia Group recommends monitoring of CMV in transplanted patients to promptly start pre-emptive antiviral therapy [113]. No clear evidence exists about herpesvirus prophylaxis in autoimmune diseases, nor in AIHA.

5.5. Vaccinations

For patients undergoing elective splenectomy, vaccination against encapsulated bacteria is recommended. Moreover, influenza virus (to be repeated annually) and VZV vaccines in over 50-year-old subjects are also advised [99-102]. Vaccines against encapsulated bacteria include the 23-valent polysaccharide pneumococcal vaccine, which covers around 70–90% of strains [18], the 13-valent conjugate pneumococcal, meningococcal vaccines against ACWY and B groups, and Haemophilus influenzae type B. Their administration and booster schedules are detailed in various guidelines [100,101]. AIHA patients who may be candidate to splenectomy should be preferably vaccinated 2–4 weeks before administration of rituximab [114,115]. Uncertainty exists about antibiotic prophylaxis after splenectomy. Most guidelines advise it for the first 2–3 years, when the infectious risk is highest, or lifelong in highly comorbid patients [100,101,115]. However, the only evidence for antibiotic prophylaxis is derived from two old studies in pediatric sickle cell disease [116,117], while no studies are available in adults. More importantly, patients should be educated in recognizing infectious symptoms, starting broad-spectrum antibiotics, and promptly referring to hospital in case of persistent fever [100]. Regarding eculizumab, quadrivalent and group B meningococcal vaccines are recommended [118,119], and antibiotic prophylaxis should be instituted until vaccinations have been performed [102]. Clinical trials with complement inhibitors in AIHA, e.g., sutimlimab and pegcetacoplan, require vaccination against all encapsulated bacteria (NCT03347396, NCT03347422, NCT03500549).

Finally, non-live vaccines can be safely administered in adult patients with autoimmune inflammatory rheumatic diseases, whereas live-attenuated vaccines should be considered with caution [120]. The Food and Drug Aministration advises against live or live attenuated vaccines in patients receiving >10 mg/day of prednisone or a cumulative dose >700 mg in 3 months, recommending to defer them at least one month after steroid discontinuation [102].

6. Infections in AIHA Secondary to Other Diseases

AIHA can develop secondarily to several conditions (Table 5): PIDs [121–123], systemic autoimmune disorders [124-126], lymphoproliferative diseases [127,128], solid cancers, solid organ, and hematopoietic stem cell transplants (HSCT) [129-131]. All these conditions are characterized by an increased infectious risk, due to the intrinsic immunodeficiency and/or immunosuppressants and chemotherapy administered. AIHA requiring immunosuppressive therapy in these contexts may further increase the infectious risk. PIDs are characterized by an increased frequency of infections, mainly upper and lower respiratory tract ones [132,133]. Regarding autoimmune diseases, infections are recognized as the major cause of death in hospitalized patients with SLE and Sjogren's syndrome [134–136]. Interestingly, Wang et al. demonstrated that AIHA is a risk factor for the development of bloodstream infections in SLE, probably due to the increased need of steroids [137]. The association of AIHA with non-Hodgkin lymphomas and CLL is well known [127], with the highest frequency in the latter (5–20%) [128]. Infections are described in up to 80% of CLL patients, cause about 60% of deaths [138], and are possibly related to hypogammaglobulinemia, abnormal T cell function, defective innate immunity, and specific therapies. Regarding transplants, the infectious risk is highly variable according to the organ transplanted, type of immunosuppressive therapy and time from transplantation. Bacterial infections

are more frequent in the first weeks, secondarily to neutropenia for HSCT and surgical intervention/hospitalization for solid organs. Later, opportunistic and viral reactivations prevail, due to long-term immunosuppression [139–141]. AIHA is known to occur in up to 15% of HSCT after a median of 3–10 months from transplant, and is usually characterized by a severe/refractory course and fatal outcome. Risk factors for AIHA secondary to HSCT include use of unrelated donor and HLA-mismatch, occurrence of graft-versus-host disease, use of cord blood, age <15 years, CMV reactivation, alemtuzumab use, and non-malignant condition pre-HSCT [142]. As for lymphoproliferative disorders, it is difficult to establish the specific contribution of AIHA treatments to the occurrence of infections, since HSCT is already marked by a profound alteration of immune homeostasis.

Table 5. Infectious risk in conditions associated with AIHA.

Condition	Frequency of AIHA	Characteristics of Infections	References
	Primary in	nmunodeficiencies	
Autoimmune lymphoproliferative syndrome	29%	Up to 30% of patients with bacterial infections related to neutropenia and splenectomy	[121]
IgA deficiency	15%	40–90% of patients experience recurrent respiratory infections, cases of Giardia lamblia infections	[122,133]
Common variable immundeficiency	2–5%	50% of subjects with upper respiratory tract infections, 50% pneumonia, 40% diarrhea, 10% VZV reactivation	[123,132]
	Autoin	nmune diseases	
Systemic lupus erythematosus	3–14%	43.1/1000 patients/year incidence of opportunistic infections; Infection is a major cause of death	[124,134,135]
Sjogren syndrome	2–3%	24.1/1000 patients/year incidence of opportunistic infections; infections are one of the main causes of death	[125,134]
Inflammatory bowel diseases	0.05%	Up to 30% of treated patients experienced infections	[126,136]
	N	eoplasms	
Chronic lymphocytic leukemia	5–20%	Frequency of drug specific infections: 57% FC; 40% FCR (mainly bacterial/opportunistic); 27% alemtuzumab (mainly CMV and fungi); 12–45% anti-CD20 (HBV, CMV, HSV, VZV, PML); 13–50% with novel drugs (ibrutinib, skin, respiratory tract, UTIs; idelalisib, PJP, CMV; venetoclax, bacterial infections)	[128,138]
	Tr	ansplants	
Hematopoietic stem cell transplant	10–15%	7–50% bacteriemia; 11–24% airways, GI tract, skin and soft tissue infection; 4–20% UTIs; 1–5% PJP (without prophylaxis); 23% invasive aspergillosis; 30–50% CMV reactivation	[131,139]
Solid organs transplant	2–3%	Frequent bacterial infections in the first month after transplant; 30–97% CMV reactivation; 2% Candida; 1–2% invasive aspergillosis; 1–2% cryptococcosis	[130,140,141]

VZV: varicella zoster virus, FC: fludarabine-cyclophosphamide, FCR: fludarabine-cyclophosphamide-rituximab, HBV: hepatitis B virus, CMV: cytomegalovirus, HSV: herpes simplex virus, PML: progressive multifocal leukoencephalopathy, UTIs: urinary tract infections, PJP: pneumocystis jirovecii pneumonia, GI: gastrointestinal.

7. AIHA Secondary to Infections

AIHA can be secondary to infections [143], particularly in children and with a prevalence of cold forms. Infectious agents can trigger AIHA through various mechanisms, including modification of erythrocyte membrane antigens, polyclonal B cell activation, innocent bystander and molecular mimicry [144,145]. Regarding viruses, parvovirus B19 [146] and hepatotropic viruses are the most frequently described [147,148]. A large population study including more than 120,000 HCV-infected American veterans concluded that the development of AIHA seems to be associated with HCV treatment with interferon [149]. Additionally, AIHA may complicate about 3% of infectious mononucleosis, with a typical onset within 1–2 weeks [150]. Paroxysmal cold hemoglobinuria, a rare form of AIHA caused by a biphasic hemolysin, more commonly occurring in children, is almost invariably preceded by a viral infection [151]. Regarding bacterial infections, Mycoplasma pneumoniae may be accompanied by severe AIHA, mainly cold but even warm forms [152]. It is worth mentioning cold AIHA secondary to Mycobacterium tuberculosis infection, with a reported efficacy of anti-tuberculosis treatment also on hemolytic anemia [153]. Moreover, case reports of AIHA have been described due to acute brucellosis [154]. Most recently, about 20 cases of AIHAs (both cold and warm forms) secondary to COVID-19 infection have been reported, with only one fatality [155]. Most of them recovered after first-line therapy with steroids +/- intravenous immunoglobulins [156,157], and some spontaneous remissions are also described [158].

8. Conclusions

The infectious burden in AIHA is considerable, consisting also in atypical and opportunistic infections, and representing a risk factor for morbidity and mortality (Figure 1). Infections are mainly associated with the load of therapy, with some peculiarity for certain treatments. Steroids encompass the widest spectrum of infections, with an underestimated risk, particularly for high doses and long-term administration, without clear indications for prophylaxis. Infections after splenectomy, mainly involving encapsulated bacteria, may be lowered by vaccination policies and antimicrobial prophylaxis/patient's education. Rituximab is generally safe, although combination therapies deserve particular attention. Among the classic immunosuppressants, a higher infectious risk exists for cyclophosphamide. Trials with novel agents have identified specific drug-related infections and recommended preventive strategies, although long-term safety data are warranted. Finally, except for HBV reactivation and post-splenectomy vaccinations, preventive strategies for other infective agents still represent an unmet need in AIHA.

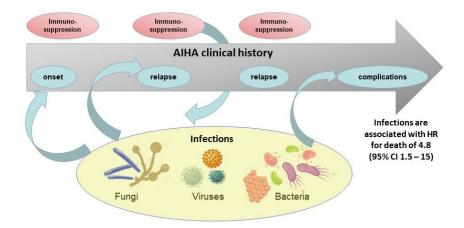


Figure 1. Relationships between autoimmune hemolytic anemia (AIHA) and infections during disease course. Infectious agents can be triggers for AIHA development at onset or at relapse (AIHA secondary to infections). On the other hand, immunosuppressive agents administered to treat AIHA expose the patient to infective events (infections secondary to AIHA), which are associated to disease complications and death.

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References

- 1. Jäger, U.; Barcellini, W.; Broome, C.M.; Gertz, M.A.; Hill, A.; Hill, Q.A.; Jilma, B.; Kuter, D.J.; Michel, M.; Montillo, M.; et al. Diagnosis and treatment of autoimmune hemolytic anemia in adults: Recommendations from the First International Consensus Meeting. *Blood Rev.* 2020, 41, 100648. [CrossRef]
- 2. Berentsen, S. How I manage patients with cold agglutinin disease. Br. J. Haematol. 2018, 181, 320–330. [CrossRef]
- 3. Barcellini, W.; Fattizzo, B.; Zaninoni, A.; Radice, T.; Nichele, I.; Di Bona, E.; Lunghi, M.; Tassinari, C.; Alfinito, F.; Ferrari, A.; et al. Clinical heterogeneity and predictors of outcome in primary autoimmune hemolytic anemia: A GIMEMA study of 308 patients. *Blood* 2014, 124, 2930–2936. [CrossRef] [PubMed]
- 4. Barcellini, W.; Zaninoni, A.; Fattizzo, B.; Giannotta, J.A.; Lunghi, M.; Ferrari, A.; Leporace, A.P.; Maschio, N.; Scaramucci, L.; Cantoni, S.; et al. Predictors of refractoriness to therapy and healthcare resource utilization in 378 patients with primary autoimmune hemolytic anemia from eight Italian reference centers. *Am. J. Hematol.* 2018, 93, E243–E246. [CrossRef]
- 5. Prabhu, R.; Bhaskaran, R.; Shenoy, V.; Sidharthan, N. Clinical characteristics and treatment outcomes of primary autoimmune hemolytic anemia: A single center study from South India. *Blood Res.* **2016**, *51*, 88–94. [CrossRef]
- 6. Giannotta, J.A.; Fattizzo, B.; Zaninoni, A.; Barcellini, W. *Infectious Complications in a Cohort of Autoommune Haemolytic Anaemia Patients*; Abstract, n. PB2401; EHA Learning Centre: Frankfurt, Germany, 2020.
- 7. Genty, I.; Michel, M.; Hermine, O.; Schaeffer, A.; Godeau, B.; Rochant, H. Characteristics of autoimmune hemolytic anemia in adults: Retrospective analysis of 83 cases. *Rev. Med. Interne* **2002**, 23, 901–909. [CrossRef]
- 8. Roumier, M.; Loustau, V.; Guillaud, C.; Languille, L.; Mahevas, M.; Khellaf, M.; Limal, N.; Noizat-Pirenne, F.; Godeau, B.; Michel, M. Characteristics and outcome of warm autoimmune hemolytic anemia in adults: New insights based on a single-center experience with 60 patients. *Am. J. Hematol.* **2014**, *89*, E150–E155. [CrossRef]
- 9. Rattarittamrong, E.; Eiamprapai, P.; Tantiworawit, A.; Rattanathammethee, T.; Hantrakool, S.; Chai-Adisaksopha, C.; Norasetthada, L. Clinical characteristics and long-term outcomes of warm-type autoimmune hemolytic anemia. *Hematology* **2016**, *21*, 368–374. [CrossRef]
- Fardet, L.; Petersen, I.; Nazareth, I. Common Infections in Patients Prescribed Systemic Glucocorticoids in Primary Care: A Population-Based Cohort Study. PLoS Med. 2016, 13, e1002024. [CrossRef]
- 11. Dixon, W.G.; Suissa, S.; Hudson, M. The association between systemic glucocorticoid therapy and the risk of infection in patients with rheumatoid arthritis: Systematic review and meta-analyses. *Arthritis Res. Ther.* **2011**, *13*, R139. [CrossRef]
- 12. Yang, Y.; Sang, J.; Pan, W.; Du, L.; Liao, W.; Chen, J.; Zhu, Y. Cryptococcal meningitis in patients with autoimmune hemolytic anemia. *Mycopathologia* **2014**, *178*, 63–70. [CrossRef] [PubMed]
- 13. Fattizzo, B.; Zaninoni, A.; Pettine, L.; Cavallaro, F.; Di Bona, E.; Barcellini, W. Low-dose rituximab in autoimmune hemolytic anemia: 10 years after. *Blood* **2019**, *133*, 996–998. [CrossRef] [PubMed]
- 14. Bohra, C.; Sokol, L.; Dalia, S. Progressive Multifocal Leukoencephalopathy and Monoclonal Antibodies: A Review. *Cancer Control* **2017**, 24, 1073274817729901. [CrossRef] [PubMed]
- 15. MacIsaac, J.; Siddiqui, R.; Jamula, E.; Li, N.; Baker, S.; Webert, K.E.; Evanovitch, D.; Heddle, N.M.; Arnold, D.M. Systematic review of rituximab for autoimmune diseases: A potential alternative to intravenous immune globulin. *Transfusion* **2018**, *58*, 2729–2735. [CrossRef] [PubMed]
- 16. Reynaud, Q.; Durieu, I.; Dutertre, M.; Ledochowski, S.; Durupt, S.; Michallet, A.-S.; Vital-Durand, D.; Lega, J.-C. Efficacy and safety of rituximab in auto-immune hemolytic anemia: A meta-analysis of 21 studies. *Autoimmun. Rev.* 2015, 14, 304–313. [CrossRef] [PubMed]
- 17. Loomba, R.; Liang, T.J. Hepatitis B reactivation associated with immune suppressive and biological modifier therapies: Current concepts, management strategies, and future directions. *Gastroenterology* **2017**, *152*, 1297–1309. [CrossRef]

- 18. Sinwar, P.D. Overwhelming post splenectomy infection syndrome—Review study. Int. J. Surg. 2014, 12, 1314–1316. [CrossRef]
- 19. Ho, G.; Brunson, A.; Keegan, T.H.M.; Wun, T. Splenectomy and the incidence of venous thromboembolism and sepsis in patients with autoimmune hemolytic anemia. *Blood Cells Mol. Dis.* **2020**, *81*, 102388. [CrossRef]
- 20. Singh, J.A.; Hossain, A.; Kotb, A.; Wells, G. Risk of serious infections with immunosuppressive drugs and glucocorticoids for lupus nephritis: A systematic review and network meta-analysis. *BMC Med.* **2016**, *14*, 137. [CrossRef]
- 21. Colombo, D.; Chimenti, S.; Grossi, P.; Marchesoni, A.; Di Nuzzo, S.; Griseta, V.; Gargiulo, A.; Parodi, A.; Simoni, L.; Bellia, G. Prevalence of past and reactivated viral infections and efficacy of cyclosporine A as monotherapy or in combination in patients with psoriatic arthritis-synergy study: A longitudinal observational study. *BioMed Res. Int.* **2014**, 2014, 941767. [CrossRef]
- 22. Salama, A. Treatment Options for Primary Autoimmune Hemolytic Anemia: A Short Comprehensive Review. *Transfus. Med. Hemother.* **2015**, 42, 294–301. [CrossRef] [PubMed]
- 23. Berentsen, S.; Hill, A.; Hill, Q.A.; Tvedt, T.H.A.; Michel, M. Novel insights into the treatment of complement-mediated hemolytic anemias. *Ther. Adv. Hematol.* **2019**, *10*, 2040620719873321. [CrossRef] [PubMed]
- 24. Socié, G.; Caby-Tosi, M.P.; Marantz, J.L.; Cole, A.; Bedrosian, C.L.; Gasteyger, C.; Mujeebuddin, A.; Hillmen, P.; Vande Walle, J.; Haller, H. Eculizumab in paroxysmal nocturnal haemoglobinuria and atypical haemolytic uraemic syndrome: 10-year pharmacovigilance analysis. *Br. J. Haematol.* **2019**, *185*, 297–310. [CrossRef]
- 25. Zinzani, P.L.; Rambaldi, A.; Gaidano, G.; Girmenia, C.; Marchetti, M.; Pane, F.; Tura, S.; Barosi, G. Infection control in patients treated for chronic lymphocytic leukemia with ibrutinib or idelalisib: Recommendations from Italian society of hematology. *Leuk. Res.* 2019, *81*, 88–94. [CrossRef]
- 26. Kunwar, S.; Devkota, A.R.; Ghimire, D.K. Fostamatinib, an oral spleen tyrosine kinase inhibitor, in the treatment of rheumatoid arthritis: A meta-analysis of randomized controlled trials. *Rheumatol. Int.* **2016**, *36*, 1077–1087. [CrossRef] [PubMed]
- 27. Ratnasingam, S.; Walker, P.A.; Tran, H.; Kaplan, Z.S.; McFadyen, J.D.; Tran, H.; The, T.C.; Fleming, S.; Catalano, J.V.; Chunilal, S.D.; et al. Bortezomib-based antibody depletion for refractory autoimmune hematological diseases. *Blood Adv.* **2016**, *1*, 31–35. [CrossRef] [PubMed]
- 28. Rossi, G.; Gramegna, D.; Paoloni, F.; Fattizzo, B.; Binda, F.; D'Adda, M.; Farina, M.; Lucchini, E.; Mauro, F.R.; Salvi, F.; et al. Short course of bortezomib in anemic patients with relapsed cold agglutinin disease: A phase 2 prospective GIMEMA study. *Blood* **2018**, *132*, 547–550. [CrossRef] [PubMed]
- Newland, A.C.; Sánchez-González, B.; Rejtő, L.; Egyed, M.; Romanyuk, N.; Godar, M.; Verschueren, K.; Gandini, D.; Ulrichts, P.; Beauchamp, J.; et al. Phase 2 study of efgartigimod, a novel FcRn antagonist, in adult patients with primary immune thrombocytopenia. Am. J. Hematol. 2020, 95, 178–187. [CrossRef]
- 30. Robak, T.; Kaźmierczak, M.; Jarque, I.; Musteata, V.; Treliński, J.; Cooper, N.; Kiessling, P.; Massow, U.; Woltering, F.; Snipes, R.; et al. Phase 2 multiple-dose study of an FcRn inhibitor, rozanolixizumab, in patients with primary immune thrombocytopenia. *Blood Adv.* 2020, 4, 4136–4146. [CrossRef]
- 31. Chew, L.-C.; Maceda-Galang, L.M.; Tan, Y.K.; Chakraborty, B.; Thumboo, J. Pneumocystis jirovecii pneumonia in patients with autoimmune disease on high-dose glucocorticoid. *J. Clin. Rheumatol.* **2015**, *21*, 72–75. [CrossRef]
- 32. Malpica, L.; van Duin, D.; Moll, S. Preventing infectious complications when treating non-malignant immune-mediated hematologic disorders. *Am. J. Hematol.* **2019**, *94*, 1396–1412. [CrossRef] [PubMed]
- 33. Youssef, J.; Novosad, S.A.; Winthrop, K.L. Infection Risk and Safety of Corticosteroid Use. *Rheum. Dis. Clin. N. Am.* **2016**, 42, 157–176. [CrossRef] [PubMed]
- 34. Danza, A.; Ruiz-Irastorza, G. Infection risk in systemic lupus erythematosus patients: Susceptibility factors and preventive strategies. *Lupus* **2013**, 22, 1286–1294. [CrossRef] [PubMed]
- 35. Hughes, M.; Trivedi, K.; Rudrappa, M. Disseminated Cryptococcal Disease with Diffuse Pulmonary Infiltrates in a Non-HIV Host. *J. La. State Med. Soc.* **2017**, *169*, 57. [PubMed]
- 36. Gea-Banacloche, J.C. Rituximab-associated infections. Semin. Hematol. 2010, 47, 187–198. [CrossRef] [PubMed]
- 37. Vidal, L.; Gafter-Gvili, A.; Leibovici, L.; Shpilberg, O. Rituximab as maintenance therapy for patients with follicular lymphoma. *Cochrane Database Syst. Rev.* **2009**, CD006552. [CrossRef]
- 38. Berger, J.R.; Malik, V.; Lacey, S.; Brunetta, P.; Lehane, P.B. Progressive multifocal leukoencephalopathy in rituximab-treated rheumatic diseases: A rare event. *J. Neurovirol.* **2018**, *24*, 323–331. [CrossRef]
- 39. Cohen, S.B.; Emery, P.; Greenwald, M.W.; Dougados, M.; Furie, R.A.; Genovese, M.C.; Keystone, E.C.; Loveless, J.E.; Burmester, G.R.; Cravets, M.W.; et al. Rituximab for rheumatoid arthritis refractory to anti-tumor necrosis factor therapy: Results of a multicenter, randomized, double-blind, placebo-controlled, phase III trial evaluating primary efficacy and safety at twenty-four weeks. *Arthritis Rheum.* 2006, 54, 2793–2806. [CrossRef]
- 40. Serris, A.; Amoura, Z.; Canouï-Poitrine, F.; Terrier, B.; Hachulla, E.; Costedoat-Chalumeau, N.; Papo, T.; Lambotte, O.; Saadoun, D.; Hié, M.; et al. Efficacy and safety of rituximab for systemic lupus erythematosus-associated immune cytopenias: A multicenter retrospective cohort study of 71 adults. *Am. J. Hematol.* 2018, 93, 424–429. [CrossRef]
- 41. Berentsen, S.; Ulvestad, E.; Gjertsen, B.T.; Hjorth-Hansen, H.; Langholm, R.; Knutsen, H.; Ghanima, W.; Shammas, F.V.; Tjønnfjord, G.E. Rituximab for primary chronic cold agglutinin disease: A prospective study of 37 courses of therapy in 27 patients. *Blood* 2004, 103, 2925–2928. [CrossRef]

42. Laribi, K.; Bolle, D.; Ghnaya, H.; Sandu, A.; Besançon, A.; Denizon, N.; Truong, C.; Pineau-Vincent, F.; de Materre, A.B. Rituximab is an effective and safe treatment of relapse in elderly patients with resistant warm AIHA. *Ann. Hematol.* **2016**, *95*, 765–769. [CrossRef]

- 43. Barcellini, W.; Zaja, W.; Zaninoni, A.; Imperiali, F.G.; Battista, M.L.; Di Bona, E.; Fattizzo, B.; Consonni, D.; Cortelezzi, A.; Fanin, R.; et al. Low-dose rituximab in adult patients with idiopathic autoimmune hemolytic anemia: Clinical efficacy and biologic studies. *Blood* 2012, 119, 3691–3697. [CrossRef] [PubMed]
- 44. Wang, H.; Yan, S.; Liu, H.; Li, L.; Song, J.; Wang, G.; Wang, H.; Wu, Y.; Shao, Z.; Fu, R. Infection risk in autoimmune hematological disorders with low-dose rituximab treatment. *J. Clin. Lab. Anal.* **2020**, *34*, e23455. [CrossRef] [PubMed]
- 45. Berentsen, S.; Randen, U.; Oksman, M.; Birgens, H.; Tvedt, T.H.A.; Dalgaard, J.; Galteland, E.; Haukås, E.; Brudevold, R.; Sørbø, J.H.; et al. Bendamustine plus rituximab for chronic cold agglutinin disease: Results of a Nordic prospective multicenter trial. *Blood* 2017, 130, 537–541. [CrossRef] [PubMed]
- 46. Berentsen, S.; Randen, U.; Vågan, A.M.; Hjorth-Hansen, H.; Vik, A.; Dalgaard, J.; Jacobsen, E.M.; Thoresen, A.S.; Beiske, K.; Tjønnfjord, G.E. High response rate and durable remissions following fludarabine and rituximab combination therapy for chronic cold agglutinin disease. *Blood* **2010**, *116*, 3180–3184. [CrossRef]
- 47. Bisharat, N.; Omari, H.; Lavi, I.; Raz, R. Risk of infection and death among post-splenectomy patients. *J. Infect.* **2001**, 43, 182–186. [CrossRef]
- 48. Thomsen, R.W.; Schoonen, W.M.; Farkas, D.K.; Riis, A.; Jacobsen, J.; Fryzek, J.P.; Sørensen, H.T. Risk for hospital contact with infection in patients with splenectomy. *Ann. Intern. Med.* **2009**, *151*, 546–555. [CrossRef]
- 49. Vianelli, N.; Palandri, F.; Polverelli, N.; Stasi, R.; Joelsson, J.; Johansson, E.; Ruggeri, M.; Zaja, F.; Cantoni, S.; Catucci, A.E.; et al. Splenectomy as a curative treatment for immune thrombocytopenia: A retrospective analysis of 233 patients with a minimum follow up of 10 years. *Haematologica* **2013**, *98*, 875–880. [CrossRef]
- 50. Waghorn, D.J. Overwhelming infection in asplenic patients: Current best practice preventive measures are not being followed. *J. Clin. Pathol.* **2001**, *54*, 214–218. [CrossRef]
- 51. Davidson, R.N.; Wall, R.A. Prevention and management of infections in patients without a spleen. *Clin. Microbiol. Infect.* **2001**, 7, 657–660. [CrossRef]
- 52. Giudice, V.; Rosamilio, R.; Ferrara, I.; Seneca, E.; Serio, B.; Selleri, C. Efficacy and safety of splenectomy in adult autoimmune hemolytic anemia. *Open Med.* **2016**, *11*, 374–380. [CrossRef]
- 53. Balagué, C.; Targarona, E.M.; Cerdán, G.; Novell, J.; Montero, O.; Bendahan, G.; García, A.; Pey, A.; Vela, S.; Diaz, M.; et al. Long-term outcome after laparoscopic splenectomy related to hematologic diagnosis. *Surg. Endosc.* **2004**, *18*, 1283–1287. [CrossRef] [PubMed]
- 54. Cortazar, F.B.; Muhsin, S.A.; Pendergraft, W.F., III; Wallace, Z.S.; Dunbar, C.; Laliberte, K.; Niles, J.L. Combination therapy with rituximab and cyclophosphamide for remission induction in ANCA vasculitis. *Kidney Int. Rep.* **2017**, *3*, 394–402. [CrossRef] [PubMed]
- 55. Teh, C.L.; Kong, K.O.; Chong, A.P.; Badsha, H. Mycobacterium haemophilum infection in a SLE patient on mycophenolate mofetil. *Lupus* **2002**, *11*, 249–252. [CrossRef] [PubMed]
- 56. Del Pozo, J.L.; de la Garza, R.G.; de Rada, P.D.; Ornilla, E.; Yuste, J.R. Listeria monocytogenes septic arthritis in a patient treated with mycophenolate mofetil for polyarteritis nodosa: A case report and review of the literature. *Int. J. Infect. Dis.* **2013**, 17, e132–e133. [CrossRef] [PubMed]
- 57. Gibson, R.H.; Evans, R.J.; Hotham, R.; Bojarczuk, A.; Lewis, A.; Bielska, E.; May, R.C.; Elks, P.M.; Renshaw, S.A.; Johnston, S.A. Mycophenolate mofetil increases susceptibility to opportunistic fungal infection independent of lymphocytes. *BioRxiv* **2017**, 131540. [CrossRef]
- 58. Bernardo, D.R.; Chahin, N. Toxoplasmic encephalitis during mycophenolate mofetil immunotherapy of neuromuscular disease. *Neurol. Neuroimmunol. Neuroinflamm.* **2015**, 2, e63. [CrossRef]
- 59. Emilia, G.; Messora, C.; Longo, G.; Bertesi, M. Long-term salvage treatment by cyclosporin in refractory autoimmune haematological disorders. *Br. J. Haematol.* **1996**, 93, 341–344. [CrossRef]
- 60. Howard, J.; Hoffbrand, A.V.; Prentice, H.G.; Mehta, A. Mycophenolate mofetil for the treatment of refractory auto-immune haemolytic anaemia and auto-immune thrombocytopenia purpura. *Br. J. Haematol.* **2002**, *117*, 712–715. [CrossRef]
- 61. Newman, K.; Owlia, M.B.; El-Hemaidi, I.; Akhtari, M. Management of immune cytopenias in patients with systemic lupus erythematosus—Old and new. *Autoimmun. Rev.* **2013**, *12*, 784–791. [CrossRef]
- 62. Langereis, J.D.; van den Broek, B.; Franssen, S.; Joosten, I.; Blijlevens, N.M.A.; de Jonge, M.I.; Langemeijer, S. Eculizumab impairs Neisseria meningitidis serogroup B killing in whole blood despite 4CMenB vaccination of PNH patients. *Blood Adv.* **2020**, *4*, 3615–3620. [CrossRef] [PubMed]
- 63. Al-Ani, F.; Chin-Yee, I.; Lazo-Langner, A. Eculizumab in the management of paroxysmal nocturnal hemoglobinuria: Patient selection and special considerations. *Ther. Clin. Risk Manag.* **2016**, *12*, 1161–1170. [PubMed]
- 64. Jäger, U.; D'Sa, S.; Schörgenhofer, C.; Bartko, J.; Derhaschnig, U.; Sillaber, C.; Jilma-Stohlawetz, P.; Fillitz, M.; Schenk, T.; Patou, G.; et al. Inhibition of complement C1s improves severe hemolytic anemia in cold agglutinin disease: A first-in-human trial. *Blood* **2019**, 133, 893–901. [CrossRef] [PubMed]
- 65. Manda, S.; Dunbar, N.; Marx-Wood, C.R.; Danilov, A.V. Ibrutinib is an effective treatment of autoimmune haemolytic anaemia in chronic lymphocytic leukaemia. *Br. J. Haematol.* **2015**, 170, 734–736. [CrossRef]

66. Molica, S.; Levato, L.; Mirabelli, R. Chronic lymphocytic leukemia, autoimmune hemolytic anemia and ibrutinib: A case report and review of the literature. *Leuk. Lymphoma.* **2016**, *57*, 735–737. [CrossRef]

- 67. Rogers, K.A.; Mousa, L.; Zhao, Q.; Bhat, S.A.; Byrd, J.C.; El Boghdadly, Z.; Guerrero, T.; Levine, L.B.; Lucas, F.; Shindiapina, P.; et al. Incidence of opportunistic infections during ibrutinib treatment for B-cell malignancies. *Leukemia* **2019**, *33*, 2527–2530. [CrossRef]
- 68. Forero-Torres, A.; Ramchandren, R.; Yacoub, A.; Wertheim, M.S.; Edenfield, W.J.; Caimi, P.; Gutierrez, M.; Akard, L.; Escobar, C.; Call, J.; et al. Parsaclisib, a potent and highly selective PI3Kδ inhibitor, in patients with relapsed or refractory B-cell malignancies. *Blood* **2019**, *133*, 1742–1752. [CrossRef]
- 69. Bussel, J.B.; Arnold, D.M.; Boxer, M.A.; Cooper, N.; Mayer, J.; Zayed, H.; Tong, S.; Duliege, A.M. Long-term fostamatinib treatment of adults with immune thrombocytopenia during the phase 3 clinical trial program. *Am. J. Hematol.* **2019**, *94*, 546–553. [CrossRef]
- 70. Scott, K.; Hayden, P.J.; Will, A.; Wheatley, K.; Coyne, I. Bortezomib for the treatment of multiple myeloma. *Cochrane Database Syst. Rev.* **2016**, *4*, CD010816. [CrossRef]
- 71. Ditto, M.C.; Parisi, S.; Varisco, V.; Talotta, R.; Batticciotto, A.; Antivalle, M.; Gerardi, M.C.; Agosti, M.; Borrelli, R.; Fusaro, E.; et al. Prevalence of hepatitis B virus infection and risk of reactivation in rheumatic population undergoing biological therapy. *Clin. Exp. Rheumatol.* **2020**, in press.
- 72. Yeo, W.; Johnson, P.J. Diagnosis, prevention and management of hepatitis B virus reactivation during anticancer therapy. *Hepatology* **2006**, 43, 209–220. [CrossRef] [PubMed]
- 73. Evens, A.M.; Jovanovic, B.D.; Su, Y.C.; Raisch, D.W.; Ganger, D.; Belknap, S.M.; Dai, M.S.; Chiu, B.C.; Fintel, B.; Cheng, Y.; et al. Rituximab-associated hepatitis B virus (HBV) reactivation in lymphoproliferative diseases: Meta-analysis and examination of FDA safety reports. *Ann. Oncol.* 2011, 22, 1170–1180. [CrossRef] [PubMed]
- 74. Torres, H.A.; Davila, M. Reactivation of hepatitis B virus and hepatitis C virus in patients with cancer. *Nat. Rev. Clin. Oncol.* **2012**, 9, 156–166. [CrossRef] [PubMed]
- 75. Ennishi, D.; Maeda, Y.; Niitsu, N.; Kojima, M.; Izutsu, K.; Takizawa, J.; Kusumoto, S.; Okamoto, M.; Yokoyama, M.; Takamatsu, Y.; et al. Hepatic toxicity and prognosis in hepatitis C virus-infected patients with diffuse large B-cell lymphoma treated with rituximab-containing chemotherapy regimens: A Japanese multicenter analysis. *Blood* **2010**, *116*, 5119–5125. [CrossRef] [PubMed]
- 76. Haggag, R.M.; Abu-Taleb, F.; Khaled, H.M. Rituximab-induced hepatitis C virus reactivation in HCV-positive patients with diffuse large Bcell lymphoma. *J. Clin. Oncol.* **2016**, *34*, e19039. [CrossRef]
- 77. Foran, J.M. Hepatitis C in the rituximab era. *Blood* **2010**, *116*, 5081–5082. [CrossRef]
- 78. de Melo Silva, J.; Pinheiro-Silva, R.; Dhyani, A.; Pontes, G.S. Cytomegalovirus and Epstein-Barr infections: Prevalence and impact on patients with hematological diseases. *BioMed Res. Int.* **2020**, 2020, 1627824. [CrossRef]
- 79. Elter, T.; Gercheva-Kyuchukova, L.; Pylylpenko, H.; Robak, T.; Jaksic, B.; Rekhtman, G.; Kyrcz-Krzemień, S.; Vatutin, M.; Wu, J.; Sirard, C.; et al. Fludarabine plus alemtuzumab versus fludarabine alone in patients with previously treated chronic lymphocytic leukaemia: A randomised phase 3 trial. *Lancet Oncol.* **2011**, *12*, 1204–1213. [CrossRef]
- 80. Park, S.S.; Cho, S.Y.; Han, E.; Min, G.J.; Park, S.; Yoon, J.H.; Lee, S.E.; Cho, B.S.; Eom, K.S.; Kim, Y.J.; et al. Reactivation and dynamics of cytomegalovirus and Epstein-Barr virus after rabbit antithymocyte globulin and cyclosporine for aplastic anemia. *Eur. J. Haematol.* **2019**, *103*, 433–441. [CrossRef]
- 81. Scheinberg, P.; Fischer, S.H.; Li, L.; Nunez, O.; Wu, C.O.; Sloand, E.M.; Cohen, J.I.; Young, N.S.; John Barrett, A. Distinct EBV and CMV reactivation patterns following antibody-based immunosuppressive regimens in patients with severe aplastic anemia. *Blood* 2007, 109, 3219–3224. [CrossRef]
- 82. Marchesi, F.; Pimpinelli, F.; Ensoli, F.; Mengarelli, A. Cytomegalovirus infection in hematologic malignancy settings other than the allogeneic transplant. *Hematol. Oncol.* **2018**, *36*, 381–391. [CrossRef] [PubMed]
- 83. Angarone, M. Epidemiology and prevention of viral infections in patients with hematologic malignancies. *Infect. Disord. Drug Targets* **2011**, *11*, 27–33. [CrossRef]
- 84. Akiyama, M.; Yoshifuji, K.; Fukuda, T.; Tohda, S.; Miki, T.; Miura, O.; Yamamoto, M. Fulminant visceral disseminated varicellazoster virus infection without skin involvement in a patient with autoimmune hemolytic anemia on prednisolone therapy. *Rinsho Ketsueki* 2016, 57, 467–471. [PubMed]
- 85. Orvain, C.; Ducancelle, A.; Eymerit-Morin, C.; Rousselet, M.-C.; Oberti, F.; Hunault-Berger, M.; Tanguy-Schmidt, A. Severe viral hepatitis in a patient with chronic lymphocytic leukemia (CLL) complicated with autoimmune hemolytic anemia (AIHA), treated with steroids. *J. Clin. Virol.* **2015**, *62*, 66–68. [CrossRef] [PubMed]
- 86. Arai, A.; Imadome, K.-I.; Fujiwara, S.; Miura, O. Autoimmune hemolytic anemia accompanied by reactivation of an Epstein-Barr virus infection with suppressed CTL response to EBV-infected cells in an elderly man. *Intern. Med.* **2010**, *49*, 325–329. [CrossRef]
- 87. Troselj-Vukic, B.; Milotic, I.; Milotic, F.; Crnic-Martinovic, M.; Grahovac, B. Cytomegalovirus reactivation after low-dose steroid treatment for hemolytic anemia in a patient with primary Epstein-Barr virus infection. *Wien. Klin. Wochenschr.* **2007**, *119*, 435–437. [CrossRef] [PubMed]
- 88. Evangelatos, G.; Koulouri, V.; Iliopoulos, A.; Fragoulis, G.E. Tuberculosis and targeted synthetic or biologic DMARDs, beyond tumor necrosis factor inhibitors. *Ther. Adv. Musculoskelet. Dis.* **2020**, *12*, 1759720X20930116. [CrossRef]
- 89. Godfrey, M.S.; Friedman, L.N. Tuberculosis and Biologic Therapies: Anti-Tumor Necrosis Factor-α and Beyond. *Clin. Chest Med.* **2019**, 40, 721–739. [CrossRef] [PubMed]
- 90. Jick, S.S.; Lieberman, E.S.; Rahman, M.U.; Choi, H.K. Glucocorticoid use, other associated factors, and the risk of tuberculosis. *Arthritis Rheum.* **2006**, *55*, 19–26. [CrossRef]

91. Tam, L.-S.; Li, E.K.; Wong, S.-M.; Szeto, C.-C. Risk factors and clinical features for tuberculosis among patients with systemic lupus erythematosus in Hong Kong. *Scand. J. Rheumatol.* **2002**, *31*, 296–300. [CrossRef]

- 92. De Martino, M.; Lodi, L.; Galli, L.; Chiappini, E. Immune response to mycobacterium tuberculosis: A narrative review. *Front. Pediatr.* **2019**, *7*, 350. [CrossRef] [PubMed]
- 93. European Association for the Study of the Liver. EASL 2017. Clinical Practice Guidelines on the management of hepatitis B virus infection. *J. Hepatol.* **2017**, *67*, 370–398. [CrossRef] [PubMed]
- 94. Reddy, K.R.; Beavers, K.L.; Hammond, S.P.; Lim, J.K.; Falck-Ytter, Y.T.; American Gastroenterological Association Institute. American Gastroenterological Association Institute guideline on the prevention and treatment of hepatitis B virus reactivation during immunosuppressive drug therapy. *Gastroenterology* 2015, 148, 215–219. [CrossRef] [PubMed]
- 95. Karadağ, Ö.; Kaşifoğlu, T.; Özer, B.; Kaymakoğlu, S.; Kuş, Y.; İnanç, M.; Keser, G.; Kiraz, S. Viral hepatitis screening guideline before biological drug use in rheumatic patients. *Eur. J. Rheumatol.* **2016**, *3*, 25–28. [CrossRef] [PubMed]
- Cohn, D.L.; O'Brien, R.J.; Geiter, L.J.; Gordin, F.; Hershfield, E.; Horsburgh, C. Targeted Tuberculin Testing and Treatment of Latent Tuberculosis Infection. American Thoracic Society. MMWR Morb. Mortal. Wkly. Rep. 2000, 49, 1–51.
- 97. Maertens, J.; Cesaro, S.; Maschmeyer, G.; Einsele, H.; Donnelly, J.P.; Alanio, A.; Hauser, P.M.; Lagrou, K.; Melchers, W.J.G.; Helweg-Larsen, J.; et al. 5th European Conference on Infections in Leukaemia (ECIL-5), a joint venture of the European Group for Blood and Marrow Transplantation (EBMT), the European Organisation for Research and Treatment of Cancer (EORTC), the Immunocompromised Host Society (ICHS) and the European LeukemiaNet (ELN). ECIL guidelines for preventing Pneumocystis jirovecii pneumonia in patients with haematological malignancies and stem cell transplant recipients. *J. Antimicrob. Chemother.* 2016, 71, 2397–2404.
- 98. Katsuyama, T.; Saito, K.; Kubo, S.; Nawata, M.; Tanaka, Y. Prophylaxis for pneumocystis pneumonia in patients with rheumatoid arthritis treated with biologics, based on risk factors found in a retrospective study. *Arthritis Res. Ther.* **2014**, *16*, R43. [CrossRef]
- 99. Rubin, L.G.; Schaffner, W.; Solomon, C.G. Care of the asplenic patient. N. Engl. J. Med. 2014, 371, 349–356. [CrossRef]
- 100. Bonanni, P.; Grazzini, M.; Niccolai, G.; Paolini, D.; Varone, O.; Bartoloni, A.; Bartalesi, F.; Santini, M.G.; Baretti, S.; Bonito, C.; et al. Recommended vaccinations for asplenic and hyposplenic adult patients. *Hum. Vaccines Immunother.* 2017, 13, 359–368. [CrossRef]
- 101. Kanhutu, K.; Jones, P.; Cheng, A.C.; Grannell, L.; Best, E.; Spelman, D. Spleen Australia guidelines for the prevention of sepsis in patients with asplenia and hyposplenism in Australia and New Zealand. *Intern. Med. J.* 2017, 47, 848–855. [CrossRef]
- 102. Lee, G.; Malpica Castillo, L.E.; Walter, J.E. Infection Risk, Immunization Recommendations, and Antimicrobial Prophylaxis Needs When Treating Non-Malignant Hematologic Disorders—Wash Your Hands and What Else? Education Program. In Proceedings of the 62nd ASH Annual Meeting and Exposition, San Diego, CA, USA, 5–8 December 2020.
- 103. Weinbaum, C.M.; Williams, I.; Mast, E.E.; Wang, S.A.; Finelli, L.; Wasley, A.; Neitzel, S.M.; Ward, J.W.; Centers for Disease Control and Prevention (CDC). Recommendations for identification and public health management of persons with chronic hepatitis B virus infection. *MMWR Recomm. Rep.* **2008**, *57*, 1–20. [CrossRef] [PubMed]
- 104. Ramsay, I.; Gorton, R.L.; Patel, M.; Workman, S.; Symes, A.; Haque, T.; Irish, D.; Seneviratne, S.L.; Burns, S.O.; Wey, E.; et al. Transmission of hepatitis B core antibody and galactomannan enzyme immunoassay positivity via immunoglobulin products: A comprehensive analysis. *Clin. Infect. Dis.* **2016**, *63*, 57–63. [CrossRef] [PubMed]
- 105. Viganò, M.; Serra, G.; Casella, G.; Grossi, G.; Lampertico, P. Reactivation of hepatitis B virus during targeted therapies for cancer and immunemediated disorders. *Expert Opin. Biol. Ther.* **2016**, *16*, 917–926. [CrossRef] [PubMed]
- 106. Pisaturo, M.; Di Caprio, G.; Calò, F.; Portunato, F.; Martini, S.; Coppola, N. Management of HBV reactivation in non-oncological patients. *Expert Rev. Anti-Infect. Ther.* **2018**, *16*, 611–624. [CrossRef]
- 107. Keane, J.; Bresnihan, B. Tuberculosis reactivation during immunosuppressive therapy in rheumatic diseases: Diagnostic and therapeutic strategies. *Curr. Opin. Rheumatol.* **2008**, *20*, 443–449. [CrossRef] [PubMed]
- 108. Mecoli, C.A.; Saylor, D.; Gelber, A.C.; Christopher-Stine, L. Pneumocystis jiroveci pneumonia in rheumatic disease: A 20-year single-centre experience. *Clin. Exp. Rheumatol.* **2017**, *35*, 671–673.
- 109. Braga, B.P.; Prieto-González, S.; Hernández-Rodríguez, J. *Pneumocystis jirovecii* pneumonia prophylaxis in immunocompromised patients with systemic autoimmune diseases. *Med. Clin.* **2019**, *152*, 502–507. [CrossRef] [PubMed]
- 110. Yahav, D.; Gafter-Gvili, A.; Muchtar, E.; Skalsky, K.; Kariv, G.; Yeshurun, M.; Leibovici, L.; Paul, M. Antiviral prophylaxis in haematological patients: Systematic review and meta-analysis. *Eur. J. Cancer* **2009**, *45*, 3131–3148. [CrossRef]
- 111. Kawamura, K.; Hayakawa, J.; Akahoshi, Y.; Harada, N.; Nakano, H.; Kameda, K.; Ugai, T.; Wada, H.; Yamasaki, R.; Ishihara, Y.; et al. Low-dose acyclovir prophylaxis for the prevention of herpes simplex virus and varicella zoster virus diseases after autologous hematopoietic stem cell transplantation. *Int. J. Hematol.* 2015, 102, 230–237. [CrossRef]
- 112. Sandherr, M.; Hentrich, M.; von Lilienfeld-Toal, M.; Massenkeil, G.; Neumann, S.; Penack, O.; Biehl, L.; Cornely, O.A. Antiviral prophylaxis in patients with solid tumours and haematological malignancies—Update of the Guidelines of the Infectious Diseases Working Party (AGIHO) of the German Society for Hematology and Medical Oncology (DGHO). *Ann. Hematol.* **2015**, *94*, 1441–1450. [CrossRef]
- 113. Ljungman, P.; de la Camara, R.; Robin, C.; Crocchiolo, R.; Einsele, H.; Hill, J.A.; Hubacek, P.; Navarro, D.; Cordonnier, C.; Ward, K.N.; et al. Guidelines for the management of cytomegalovirus infection in patients with haematological malignancies and after stem cell transplantation from the 2017 European Conference on Infections in Leukaemia (ECIL 7). *Lancet Infect. Dis.* **2019**, *19*, e260–e272. [CrossRef]

114. Moulis, G.; Lapeyre-Mestre, M.; Mahévas, M.; Montastruc, J.-L.; Sailler, L. Need for an improved vaccination rate in primary immune thrombocytopenia patients exposed to rituximab or splenectomy. A nationwide population-based study in France. *Am. J. Hematol.* **2015**, *90*, 301–305. [CrossRef] [PubMed]

- 115. Davies, J.M.; Barnes, R.; Milligan, D.; British Committee for Standards in Haematology. Working Party of the Haematology/Oncology Task Force. Update of guidelines for the prevention and treatment of infection in patients with an absent or dysfunctional spleen. *Clin. Med.* 2002, 2, 440–443. [CrossRef] [PubMed]
- 116. Gaston, M.H.; Verter, J.I.; Woods, G.; Pegelow, C.; Kelleher, J.; Presbury, G.; Zarkowsky, H.; Vichinsky, E.; Iyer, R.; Lobel, J.S.; et al. Prophylaxis with oral penicillin in children with sickle cell anemia. A randomized trial. *N. Engl. J. Med.* **1986**, *314*, 1593. [CrossRef]
- 117. Salamah, M.M. Oral penicillin prophylaxis in children with sickle cell anaemia in Saudi Arabia. N. Engl. J. Med. 1987, 316, 274.
- 118. Cohn, A.C.; MacNeil, J.R.; Clark, T.A.; Ortega-Sanchez, I.R.; Briere, E.Z.; Meissner, H.C.; Baker, C.J.; Messonnier, N.E.; Centers for Disease Control and Prevention (CDC). Prevention and control of meningococcal disease: Recommendations of the Advisory Committee on Immunization Practices (ACIP). MMWR Recomm. Rep. 2013, 62, 1–28.
- 119. McNamara, L.A.; Topaz, N.; Wang, X.; Hariri, S.; Fox, L.; MacNeil, J.R. High risk for invasive Meningococcal disease among patients receiving Eculizumab (Soliris) despite receipt of Meningococcal vaccine. *MMWR Morb. Mortal. Wkly. Rep.* **2017**, *66*, 734–737. [CrossRef]
- 120. Furer, V.; Rondaan, C.; Heijstek, M.W.; Agmon-Levin, N.; van Assen, S.; Bijl, M.; Breedveld, F.C.; D'Amelio, R.; Dougados, M.; Kapetanovic, M.C.; et al. 2019 update of EULAR recommendations for vaccination in adult patients with autoimmune inflammatory rheumatic diseases. *Ann. Rheum. Dis.* 2020, 79, 39–52. [CrossRef]
- 121. Oliveira, J.B. The expanding spectrum of the autoimmune lymphoproliferative syndromes. *Curr. Opin. Pediatr.* **2013**, 25, 722–729. [CrossRef]
- 122. Odineal, D.D.; Gershwin, M.E. The Epidemiology and Clinical Manifestations of Autoimmunity in Selective IgA Deficiency. *Clin. Rev. Allergy Immunol.* **2020**, *58*, 107–133. [CrossRef]
- 123. Feuille, E.J.; Anooshiravani, N.; Sullivan, K.E.; Fuleihan, R.L.; Cunningham-Rundles, C. Autoimmune Cytopenias and Associated Conditions in CVID: A Report From the USIDNET Registry. *J. Clin. Immunol.* 2018, *38*, 28–34. [CrossRef] [PubMed]
- 124. Gormezano, N.W.; Kern, D.; Pereira, O.L.; Esteves, G.C.; Sallum, A.M.; Aikawa, N.E.; Pereira, R.M.; Silva, C.A.; Bonfá, E. Autoimmune hemolytic anemia in systemic lupus erythematosus at diagnosis: Differences between pediatric and adult patients. *Lupus* 2017, 26, 426–430. [CrossRef] [PubMed]
- 125. Wen, W.; Liu, Y.; Zhao, C.; Sun, X.; Zhang, C.; Li, Z. Clinical and serologic features of primary Sjögren's syndrome concomitant with autoimmune hemolytic anemia: A large-scale cross-sectional study. *Clin. Rheumatol.* **2015**, *34*, 1877–1884. [CrossRef] [PubMed]
- 126. Uzzan, M.; Galicier, L.; Gornet, J.M.; Oksenhendler, E.; Fieschi, C.; Allez, M.; Bouhnik, Y.; Kirchgesner, J.; Boutboul, D.; Treton, X.; et al. Autoimmune cytopenias associated with inflammatory bowel diseases: Insights from a multicenter retrospective cohort. *Dig. Liver Dis.* 2017, 49, 397–404. [CrossRef] [PubMed]
- 127. Sallah, S.; Sigounas, G.; Vos, P.; Wan, J.Y.; Nguyen, N.P. Autoimmune hemolytic anemia in patients with non-Hodgkin's lymphoma. *Ann. Oncol.* **2000**, *11*, 1571–1577. [CrossRef]
- 128. Visco, C.; Barcellini, W.; Maura, F.; Neri, A.; Cortelezzi, A.; Rodeghiero, F. Autoimmune cytopenias in chronic lymphocytic leukemia. *Am. J. Hematol.* **2014**, *89*, 1055–1062. [CrossRef]
- 129. De Keyzer, K.; Peeters, P.; Verhelst, C.; Dendooven, A.; Vonck, A.; Vanholder, R. Autoimmune haemolytic anaemia associated with a thymoma: Case report and review of the literature. *Acta Clin. Belg.* **2009**, *64*, 447–451. [CrossRef]
- 130. Li, M.; Goldfinger, D.; Yuan, S. Autoimmune hemolytic anemia in pediatric liver or combined liver and small bowel transplant patients: A case series and review of the literature. *Transfusion* **2012**, *52*, 48–54. [CrossRef]
- 131. González-Vicent, M.; Sanz, J.; Fuster, J.L.; Cid, J.; de Heredia, C.D.; Morillo, D.; Fernández, J.M.; Pascual, A.; Badell, I.; Serrano, D.; et al. Autoimmune hemolytic anemia (AIHA) following allogeneic hematopoietic stem cell transplantation (HSCT): A retrospective analysis and a proposal of treatment on behalf of the Grupo Español De Trasplante de Medula Osea en Niños (GETMON) and the Grupo Español de Trasplante Hematopoyetico (GETH). *Transfus. Med. Rev.* 2018, 32, 179–185.
- 132. Saikia, B.; Gupta, S. Common Variable Immunodeficiency. Indian J. Pediatr. 2016, 83, 338–344. [CrossRef]
- 133. Yazdani, R.; Azizi, G.; Abolhassani, H.; Aghamohammadi, A. Selective IgA Deficiency: Epidemiology, Pathogenesis, Clinical Phenotype, Diagnosis, Prognosis and Management. *Scand. J. Immunol.* **2017**, *85*, 3–12. [CrossRef] [PubMed]
- 134. Hsu, C.Y.; Ko, C.H.; Wang, J.L.; Hsu, T.C.; Lin, C.Y. Comparing the burdens of opportunistic infections among patients with systemic rheumatic diseases: A nationally representative cohort study. *Arthritis Res. Ther.* **2019**, *21*, 211. [CrossRef] [PubMed]
- 135. Barber, R.M.W.; Clarke, A.E. Systemic lupus erythematosus and risk of infection. *Expert Rev. Clin. Immunol.* **2020**, *16*, 527–538. [CrossRef] [PubMed]
- 136. Wheat, C.L.; Ko, C.W.; Clark-Snustad, K.; Grembowski, D.; Thornton, T.A.; Devine, B. Inflammatory Bowel Disease (IBD) pharmacotherapy and the risk of serious infection: A systematic review and network meta-analysis. *BMC Gastroenterol.* **2017**, 17, 52. [CrossRef] [PubMed]
- 137. Wang, M.; Zhang, H.; Yang, X.; Li, W.; Li, T.; Liu, S. Laboratory-confirmed bloodstream infection in systemic lupus erythematosus: Risk profiling and short-term mortality. *Lupus* **2020**, *24*, 961203320948964. [CrossRef] [PubMed]
- 138. Morrison, V.A. Infections in patients with leukemia and lymphoma. Cancer Treat. Res. 2014, 161, 319-349.

139. Ullmann, A.J.; Schmidt-Hieber, M.; Bertz, H.; Heinz, W.J.; Kiehl, M.; Krüger, W.; Mousset, S.; Neuburger, S.; Neumann, S.; Penack, O.; et al. Infectious Diseases Working Party of the German Society for Hematology and Medical Oncology (AGIHO/DGHO) and the DAG-KBT (German Working Group for Blood and Marrow Transplantation). Infectious diseases in allogeneic haematopoietic stem cell transplantation: Prevention and prophylaxis strategy guidelines 2016. *Ann. Hematol.* 2016, 95, 1435–1455.

- 140. Fishman, J.A. Infection in solid-organ transplant recipients. N. Engl. J. Med. 2007, 357, 2601–2614. [CrossRef]
- 141. Gavaldà, J.; Meije, Y.; Fortún, J.; Roilides, E.; Saliba, F.; Lortholary, O.; Muñoz, P.; Grossi, P.; Cuenca-Estrella, M.; ESCMID Study Group for Infections in Compromised Hosts. Invasive fungal infections in solid organ transplant recipients. *Clin. Microbiol. Infect.* **2014**, 20, 27–48. [CrossRef]
- 142. Barcellini, W.; Fattizzo, B.; Zaninoni, A. Management of refractory autoimmune hemolytic anemia after allogeneic hematopoietic stem cell transplantation: Current perspectives. *J. Blood Med.* **2019**, *10*, 265–278. [CrossRef]
- 143. Valent, P.; Lechner, K. Diagnosis and treatment of autoimmune haemolytic anaemias in adults: A clinical review. *Wien. Klin. Wochenschr.* **2008**, *120*, 136–151. [CrossRef] [PubMed]
- 144. Rose, N.R. Predictors of autoimmune disease: Autoantibodies and beyond. Autoimmunity 2008, 41, 419-428. [CrossRef] [PubMed]
- 145. Barcellini, W. New Insights in the Pathogenesis of Autoimmune Hemolytic Anemia. *Transfus. Med. Hemother.* **2015**, 42, 287–293. [CrossRef] [PubMed]
- 146. Giovannetti, G.; Pauselli, S.; Barrella, G.; Neri, A.; Antonetti, L.; Gentile, G.; Iacobini, M.; Girelli, G.; Coluzzi, S. Severe warm autoimmune haemolytic anaemia due to anti-Jk(a) autoantibody associated with Parvovirus B19 infection in a child. *Blood Transfus*. 2013, 11, 634–635. [PubMed]
- 147. Ramos-Casals, M.; García-Carrasco, M.; López-Medrano, F.; Trejo, O.; Forns, X.; López-Guillermo, A.; Muñoz, C.; Ingelmo, M.; Font, J. Severe autoimmune cytopenias in treatment-naive hepatitis C virus infection: Clinical description of 35 cases. *Medicine* 2003, 82, 87–96. [CrossRef] [PubMed]
- 148. Khawaja, S.; Abdul Muqtadir, K.; Taj, Y. Warm autoimmune haemolytic anaemia and autoimmune hepatitis in an asymptomatic carrier of hepatitis B virus. *J. Pak. Med. Assoc.* **2011**, *61*, 512–515.
- 149. Chiao, E.Y.; Engels, E.A.; Kramer, J.R.; Pietz, K.; Henderson, L.; Giordano, T.P.; Landgren, O. Risk of immune thrombocytopenic purpura and autoimmune hemolytic anemia among 120 908 US veterans with hepatitis C virus infection. *Arch. Intern. Med.* **2009**, 169, 357–363. [CrossRef]
- 150. Wilkinson, L.S.; Petz, L.D.; Garratty, G. Reappraisal of the role of anti-i in haemolytic anaemia in infectious mononucleosis. *Br. J. Haematol.* **1973**, 25, 715–722. [CrossRef]
- 151. Zeller, M.P.; Arnold, D.M.; Al Habsi, K.; Cserti-Gazdewich, C.; Delage, G.; Lebrun, A.; Heddle, N.M. Paroxysmal cold hemoglobinuria: A difficult diagnosis in adult patients. *Transfusion* **2017**, *57*, 137–143. [CrossRef]
- 152. Atta, M.; Brannigan, E.T.; Bain, B.J. Cold autoimmune hemolytic anemia secondary to atypical pneumonia. *Am. J. Hematol.* **2017**, 92, 109. [CrossRef]
- 153. Ramagopalan, S.V.; Goldacre, R.; Skingsley, A.; Conlon, C.; Goldacre, M.J. Associations between selected immune-mediated diseases and tuberculosis: Record-linkage studies. *BMC Med.* **2013**, *11*, 97. [CrossRef] [PubMed]
- 154. Eskazan, A.E.; Dal, M.S.; Kaya, S.; Dal, T.; Ayyildiz, O.; Soysal, T. Two cases of autoimmune hemolytic anemia secondary to brucellosis: A review of hemolytic disorders in patients with brucellosis. *Intern. Med.* **2014**, *53*, 1153–1158. [CrossRef] [PubMed]
- 155. Maslov, D.V.; Simenson, V.; Jain, S.; Badari, A. COVID-19 and Cold Agglutinin Hemolytic Anemia. *TH Open* **2020**, *4*, e175–e177. [CrossRef] [PubMed]
- 156. Lazarian, G.; Quinquenel, A.; Bellal, M.; Siavellis, J.; Jacquy, C.; Re, D.; Merabet, F.; Mekinian, A.; Braun, T.; Damaj, G. Autoimmune haemolytic anaemia associated with COVID-19 infection. *Br. J. Haematol.* **2020**, 190, 29–31. [CrossRef]
- 157. Lopez, C.; Kim, J.; Pandey, A.; Huang, T.; DeLoughery, T.G. Simultaneous onset of COVID-19 and autoimmune haemolytic anaemia. *Br. J. Haematol.* **2020**, *190*, 31–32. [CrossRef]
- 158. Patil, N.R.; Herc, E.S.; Girgis, M. Cold agglutinin disease and autoimmune hemolytic anemia with pulmonary embolism as a presentation of COVID-19 infection. *Hematol. Oncol. Stem Cell Ther.* **2020**. [CrossRef]