

## Article

# Chitosan–sEPDM and Melatonin–Chitosan–sEPDM Composite Membranes for Melatonin Transport and Release

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**Abstract:** Melatonin is the hormone that focuses the attention of the researchers in the medical, pharmaceutical, materials, and membranes fields due to its multiple biomedical implications. The variety of techniques and methods for the controlled release of melatonin is linked to the multitude of applications, among which sports medicine occupies a special place. This paper presents the preparation and characterization of composite membranes based on chitosan (Chi) and sulfonated ethylene-propylene-diene terpolymer (sEPDM). The membranes were obtained by controlled vacuum evaporation from an 8% sEPDM solution in toluene (*w/w*), in which chitosan was dispersed in an ultrasonic field (sEPDM:Chi = 1:1, *w/w*). For the comparative evaluation of the membranes' performances, a melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel:Chi:sEPDM = 0.5:0.5:1.0, *w/w/w*) test membrane was made. The prepared membranes were morphologically and structurally characterized by scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), energy-dispersive spectroscopy analysis (EDAX), thermal analysis (TG, DSC), thermal analysis coupled with chromatography and infrared analysis, and contact angle measurements, but also from the point of view of performance in the process of transport and release of melatonin in dedicated environments (aqueous solutions with controlled pH and salinity). The prepared membranes can release melatonin in amounts between 0.4 mg/cm<sup>2</sup>·per day (sEPDM), 1.6 mg/cm<sup>2</sup>·per day (Chi/sEPDM), and 1.25 mg/cm<sup>2</sup>·per day (Mel/Chi/sEPDM).

**Keywords:** melatonin; composite membranes; chitosan; sEPDM; melatonin transport and release



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## 1. Introduction

The biomedical implications of melatonin, the hormone secreted by the pineal gland, are so diverse and of particular importance that researchers have devoted extensive studies to it [1–3].

One of the current medical concerns is ensuring the daily amount of melatonin in the body, because the secretion of the pineal gland can be affected by various dysfunctions of the human body [4,5]. Even getting older is a problem in reducing the amount of melatonin generated in the body [6].

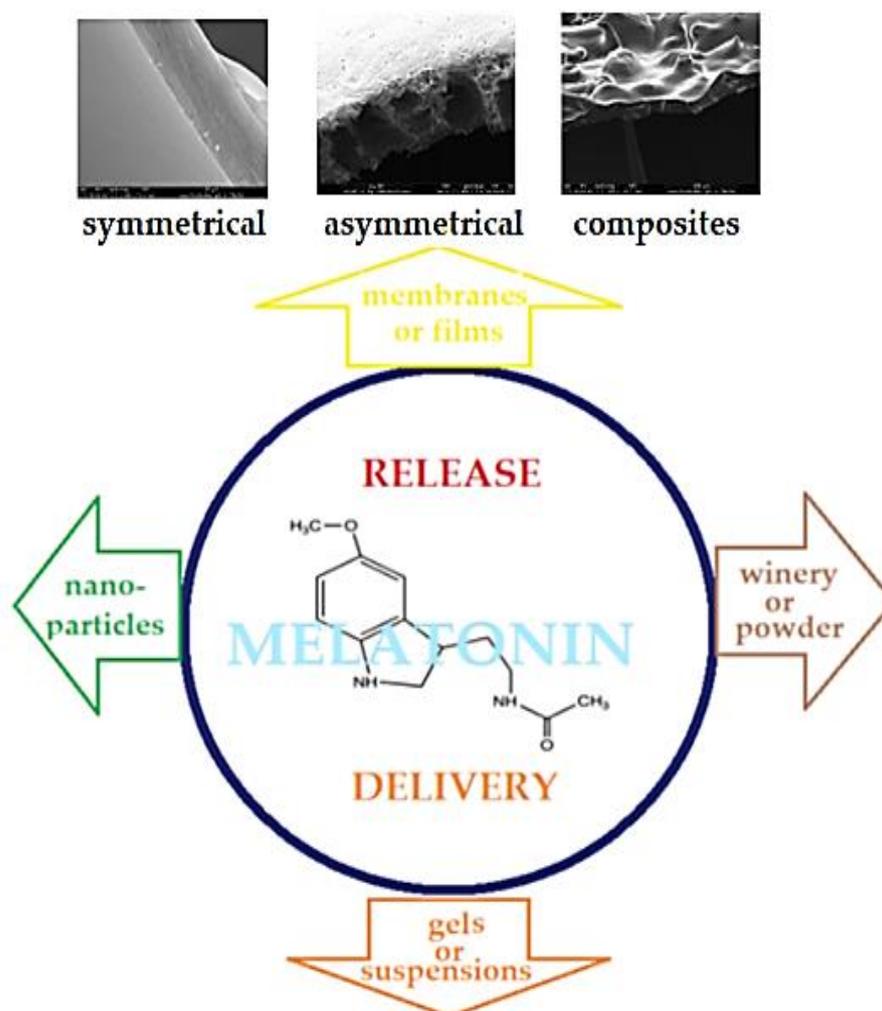
Starting with the circadian cycle [7], melatonin is involved in a series of biochemical processes related to: reducing oxidative stress [8], increasing aerobic performances [9],

tissue repair [10], muscle adaptation and skeletal muscle capacity [11], sleep and neuroprotection [12], blood pressure [13], metabolic diseases [14], cancer [15], rheumatic diseases [16], trauma and accidents [17], stimulation of cartilage matrix development [18], cardiovascular diseases [19], arthritis [20], and depression [21]. In order to have an overview of all of melatonin's connections with the body's health level, one can imagine a spiral scheme of its effects on some diseases or conditions of the body (Figure S1, see supplementary material).

The diversity of melatonin's implications in human body conditions has led to a variety of administration methods. The oral one predominates in current treatments because melatonin is easily adsorbed and desorbed from the digestive system, allowing the use of multiple materials and forms of conditioning [22–24].

However, there are more and more cases: trauma following various accidents [25], contusions [26], dislocation of bones from the joints [27], broken or displaced teeth in contact sports or that use special equipment or devices [28], blows to the head and spine [29], fractures and various accidents in the practice of motorized sports [30], which require the application and delivery of melatonin locally, alone or accompanying other drugs or pharmaceutical preparations [31].

The representative ways of delivery, controlled release of the various chemical species of interest, many of which have been studied for melatonin as well, are suggested in Figure 1 [32–44].



**Figure 1.** Schematic presentation of interest for chemical species' (melatonin) release or delivery techniques.

Among the recent applications of melatonin with significant results are those in sports medicine, while among the methods of controlled release, the attention given to the involvement of chitosan in various formulations can be highlighted [35].

Chitosan ensures a controlled release of melatonin, especially through ingestion, but for applications in sports medicine, orthopaedics, or dentistry, a reduction in the solubility of this biopolymer and an improvement in physical stability are necessary [36]. By embedding in various organic or inorganic nanoparticles, films, and membranes from biodegradable polymers, it is possible to reduce the solubility of chitosan [37].

Composite membranes are effective means both for the transport and the delivery or controlled release of melatonin [38], which justifies the expansion research in the field [39].

Studies related to improving the performance of fuel cell membranes have shown that an effective means of chitosan reticulation can be performed with ionophores with sulfonic groups of the polyether–ether sulfonated ketone type [40].

The study presented in this paper refers to the preparation and characterization of a composite membrane based on chitosan (Chi) and sulfonated ethylene-propylene-diene terpolymer (sEPDM) and its controlled melatonin release performance in synthetic aqueous solutions. The study addressed the transport and release of melatonin through the composite membrane (Chi-sEPDM) compared to the integral membrane from sEPDM. On the other hand, the results of the controlled release through the composite membrane (Chi-sEPDM) were compared to the parallel measurements of melatonin release from a melatonin/chitosan/sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) test composite membrane. The operational parameters of the melatonin transport and release experiments from a saturated source phase (SP) or from the test membrane were the pH and salinity (NaCl) of some synthetic receiving aqueous solutions (RP).

## 2. Materials and Methods

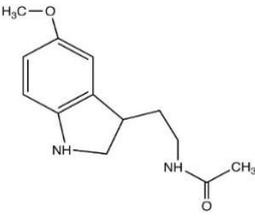
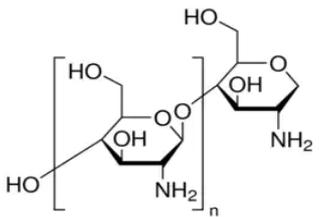
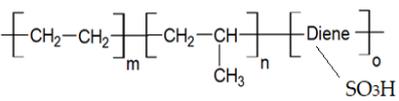
### 2.1. Reagents and Materials

All reagents and organic compounds used in the presented work were of analytical grade. They were purchased from Merck (Merck KGaA, Darmstadt, Germany): hydrochloric acid, sodium chloride, and sodium hydroxide.

Melatonin (Mel-Sigma-Aldrich, Merck KGaA, Darmstadt, Germany) and the basic polymeric materials for making the membranes, i.e., chitosan (Chi) (Sigma-Aldrich Chemie GmbH, Steinheim, Germany) and sulfonated ethylene-propylene-diene terpolymer (sEPDM), were recently used in our research group for ionic and molecular separations [41]. Their main characteristics are given in Table 1.

The purified water characterized by 18.2  $\mu\text{S}/\text{cm}$  conductivity was obtained with a RO Millipore system (MilliQ<sup>®</sup> Direct 8 RO Water Purification System, Merck, Darmstadt, Germany) [42].

**Table 1.** The characteristics of the used organic compounds.

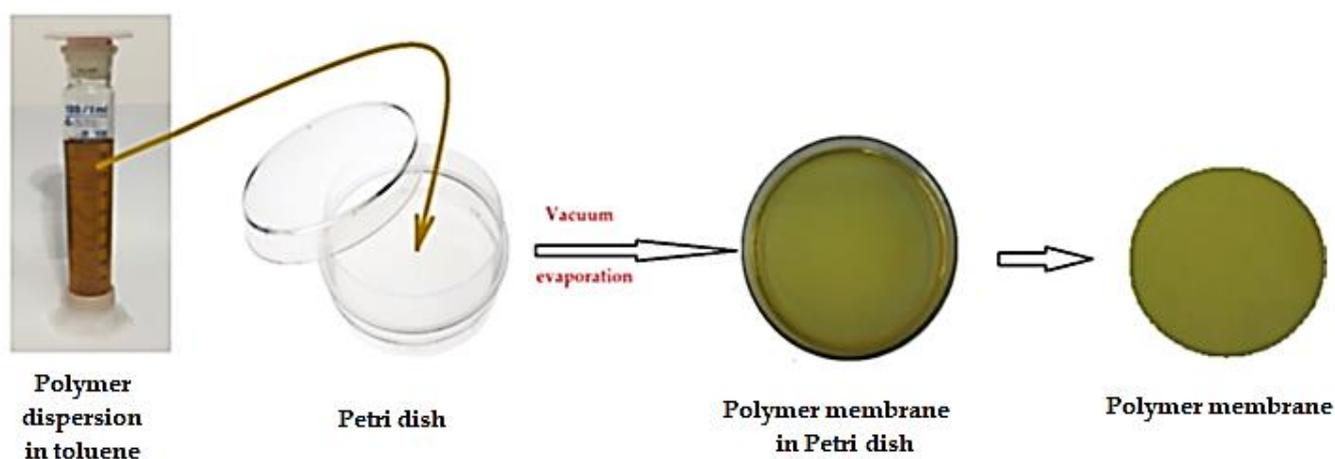
Organic Compounds	Name and Symbol	Molar Mass (g/mol)	Solubility in Water (g/L)	pKa
	Melatonin (Mel)	232.28	2g/L; max. $3 \cdot 10^{-3}$ mol/L	5.7 and 10.2
	Chitosan (Chi)	1526.5	soluble in acid media (0.5 M HCl: 50 mg/mL)	6.2 to 7.0
	sulfonated ethylene-propylene-diene terpolymer (sEPDM)	2500–6500	soluble in toluene	1.9 to 2.2

## 2.2. Methods and Procedures

### 2.2.1. Obtaining the Composite Membranes Based on SEPDM

Obtaining the membranes from sEPDM (M1), chitosan–sEPDM (M2), and melatonin–chitosan–sEPDM (M3) was carried out by phase inversion method [41] and by controlled evaporation technique [43].

The polymer solution (sEPDM 8%, *w/w*) in toluene is introduced in a Petri dish and evaporated in a vacuum oven at a temperature of 60 °C, thus obtaining the polymer membrane from sEPDM (M1) (Figure 2).

**Figure 2.** Schematic representation of the procedure for obtaining sEPDM based membranes.

To obtain the chitosan with sEPDM (M2) composite membrane, a dispersion of chitosan in toluene solution of sEPDM was made, by introducing 1 g of chitosan in 12.5 g of sEPDM solution (8% in toluene) so that the mass ratio of the two polymers is 1:1. To disperse the chitosan, the glass bottle was placed in an ultrasonic bath (Elmasonic S, Elma

Schmidbauer GmbH, Singen, Germany) for three hours, observing the complete dispersion. The dispersion is introduced into a Petri dish and evaporated under controlled conditions at 60 °C, under vacuum.

The preparation of the melatonin–chitosan–sEPDM composite membrane (M3) is carried out by using a mixture of melatonin and chitosan powders (0.5 g:0.5 g), which is homogenized in a dry way, for two hours, at 100 rpm, in a mill with spherical ceramic bodies (Retsch PM100 mill form Viola Schimadzu, Bucharest, Romania). The obtained powder is dispersed in 12.5 g toluene solution of sEPDM (8%, *w/w*), in the Elmasonic S ultrasonic bath, for three hours, obtaining a dispersion in toluene with a melatonin:chitosan:sEPDM ratio = 0.5 g:0.5 g:1.0 g. The dispersion is then introduced into a Petri dish and evaporated under controlled conditions at 60 °C, under vacuum.

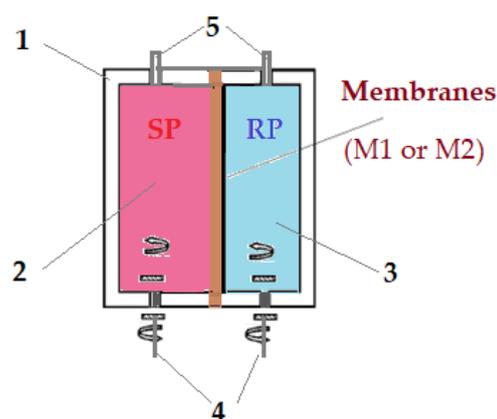
In all three cases, the complete evaporation of toluene was followed, by bringing the STERIPLAN ROTH Petri bottles to constant mass.

The membranes were cut into 1.2 cm and 3.3 cm diameter disks, intended for morphological, structural, transport, and release characterizations. The prepared membranes were morphologically and structurally characterized by scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), energy-dispersive spectroscopy analysis (EDAX), thermal analysis (TG, DSC), thermal analysis coupled with chromatography and analysis in infrared, and contact angle measurements, but also from the point of view of performance in the process of transport and release of melatonin in dedicated environments (aqueous solutions with controlled pH and salinity).

### 2.2.2. Transport and Release of the Melatonin through/from Composite Membranes Based on SEPDM

#### Transport Performance of the Obtained Membranes

To determine the transport performances of the prepared membranes (M1 or M2), a permeation module (Figure 3) with two compartments separated by a disk with a free membrane diameter of 3.3 cm was used [44]. Both compartments have a stirring magnetic bar (50 rpm) placed at the base. In one compartment, a 100 mL source solution of 2.0 g/L melatonin in ultra-pure water is introduced (the source phase, SP), and, in the second compartment, 100 mL synthetic receiving solutions of imposed pH made with hydrochloric acid or sodium hydroxide are introduced, in a range close to the biological pH (the receiving phase, RP).



**Figure 3.** Schematic representation of pertraction installation for the melatonin transport through prepared membranes (M1 and M2): 1-pertraction module; 2-source compartment; 3-receiving compartment; 4-magnetic stirrer system; 5-pipes; source phase (SP – 2g/L melatonin neutral aqueous solution); receiving phase (RP – pH and salinity-controlled solution).

In another set of experiments, 1–5% NaCl salinity solutions in ultra-pure water were used as the receiving phase. The experiments were carried out in five identical bipartite modules, with a volume of 100 mL melatonin solution and an imposed pH or salinity

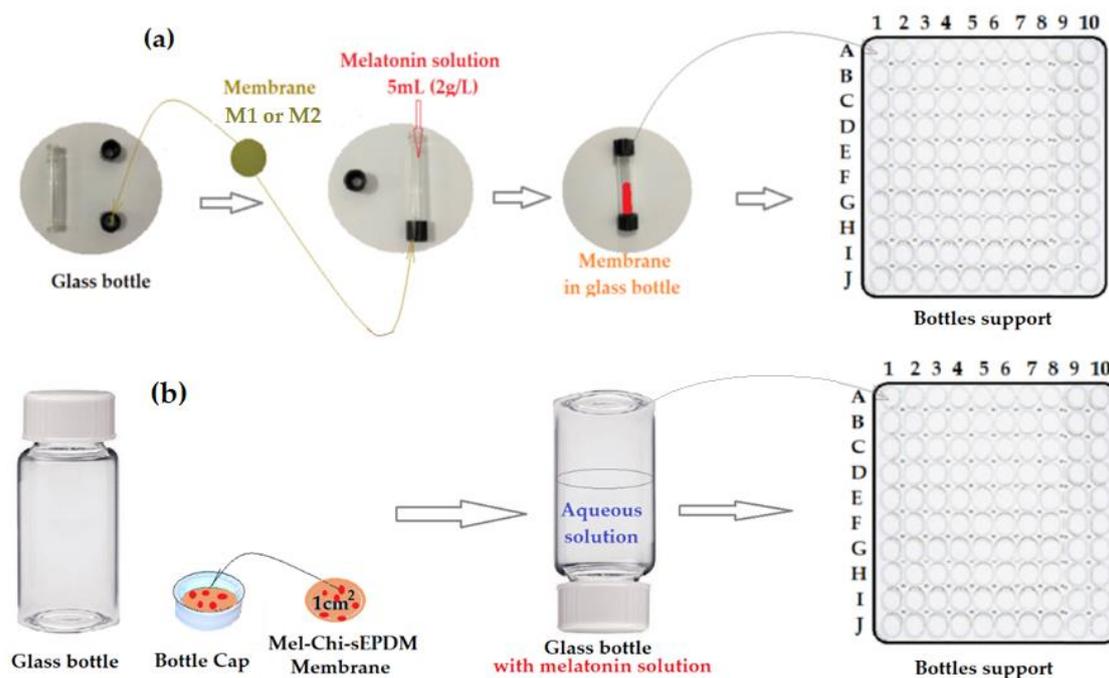
solution of the same volume, so that the results can be averaged. The five membranes, dedicated to each set of tests, were kept for 48 h in the 2 g/L melatonin solution and wiped by gentle pressing between two filter paper discs (Whatman® Filter Paper, Merck KGaA, Darmstadt, Germany) and then were fixed by silicone rubber gaskets in the permeation modules. The spectrophotometric analyses were performed daily, at two wavelengths, 278 nm, and 285 nm, for ten days, collecting 1.0 mL of solution from the source phase. The analyses were performed on two different spectrometers, by the same operator and repeated by an independent operator. The analysis laboratory works and respects the specific recommendations and guidelines of EURACHEM [45]. The validation of the analysis method was carried out by a fast and sensitive electrochemical method, developed and reported previously [46].

#### Release Performance of the Obtained Membranes

The controlled release experiments were carried out according to a previously described procedure [47].

In this study, two types of melatonin release are experimented:

1. The release of melatonin from a saturated solution “through” membranes prepared in synthetic solutions of controlled pH and salinity (Figure 4a);
2. The release of melatonin “from a” test composite membrane (Mel-Chi-sEPDM, M3) in synthetic solutions of controlled pH and salinity (Figure 4b).



**Figure 4.** Release schematic procedure “through” prepared membranes (M1 and M2) (a); and release schematic procedure “from” composite membrane (M3) (b).

In the first case, the membrane discs were placed in the lids of 10 cm<sup>3</sup> glass bottles. Then, 5.0 mL of controlled 2 g/L melatonin aqueous solution was introduced into the glass bottles, and the perforated bottle cap was sealed with a membrane and placed with the cap down in a cup in which 100 bottles could be inserted simultaneously (Figure 4a). The entire assembly is placed in a vessel (10.0 L, simulating a high, quasi-constant dilution, and to maintain the concentration gradient for melatonin), with controlled pH or salinity solution, which is recirculated with a constant flow rate (200 mL/min). Seven bottles were retrieved daily for analysis so that the results of the melatonin analysis could be averaged, and three bottles were stored as control samples.

In the second case, the composite membranes (Mel-Chi-sEPDM, M3) are placed in the lid of the 100 bottles containing 5.0 mL synthetic solution of imposed pH and salinity, and melatonin from the membranes is released in contact with them (through overturning) (Figure 4b).

As in the previous case, seven bottles were retrieved daily for analysis so that the results of the melatonin analysis could be averaged, and three bottles were stored as control samples.

The validation of the results was performed periodically by electrochemical and/or UV-Vis methods at an independent laboratory [47,48].

The flows of the melatonin derivatives from the source phase were determined at specific time intervals, using the relation (1) [49]:

$$J = \frac{M}{S \cdot \Delta t} \text{ (mg/(m}^2 \text{ s)) or mol/(m}^2 \text{ s))} \quad (1)$$

$M$  being the permeate mass (g or mol),  $S$  being the effective surface of the membrane (m<sup>2</sup>), and  $\Delta t$  the time interval (s).

The release (or pertraction) efficiency RE% (or PE%) for the melatonin derivatives was calculated as follows [50], based on melatonin solution concentration:

$$RE \text{ or } PE(\%) = \frac{(c_0 - c_f)}{c_0} \cdot 100 \quad (2)$$

$c_f$  being the final concentration of the solute (melatonin) and  $c_0$  the initial concentration of solute (melatonin).

The same release efficiency can also be obtained based directly upon the absorbance of the considered solutions (melatonin) [51], as in (3):

$$RE \text{ or } PE(\%) = \frac{(A_0 - A_s)}{A_0} \cdot 100 \quad (3)$$

$A_0$  being the initial absorbance of the sample melatonin solution and  $A_s$  the current absorbance of the sample.

### 2.3. Equipment

The surface and cross-section characteristics of the membranes were determined with a scanning electron microscopy (SEM) equipped with a probe for energy dispersive spectroscopy analysis (EDX). A Hitachi S4500 system was used (Hitachi High-Technologies Europe GmbH, Krefeld, Germany) [52].

Thermal analysis (TG-DSC) was performed with a STA 449C Jupiter apparatus, from Netzsch (NETZSCH-Gerätebau GmbH, Selb, Germany). Each sample weighed approximately 10 mg. The samples were placed in an open alumina crucible and heated up to 900 °C with 10 K·min<sup>-1</sup> rate, under flow of 50 mL·min<sup>-1</sup> dried air. As reference, we used an empty alumina crucible. The evolved gases were analysed with a FTIR Tensor 27 from Bruker (Bruker Co., Ettlingen, Germany), equipped with a thermostat gas cell [53].

FTIR 2D maps were recorded with a Nicolet iS50R FTIR microscope (Thermo Fisher Scientific Inc., Waltham, MA, USA), with a deuterated triglycine sulphate (DTGS) detector, in the wavenumber range 4000–600 cm<sup>-1</sup>. The FTIR 2D maps were used to obtain information about the spatial distribution of the components [54].

Determination and monitoring of pH and salinity for every stock solution was achieved using a conductance cell or combined selective electrode (HI 4107, Hanna Instruments Ltd., Leighton Buzzard, UK) and a multi-parameter system (HI 5522, Hanna Instruments Ltd., Leighton Buzzard, UK) [55].

The UV-Vis spectra of the melatonin samples were recorded for a wavelength ranging from 200 to 800 nm, at room temperature, using 10 mm quartz cells on CamSpec M550

spectrometer (Spectronic CamSpec Ltd., Leeds, UK), and, for the daily determinations, two wavelengths were chosen, 278 nm and 285 nm [56,57].

Additionally, the UV–Vis validation analysis of the melatonin solutions was performed on dual-beam UV equipment—Varian Cary 50 (Agilent Technologies Inc., Santa Clara, CA, USA)—at a resolution of 1 nm, spectral bandwidth of 1.5 nm, and a scan rate of 300 nm/s [57].

Contact angle measurements for the considered spheres materials (with distilled water or melatonin derivatives solution) [58] were carried out with a horizontal microscope with video camera (Viola–Shimadzu, Bucharest, Romania).

### 3. Results and Discussion

The controlled release of pharmaceutical preparations is an important aspect that doctors take into account both when prescribing drug doses and when administering food supplements [59–62].

In the case of melatonin, the possibility of oral administration allows its inclusion in powdery materials, tablets, or cassettes, which, by ingestion, ensure the release of controlled amounts in the body [63]. If a localized administration is desired (injuries, trauma, areas of the oral cavity), as is the case in sports accidents, creams, gels, patches, or films (membranes) can be used [64–69].

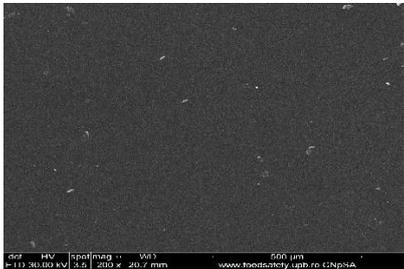
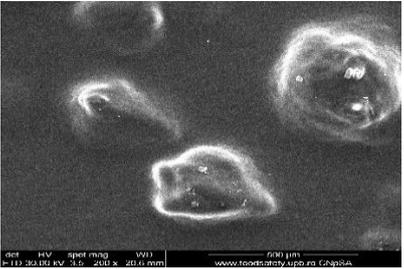
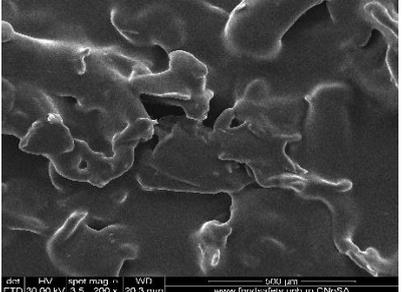
In the present study, the controlled transport or release of melatonin through a chitosan (Chi)-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) composite membrane, with possible applications in sports medicine, were considered. The results of the controlled release through the composite membrane (Chi-sEPDM) were compared to the parallel measurements of melatonin release from a melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer composite test membrane (Mel-Chi-sEPDM).

The membrane prepared by controlled evaporation from a chitosan or a melatonin–chitosan dispersion in sEPDM solution in toluene was characterized morphologically and structurally by scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), energy-dispersive spectroscopy analysis (EDAX), thermal analysis (TG, DSC), thermal analysis coupled with chromatography and infrared analysis, but also from the point of view of melatonin transport to solutions of controlled pH and salinity.

#### 3.1. Morphological and Structural Membrane Characteristics

The obtained membranes have the macroscopic characteristics illustrated in Table 2: the thickness, the general aspects (photo), low resolution scanning electron microscopy, and the contact angle with distilled water. The membranes have very different appearance, roughness, morphology, and wetting characteristics, which can be found in the performance of transport and/or release of melatonin in aqueous solutions of variable pH and salinity. Although the thickness of the three membranes is relatively similar, in the micrometric determinations, it is still affected by the very different roughness [70]. The hydrophilicity of the membranes increases sharply in the series  $M1 < M2 < M3$ , both through the influence of the composition and the roughness found in low resolution scanning electron microscopy (Table 2).

**Table 2.** The macroscopic characteristics for the obtained membranes.

Material	Membrane Symbols	Thickness ( $\mu\text{m}$ )	Membrane View (Photo)	Low Magnitude SEM (Membrane Surface)	Contact Angle ( $\theta^\circ$ )
sEPDM	M1	$50 \pm 2$			$73 \pm 3$
Chi-sEPDM	M2	$51 \pm 4$			$42 \pm 5$
Mel-Chi-sEPDM	M3	$49 \pm 5$			$33 \pm 5$

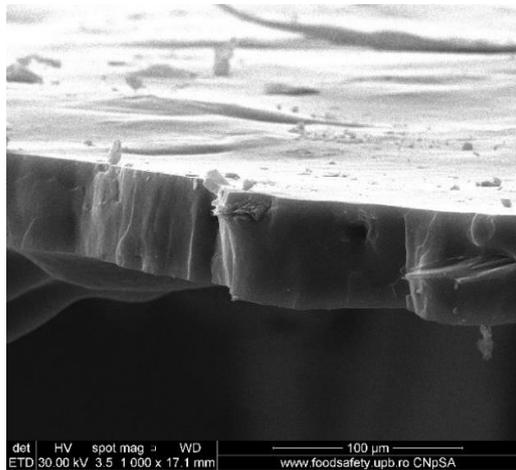
### 3.1.1. Scanning Electron Microscopy (SEM)

The membrane samples based on sulfonated ethylene-propylene-diene terpolymer (sEPDM) (M1), chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) (M2), and melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) (M3) with a size of  $10 \text{ cm}^2$ , were fractured in liquid nitrogen and metallized with a superficial layer of gold, to be able to examine the section of the membranes (scanning electron microscopy, SEM) and the elemental distribution on the surface (energy-dispersive spectroscopy analysis, EDAX), analyses available on a Hitachi S4500 system.

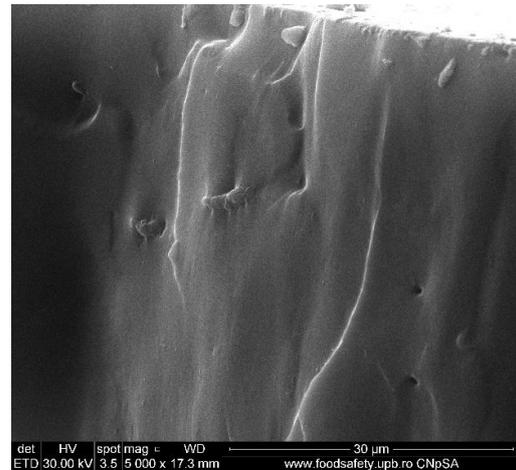
Figure 5 shows the images obtained for the three membranes (M1, M2 and M3), at a magnification of  $\times 1000$  for the cross-section (Figure 5a,c,e), and the cross-section details  $\times 5000$  (Figure 5b,d,f). Figure 6 illustrates the elemental composition.

The uniformity of the sEPDM membrane surface (M1, Figure 5a) is specific to an integral membrane obtained by phase inversion through evaporation, both on the surface and in the section. The electrostatic charge causes the adhesion of several polymer fragments on the surface, and isolated crystallites are observed in the section (Figure 5a,b).

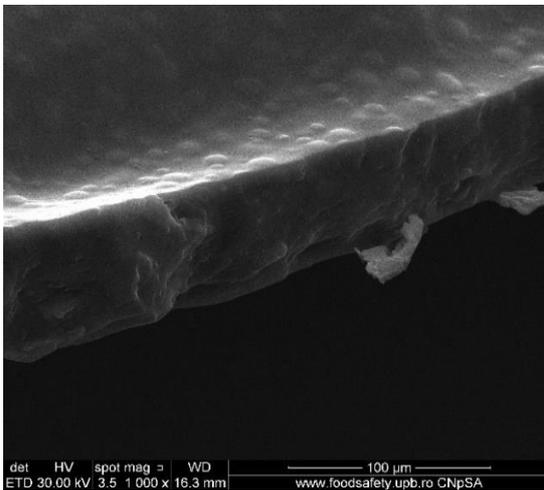
The images of the two composite membranes (M2 and M3) (Figure 5b–f) highlight the heterogenous nature, the micro-aggregates of chitosan or melatonin being clearly observed especially on the surfaces (Figure 5c,d) but also in sections (Figure 5d,f). These superficial agglomerations amplify to a microscopic level the roughness of the two composite membranes, also observed in the assembly presented in Table 2.



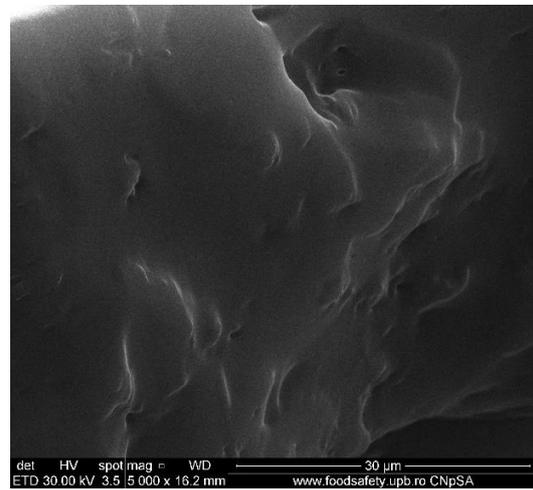
(a)



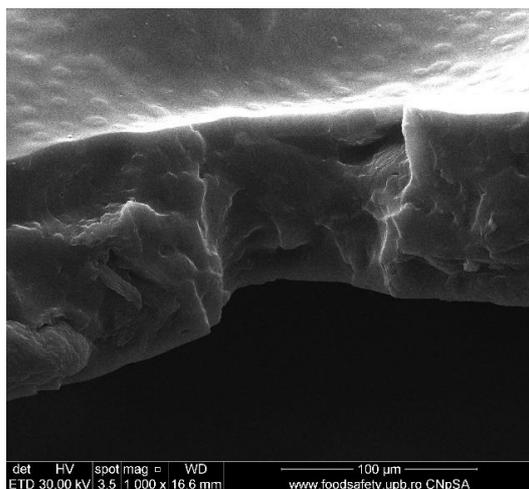
(b)



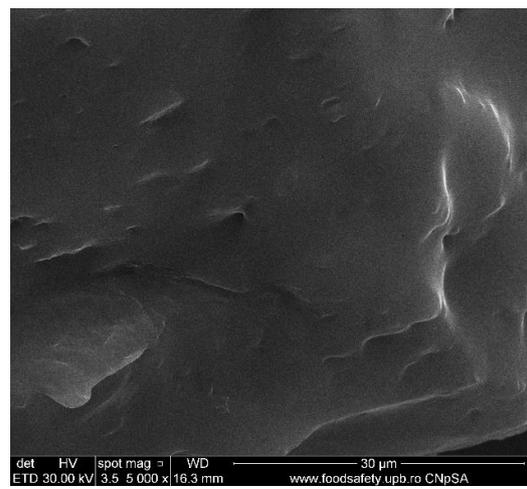
(c)



(d)

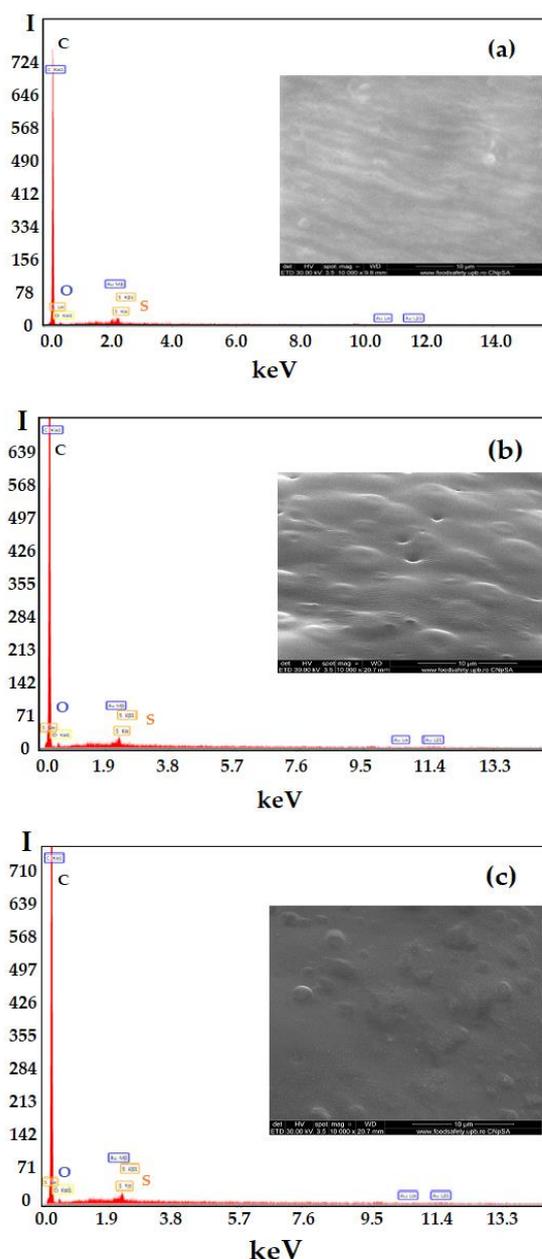


(e)



(f)

**Figure 5.** Scanning electron microscopy (SEM) images for: sulfonated ethylene-propylene-diene terpolymer (sEPDM) (M1)—(a,c,e); chitosan (Chi)-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) (M2)—(b,d,f).



**Figure 6.** Energy-dispersive spectroscopy analysis (EDAX) diagram for: sulfonated ethylene-propylene-diene terpolymer (sEPDM) (M1) (a); chitosan (Chi)-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) (M2) (b), melatonin/chitosan (Chi)-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) (M3) (c).

The elemental analysis on the surface (EDAX) allows highlighting of carbon (C) elements, the majority, but also oxygen (O) and sulphur (S) on all membranes: the sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) (Figure 6a) and in the chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) composite membrane (M2) (Figure 5b), and melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) composite membrane (M3) (Figure 5c).

The surface elemental concentration is slightly different for all membranes (Table 3). However, one observes a remarkable reduction with almost half of the sulphur concentration and with a third of the oxygen concentration in the case of the Chi-sEPDM membrane (M2), compared to the melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) composite membrane (M3) in the examined areas.

**Table 3.** Energy dispersive spectroscopy analysis (EDAX) for the prepared membranes.

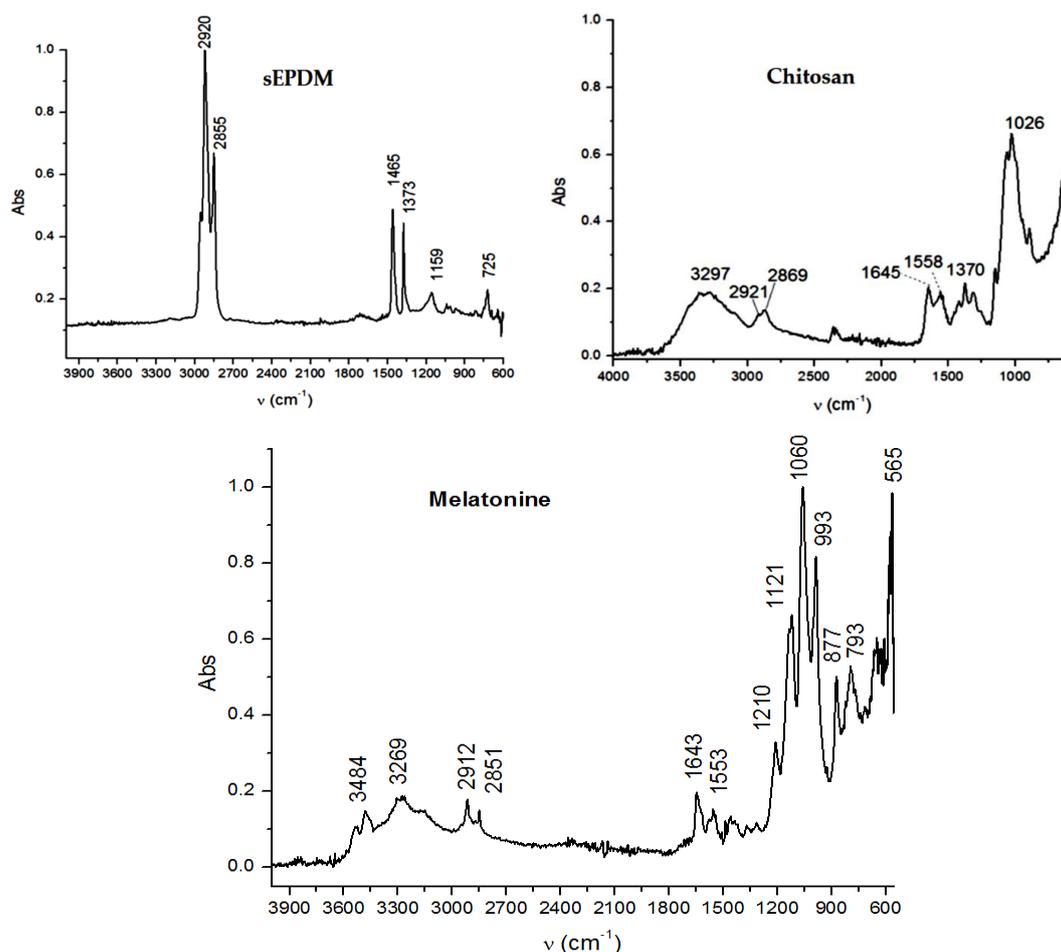
Membranes	M1			M2			M3		
Surface Composition	Weight (%)	Atomic (%)	Error (%)	Weight (%)	Atomic (%)	Error (%)	Weight (%)	Atomic (%)	Error (%)
C K	94.48	95.88	3.07	96.37	97.3	2.29	93.16	94.92	2.88
O K	5.28	4.02	29.5	3.49	2.64	30.79	6.44	4.93	20.94
S K	0.24	0.09	62.31	0.15	0.06	61.64	0.4	0.15	16.27

Although the difference in elemental surface composition is relatively small, the hydrophilicity of the membranes' surfaces changes dramatically (Table 2): the composite membrane (M3) being much more hydrophilic ( $\theta = 35^\circ$ ), comparatively with the composite membrane (M2) being more hydrophilic ( $\theta = 45^\circ$ ), and much more than the sEPDM membrane (M1,  $\theta = 75^\circ$ ). This observation shows, for the present case, that the surface roughness is decisive for the hydrophilicity of the membranes.

### 3.1.2. Fourier Transform InfraRed Spectroscopy (FTIR) Membrane Characteristics

The data obtained from the elemental analysis (EDAX) required a study in the infrared domain both spectrally (FTIR) and by interference reflection microscopy (IRM), which would provide more structural information and surface composition.

Figure 6 shows the spectra of the base materials: sulfonated ethylene-propylene-diene terpolymer (sEPDM), chitosan (Chi), and melatonin (Mel) (Figure 7).



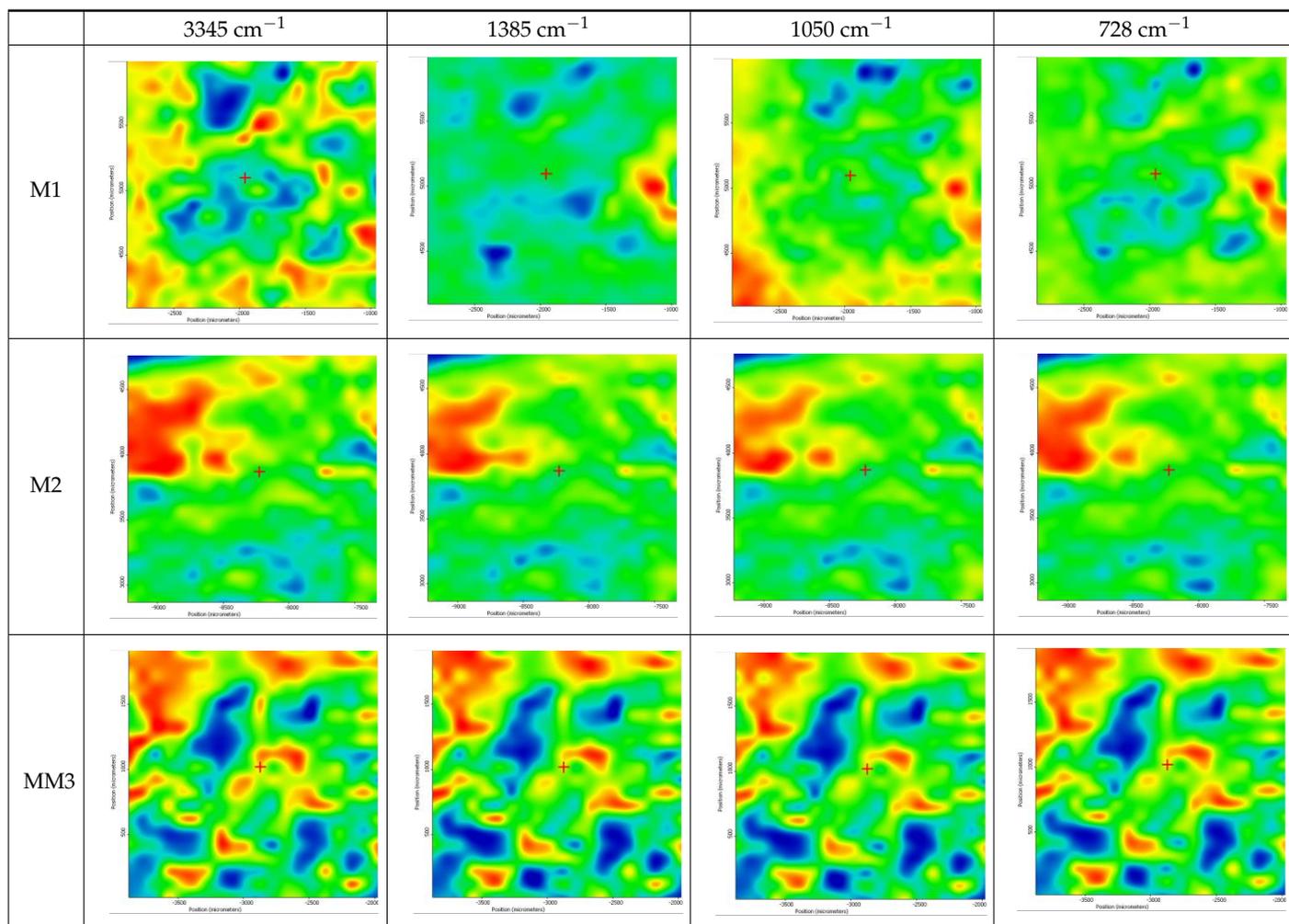
**Figure 7.** Fourier transform infrared spectra for: sulfonated ethylene-propylene-diene terpolymer (sEPDM); chitosan (Chi), and melatonin (Mel).

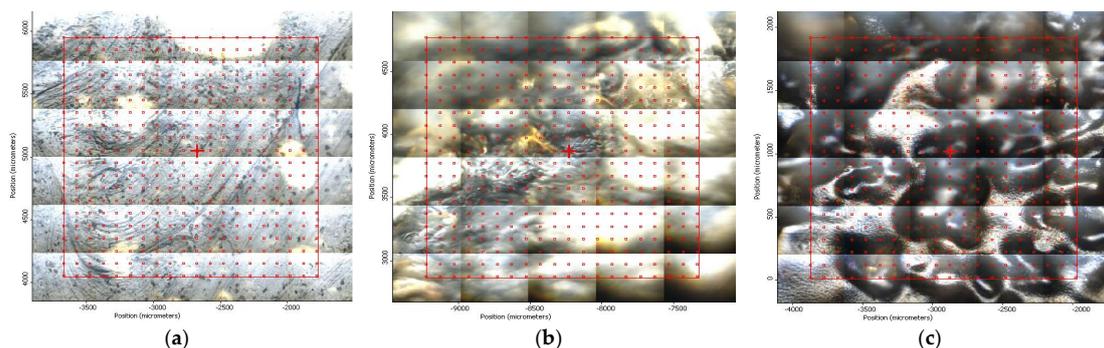
The spectra obtained were used to select the wave numbers for which the infrared microscopy map (FTIR, 2D) was made, for the three membranes obtained.

Most of the specific wave numbers of the three materials are located in very close areas and therefore cannot be used as safe specific values for the FTIR microscopy study. It should also be emphasized that the sEPDM film subjected to FTIR analysis was obtained from toluene solution, which did not favour the highlighting of hydrogen bonds of the sulfonic group (Figure 7). The examined chitosan and melatonin were presented as a powder, and the obtained spectra are compatible with the literature data.

In another train of thought, the three materials used to obtain the composite membrane interact. Thus, the sulfonic groups in sEPDM give a neutralization reaction with primary amino groups from chitosan and secondary amino groups from melatonin, but there are also other possible interactions such as hydrogen bonds, ionic bonds, and hydrophobic bonds (Table 1). Figure 8 shows the images of the selected areas (M1 in Figure 8a, M2 in Figure 8b, and M3 in Figure 8c), and Table 4 shows FTIR 2D maps, at randomly selected wave numbers, but targeting each representative range of the spectra:  $3345\text{ cm}^{-1}$ ,  $1385\text{ cm}^{-1}$ ,  $1050\text{ cm}^{-1}$ , and  $728\text{ cm}^{-1}$ .

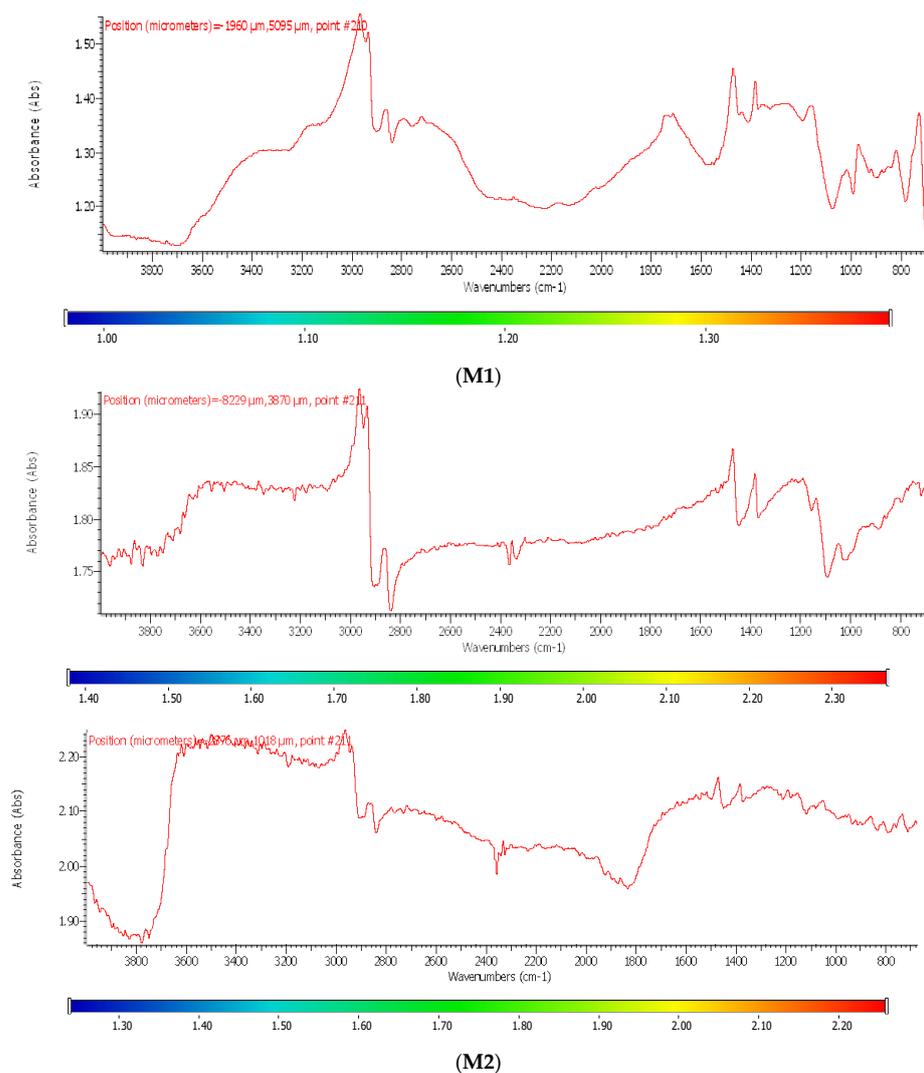
**Table 4.** The FTIR 2D maps for sEPDM membrane (M1) and composite membrane (M2 and M3).



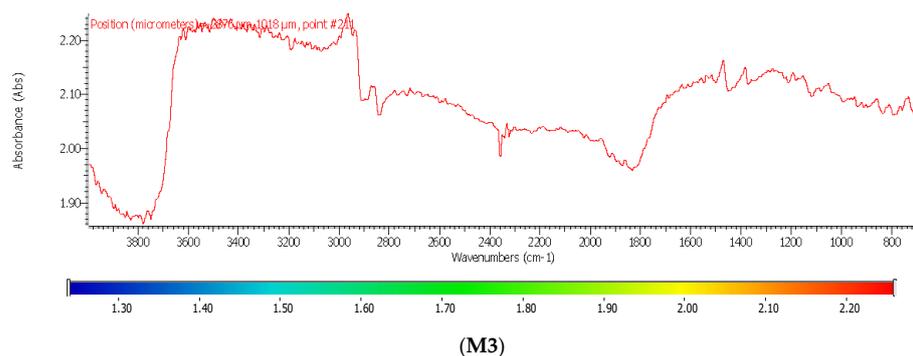


**Figure 8.** Video images for; (a) sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1); (b) chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2); and (c) melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) membrane (M3).

The associated spectra and the colour scale used are shown in Figure 9, showing significant differences that can largely justify the differences in hydrophilicity presented by the prepared membranes (Tables 1 and 4).



**Figure 9.** Cont.



**Figure 9.** Infrared associated spectrum and colour scales for sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1); chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2), and melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) membrane (M3).

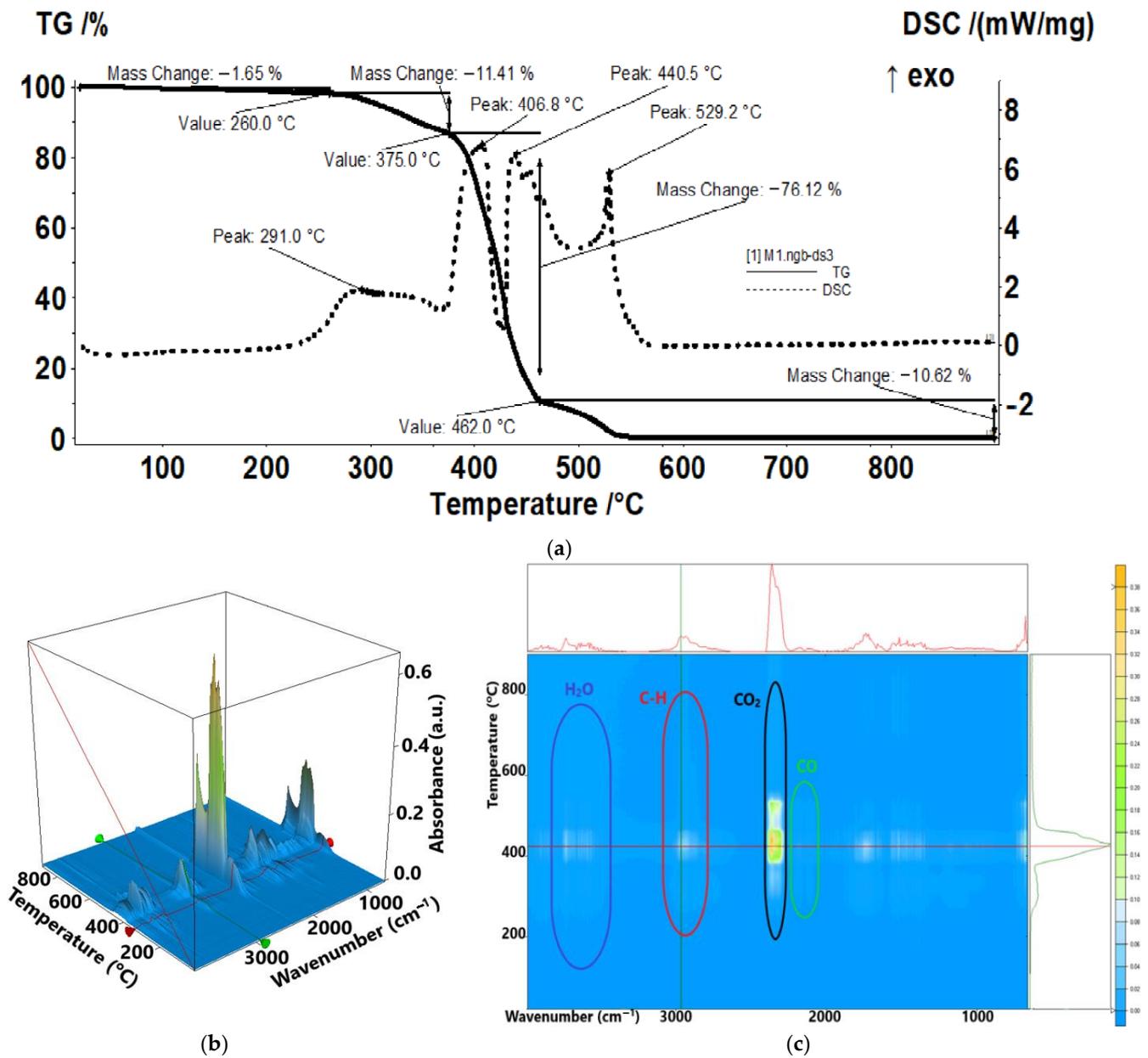
The HD-IR obtained maps show a relatively uniform, regular, and repeatable distribution of the surface of the obtained membranes, especially for the sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1). The composite chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2), and melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) membrane (M3) show areas that, with the greatest probability, are due to the agglomeration of chitosan or melatonin-chitosan (upper left corner of the images), being more obvious for a wave number of  $3345\text{ cm}^{-1}$ , but also present for all other wave numbers (see Table 4). This agglomeration was also highlighted in the scanning electron microscopy details (Figure 5c,e).

### 3.1.3. Thermal Characteristics of the Prepared Membranes

The complex thermal analysis had both the role of highlighting the thermal behaviour of the membranes at relatively low temperatures (up to  $300\text{ }^{\circ}\text{C}$ ) and their composition through gas chromatographic analysis coupled with infrared spectrometry of combustion gases (up to  $800\text{ }^{\circ}\text{C}$ ).

The sample M1 (Figure 10) can be considered stable up to  $260\text{ }^{\circ}\text{C}$ , losing only 1.65% of its mass, mainly residual solvent but also some sulphur is removed as  $\text{SO}_2$ , as indicated by the FTIR analysis of the evolved gases. Between  $260\text{--}375\text{ }^{\circ}\text{C}$ , the sample is losing 11.41% of its mass, the process being accompanied by a broad exothermic effect with peak at  $291\text{ }^{\circ}\text{C}$ . The main degradation process takes place between  $375\text{--}462\text{ }^{\circ}\text{C}$ , when the sample is losing 76.12% of its mass (Figure 10a). The DSC curve indicates two strong exothermic effects, but the FTIR of the evolved gases indicates a quasi-continuous production of  $\text{CO}_2$ ,  $\text{H}_2\text{O}$ , or hydrocarbon fragments, which means that any backbone breaking in smaller fragments is also accompanied by the combustion of those fragments (Figure 10a–c). The FTIR spectrum at  $429\text{ }^{\circ}\text{C}$ , in the middle of the strongest degradation process, indicates the evolving of  $\text{H}_2\text{O}$ ,  $\text{CO}_2$ , and  $\text{CO}$  as combustion products, but also of saturated hydrocarbon fragments from pyrolysis of the polymer backbone and  $\text{SO}_2$ . The residual carbonaceous mass is burned after  $460\text{ }^{\circ}\text{C}$ , the process being accompanied by a strong exothermic peak at  $529.2\text{ }^{\circ}\text{C}$ . The FTIR analysis of evolved gases at  $529\text{ }^{\circ}\text{C}$  indicates that the product is mainly  $\text{CO}_2$ . It can be seen that some desulfurization processes also take place under  $200\text{ }^{\circ}\text{C}$ .

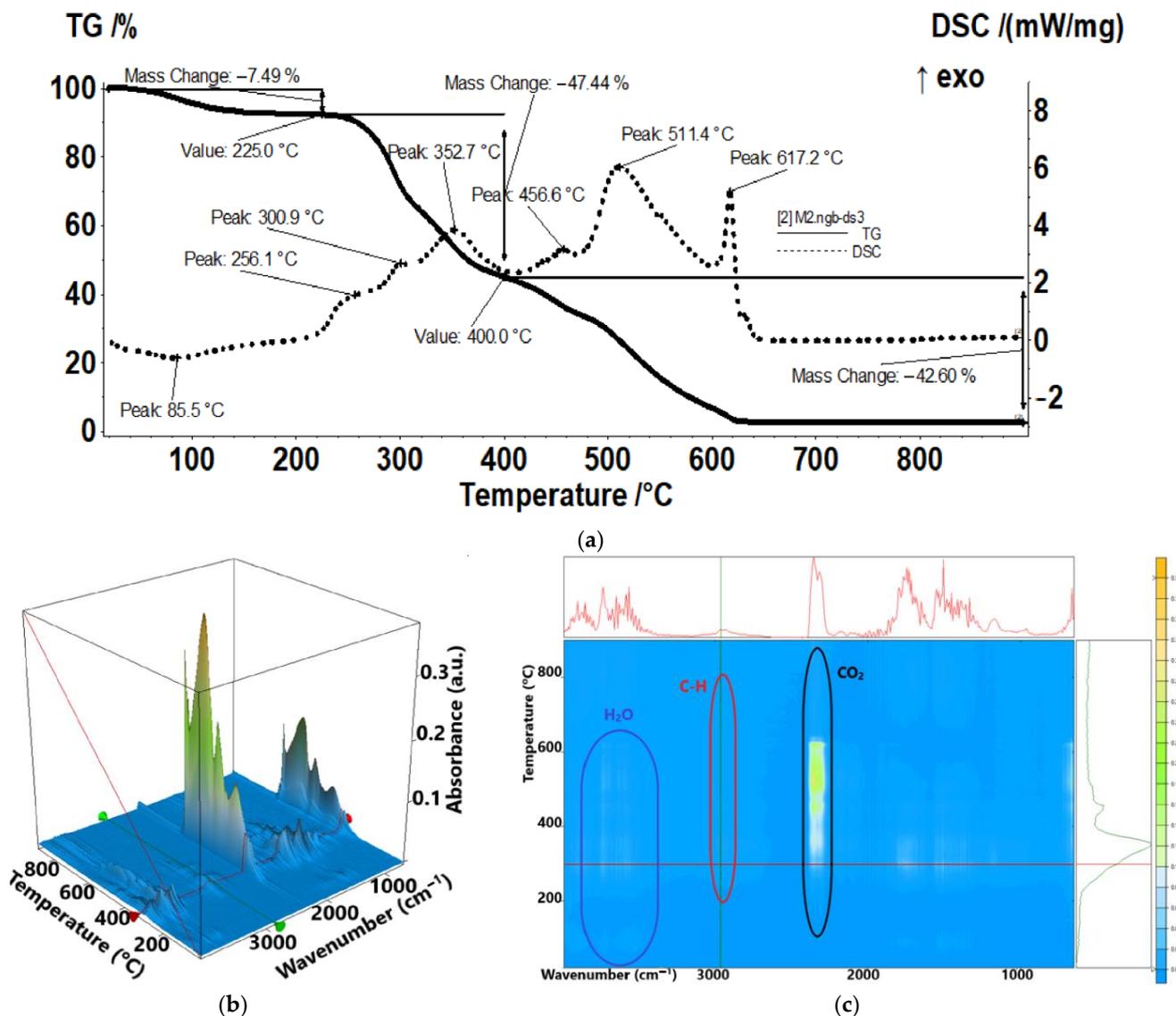
The traces for evolving  $\text{SO}_2$  ( $1367\text{ cm}^{-1}$ ) vs. temperature and traces for evolving  $\text{CO}_2$  ( $2355\text{ cm}^{-1}$ ) vs. temperature were detailed in the supplementary material (Figure S2, see supplementary material).



**Figure 10.** Thermal characteristics of the sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1): (a) thermal diagram; (b) 3D complex analysis; (c) 2D complex analysis.

The sample M2 (Figure 11) is losing 7.49% in the temperature interval RT–225 °C, the associated effect being weak and endothermic with minimum at 85.5 °C. The sample is losing some residual water molecules in this interval, but also the desulfurization processes start (Figure 11a), as indicated by the FTIR of the evolved gases and the traces for individual wavenumbers vs. temperature (Figure 11b). In the interval 225–400 °C, the sample begins to suffer an oxidative degradation, with multiple exothermic peaks visible on the DSC curve, partially overlapped. The FTIR of evolved gases allows identification of combustion products such as CO<sub>2</sub>, CO, and H<sub>2</sub>O, but also saturated hydrocarbons from polymer backbone fragmenting and SO<sub>2</sub>, indicating the complexity of the thermal degradation (Figure 10b,c). The majority of SO<sub>2</sub> is evolving in this interval, with only minor peaks being identified on the compound trace after 400 °C (Figure 10a). The same can be stated for the saturated hydrocarbon fragments: after 400 °C, only a small peak is observable on the trace line (Figure 10c). After 400 °C, the sample suffers mostly oxidation processes, as indicated by the evolving of CO<sub>2</sub> and H<sub>2</sub>O in larger quantities, culminating with the burning of the

residual carbonaceous mass, which is accompanied by the strong and sharp exothermic effect from 617.2 °C. The recorded mass loss after 400 °C is 42.60%.

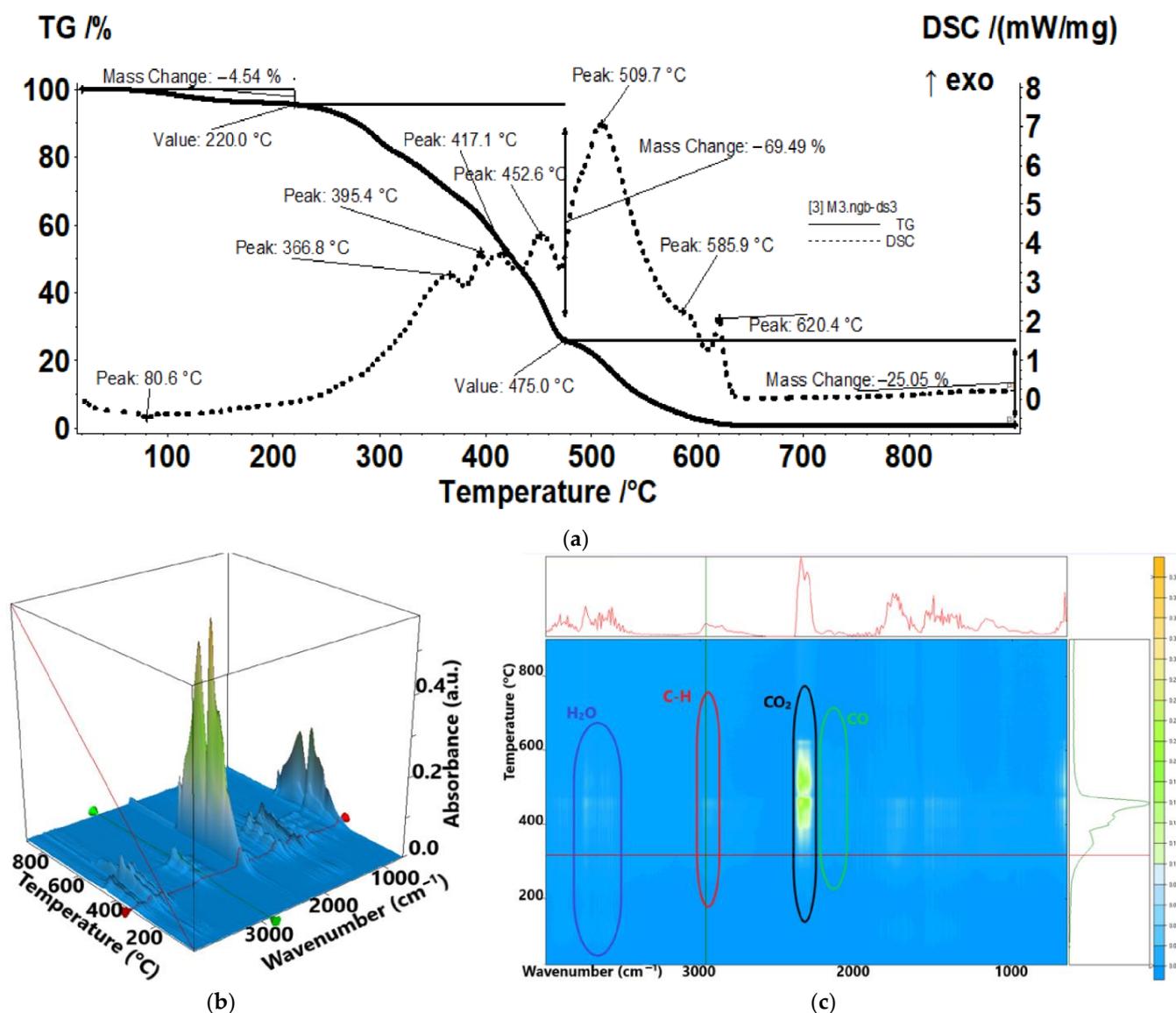


**Figure 11.** Thermal characteristics of the chitosan–sulfonated ethylene-propylene-diene terpolymer membranes (Chi-SEPDM) (M2): (a) thermal diagram, (b) 3D complex analysis; (c) 2D complex analysis.

The traces for evolving SO<sub>2</sub> (1367 cm<sup>-1</sup>), CO<sub>2</sub> (2355 cm<sup>-1</sup>), hydrocarbons (2964 cm<sup>-1</sup>), and H<sub>2</sub>O (3566 cm<sup>-1</sup>) vs. temperature were detailed in the supplementary material (Figure S3, see supplementary material).

Sample M3 (Figure 12) is losing 4.54% up to 220 °C, mostly water molecules and some traces of SO<sub>2</sub>, as indicated by the FTIR spectra. The principal degradation processes take place between 220 °C and 475 °C, when a series of exothermic effects are seen on the DSC curve, indicating multiple oxidation reactions. The bulk of gaseous products CO, H<sub>2</sub>O, hydrocarbon fragments, and SO<sub>2</sub> is eliminated in this interval. The recorded mass loss is 69.49%. After 475 °C, the residual carbonaceous mass is burned, the principal degradation product identified by FTIR being CO<sub>2</sub>.

The traces for evolving SO<sub>2</sub> (1367 cm<sup>-1</sup>), CO<sub>2</sub> (2355 cm<sup>-1</sup>), hydrocarbons (2964 cm<sup>-1</sup>), and H<sub>2</sub>O (3566 cm<sup>-1</sup>) vs. temperature were detailed in the supplementary materials (Figure S4, see supplementary material).



**Figure 12.** Thermal characteristics of the melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer membranes (Mel-Chi-sEPDM) (M3): (a) thermal diagram, (b) 3D complex analysis; (c) 2D complex analysis.

The results of complex thermal analysis, coupled with the gas chromatography and infrared spectrometry, provide the stability range of each prepared membrane but, above all, confirm their composition.

### 3.2. Transport and Release of the Melatonin through Prepared Membranes

A natural hormone, synthesized in the body, melatonin can be administered orally most of the time, and the main concern of the researchers was to find the most suitable methods of controlled release [71].

However, there are some specific aspects that make melatonin remain constantly in the attention of researchers in order to design new methods of delivery in the body:

- A universal dose of melatonin cannot be prescribed because each body has its own production [72];
- Age and health greatly affect the production of the pineal gland [73];
- The time of day is very important because the production of melatonin in the body is cyclical [74];

- In the case of accidents, especially those from various sports competitions, local administration is necessary (oral cavity, skin, bones, joints) [75,76].

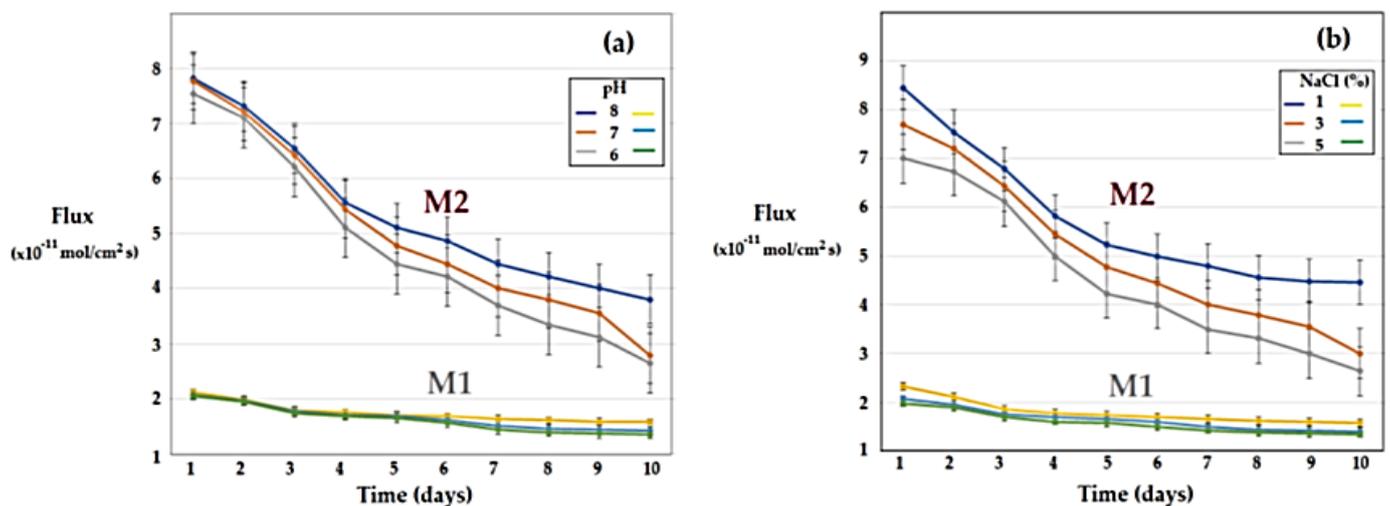
All these considerations have encouraged experimental research on the transport and/or release of melatonin through composite membranes of chitosan-sulfonated ethylene-propylene-diene terpolymer membranes (Chi-sEPDM), and the melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer composite membranes (Mel-Chi-sEPDM), even if recently a mathematical model of the release of various active substances has been proposed [66].

In this study, the transport “through” the composite membrane obtained in a two-compartment membrane system and the release of melatonin “through” the composite membrane in an open system (the receiving solution is renewed) were followed.

For comparison, melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer (Mel-Chi-sEPDM) composite membrane was tested in a system for releasing melatonin “from inside” the membrane to a receiving phase consisting of synthetic solutions with imposed pH and salinity.

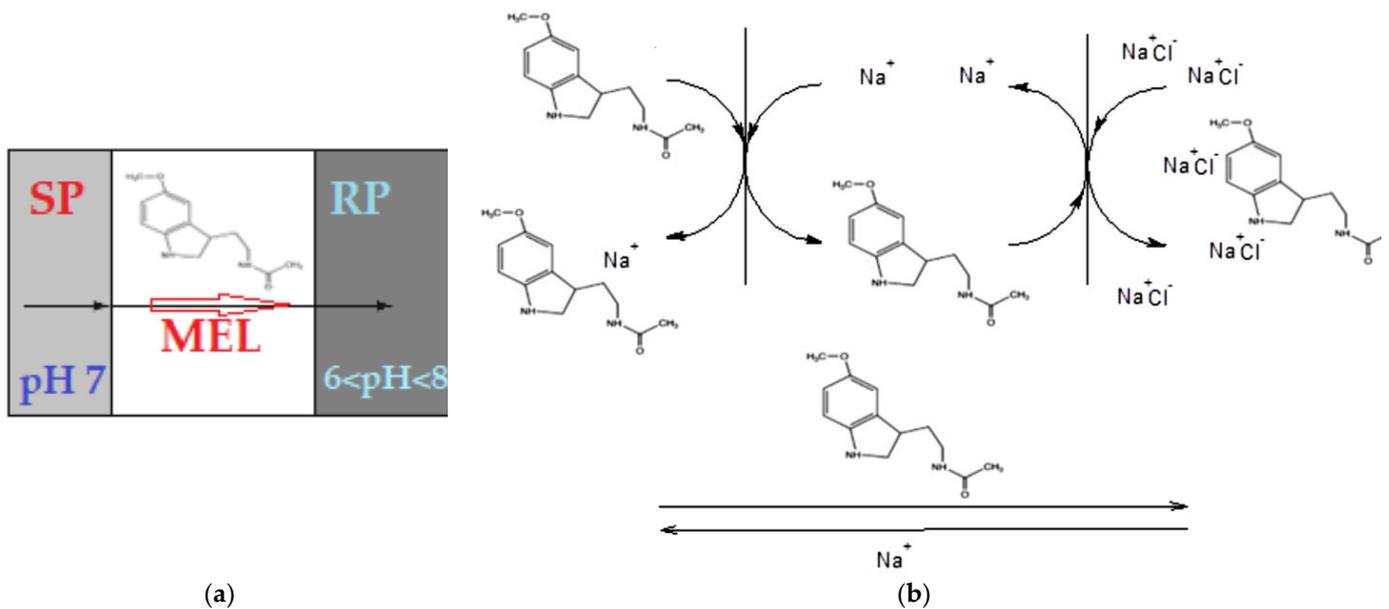
### 3.2.1. Transport of Melatonin Transport through the Obtained Membranes (M1 and M2)

The melatonin transport experiments through sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) and chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2) were carried out from a 100 mL source phase, with a concentration of 2 g/L, and imposed pH (6.0, 7.0 and 8.0) or salinity (1.0%, 3.0% and 5.0% NaCl) 100 mL receiving phases. The compartments of the membrane system were constantly stirred. The results obtained (Figure 13) show that pH influences the transport, especially in the second part of the studied interval (Figure 13a), while sodium chloride has an effect on the transport from the beginning of the range, especially at lower concentrations (Figure 13b). The pronounced increase in salinity disfavours the transport, most probably by reducing the solubility of melatonin in the receiving phase.



**Figure 13.** Time variation of melatonin flow through sulfonated ethylene-propylene-diene terpolymer (sEPDM) membranes (M1) and chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membranes (M2), depending on pH (a) and salinity (b).

The transport of melatonin in the system with the receiving phase of variable pH is mainly determined by the difference in concentration between the aqueous phases, so by the solubility of melatonin in the aqueous phases of relatively close composition (Figure 13a). On the other hand, the transport of melatonin to the receiving phase of controlled salinity corresponds to a coupled transfer mechanism (Figure 13b), in which the melatonin transport from source phase to the receiving phase is coupled with the transport of sodium ions in the opposite direction (Figure 14).

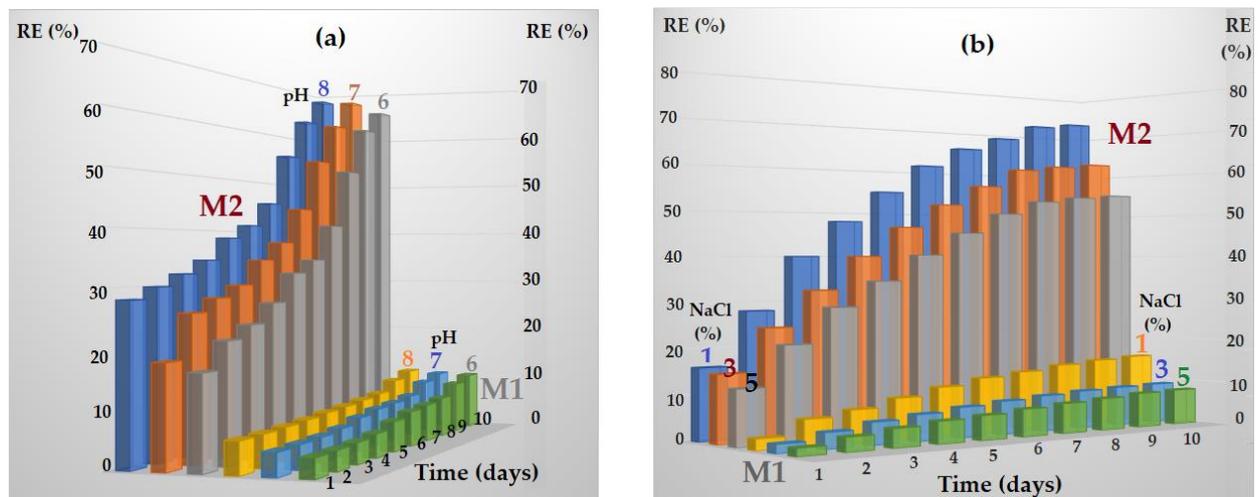


**Figure 14.** Transport schemes in the case of the receiving phase of controlled pH (a) or imposed salinity (b).

3.2.2. Release of Melatonin through the Obtained Membranes

The study of the controlled release of melatonin was carried out for a source phase of 5 mL, with a concentration of 2 g/L, and receiving phases of a much larger volume of 5 L with imposed pH (6, 7.0 and 8) or salinity (1%, 3% and 5% NaCl). Basically, a set of 100 vials containing the source phase is immersed in a vessel with 10.0 L of recirculated receiving solution, with a flow rate of 100 mL/min. Thus, it can be appreciated that the receiving phase will remain at the imposed pH and salinity. Ten of the vials from the set are taken out daily for analysis, for 10 days, seven of them for averaging the results and three to be sent for validation of the analyses.

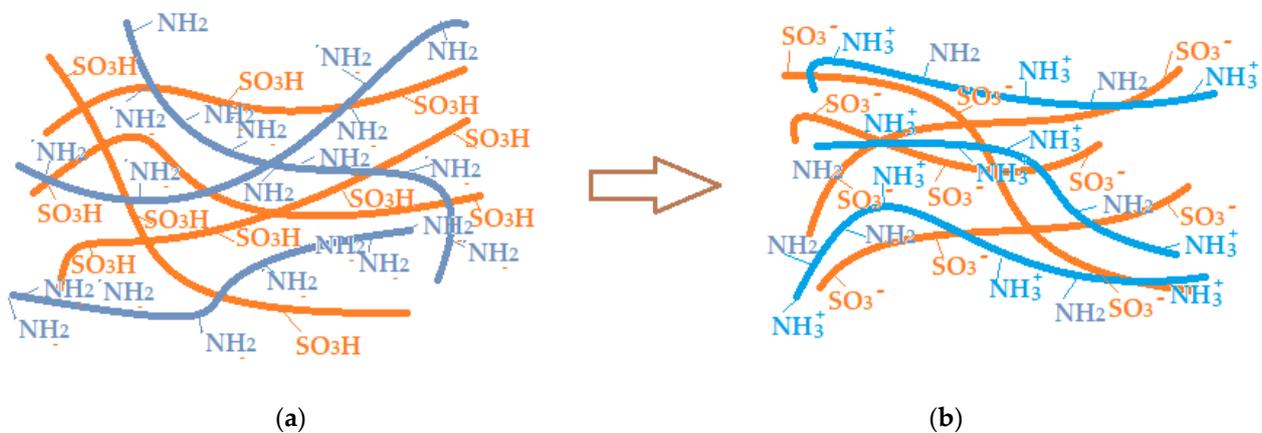
The delivery results of melatonin for ten consecutive days through sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) and chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2) are presented in Figure 15.



**Figure 15.** Time variation of melatonin cumulative release through sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) and chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2), depending on pH (a) and salinity (b).

The results obtained for the release of melatonin show that chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2) allows a faster transfer and in an amount that approaches the data from the literature, while sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) has a low but relatively constant release over time.

For the sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) that contains sulfonic reactive functional groups ( $\text{SO}_3\text{H}$ ), the interactions with melatonin during its transport and release are predictable, since at the working pH they are in sulfonate form ( $\text{SO}_3^-$ ). In the case of chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) composite membrane (M2), the interactions with melatonin in transport and release are complex because at the working pH the amino groups can be free or ionized (ammonium) (Figure 16).



**Figure 16.** Schematic representation of the polymer mixture: before (a) and after (b) the formation of the chitosan-sulfonated ethylene-propylene-diene terpolymer membrane (Chi-sEPDM) (M2).

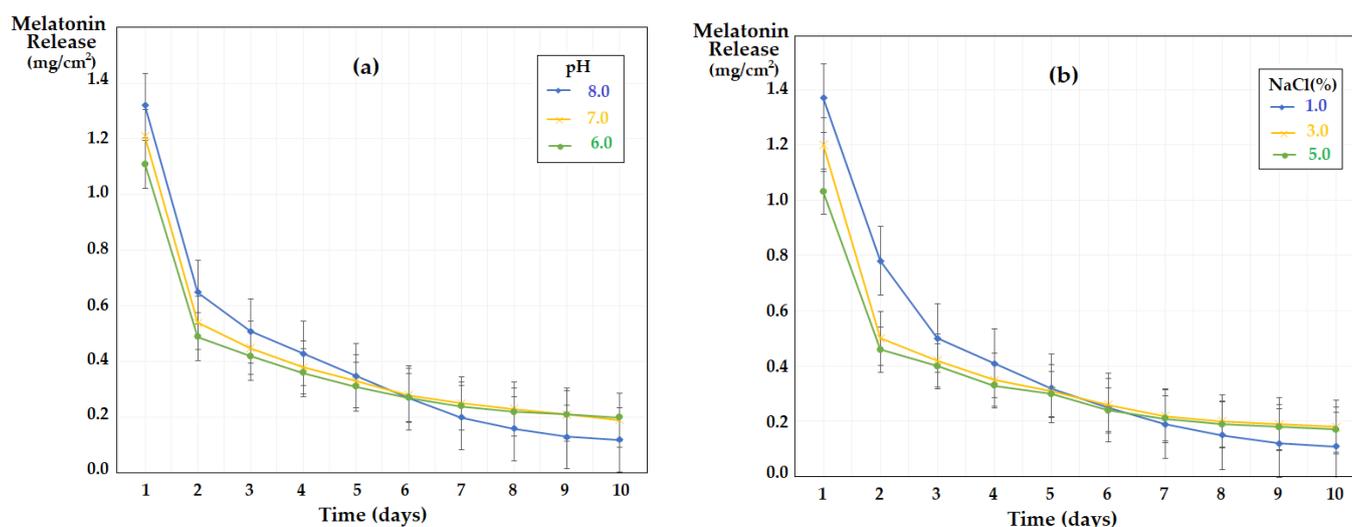
The ionic situation presented in Figure 16b is close to reality in the case of acidic pH, but at  $\text{pH} = 7$  or higher, the ammonium groups will change to the amino form, and the membrane charge will be slightly negative (due to the sulfonate groups). All these considerations explain to a good extent not only the large difference between the hydrophilicity of the prepared membranes but also the influence of pH and salinity on the transport and release of melatonin.

The study of the sensitivity to pH variation for the case of chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) composite membrane (M2) requires a greater depth compared to the experiments carried out so far, by widening the range both towards strongly acidic and towards strongly basic environments.

### 3.2.3. Melatonin Release from Melatonin-chitosan-sulfonated Ethylene-propylene-diene Terpolymer Composite Membranes (Mel-Chi-sEPDM, M3)

Melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer composite membranes (Mel-Chi-sEPDM, M3) were tested for the release of melatonin “from” their composition in synthetic solutions with pH and salinity similar to those of the previous experiments. The surface of the membranes in contact with 5.0 mL synthetic solution, from each of the 100 test bottles, is  $1 \text{ cm}^2$ , and the mass of a membrane is  $24.5 \pm 0.5 \text{ mg}$ , so the amount of melatonin in each membrane (available for release) is approx. 6.25 mg.

The results obtained (Figure 17) show that the release rate of melatonin starts with approx.  $1.25 \pm 0.15 \text{ mg/cm}^2$  per day, for the first day, and decreases constantly, reaching  $0.1 \pm 0.05 \text{ mg/cm}^2$  per day, on the tenth day. In this case as well, the increase in pH slightly favours the release of melatonin (Figure 17a), and the increase in salinity causes a slower release of this hormone (Figure 17b).



**Figure 17.** The variation of the amount of melatonin delivered daily per surface unit (cm<sup>2</sup>) of melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer composite membranes (Mel-Chi-sEPDM, M3), over a period of 10 days: (a) depending on the pH and (b) the salinity of the receiving phase.

What must be emphasized, from the perspective of the applications, is that after the fifth day, the amount of melatonin delivered drops almost five times, after which the level remains low and relatively constant.

These observations are useful for the administration of melatonin, by means of this type of composite membrane, in areas of the body with a higher pH, or in narrow places with profuse perspiration, but taking it to refresh the melatonin content of the membrane or to change the whole membrane after 5 days of application.

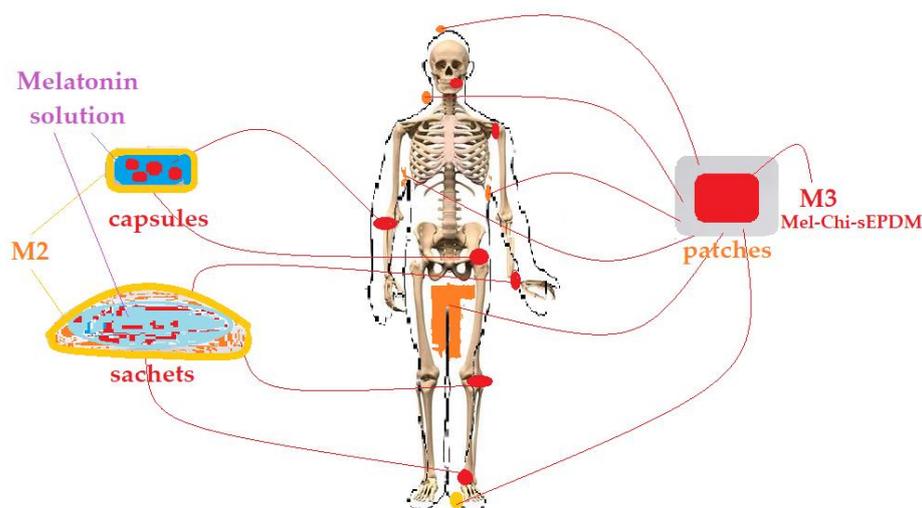
### 3.3. Aspects Regarding the Administration Method of Melatonin Using the Principle of the Studied Systems

The delivery of melatonin orally, in doses of 1 mg to 10.0 mg per day, is widespread and involves ingestion as pure powder or conditioned (formulated) powder using various compounds that ensure delayed release, in the form of cachets, capsules, or tablets [77]. Although other techniques and delivery procedures have been developed (gels, bandages, creams, patches) [78], the approach to the direction of release or delivery with membranes is of special interest, especially for cases of accidents resulting in scratches, punctures, tears, trauma, fractures, injuries caused by practicing various contact sports, the use of sports equipment, or motorized sports [79].

In the case of the present study, the development of composite membranes, for the transport and delivery of melatonin, based on an ion polymer (sEPDM) with excellent mechanical, thermal, and chemical (pH, oxidative potential, bacterial attack) properties and a biopolymer (chitosan) that is biocompatible and interacts excellently with melatonin, ensures the diversification of delivery forms of this hormone in the case of sports biomedicine applications (Figure 18).

Thus, the M2 composite membrane, which allows the transport and delivery of melatonin from a quasi-saturated source phase toward the affected tissue, can be applied in the case of sprains, dislocations, joint fractures, broken or displaced teeth, as soft bags or soft capsules that, at certain intervals, can be “refuelled” without removing them from the place where they were placed (Figure 18).

For cases of superficial injuries (skin, head, fingers, spine), the composite membrane M3 is indicated to be applied as patches or adhesive dressings, the most suitable places being the narrow areas of the body.



**Figure 18.** Possible ways to deliver melatonin using chitosan-sEPDM (M2) or melatonin-chitosan-sEPDM (M3) composite membranes.

Both types of membranes are included in the non-resorbable category, being removed after the lesions have either improved or healed.

Unlike oral administration, when melatonin comes into contact with a strongly acidic (pH 2–3, in the stomach) or basic (intestines) environment, in the case of the skin and oral cavity, the pH is close to neutral but highly dependent on the external contact environment, which involves studying transport and delivery over the entire pH scale.

#### 4. Conclusions

This paper presents the preparation and characterization of chitosan and sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) composite membrane, and melatonin-chitosan-sulfonated ethylene-propylene-diene terpolymer composite membranes (Mel-Chi-sEPDM), and its controlled release performance in synthetic aqueous solutions.

The membranes were obtained from an 8% sEPDM solution in toluene (*w/w*), in which chitosan or melatonin-chitosan (1:1, *w/w*) powders were dispersed in an ultrasonic field (sEPDM:Chi=1:1, *w/w* and Mel:Chi:sEPDM=0.5:0.5:1, *w/w/w*), through controlled vacuum evaporation. They were morphologically and structurally characterized by scanning electron microscopy (SEM), Fourier transform infrared spectroscopy (FTIR), energy-dispersive spectroscopy analysis (EDAX), thermal analysis (TG, DSC), thermal analysis coupled with infrared chromatography and analysis, and contact angle measurements, but also from the perspective of performance in the processes of transport and release of melatonin in dedicated environments (aqueous solution with controlled pH and salinity).

The transport of melatonin in the system with the receiving phase of variable pH is mainly determined by the difference in concentration between the aqueous phases, so by the solubility of melatonin in aqueous phases of relatively close composition, while the transport of melatonin to the receiving phase of controlled salinity corresponds to a coupled transfer mechanism in which the transport of melatonin from the source phase to the receptor phase is coupled with the transport of sodium ions in the opposite direction.

The results obtained for the release of melatonin show that chitosan-sulfonated ethylene-propylene-diene terpolymer (Chi-sEPDM) membrane (M2) allows a faster transfer and in a quantity that approaches the data from the literature, while sulfonated ethylene-propylene-diene terpolymer (sEPDM) membrane (M1) has a low but relatively constant release over time.

The prepared membranes can release melatonin in amounts between 0.4 mg/cm<sup>2</sup>·per day (M1), 1.6 mg/cm<sup>2</sup>·per day (M2), and 1.25 mg/cm<sup>2</sup>·per day (M3).

**Supplementary Materials:** The following are available online at <https://www.mdpi.com/article/10.3390/membranes13030282/s1>, Figure S1: Schematic presentation of the melatonin biomedical implication, Figure S2: Trace for evolving SO<sub>2</sub> (1367 cm<sup>-1</sup>) vs. temperature (a); and trace for evolving CO<sub>2</sub> (2355 cm<sup>-1</sup>) vs. temperature (b), Figure S3: Trace for evolving (a) SO<sub>2</sub> (1367 cm<sup>-1</sup>); (b) CO<sub>2</sub> (2355 cm<sup>-1</sup>); (c) hydrocarbons (2964 cm<sup>-1</sup>); and (d) H<sub>2</sub>O (3566 cm<sup>-1</sup>) vs temperature, Figure S4: Trace for evolving (a) SO<sub>2</sub> (1367 cm<sup>-1</sup>); (b) CO<sub>2</sub> (2355 cm<sup>-1</sup>); (c) hydrocarbons (2964 cm<sup>-1</sup>); and (d) H<sub>2</sub>O (3566 cm<sup>-1</sup>) vs. temperature.

**Author Contributions:** Conceptualization, F.M.P., A.A.K.K.R., O.C.O., S.G.B., A.C.N., A.R.G. and G.N.; methodology, O.C.O., A.A.K.K.R., G.N., A.R.G., F.D. and V.-A.G.; validation, G.N., S.G.B., O.C.O. and A.C.N.; formal analysis F.M.P., A.A.K.K.R., O.C.O., S.-K.T., V.-A.G., A.C.N. and G.N.; investigation, F.M.P., A.A.K.K.R., O.C.O., S.-K.T., F.D. and A.R.G.; resources, A.C.N., S.G.B. and G.N.; data curation, A.R.G. and V.-A.G.; writing—original draft preparation, F.M.P., O.C.O., G.N., A.A.K.K.R. and A.C.N.; writing—review and editing, F.M.P., V.-A.G. and G.N.; supervision, S.G.B., A.C.N. and A.R.G. All authors have read and agreed to the published version of the manuscript.

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## References

1. Minich, D.M.; Henning, M.; Darley, C.; Fahoum, M.; Schuler, C.B.; Frame, J. Is Melatonin the “Next Vitamin D”? A Review of Emerging Science, Clinical Uses, Safety, and Dietary Supplements. *Nutrients* **2022**, *14*, 3934. [[CrossRef](#)] [[PubMed](#)]
2. Tan, D.X.; Xu, B.; Zhou, X.; Reiter, R.J. Pineal Calcification, Melatonin Production, Aging, Associated Health Consequences and Rejuvenation of the Pineal Gland. *Molecules* **2018**, *23*, 301. [[CrossRef](#)]
3. Chitimus, D.M.; Popescu, M.R.; Voiculescu, S.E.; Panaitescu, A.M.; Pavel, B.; Zagrean, L.; Zagrean, A.-M. Melatonin’s Impact on Antioxidative and Anti-Inflammatory Reprogramming in Homeostasis and Disease. *Biomolecules* **2020**, *10*, 1211. [[CrossRef](#)] [[PubMed](#)]
4. Hardeland, R. Aging, Melatonin, and the Pro- and Anti-Inflammatory Networks. *Int. J. Mol. Sci.* **2019**, *20*, 1223. [[CrossRef](#)] [[PubMed](#)]
5. Fowler, S.; Hoedt, E.C.; Talley, N.J.; Keely, S.; Burns, G.L. Circadian Rhythms and Melatonin Metabolism in Patients with Disorders of Gut-Brain Interactions. *Front. Neurosci.* **2022**, *16*, 825246. [[CrossRef](#)]
6. Iguchi, H.; Kato, K.-I.; Ibayashi, H. Age-Dependent Reduction in Serum Melatonin Concentrations in Healthy Human Subjects. *J. Clin. Endocrinol. Metab.* **1982**, *55*, 27–29. [[CrossRef](#)]
7. Martínez-Águila, A.; Martín-Gil, A.; Carpena-Torres, C.; Pastrana, C.; Carracedo, G. Influence of Circadian Rhythm in the Eye: Significance of Melatonin in Glaucoma. *Biomolecules* **2021**, *11*, 340. [[CrossRef](#)]
8. Ramos, E.; Gil-Martín, E.; Ríos, C.D.L.; Egea, J.; López-Muñoz, F.; Pita, R.; Juberías, A.; Torrado, J.J.; Serrano, D.R.; Reiter, R.J.; et al. Melatonin as Modulator for Sulfur and Nitrogen Mustard-Induced Inflammation, Oxidative Stress and DNA Damage: Molecular Therapeutics. *Antioxidants* **2023**, *12*, 397. [[CrossRef](#)]
9. Beck, W.R.; Scariot, P.P.M.; Gobatto, C.A. Melatonin is an ergogenic aid for exhaustive aerobic exercise only during the wakefulness period. *Int. J. Sport. Med.* **2016**, *37*, 71–76. [[CrossRef](#)]
10. Luchetti, F.; Balduini, W.; Carloni, S.; Nasoni, M.; Reiter, R. Melatonin, tunneling nanotubes, mesenchymal cells, and tissue regeneration. *Neural Regen. Res.* **2023**, *18*, 760. [[CrossRef](#)]
11. Juliana, N.; Azmi, L.; Effendy, N.M.; Teng, N.I.M.F.; Abu, I.F.; Abu Bakar, N.N.; Azmani, S.; Abu Yazit, N.A.; Kadiman, S.; Das, S. Effect of Circadian Rhythm Disturbance on the Human Musculoskeletal System and the Importance of Nutritional Strategies. *Nutrients* **2023**, *15*, 734. [[CrossRef](#)]
12. Lee, J.G.; Woo, Y.S.; Park, S.W.; Seog, D.-H.; Seo, M.K.; Bahk, W.-M. The Neuroprotective Effects of Melatonin: Possible Role in the Pathophysiology of Neuropsychiatric Disease. *Brain Sci.* **2019**, *9*, 285. [[CrossRef](#)]
13. Arioz, B.; Taracioglu, E.; Olcum, M.; Genc, S. The Role of Melatonin on NLRP3 Inflammasome Activation in Diseases. *Antioxidants* **2021**, *10*, 1020. [[CrossRef](#)]
14. Lauritzen, E.S.; Kampmann, U.; Smedegaard, S.B.; Støy, J. Effects of daily administration of melatonin before bedtime on fasting insulin, glucose and insulin sensitivity in healthy adults and patients with metabolic diseases. A systematic review and meta-analysis. *Clin. Endocrinol.* **2021**, *95*, 691–701. [[CrossRef](#)]

15. Shafabakhsh, R.; Reiter, R.J.; Mirzaei, H.; Teymoordash, S.N.; Asemi, Z. Melatonin: A new inhibitor agent for cervical cancer treatment. *J. Cell Physiol.* **2019**, *234*, 21670–21682. [[CrossRef](#)]
16. Xie, L.-L.; Li, S.-S.; Fan, Y.-J.; Qi, M.-M.; Li, Z.-Z. Melatonin alleviates traumatic brain injury-induced anxiety-like behaviors in rats: Roles of the protein kinase A/cAMP-response element binding signaling pathway. *Exp. Ther. Med.* **2022**, *23*, 11173. [[CrossRef](#)]
17. Lissoni, P.; Barni, S.; Cattaneo, G.; Tancini, G.; Esposti, G.; Frascini, F. Clinical Results with the Pineal Hormone Melatonin in Advanced Cancer Resistant to Standard Antitumor Therapies. *Oncology* **1991**, *48*, 448–450. [[CrossRef](#)]
18. Srinivasan, V.; Spence, D.W.; Pandi-Perumal, S.R.; Trakht, I.; Cardinali, D.P. Jet lag: Therapeutic use of melatonin and possible application of melatonin analogs. *Travel Med. Infect. Dis.* **2008**, *6*, 17–28. [[CrossRef](#)]
19. Zarezadeh, M.; Khorshidi, M.; Emami, M.; Janmohammadi, P.; Kord-Varkaneh, H.; Mousavi, S.M.; Mohammed, S.H.; Saedisomeolia, A.; Alizadeh, S. Melatonin supplementation and pro-inflammatory mediators: A systematic review and meta-analysis of clinical trials. *Eur. J. Nutr.* **2019**, *59*, 1803–1813. [[CrossRef](#)]
20. Reiter, R.J.; Sharma, R.; Simko, F.; Dominguez-Rodriguez, A.; Tesarik, J.; Neel, R.L.; Slominski, A.T.; Kleszczynski, K.; Martin-Gimenez, V.M.; Manucha, W.; et al. Melatonin: Highlighting its use as a potential treatment for SARS-CoV-2 infection. *Cell Mol. Life Sci.* **2022**, *79*, 143. [[CrossRef](#)]
21. Won, E.; Na, K.-S.; Kim, Y.-K. Associations between Melatonin, Neuroinflammation, and Brain Alterations in Depression. *Int. J. Mol. Sci.* **2021**, *23*, 305. [[CrossRef](#)]
22. Schrire, Z.M.; Phillips, C.L.; Chapman, J.L.; Duffy, S.L.; Wong, G.; D’Rozario, A.L.; Comas, M.; Raisin, I.; Saini, B.; Gordon, C.J.; et al. Safety of higher doses of melatonin in adults: A systematic review and meta-analysis. *J. Pineal Res.* **2021**, *72*, 12782. [[CrossRef](#)]
23. Moroni, I.; Garcia-Bennett, A.; Chapman, J.; Grunstein, R.R.; Gordon, C.J.; Comas, M. Pharmacokinetics of exogenous melatonin in relation to formulation, and effects on sleep: A systematic review. *Sleep Med. Rev.* **2021**, *57*, 101431. [[CrossRef](#)]
24. Balaji, T.; Varadarajan, S.; Jagannathan, R.; Mahendra, J.; Fageeh, H.; Fageeh, H.; Mushtaq, S.; Baeshen, H.; Bhandi, S.; Gupta, A.; et al. Melatonin as a Topical/Systemic Formulation for the Management of Periodontitis: A Systematic Review. *Materials* **2021**, *14*, 2417. [[CrossRef](#)]
25. Atkinson, G.; Drust, B.; Reilly, T.; Waterhouse, J. The relevance of melatonin to sports medicine and science. *Sport. Med.* **2003**, *33*, 809–831. [[CrossRef](#)]
26. Gancitano, G.; Reiter, R.J. The Multiple Functions of Melatonin: Applications in the Military Setting. *Biomedicines* **2022**, *11*, 5. [[CrossRef](#)]
27. Teitz, C.C.; Hu, S.S.; Arendt, E.A. The Female Athlete: Evaluation and Treatment of Sports-Related Problems. *J. Am. Acad. Orthop. Surg.* **1997**, *5*, 87–96. [[CrossRef](#)]
28. Pramusita, A.; Nugraha, A.P.; Yuliyanasari, N.; Ardani, I.G.A.W.; Triwardhani, A. The potential capability of melatonin to anticipate postorthodontic treatment relapse: A literature review. *Biochem. Cell. Arch.* **2020**, *20*, 3061–3066.
29. Schröder, A.; Alefeld, A.; Forneck, A.; Spanier, G.; Deschner, J.; Proff, P.; Kirschnack, C. Impact of melatonin on periodontal ligament fibroblasts during mechanical strain. *Eur. J. Orthod.* **2022**, *44*, 659–668. [[CrossRef](#)]
30. Souissi, A.; Dergaa, I. An Overview of the Potential Effects of Melatonin Supplementation on Athletic Performance. *Int. J. Sport Stud. Health* **2022**, *4*, 121714. [[CrossRef](#)]
31. Chuffa, L.; Seiva, F.; Novais, A.; Simão, V.; Giménez, V.M.; Manucha, W.; Zuccari, D.; Reiter, R. Melatonin-Loaded Nanocarriers: New Horizons for Therapeutic Applications. *Molecules* **2021**, *26*, 3562. [[CrossRef](#)]
32. Mirza-Aghazadeh-Attari, M.; Mihanfar, A.; Yousefi, B.; Majidinia, M. Nanotechnology-based advances in the efficient delivery of melatonin. *Cancer Cell Int.* **2022**, *22*, 43. [[CrossRef](#)] [[PubMed](#)]
33. Lee, B.-J.; Parrott, K.A.; Ayres, J.W.; Sack, R.L. Design and evaluation of an oral controlled release delivery system for melatonin in human subjects. *Int. J. Pharm.* **1995**, *124*, 119–127. [[CrossRef](#)]
34. Flo, A.; Calpena, A.C.; Halbaut, L.; Araya, E.I.; Fernández, F.; Clares, B. Melatonin Delivery: Transdermal and Transbuccal Evaluation in Different Vehicles. *Pharm. Res.* **2016**, *33*, 1615–1627. [[CrossRef](#)] [[PubMed](#)]
35. Duttagupta, D.S.; Jadhav, M.V.; Kadam, J.V. Chitosan: A propitious biopolymer for drug delivery. *Curr. Drug Deliv.* **2015**, *12*, 369–381. [[CrossRef](#)]
36. Hafner, A.; Lovrić, J.; Pepić, I.; Filipović-Grčić, J. Lecithin/chitosan nanoparticles for transdermal delivery of melatonin. *J. Microencapsul.* **2011**, *28*, 807–815. [[CrossRef](#)]
37. Hidayati, N.; Harmoko, T.; Mujiburohman, M.; Purnama, H. Characterization of sPEEK/chitosan membrane for the direct methanol fuel cell. In Proceedings of the AIP Conference Proceedings, Leuven, Belgium, 8–10 April 2019; AIP Publishing LLC.: Melville, NY, USA, 2019; Volume 2114, p. 060008. [[CrossRef](#)]
38. Nalika, N.; Waseem, M.; Kaushik, P.; Salman, M.; Andrabi, S.S.; Parvez, S. Role of melatonin and quercetin as countermeasures to the mitochondrial dysfunction induced by titanium dioxide nanoparticles. *Life Sci.* **2023**, 121403. [[CrossRef](#)] [[PubMed](#)]
39. Alahri, M.B.; Ibrahim, A.J.; Barani, M.; Arkaban, H.; Shadman, S.M.; Salarpour, S.; Zarrintaj, P.; Jaberi, J.; Jalil, A.T. Management of Brain Cancer and Neurodegenerative Disorders with Polymer-Based Nanoparticles as a Biocompatible Platform. *Molecules* **2023**, *28*, 841. [[CrossRef](#)]
40. Cimbru, A.M.; Rikabi, A.A.K.K.; Oprea, O.; Grosu, A.R.; Tanczos, S.-K.; Simonescu, M.C.; Pașcu, D.; Grosu, V.-A.; Dumitru, F.; Nechifor, G. *pH* and *pCl* Operational Parameters in Some Metallic Ions Separation with Composite Chitosan/Sulfonated Polyether Ether Ketone/Polypropylene Hollow Fibers Membranes. *Membranes* **2022**, *12*, 833. [[CrossRef](#)]

41. Nechifor, A.; Goran, A.; Grosu, V.-A.; Bungău, C.; Albu, P.; Grosu, A.; Oprea, O.; Păncescu, F.; Nechifor, G. Improving the Performance of Composite Hollow Fiber Membranes with Magnetic Field Generated Convection Application on pH Correction. *Membranes* **2021**, *11*, 445. [[CrossRef](#)]
42. Urducea, C.B.; Nechifor, A.C.; Dimulescu, I.A.; Oprea, O.; Nechifor, G.; Totu, E.E.; Isildak, I.; Albu, P.C.; Bungău, S.G. Control of Nanostructured Polysulfone Membrane Preparation by Phase Inversion Method. *Nanomaterials* **2020**, *10*, 2349. [[CrossRef](#)] [[PubMed](#)]
43. Nechifor, A.; Panait, V.; Naftanaila, L.; Batalu, D.; Voicu, S.I. Symmetrically polysulfone membranes obtained by solvent evaporation using carbon nanotubes as additives. Synthesis, characterization and applications. *Dig. J. Nanomater. Biostruc.* **2013**, *8*, 875–884.
44. Zaharia, I.; Aboul-Enein, H.Y.; Diaconu, I.; Ruse, E.; Bunaciu, A.A.; Nechifor, G. Facilitated transport of 5-aminosalicylic acid through bulk liquid membrane. *J. Iran. Chem. Soc.* **2013**, *10*, 1129–1136. [[CrossRef](#)]
45. AMCTB No. 96; Analytical Methods Committee. What's novel in the new Eurachem guide on uncertainty from sampling? *Anal. Methods* **2020**, *12*, 2295–2297. [[CrossRef](#)] [[PubMed](#)]
46. Miccoli, A.; Restani, P.; Floroian, L.; Taus, N.; Badea, M.; Cioca, G.; Bungau, S. Sensitive Electrochemical Detection Method of Melatonin in Food Supplements. *Rev. Chim.* **2018**, *69*, 854–859. [[CrossRef](#)]
47. Nechifor, G.; Grosu, A.R.; Dinu, A.F.; Tanczos, S.-K.; Goran, A.; Grosu, V.-A.; Bungău, S.G.; Păncescu, F.M.; Albu, P.C.; Nechifor, A.C. Simultaneous Release of Silver Ions and 10-Undecenoic Acid from Silver Iron–Oxide Nanoparticles Impregnated Membranes. *Membranes* **2022**, *12*, 557. [[CrossRef](#)]
48. Sorouraddin, M.-H.; Rashidi, M.-R.; Ghorbani-Kalhor, E.; Asadpour-Zeynali, K. Simultaneous spectrofluorimetric and spectrophotometric determination of melatonin and pyridoxine in pharmaceutical preparations by multivariate calibration methods. *Il Farm.* **2005**, *60*, 451–458. [[CrossRef](#)]
49. Szczepański, P.; Diaconu, I. Transport of *p*-Nitrophenol through an Agitated Bulk Liquid Membrane. *Sep. Sci. Technol.* **2012**, *47*, 1725–1732. [[CrossRef](#)]
50. Szczepański, P.; Tanczos, S.K.; Ghindeanu, L.D.; Wódzki, R. Transport of *p*-nitrophenol in an agitated bulk liquid membrane system—Experimental and theoretical study by network analysis. *Sep. Purif. Technol.* **2014**, *132*, 616–626. [[CrossRef](#)]
51. Diaconu, I.; Nechifor, G.; Nechifor, A.C.; Totu, E.E.; Ruse, E. The transport of nitrophenols through liquid membranes. *Rev. Chim.* **2009**, *60*, 1243–1246.
52. Nechifor, A.C.; Pîrțac, A.; Albu, P.C.; Grosu, A.R.; Dumitru, F.; Dimulescu (Nica), I.A.; Oprea, O.; Pașcu, D.; Nechifor, G.; Bungău, S.G. Recuperative Amino Acids Separation through Cellulose Derivative Membranes with Microporous Polypropylene Fiber Matrix. *Membranes* **2021**, *11*, 429. [[CrossRef](#)] [[PubMed](#)]
53. Motelica, L.; Ficaș, D.; Oprea, O.-C.; Ficaș, A.; Ene, V.-L.; Vasile, B.-S.; Andronescu, E.; Holban, A.-M. Antibacterial Biodegradable Films Based on Alginate with Silver Nanoparticles and Lemongrass Essential Oil—Innovative Packaging for Cheese. *Nanomaterials* **2021**, *11*, 2377. [[CrossRef](#)] [[PubMed](#)]
54. Bartlam, C.; Morsch, S.; Heard, K.W.; Quayle, P.; Yeates, S.G.; Vijayaraghavan, A. Nanoscale infrared identification and mapping of chemical functional groups on graphene. *Carbon* **2018**, *139*, 317–324. [[CrossRef](#)]
55. Dimulescu (Nica), I.; Nechifor, A.; Urducea, C.B.; Oprea, O.; Pașcu, D.; Totu, E.; Albu, P.; Nechifor, G.; Bungău, S. Accessible Silver-Iron Oxide Nanoparticles as a Nanomaterial for Supported Liquid Membranes. *Nanomaterials* **2021**, *11*, 1204. [[CrossRef](#)]
56. Nechifor, G.; Totu, E.E.; Nechifor, A.C.; Isildak, I.; Oprea, O.; Cristache, C.M. Non-Resorbable Nanocomposite Membranes for Guided Bone Regeneration Based on Polysulfone-Quartz Fiber Grafted with Nano-TiO<sub>2</sub>. *Nanomaterials* **2019**, *9*, 985. [[CrossRef](#)]
57. Nechifor, A.C.; Goran, A.; Tanczos, S.-K.; Păncescu, F.M.; Oprea, O.-C.; Grosu, A.R.; Matei, C.; Grosu, V.-A.; Vasile, B.; Albu, P.C. Obtaining and Characterizing the Osmium Nanoparticles/*n*-Decanol Bulk Membrane Used for the *p*-Nitrophenol Reduction and Separation System. *Membranes* **2022**, *12*, 1024. [[CrossRef](#)]
58. Florea-Spiroiu, M.; Olteanu, M.; Stanescu, V.; Nechifor, G. Surface tension components of plasma treated polysulphone membranes. *An. Univ. București–Chim. Anul XVII* **2008**, *2*, 13–18.
59. Tirla, A.; Islam, F.; Islam, R.; Vicas, S.I.; Cavalu, S. New Insight and Future Perspectives on Nutraceuticals for Improving Sports Performance of Combat Players: Focus on Natural Supplements, Importance and Advantages over Synthetic Ones. *Appl. Sci.* **2022**, *12*, 8611. [[CrossRef](#)]
60. Nikolaev, G.; Robeva, R.; Konakchieva, R. Membrane Melatonin Receptors Activated Cell Signaling in Physiology and Disease. *Int. J. Mol. Sci.* **2021**, *23*, 471. [[CrossRef](#)]
61. Farjallah, M.; Graja, A.; Mahmoud, L.; Ghattassi, K.; Boudaya, M.; Driss, T.; Jamoussi, K.; Sahnoun, Z.; Souissi, N.; Hammouda, O. Effects of melatonin ingestion on physical performance and biochemical responses following exhaustive running exercise in soccer players. *Biol. Sport* **2022**, *39*, 473–479. [[CrossRef](#)]
62. Kalra, S.; Banderwal, R.; Arora, K.; Kumar, S.; Singh, G.; Chawla, P.A.; Behl, T.; Sehgal, A.; Singh, S.; Bhatia, S.; et al. An update on pathophysiology and treatment of sports-mediated brain injury. *Environ. Sci. Pollut. Res.* **2022**, *29*, 16786–16798. [[CrossRef](#)]
63. Gonçalves, A.L.; Ferreira, A.M.; Ribeiro, R.T.; Zukerman, E.; Cipolla-Neto, J.; Peres, M.F.P. Randomised clinical trial comparing melatonin 3 mg, amitriptyline 25 mg and placebo for migraine prevention. *J. Neurol. Neurosurg. Psychiatry* **2016**, *87*, 1127–1132. [[CrossRef](#)] [[PubMed](#)]
64. Reid, K.; Heuvel, C.J.V.D.; Dawson, D. Day-time melatonin administration: Effects on core temperature and sleep onset latency. *J. Sleep Res.* **1996**, *5*, 150–154. [[CrossRef](#)]

65. Rusanova, I.; Martínez-Ruiz, L.; Florido, J.; Rodríguez-Santana, C.; Guerra-Librero, A.; Acuña-Castroviejo, D.; Escames, G. Protective Effects of Melatonin on the Skin: Future Perspectives. *Int. J. Mol. Sci.* **2019**, *20*, 4948. [[CrossRef](#)] [[PubMed](#)]
66. Marón, F.J.M.; Ferder, L.; Reiter, R.J.; Manucha, W. Daily and seasonal mitochondrial protection: Unraveling common possible mechanisms involving vitamin D and melatonin. *J. Steroid Biochem. Mol. Biol.* **2020**, *199*, 105595. [[CrossRef](#)]
67. Favero, G.; Franceschetti, L.; Bonomini, F.; Rodella, L.F.; Rezzani, R. Melatonin as an Anti-Inflammatory Agent Modulating Inflammasome Activation. *Int. J. Endocrinol.* **2017**, *2017*, 1835195. [[CrossRef](#)]
68. D'Angelo, G.; Chimenz, R.; Reiter, R.J.; Gitto, E. Use of Melatonin in Oxidative Stress Related Neonatal Diseases. *Antioxidants* **2020**, *9*, 477. [[CrossRef](#)]
69. Walrand, S.; Gaulmin, R.; Aubin, R.; Sapin, V.; Coste, A.; Abbot, M. Nutritional factors in sport-related concussion. *Neurochirurgie* **2021**, *67*, 255–258. [[CrossRef](#)]
70. Miricioiu, M.; Niculescu, V.-C.; Filote, C.; Raboaca, M.; Nechifor, G. Coal Fly Ash Derived Silica Nanomaterial for MMMs—Application in CO<sub>2</sub>/CH<sub>4</sub> Separation. *Membranes* **2021**, *11*, 78. [[CrossRef](#)]
71. Ochoa, J.J.; Díaz-Castro, J.; Kajarabille, N.; García, C.; Guisado, I.M.; De Teresa, C.; Guisado, R. Melatonin supplementation ameliorates oxidative stress and inflammatory signaling induced by strenuous exercise in adult human males. *J. Pineal Res.* **2011**, *51*, 373–380. [[CrossRef](#)]
72. Gitto, E.; Tan, D.-X.; Reiter, R.J.; Karbownik-Lewinska, M.; Manchester, L.C.; Cuzzocrea, S.; Fulia, F.; Barberi, I. Individual and synergistic antioxidative actions of melatonin: Studies with vitamin E, vitamin C, glutathione and desferrioxamine (desferoxamine) in rat liver homogenates. *J. Pharm. Pharmacol.* **2001**, *53*, 1393–1401. [[CrossRef](#)] [[PubMed](#)]
73. Zhao, Y.; Shao, G.; Liu, X.; Li, Z. Assessment of the Therapeutic Potential of Melatonin for the Treatment of Osteoporosis through a Narrative Review of Its Signaling and Preclinical and Clinical Studies. *Front. Pharmacol.* **2022**, *13*, 866625. [[CrossRef](#)] [[PubMed](#)]
74. Stacchiotti, A.; Favero, G.; Rodella, L.F. Impact of Melatonin on Skeletal Muscle and Exercise. *Cells* **2020**, *9*, 288. [[CrossRef](#)] [[PubMed](#)]
75. Bantounou, M.; Plasevic, J.; Galley, H.F. Melatonin and Related Compounds: Antioxidant and Anti-Inflammatory Actions. *Antioxidants* **2022**, *11*, 532. [[CrossRef](#)]
76. Miricioiu, C.; Voicu, V.; Anuta, V.; Tudose, A.; Celia, C.; Paolino, D.; Fresta, M.; Sandulovici, R.; Miricioiu, I. Mathematical Modeling of Release Kinetics from Supramolecular Drug Delivery Systems. *Pharmaceutics* **2019**, *11*, 140. [[CrossRef](#)]
77. Jafari, H.; Hassanpour, M.; Akbari, A.; Rezaie, J.; Gohari, G.; Mahdavinia, G.R.; Jabbari, E. Characterization of pH-sensitive chitosan/hydroxypropyl methylcellulose composite nanoparticles for delivery of melatonin in cancer therapy. *Mater. Lett.* **2020**, *282*, 128818. [[CrossRef](#)]
78. Blažević, F.; Milekić, T.; Romić, M.D.; Juretić, M.; Pepić, I.; Filipović-Grčić, J.; Lovrić, J.; Hafner, A. Nanoparticle-mediated interplay of chitosan and melatonin for improved wound epithelialisation. *Carbohydr. Polym.* **2016**, *146*, 445–454. [[CrossRef](#)]
79. Musazzi, U.M.; Dolci, L.S.; Albertini, B.; Passerini, N.; Cilurzo, F. A new melatonin oral delivery platform based on orodispersible films containing solid lipid microparticles. *Int. J. Pharm.* **2019**, *559*, 280–288. [[CrossRef](#)]

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