

Supporting Information

Table S1. Cultural characteristics of strain NEAU-H2^T, *S. rhizosphaerihabitans* NBRC 109807^T, *S. populi* A249^T and *S. siamensis* NBRC 108799^T. Abbreviations: BA, Bennett's agar; CA, Czapek's agar; NA, Nutrient agar.

Characteristic	NEAU-H2 ^T	NBRC 109807 ^T	A249 ^T	NBRC 108799 ^T . 111515532 15532 ^T
Growth on ISP1				
Aerial mycelium	White	Pinkish White	Pinkish White	White
Substrate mycelium	White	Pinkish White	Pinkish White	Grayish Greenish Yellow
Growth on ISP2				
Aerial mycelium	Moderate Yellow	Pale Violet	White	White
Substrate mycelium	Moderate Yellow	Pale Violet	White	Medium Gray
Growth on ISP3				
Aerial mycelium	White	Pale Blue	Pale Blue	White
Substrate mycelium	Light Yellow	Pale Blue	Pale Blue	Light Bluish Gray
Growth on ISP4				
Aerial mycelium	Light Yellow	White	White	White
Substrate mycelium	Light Yellow GreenGreenGreen	Pale Yellow	White	White
Growth on ISP5				
Aerial mycelium	Pale Yellow	White	Pale Green	Strong Greenish Yellow
Substrate mycelium	Pale Yellow	White	Pale Green	Strong Greenish Yellow
Growth on ISP6				
Aerial mycelium	Light Olive Gray	Very Pale Blue	Very Pale Blue	White
Substrate mycelium	Light Olive Gray	Very Pale Blue	Very Pale Blue	White
Diffusible pigment	Dark Greenish Olive	None	None	None
Growth on ISP7				
Aerial mycelium	Yellow White	White	White	White
Substrate mycelium	Yellow White	White	White	White
Growth on NA				
Aerial mycelium	Greenish White	Light Greenish Gray	Bluish White	White
Substrate mycelium	Greenish White	Light Greenish Gray	Bluish White	White
Growth on CA				
Aerial mycelium	White	White	White	White
Substrate mycelium	White	White	White	White
Growth on MBA				
Aerial mycelium colour	Pale Greenish Yellow	White	Pale Greenish Yellow	Pale Yellow
Substrate mycelium colour	Pale Greenish Yellow	Pale Greenish Yellow	Pale Greenish Yellow	Pale Yellow

Table S3. Genome sequence features of strain NEAU-H2^T and *S. populi* A249^T.

Features	NEAU-H2 ^T	<i>S. populi</i> A249 ^T
Bioproject	PRJNA574783	PRJNA421064
Accession No.	WBKG00000000	PJOS00000000
Sequencing Technology	Illumina HiSeq	Illumina HiSeq
Assembly method	SOAP denovo v. 2.04	SOAP denovo v. 2.04
Genome coverage	152.0x	310.0x
N50	167,996	75,083
Contigs	135	279
Genome size (bp)	9,921,301	9,587,301
DNA GC content (%)	71.5	71.7
Number of genes	8,929	8,603
Protein coding genes	8,287	7,911
rRNAs	12	7
tRNAs	73	70
ncRNAs	3	3

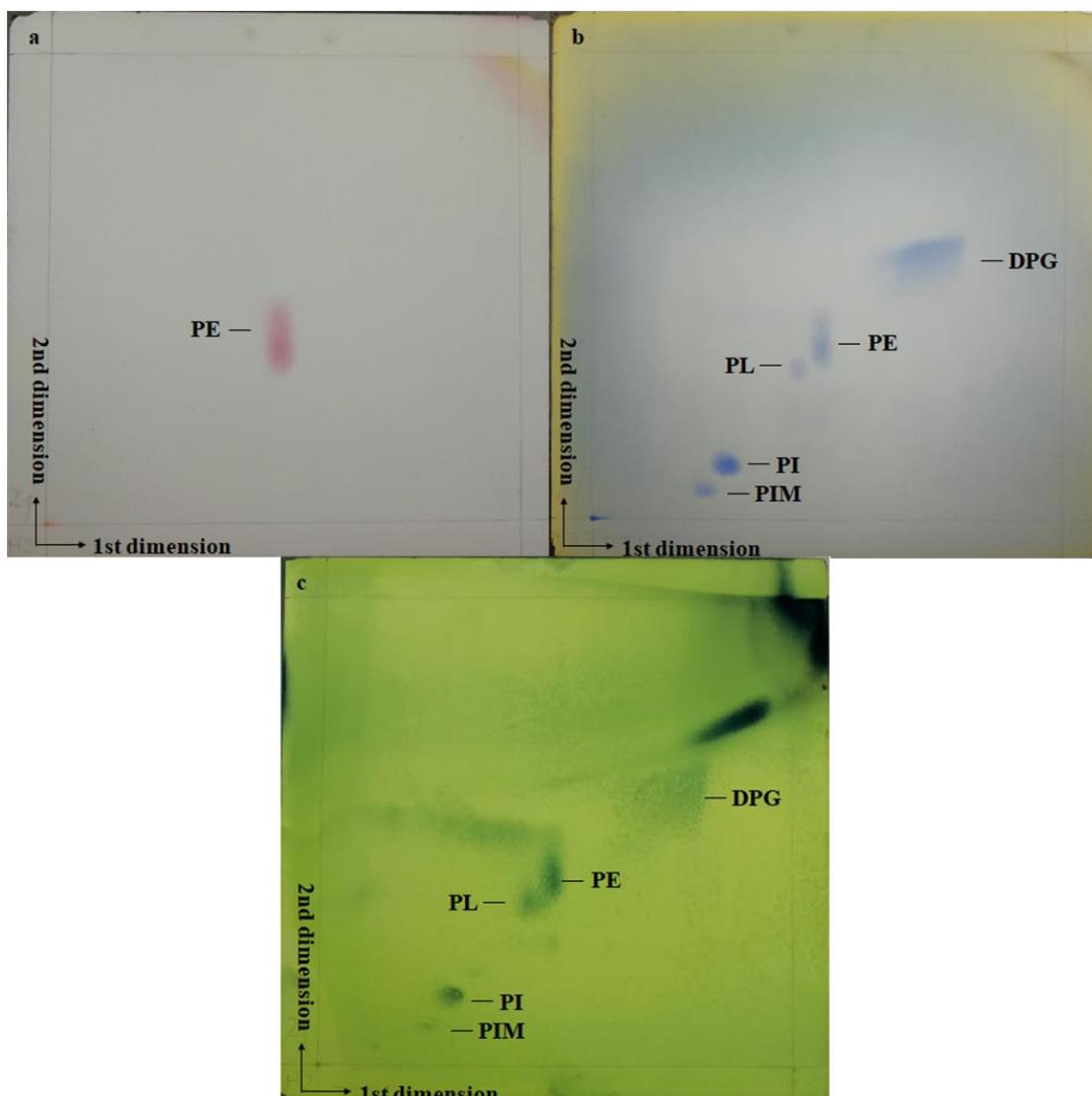


Figure S1. The phospholipids of strain NEAU-H2^T. **a**, Using ninhydrin reagent; **b**, Using molybdenum blue reagent; **c**, Using molybdophosphoric acid reagent. 1st dimension:Chloroform:Methanol:Water (65:25:4, v/v); 2nd dimension: Chloroform:Acetic acid:Methanol:Water (80:18:12:5, v/v). Abbreviations: DPG, diphosphatidylglycerol; PE,

phosphatidylethanolamine; PI, phosphatidylinositol; PIM, phosphatidylinositol mannoside; PL, unidentified phospholipid.

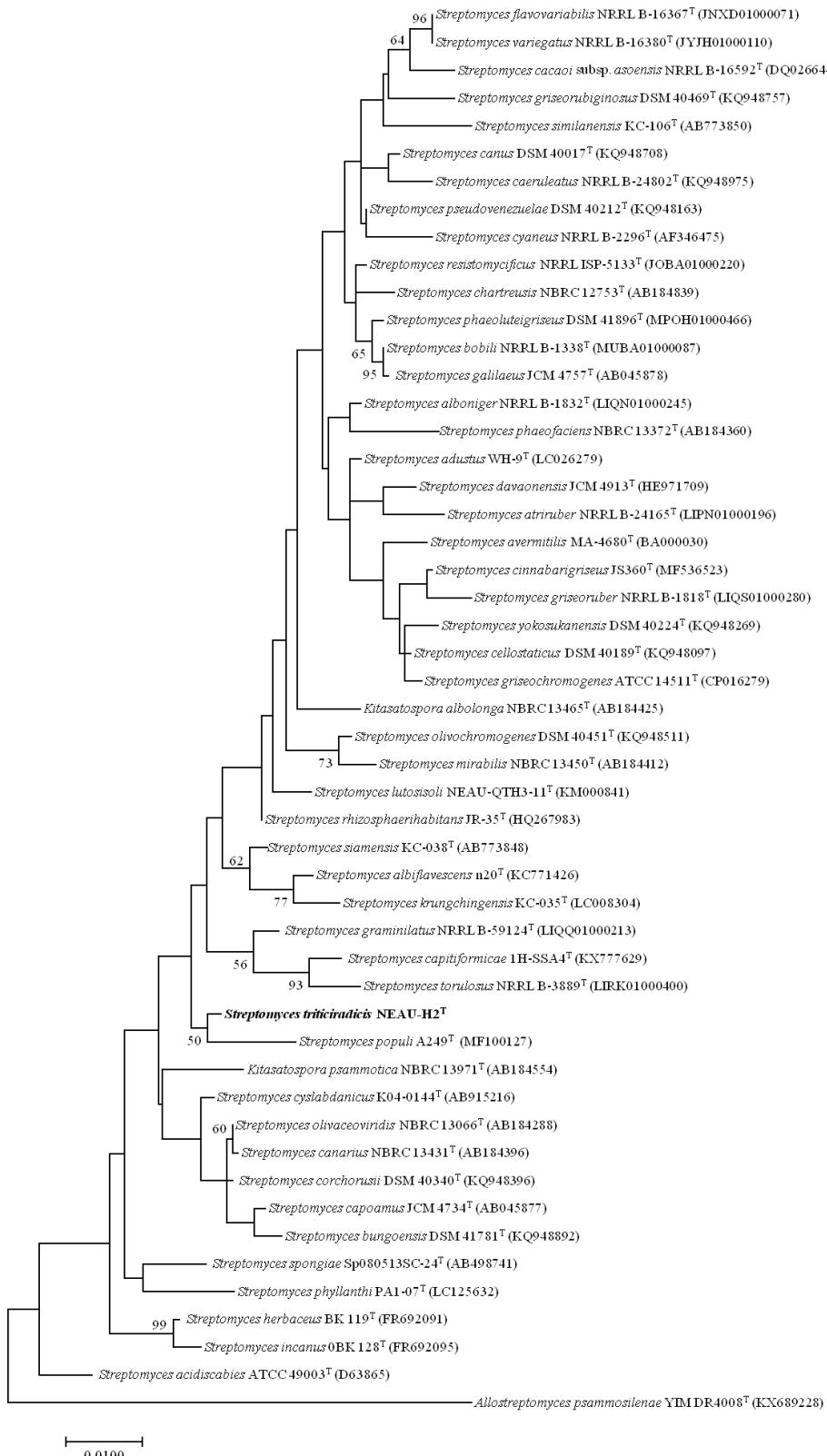


Figure S2. Maximum-likelihood tree based on 16S rRNA gene sequences (1418 bp) showing relationships of NEAU-H2^T (in bold) with related taxa which are the top 50 type strains of *Streptomyces* species of gene sequence similarities based on analysis using EzTaxon-e. Only bootstrap

values above 50% (percentages of 1000 replications) are indicated. *Allostreptomyces psammosilena* YIM DR4008^T (KX689228) was used as an outgroup. Bar, 0.01 nucleotide substitutions per site.

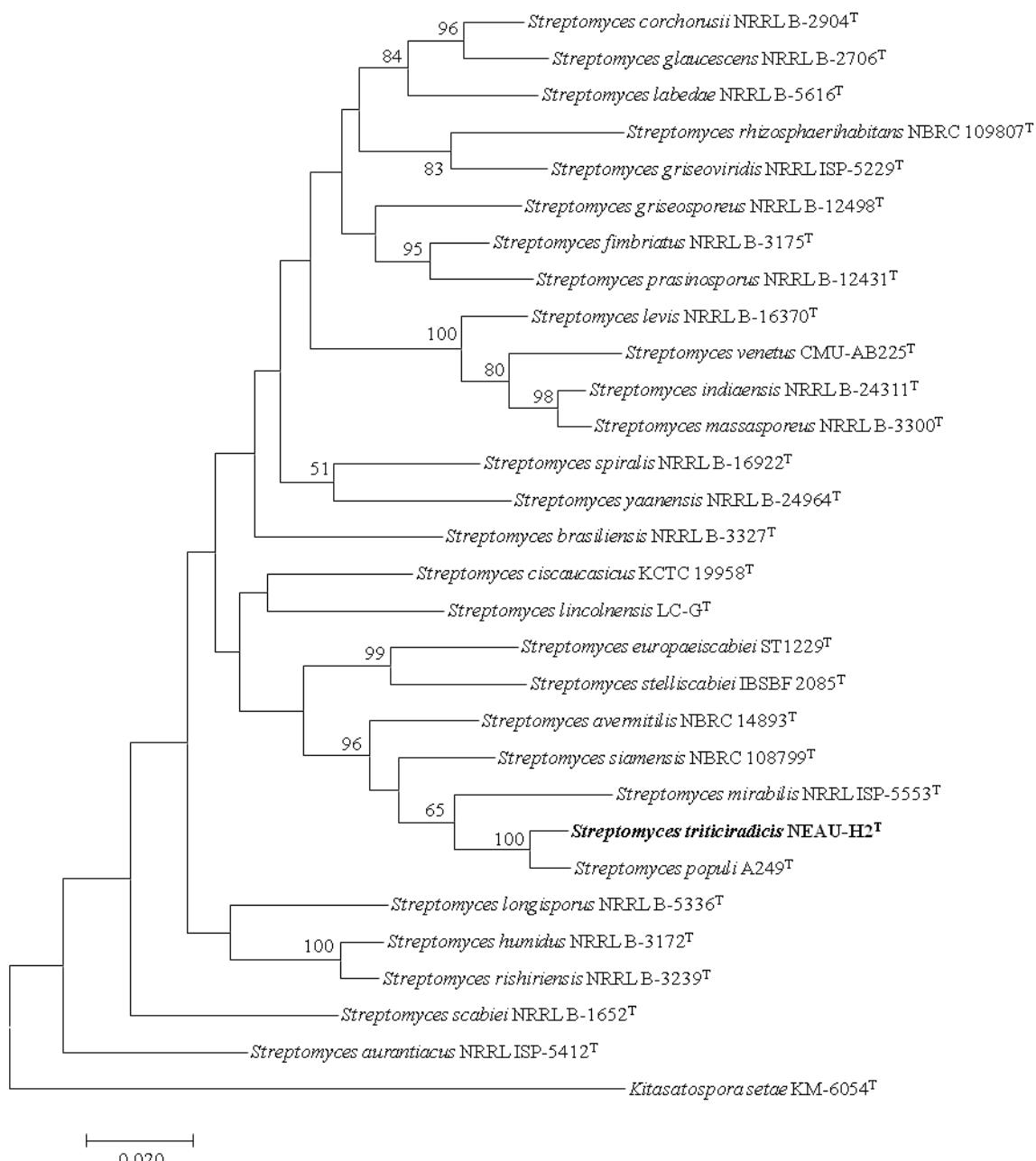


Figure 3. Maximum-likelihood tree based on MLSA analysis of the concatenated partial sequences (2060 bp) from five housekeeping genes (*atpD*, *gyrB*, *recA*, *rpoB* and *trpB*) of strain NEAU-H2^T (in bold) with related taxa. Only bootstrap values above 50% (percentages of 1000 replications) are indicated. *Kitasatospora setae* KM-6054^T was used as an outgroup. Bar, 0.02 nucleotide substitutions per site.

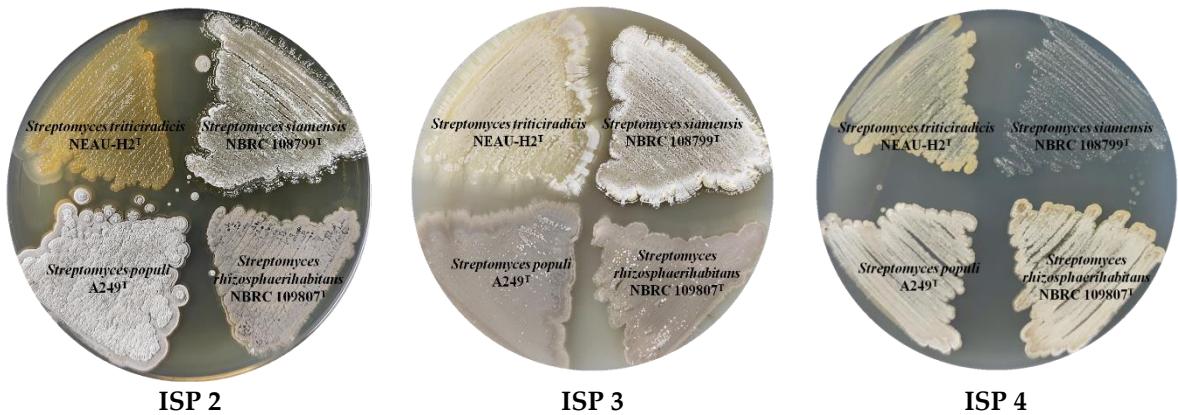
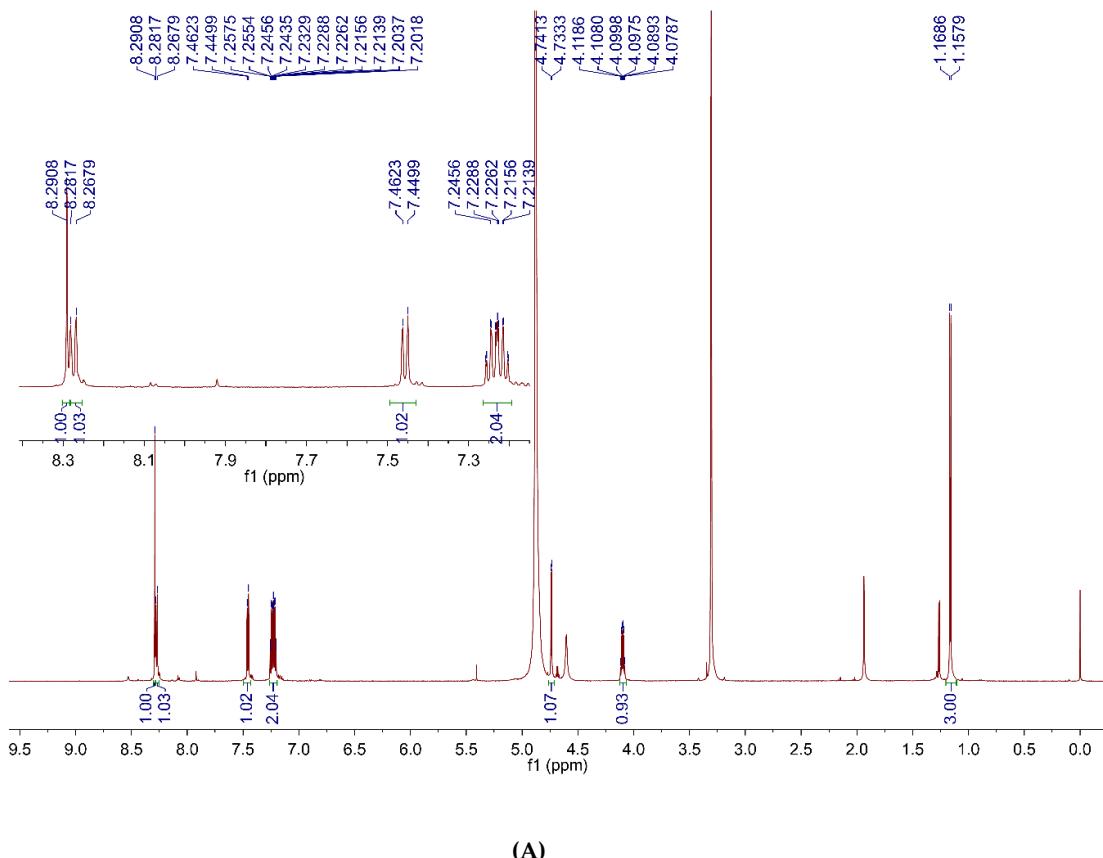
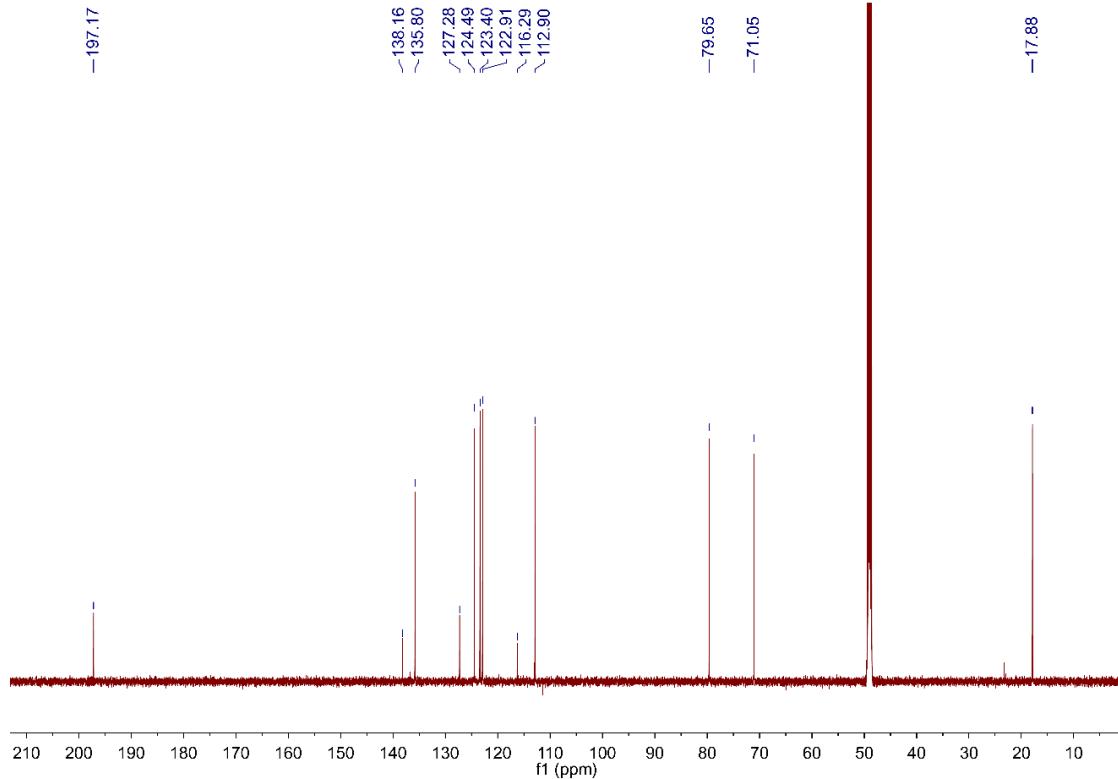
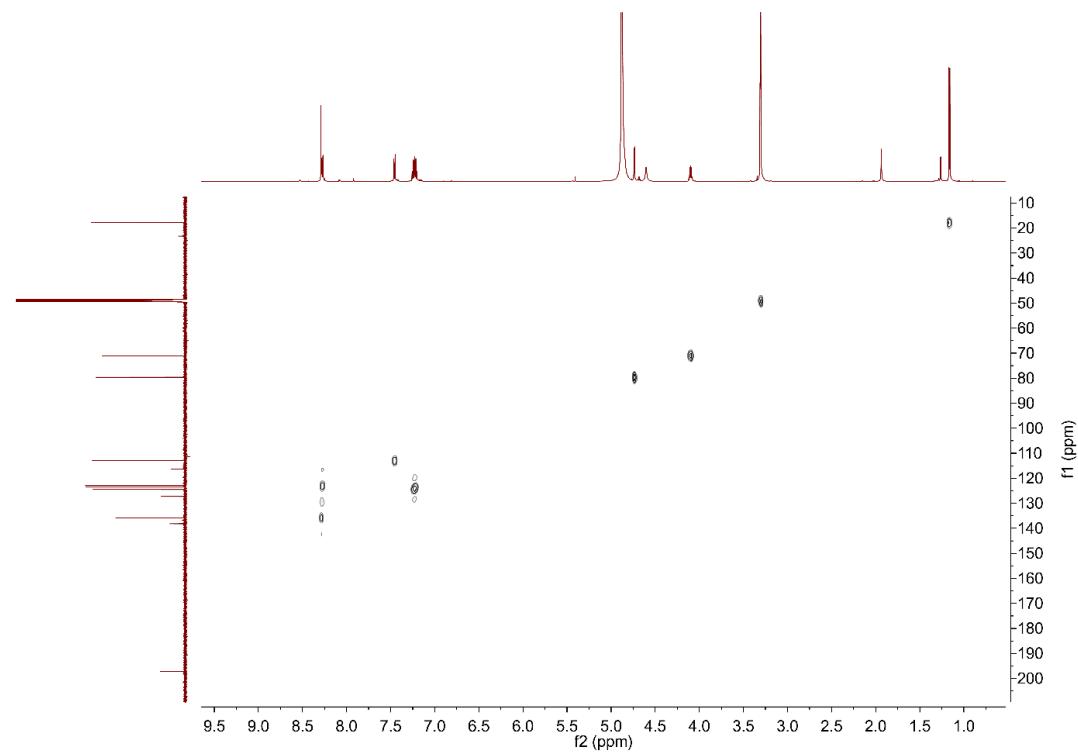


Figure S4. Cultural characteristics of strain NEAU-H2^T, *Streptomyces rhizosphaerihabitans* NBRC 109807^T, *Streptomyces populi* A249^T and *Streptomyces siamensis* NBRC 108799^T observed on ISP 2, ISP 3 and ISP 4 media at 28 °C for 2 weeks.

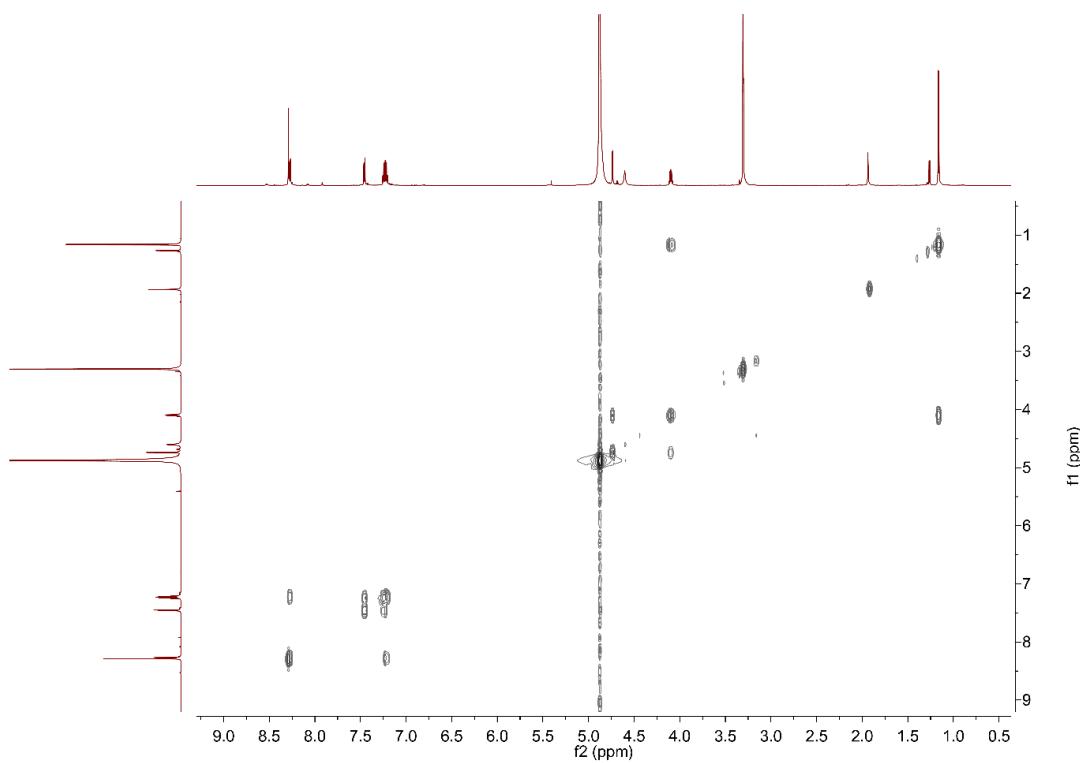




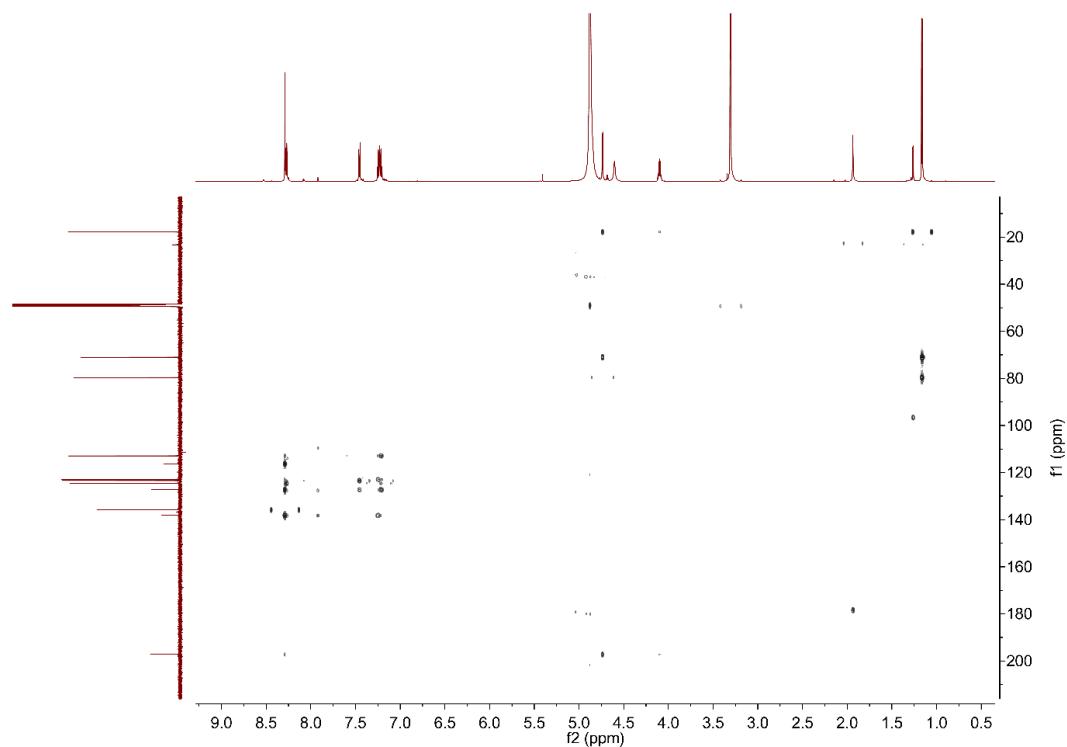
(B)



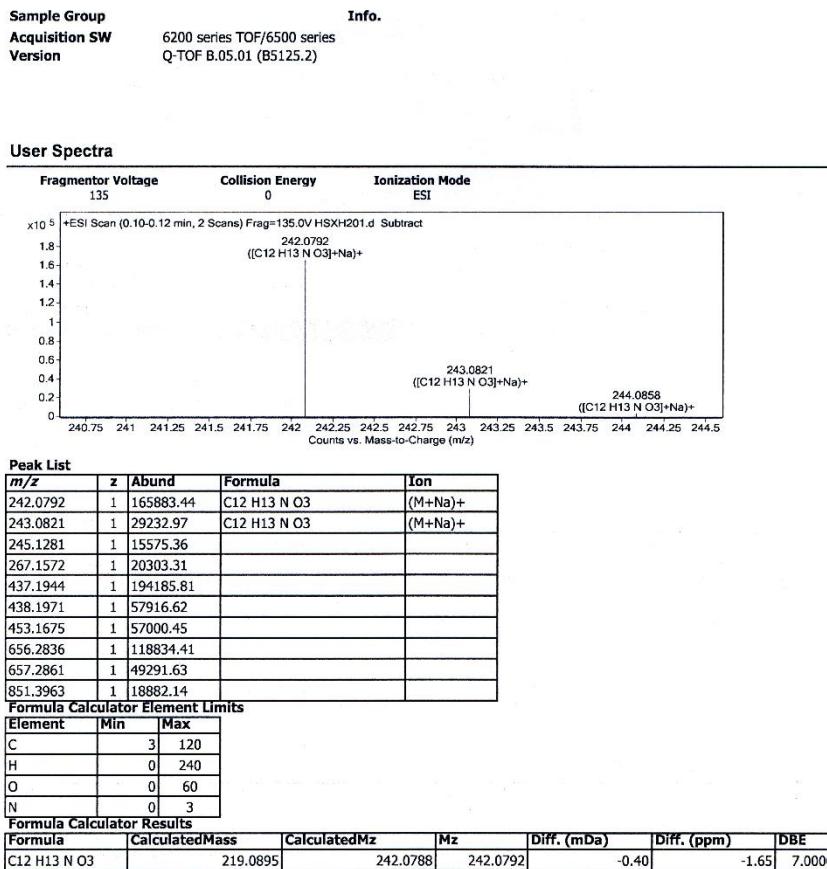
(C)



(D)

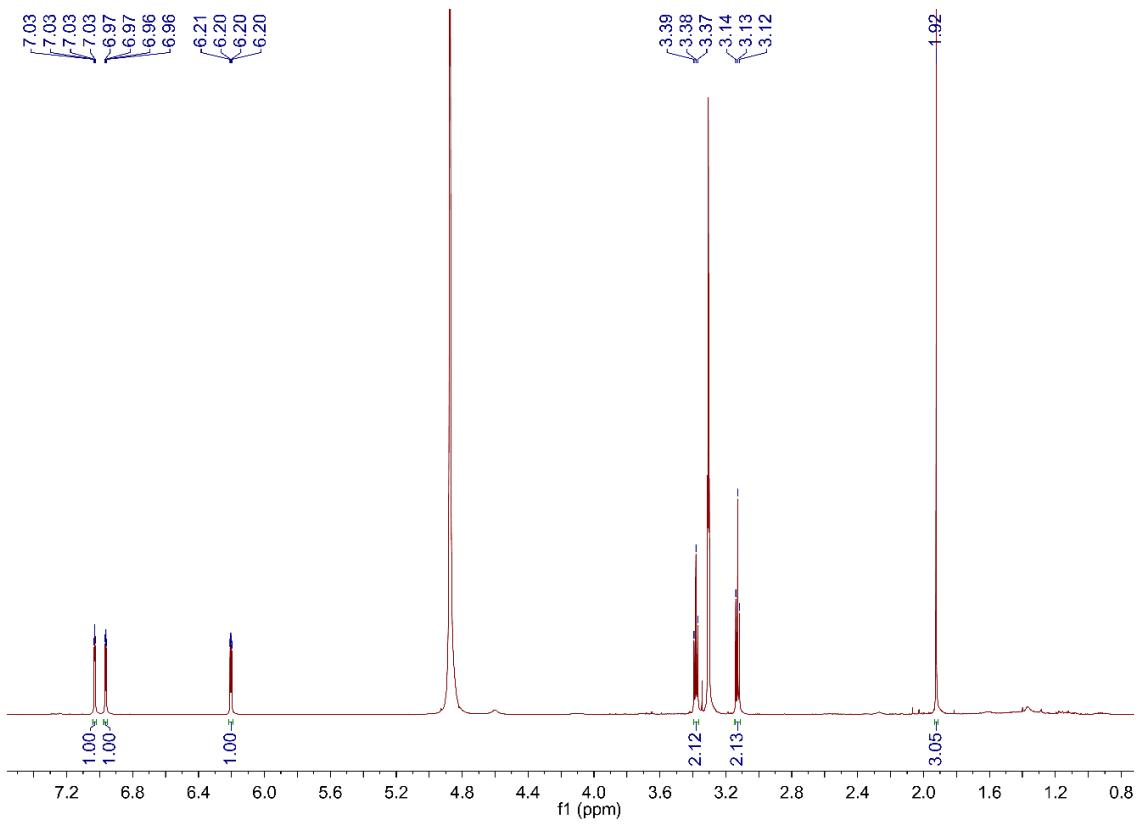


(E)

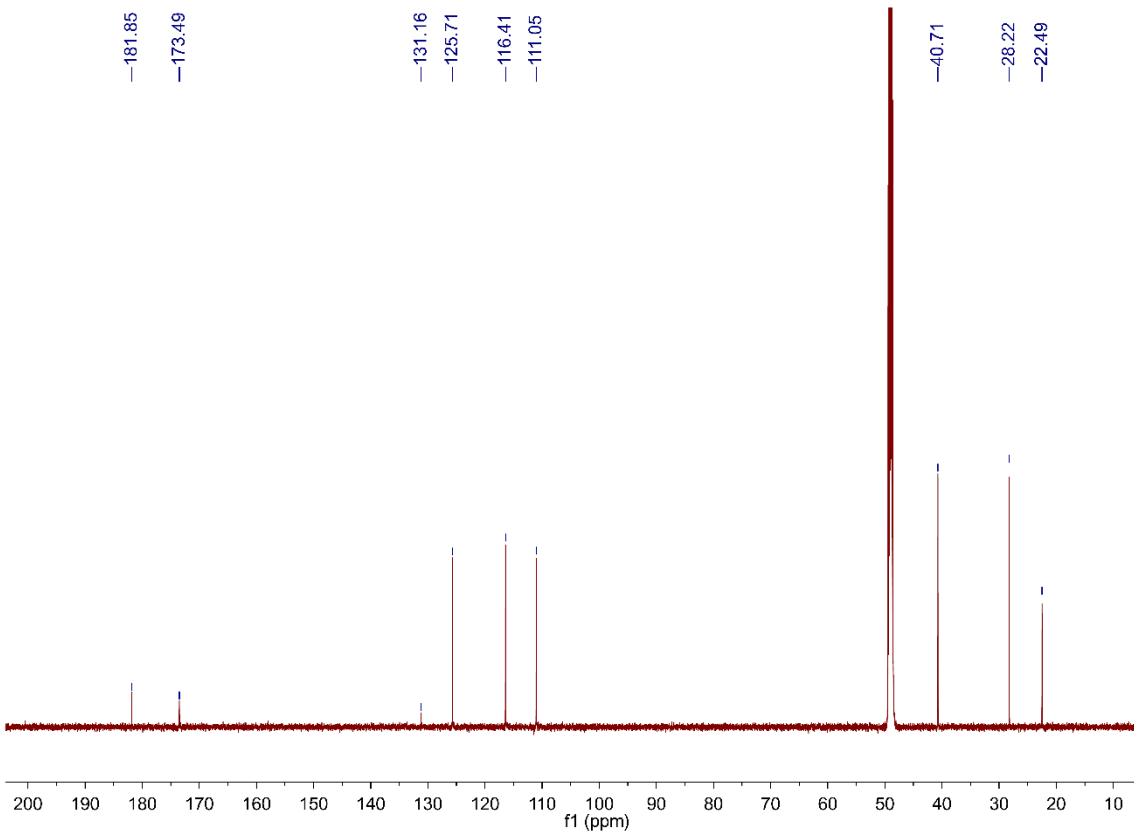


(F)

Figure S5. (A) ¹H NMR (600 MHz) spectrum of compound **1** in methanol-*d*₄; (B) ¹³C NMR (600 MHz) spectrum of compound **1** in methanol-*d*₄; (C) ¹H-¹H COSY (600 MHz) spectrum of compound **1** in methanol-*d*₄; (D) HSQC (600 MHz) spectrum of compound **1** in methanol-*d*₄; (E) HMBC (600 MHz) spectrum of compound **1** in methanol-*d*₄; (F) HRESI spectrum of compound **1**.



(A)

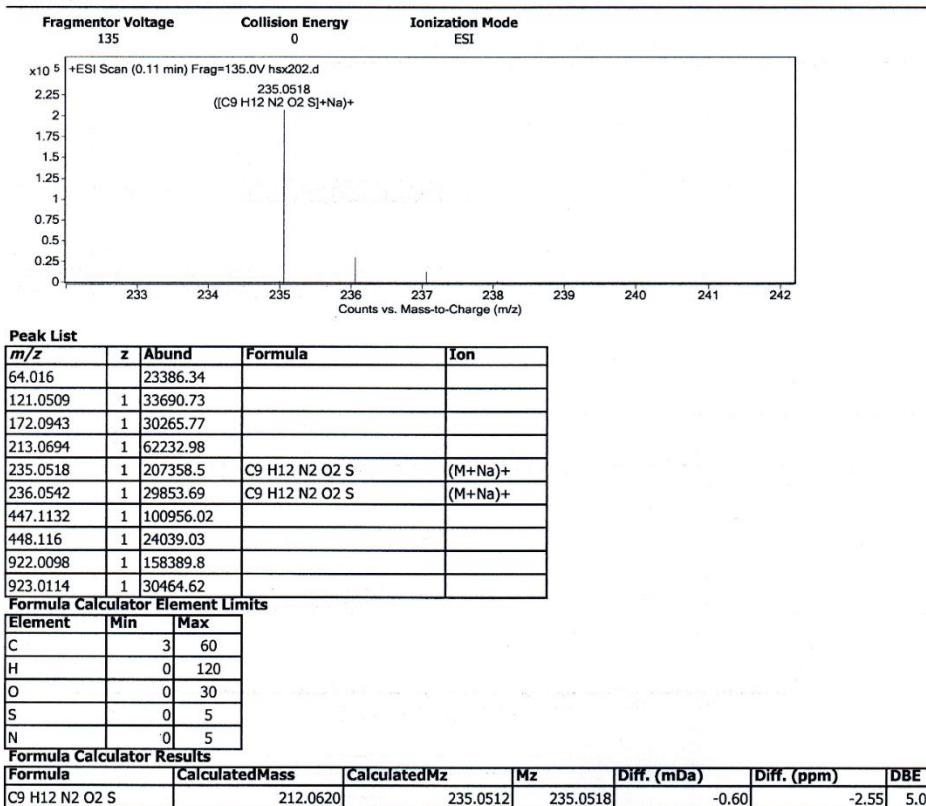


(B)

Qualitative Analysis Report

Data Filename	hsx202.d	Sample Name	hsx202
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Acq Method	s.m	Acquired Time	6/28/2019 2:19:59 PM
IRM Calibration Status	Success	DA Method	Default.m
Comment			
Sample Group	Info.		
Acquisition SW	6200 series TOF/6500 series		
Version	Q-TOF B.05.01 (B5125.2)		

User Spectra



--- End Of Report ---

(C)

Figure S6. (A)¹H-NMR (600 MHz) spectrum of compound **2** in methanol-*d*₄; (B)¹³C-NMR (600 MHz) spectrum of compound **2** in methanol-*d*₄; (C)HRESIMS spectrum of compound **2**.

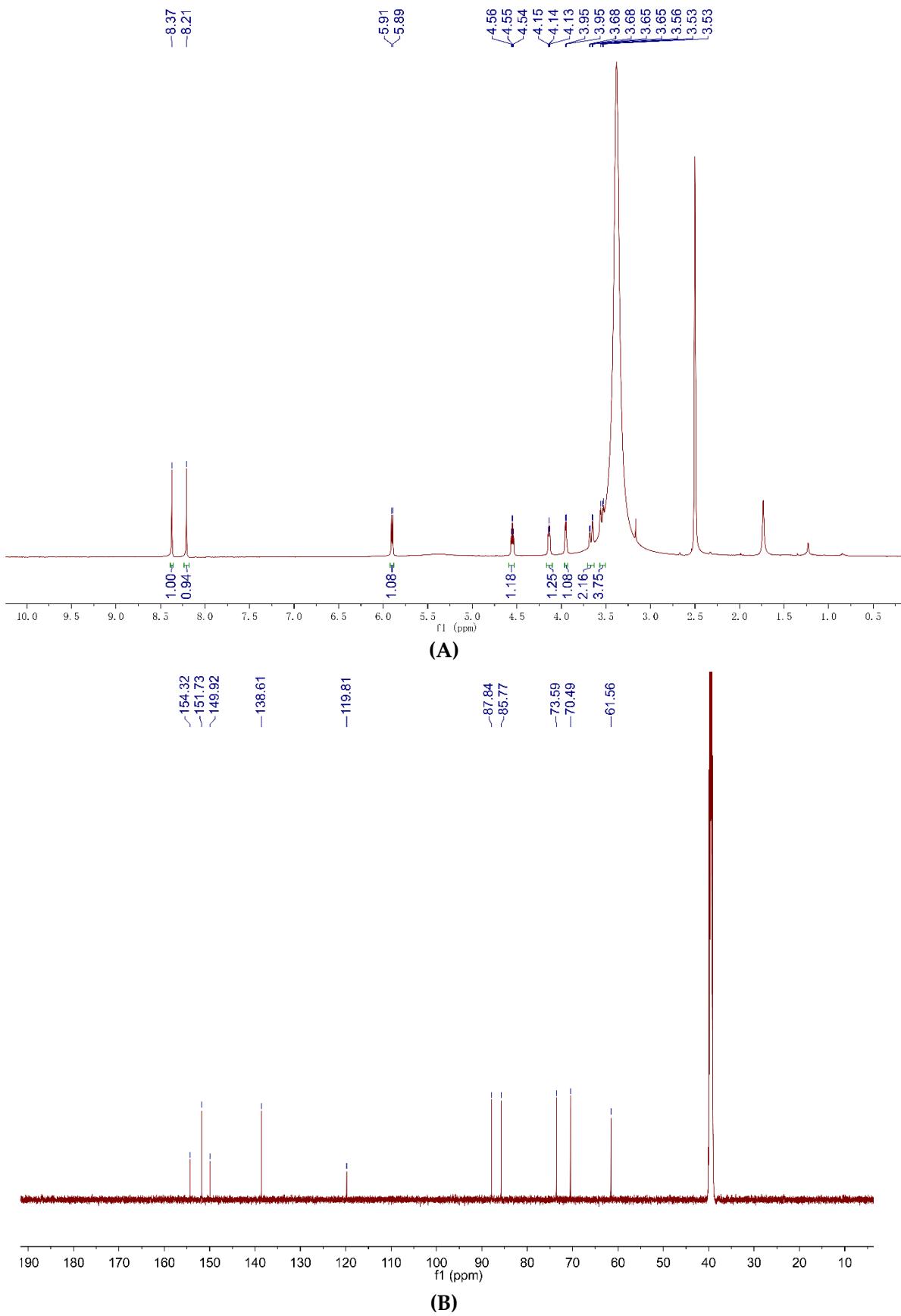


Figure S7. (A) ^1H -NMR (400 MHz) spectrum of compound 3 in $\text{DMSO}-d_6$; (B) ^{13}C -NMR (150 MHz) spectrum of compound 3 in $\text{DMSO}-d_6$.

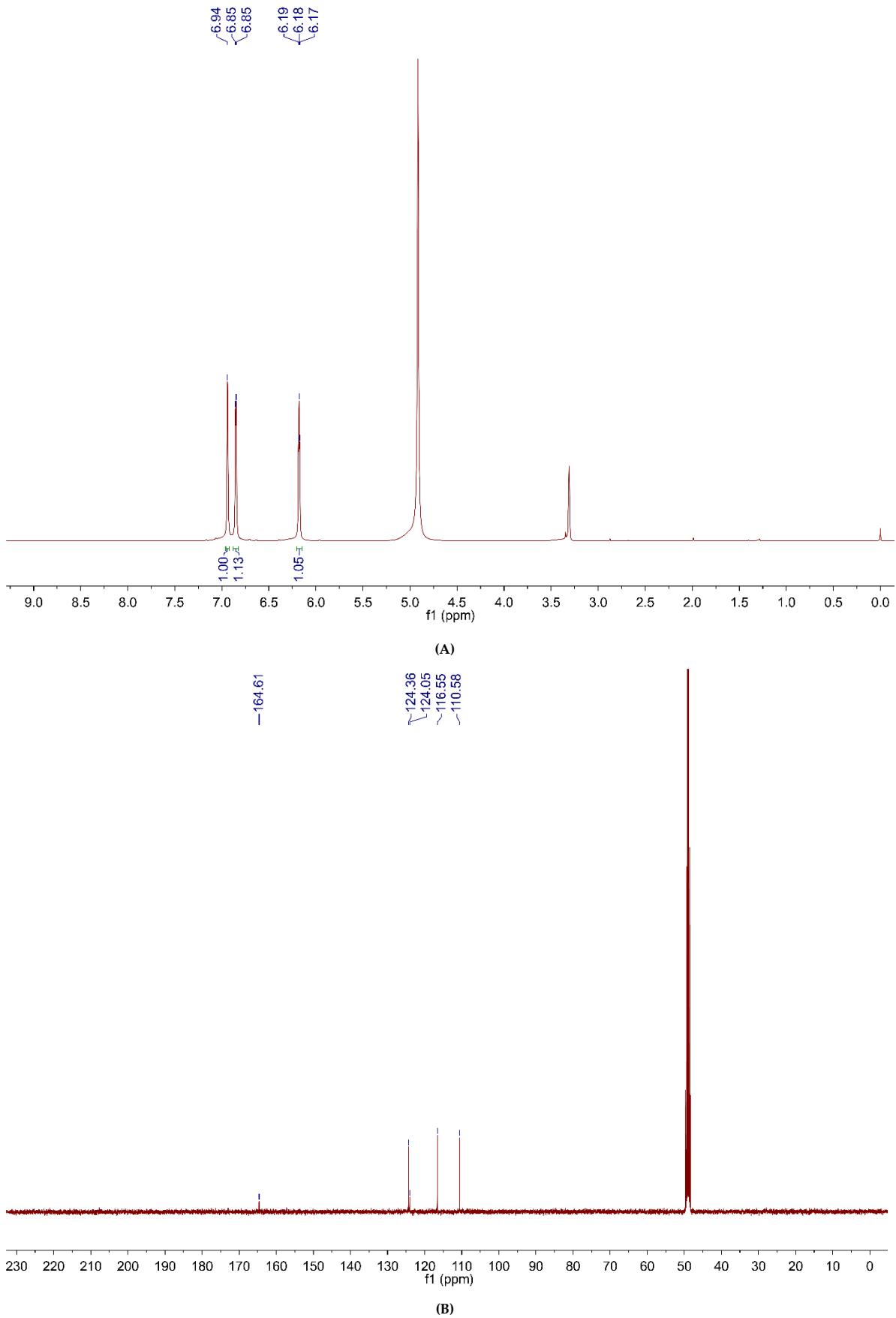


Figure S8. (A) ¹H NMR (400 MHz) spectrum of compound 4 in methanol-*d*₄; (B) ¹³C (100 MHz) spectrum of compound 4 in methanol-*d*₄.