

Table S3. Predicted aromatic central pathways in *C. metallidurans* CH34

Central pathway	Gene	ORF (aa)	CDS	Function	Organism	Id (%)	Cov (%)	UniProtKB Accession (aa)
Catechol (extradiol)	<i>tomR</i>	RMET_RS06580 (614)	TomR	Transcriptional activator of aromatic catabolism, XylR/NtrC type	<i>B. vietnamiensis</i> G4	97	98	A4JW32 (611)
	<i>tomD</i>	RMET_RS06585 (275)	TomD	Semialdehyde-2-hydroxymuconate hydrolase (HMSH)	<i>B. vietnamiensis</i> G4	100	100	A4JW31 (275)
	<i>tomI</i>	RMET_RS06645 (63)	TomI	4-oxalocrotonate tautomerase	<i>B. vietnamiensis</i> G4	100	100	A4JW27 (63)
	<i>tomH</i>	RMET_RS06650 (262)	TomH	4-oxalocrotonate decarboxylase	<i>B. vietnamiensis</i> G4	99	99	A4JW26 (260)
	<i>tomG</i>	RMET_RS06655 (348)	TomG	4-hydroxy-2-ketovalerate aldolase	<i>B. vietnamiensis</i> G4	99	100	A4JW25 (343)
	<i>tomF</i>	RMET_RS06660 (303)	TomF	Acetaldehyde-CoA dehydrogenase II	<i>B. vietnamiensis</i> G4	99	100	A4JW24 (299)
	<i>tomE</i>	RMET_RS06665 (262)	TomE	2-hydroxypent-2,4-dienoate hydratase	<i>B. vietnamiensis</i> G4	99	100	A4JW23 (258)
	<i>tomC</i>	RMET_RS06670 (503)	TomC	Semialdehyde-2-hydroxymuconate dehydrogenase (HMSD)	<i>B. vietnamiensis</i> G4	98	100	A4JW22 (493)
	<i>tomB</i>	RMET_RS06680 (314)	TomB	Catechol-2,3-dioxygenase (C23O)	<i>B. vietnamiensis</i> G4	100	100	A4JW20 (311)
Catechol (intradiol β -ketoadipate)	<i>catA1</i>	RMET_RS08920 (305)	CatA1	Catechol-1,2-dioxygenase (C12O)	<i>A. lwoffii</i> K24	71	84	O33950 (275)
	<i>catD</i>	RMET_RS25015 (258)	CatD	3-oxoadipate enol-lactonase	<i>A. baylyi</i> ADP1	44	96	P00632 (267)
	<i>catC</i>	RMET_RS25025 (92)	CatC	Muconolactone delta-isomerase	<i>C. pinatubonensis</i> JMP134	84	100	P80573 (92)
	<i>catB</i>	RMET_RS25030 (374)	CatB	Muconate cycloisomerase	<i>C. pinatubonensis</i> JMP134	93	98	P08310 (375)
	<i>catM</i>	RMET_RS25035 (291)	CatM	LysR family transcriptional regulator	<i>P. putida</i> PSR2000	58	100	P20667 (289)
	<i>benM</i>	RMET_RS25040 (305)	BenM	LysR family transcriptional regulator	<i>A. baylyi</i> ADP1	48	98	O68014 (304)
	<i>catA2</i>	RMET_RS25045 (307)	CatA2	Catechol-1,2-dioxygenase	<i>A. lwoffii</i> K24	65	93	O33948 (275)
Protocatechuate (intradiol β -ketoadipate)	<i>pcaI</i>	RMET_RS19060 (229)	PcaI	3-oxoacid CoA-transferase subunit A	<i>P. putida</i> PRS2000	74	93	Q01103 (231)
	<i>pcaJ</i>	RMET_RS19065 (215)	PcaJ	3-oxoacid CoA-transferase subunit B	<i>P. putida</i> KT2440	75	96	P0A101 (213)
	<i>pcaF</i>	RMET_RS19070 (400)	PcaF	3-oxoadipyl-CoA thiolase	<i>A. baylyi</i> ADP1	68	100	Q43974 (401)
	<i>pcaK</i>	RMET_RS20760 (459)	PcaK	4-hydroxybenzoate transporter	<i>P. putida</i> PRS2000	57	95	Q51955 (448)
	<i>pcaQ</i>	RMET_RS20765 (317)	PcaQ	LysR family transcriptional regulator	<i>E. coli</i> K-12	42	94	P77171 (307)
	<i>pcaH</i>	RMET_RS20770 (237)	PcaH	Protocatechuate 3,4-dioxygenase subunit beta	<i>B. cepacia</i> DBO1	63	93	P15110 (235)
	<i>pcaG</i>	RMET_RS20775 (189)	PcaG	Protocatechuate 3,4-dioxygenase subunit alpha	<i>P. putida</i> KT2440	43	100	P00436 (201)
	<i>pcaB</i>	RMET_RS20780 (490)	PcaB	3-carboxy- <i>cis,cis</i> -muconate cycloisomerase	<i>A. baylyi</i> ADP1	49	96	Q59092 (451)
	<i>pcaL</i>	RMET_RS20785 (392)	PcaL	Enol-lactone hydrolase/4-CML decarboxylase	<i>R. jostii</i> RHA1	42	95	Q0SH24 (400)
Gallate	<i>pmdE</i>	RMET_RS19250 (341)	PmdE	4-oxalomesaconate hydratase	<i>C. testosteroni</i> BR6020	76	99	Q93PS9 (342)
	<i>galD</i>	RMET_RS19255 (382)	GalD	4-oxalomesaconate tautomerase	<i>P. putida</i> KT2440	51	99	Q88JY0 (361)
	<i>galR</i>	RMET_RS19260 (406)	GalR	LysR family transcriptional regulator	<i>P. putida</i> KT2440	43	98	Q88JX7 (397)

	<i>galT</i>	RMET_RS19265 (441)	GalT	Aromatic acid/H ⁺ symport family MFS transporter	<i>P. putida</i> KT2440	55	97	E8ZB61 (449)
	<i>galA</i>	RMET_RS19270 (430)	GaIA	Gallate 2,3-dioxygenase	<i>P. putida</i> KT2440	64	99	Q88JX5 (420)
	<i>galB</i>	RMET_RS21720 (245)	GalB	4-oxalomesaconate hydratase	<i>P. putida</i> KT2440	77	100	Q88JX8 (258)
	<i>galC</i>	RMET_RS21725 (237)	GalC	4-carboxy-4-hydroxy-2-oxoadipate aldolase	<i>P. putida</i> KT2440	60	96	Q88JX9 (238)
Homogentisate	<i>hmgC</i>	RMET_RS01425 (215)	HmgC	Maleylacetoacetate isomerase	<i>P. xenovorans</i> LB400	64	100	Q140J8 (214)
	<i>hmgA</i>	RMET_RS22545 (442)	HmgA	Homogentisate 1,2-dioxygenase	<i>P. xenovorans</i> LB400	81	98	Q140K0 (439)
	<i>hmgB</i>	RMET_RS22550 (420)	HmgB	Fumarylacetoacetase	<i>P. xenovorans</i> LB400	65	96	Q144Z1 (419)
Hydroxyquinol	<i>pnpC</i>	RMET_RS25940 (296)	PnpC	Hydroxyquinol 1,2-dioxygenase	<i>P. putida</i> DDL-E4	68	95	C6FI44 (290)
	<i>pnpE</i>	RMET_RS25945 (355)	PnpE	Maleylacetate reductase	<i>P. putida</i> DDL-E4	73	100	C6FI43 (355)
Benzoyl-CoA	<i>boxA</i>	RMET_RS06160 (415)	BoxA	Benzoyl-CoA oxygenase component A	<i>A. Evansii</i> KB740	62	99	Q9AIX6 (414)
	<i>boxB</i>	RMET_RS06165 (474)	BoxB	Benzoyl-CoA oxygenase component B	<i>A. Evansii</i> KB740	72	99	Q9AIX7 (473)
	<i>boxC</i>	RMET_RS06170 (558)	BoxC	Benzoyl-CoA-dihydrodiol lyase	<i>A. Evansii</i> KB740	70	97	Q84HH6 (555)
	<i>boxR</i>	RMET_RS06175 (316)	BoxR	Transcriptional regulator	<i>Azoarcus</i> sp. CIB	50	91	G8B2G2 (300)
Phenylacetyl-CoA	<i>paaH</i>	RMET_RS04900 (507)	PaaH	3-hydroxybutyryl-CoA dehydrogenase	<i>B. cenocepacia</i> J2315	41	94	B4EL90 (518)
	<i>paaF</i>	RMET_RS15855 (258)	PaaF	Enoyl-CoA hydratase	<i>B. cenocepacia</i> J2315	84	100	B4E7C0 (258)
	<i>paaG</i>	RMET_RS15860 (280)	PaaG	2-(1,2-epoxy-1,2-dihydrophenyl)acetyl-CoA isomerase	<i>B. cenocepacia</i> J2315	75	93	B4E7B7 (263)
	<i>paaI</i>	RMET_RS15865 (146)	PaaI	Hydroxyphenylacetyl-CoA thioesterase PaaI	<i>B. cenocepacia</i> J2315	73	94	B4E7B6 (150)
	<i>paaA</i>	RMET_RS16925 (335)	PaaA	1,2-phenylacetyl-CoA epoxidase subunit A	<i>B. cenocepacia</i> J2315	81	97	B4E5A2 (332)
	<i>paaB</i>	RMET_RS16930 (95)	PaaB	1,2-phenylacetyl-CoA epoxidase subunit B	<i>B. cenocepacia</i> J2315	86	98	B4E5A1 (94)
	<i>paaC</i>	RMET_RS16935 (285)	PaaC	1,2-phenylacetyl-CoA epoxidase subunit C	<i>B. cenocepacia</i> J2315	67	97	B4E5A0 (267)
	<i>paaD</i>	RMET_RS16940 (202)	PaaD	Phenylacetate-CoA oxygenase subunit PaaD	<i>B. cenocepacia</i> J2315	69	91	B4E599 (184)
	<i>paaE</i>	RMET_RS16945 (361)	PaaE	Phenylacetate-CoA oxygenase/reductase subunit PaaE	<i>B. cenocepacia</i> J2315	59	99	B4E598 (362)
2-aminobenzoyl-CoA	<i>paaJ</i>	RMET_RS19070 (400)	PaaJ	β -ketoadipyl-CoA thiolase	<i>B. cenocepacia</i> J2315	88	100	B4E7B8 (400)
	<i>abmE</i>	RMET_RS11025 (137)	AbmE	Translation inhibitor protein	<i>A. Evansii</i> KB740	60	96	Q93FB4 (132)
	<i>abmD</i>	RMET_RS11040 (388)	AbmD	Acyl-CoA dehydrogenase	<i>A. Evansii</i> KB740	70	94	Q93FB5 (401)
	<i>abmC</i>	RMET_RS11045 (283)	AbmC	Enoyl-CoA hydratase/isomerase	<i>A. Evansii</i> KB740	70	98	Q93FB6 (281)
	<i>abmB</i>	RMET_RS11050 (264)	AbmB	3-Hydroxyacyl-CoA dehydrogenase	<i>A. Evansii</i> KB740	64	94	Q93FB7 (270)
	<i>abmA</i>	RMET_RS11055 (791)	AbmA	2-Aminobenzoyl-CoA monooxygenase/reductase	<i>A. Evansii</i> KB740	66	99	Q93FB8 (773)
3-hydroxyanthranilate	<i>onbC</i>	RMET_RS26575 (174)	OnbC	3-hydroxyanthranilate 3,4-dioxygenase	<i>P. phytofirmans</i> PsJN	88	100	B2T2S5 (174)
	<i>onbR</i>	RMET_RS26580 (354)	OnbR	LysR family transcriptional regulator	<i>Cupriavidus</i> sp. ST-14	63	98	A0A0K0PM17 (320)

<i>onbE</i>	<i>RMET_RS26640 (487)</i>	OnbE	2-aminomuconic 6-semialdehyde dehydrogenase	<i>Cupriavidus</i> sp. ST-14	71	97	A0A0K0PM01 (496)
<i>onbH</i>	<i>RMET_RS26645 (273)</i>	OnbH	2-oxopent-4-dienoate hydratase	<i>Cupriavidus</i> sp. ST-14	67	90	A0A0K0PLY5 (275)
<i>onbJ</i>	<i>RMET_RS26650 (314)</i>	OnbJ	Acetaldehyde dehydrogenase	<i>Cupriavidus</i> sp. ST-14	73	94	A0A0K0PN84 (316)
<i>onbI</i>	<i>RMET_RS26655 (345)</i>	OnbI	4-hydroxy-2-oxovalerate aldolase	<i>Cupriavidus</i> sp. ST-14	79	98	A0A0K0PMH1 (346)
<i>onbG</i>	<i>RMET_RS26660 (254)</i>	OnbG	4-oxalocrotonate decarboxylase	<i>P. fluorescens</i> KU-7	73	77	Q83V28 (251)
<i>onbF</i>	<i>RMET_RS26665 (145)</i>	OnbF	2-aminomuconate deaminase	<i>P. fluorescens</i> KU-7	75	98	Q83V27 (143)
<i>onbD</i>	<i>RMET_RS26670 (333)</i>	OnbD	2-amino-3-carboxymuconate 6-semialdehyde decarboxylase	<i>Cupriavidus</i> sp. ST-14	71	96	A0A0K0PM06 (340)
