

## Supplementary Information

**Table 1.** Haemosporidian parasite lineages identified in this study, and records of their occurrence on MalAvi Database. M, migratory; R, resident.

This Study		Malavi Database		
Lineage (Genbank n°)	Host	Hosts	Country	Reference Name
pBASCUL01 (MT724400)	R: <i>Basileuterus culicivorus</i>			
pCOLL4 (DQ368374)	M: <i>Elaenia albiceps</i>	<i>Carduelis spinus</i> , <i>Lanius collurio</i> , <i>Loxia curvirostris</i> , <i>Sturnus vulgaris</i>	Russia	[1], [2]
		<i>Coryphistera alaudina</i> , <i>Gnorimopsar chopi</i> , <i>Limnornis curvirostris</i> , <i>Mimus saturninus</i> , <i>Poospiza lateralis</i> , <i>Stephanophorus diadematus</i>	Uruguay	[3]
		<i>Ficedula albicollis</i>	Sweden Hungary	[4], [5] [6]
		<i>Cyclarhis gujanensis</i> <i>Dolichonyx oryzivorus</i> <i>Foudia omissa</i> , <i>Hartlaubius auratus</i> <i>Pycnonotus barbatus</i>	Brazil United States Madagascar Benin	[7] [8] [9] [10]
pCONLIN16 (JX021452)	R: <i>Conopophaga lineata</i>	<i>Basileuterus flaveolus</i> , <i>Conopophaga lineata</i>	Brazil	[11]
pCURCUR01 <i>P. homopolare</i> (pTURFAL01) (MT724472)	R: <i>Lochmias nematura</i>	<i>Curaeus curaeus</i> , <i>Turdus falcklandii</i>	Argentina	[7]
pDENPET03 (KU562464) <i>P. nucleophilum</i>	M: <i>Elaenia albiceps</i> R: <i>Troglodytes aedon</i> M: <i>Turdus flavipes</i> M: <i>Vireo olivaceus</i> R: <i>Zonotrichia capensis</i>	<i>Alopochen aegyptiacus</i> , <i>Ammodramus humeralis</i> , <i>Arremon taciturnus</i> , <i>Basileuterus flaveolus</i> , <i>Basileuterus leucoblepharus</i> , <i>Cacicus solitarius</i> , <i>Cantorchilus longirostris</i> , <i>Coereba flaveola</i> , <i>Colaptes melanochloros</i> , <i>Coryphistera alaudina</i> , <i>Coryphospingus pileatus</i> , <i>Cyclarhis gujanensis</i> , <i>Cygnus atratus</i> , <i>Cypsnagra hirundinacea</i> , <i>Formicivora melanogaster</i> , <i>Guaruba guarouba</i> , <i>Gubernatrix cristata</i> , <i>Hemithraupis guira</i> , <i>Mimus saturninus</i> , <i>Myiarchus swainsoni</i> , <i>Myiothlypis flaveola</i> , <i>Neothraupis fasciata</i> , <i>Netta erythrophthalma</i> , <i>Pachyrhamphus polychopterus</i> , <i>Parula pitiayumi</i> , <i>Passer</i>	Brazil	[7], [11], [12], [13], [14], [15], [16], [17]; [18], [19]

		<i>domesticus</i> , <i>Phoenicopterus chilensis</i> , <i>Pipile jacutinga</i> , <i>Psarocolius decumanus</i> , <i>Ramphocelus carbo</i> , <i>Ramphastos toco</i> , <i>Ramphastos vitellinus</i> , <i>Rynchops niger</i> , <i>Saltator coerulescens</i> , <i>Schoeniophylax phryganophilus</i> , <i>Spheniscus magellanicus</i> , <i>Synallaxis frontalis</i> , <i>Thamnophilus nigrocinereus</i> , <i>Thryothorus longirostris</i> , <i>Trichothraupis melanops</i> , <i>Turdus albicollis</i> , <i>Turdus leucomelas</i> , <i>Turdus rufoventris</i> , <i>Volatinia jacarina</i> , <i>Zonotrichia capensis</i>		
		<i>Anas discors</i> , <b><i>Dendroica petechia</i></b> , <i>Dolichonyx oryzivorus</i> , <i>Dumetella carolinensis</i> , <i>Geothlypis trichas</i> , <i>Petrochelidon pyrrhonota</i> , <i>Riparia riparia</i> , <i>Setophaga petechia</i> , <i>Turdus migratorius</i> , <i>Vireo griseus</i>	United States	[8], [20], [21], [22], [23], [24], [25]
		<i>Basileuterus culicivorus</i> , <i>Basileuterus leucoblepharus</i> , <i>Craniolaema pyrrhophia</i> , <i>Gnorimopsar chopi</i> , <i>Zonotrichia capensis</i>	Uruguay	[3]
		<i>Automolus rufipileatus</i> , <i>Arremon taciturnus</i> , <i>Cyporhinus arada</i> , <i>Hypocnemis subflava</i> , <i>Pipra fasciicauda</i> , <i>Ramphocelus carbo</i> , <i>Turdus hauxwelli</i> , <i>Xiphorhynchus ocellatus</i>	Peru	[16]
		<i>Carduelis barbata</i> <i>Dendroica coronata</i>	Argentina Canada	[7] [26]
		<i>Cacicus cela</i> , <i>Cacicus haemorrhous</i> , <i>Diopsittaca nobilis</i> , <i>Volatinia jacarina</i>	Guyana	[3]
pELAALB07 (MT724471)	<b>M: <i>Elaenia albiceps</i></b>			
pGEOTRI01 (MF817777)	R: <i>Geothlypis aequinoctialis</i>	<i>Catharus ustulatus</i> , <i>Dendroica magnolia</i> , <i>Dendroica palmarum</i> , <b><i>Geothlypis trichas</i></b> , <i>Helmitheros vermivorum</i> , <i>Melospiza lincolni</i>	United States	[20], [23], [25], [27]
		<i>Atlapetes albinucha</i> <i>Euneornis campestris</i> <i>Melospiza melodia</i> , <i>Zonotrichia albicollis</i> <i>Seiurus noveboracensis</i>	Colombia Unknown Canada Venezuela	[28] [29] [30], [31] [23], [32], [33],
pLEAMA01 (JX021454)	R: <i>Conopophaga lineata</i>	<b><i>Leptopogon amaurocephalus</i></b> <i>Conopophaga lineata</i>	Brazil	[11]

pLEPCOR05 (pLECOR04) (KU236434) (pVOLJAC03)	R: <i>Tachyphonus coronatus</i> M: <i>Elaenia albiceps</i>	<i>Coryphospingus pileatus</i> , <b><i>Lepidothrix coronata</i></b> , <i>Volatinia jacarina</i> <i>Tangara schrankii</i>	Brazil  Peru	[7], [17], [34], [35]  [7]
pPADOM09 (AF069611) <i>P. elongatum</i>	M: <i>Elaenia albiceps</i>	<i>Ammodramus humeralis</i> , <i>Anser cygnoides</i> , <i>Asthenes pyrrholeuca</i> , <i>Basileuterus culicivorus</i> , <i>Carduelis barbata</i> , <i>Cnemotriccus fuscatus</i> , <i>Coryphospingus cucullatus</i> , <i>Coryphospingus pileatus</i> , <i>Dendrocolaptes certhia</i> , <i>Donacobius atricapilla</i> , <i>Elaenia albiceps</i> , <i>Elaenia cristata</i> , <i>Elaenia spectabilis</i> , <i>Furnarius leucopus</i> , <i>Haplospiza unicolor</i> , <i>Lathrotriccus euleri</i> <i>Myiarchus swainsoni</i> , <i>Myiarchus tyrannulus</i> , <i>Myiopagis viridicata</i> , <i>Myiophobus fasciatus</i> , <i>Neothraupis fasciata</i> , <i>Paroaria capitata</i> , <i>Passer domesticus</i> , <i>Pheugopedius genibarbis</i> , <i>Pitangus sulphuratus</i> , <i>Puffinus puffinus</i> , <i>Ramphotrigon ruficauda</i> , <i>Rhytipterna simplex</i> , <i>Saltator coerulescens</i> , <i>Spheniscus magellanicus</i> , <i>Synallaxis scutata</i> , <i>Tachyphonus coronatus</i> , <i>Tachyphonus cristatus</i> , <i>Tachyphonus phoenicius</i> , <i>Thraupis palmarum</i> , <i>Thryothorus genibarbis</i> , <i>Trichothraupis melanops</i> , <i>Troglodytes aedon</i> , <i>Troglodytes musculus</i> , <i>Tyrannus melancholicus</i> , <i>Zonotrichia capensis</i>	Brazil	[7], [11], [12], [13], [15], [16], [17], [18], [36]
		<i>Dendroica coronata</i> , <i>Dolichonyx oryzivorus</i> , <i>Geothlypis trichas</i> , <i>Larosterna inca</i> , <i>Mniotilta varia</i> , <i>Passer domesticus</i> , <i>Tachycineta thalassina</i>	United States	[8], [22], [27], [37], [38]
		<i>Gnorimopsar chopi</i> , <i>Pseudoleistes guirahuro</i> , <i>Stephanophorus diadematus</i> , <i>Tangara preciosa</i> , <i>Troglodytes aedon</i> , <i>Turdus rufiventris</i>	Uruguay	[3], [37]
		<i>Basileuterus culicivorus</i> , <i>Coryphospingus cucullatus</i> , <i>Elaenia albiceps</i> , <i>Elaenia mesoleuca</i> , <i>Troglodytes aedon</i> , <i>Zonotrichia capensis</i>	Argentina	[7], [39]
		<i>Elaenia albiceps</i> , <i>Troglodytes musculus</i>	Chile	[40]
		<i>Campylorhynchus yucatanicus</i>	Unknown	[29]
		<i>Troglodytes aedon</i>	Peru	[41]
pPADOM11 (HM146899)	M: <i>Elaenia albiceps</i>	<i>Aegolius acadicus</i> , <i>Agelaius phoeniceus</i> , <i>Baeolophus bicolor</i> , <i>Cardinalis cardinalis</i> , <i>Carduelis</i>	United States	[8], [12], [25], [27], [38], [42], [43], [44], [45], [46], [47],

		<i>tristes</i> , <i>Carpodacus mexicanus</i> , <i>Colaptes auratus</i> , <i>Dolichonyx oryzivorus</i> , <i>Dumetella carolinensis</i> , <i>Gavia immer</i> , <i>Quiscalus quiscula</i> , <i>Melospiza melodia</i> , <b><i>Passer domesticus</i></b> , <i>Passerina cyanea</i> , <i>Pheucticus ludovicianus</i> , <i>Sialia sialis</i> , <i>Somateria</i> spp, <i>Spheniscus demersus</i> , <i>Spizella passerina</i> , <i>Spizella pusilla</i> , <i>Strix varia</i> , <i>Turdus migratorius</i>		
		<i>Basileuterus flaveolus</i> , <i>Camptostoma obsoletum</i> , <i>Campylorhamphus trochilirostris</i> , <i>Coereba flaveola</i> , <i>Coryphospingus pileatus</i> , <i>Dacnis cayana</i> , <i>Neothraupis fasciata</i> , <i>Passer domesticus</i> , <i>Polioptila plumbea</i> , <i>Saltator similis</i> , <i>Sittasomus griseicapillus</i> , <i>Tachyphonus phoenicius</i> , <i>Tachyphonus rufus</i> , <i>Thlypopsis sordida</i> , <i>Trichothraupis melanops</i> , <i>Volatinia jacarina</i>	Brazil	[7], [11], [12], [16], [17],
		<i>Cacicus cela</i> , <i>Cyanocompsa cyanoides</i> , <i>Saltator grossus</i> , <i>Saltator maximus</i>	Guyana	[3]
		<i>Melospiza melodia</i> , <i>Tachycineta bicolor</i>	Canada	[30], [49]
		<i>Catharus minimus</i> <i>Chrysomus ruficapillus</i> , <i>Polioptila dumicola</i> <i>Mniotilta varia</i> , <i>Turdus fumigatus</i> , <i>Vireo griseus</i> <i>Volatinia jacarina</i>	Colombia Uruguay Unknown Peru	[49] [3] [29] [16]
pPYLEU01 (JX021484)	R: <i>Dysithamnus mentalis</i>	<i>Dysithamnus plumbeus</i> , <b><i>Pyriglena leucoptera</i></b>	Brazil	[11]
pRAMCAR05 (KU562679)	M: <i>Haplospiza unicolor</i>	<b><i>Ramphocelus carbo</i></b>	Brazil	[16]
pSPMAG06 (HM031936) <i>P. lutzi</i>	R: <i>Turdus leucomelas</i>	<i>Musophaga violacea</i> , <b><i>Spheniscus magellanicus</i></b> <i>Turdus rufiventris</i>	Brazil	[13], [50], [51], [7], [52]
pTARUF01 (JX021475)	R: <i>Tachyphonus coronatus</i>	<i>Basileuterus flaveolus</i> , <i>Saltator similis</i> , <b><i>Tachyphonus rufus</i></b> , <i>Tangara cayana</i> , <i>Thraupis palmarum</i>	Brazil	[7], [11],
pTRMEL02 (JX021468)	M: <i>Elaenia mesoleuca</i>	<i>Zonotrichia capensis</i>	Argentina	[39]
		<i>Anabazenops fuscus</i> , <b><i>Trichothraupis melanops</i></b> , <i>Vireo olivaceus</i> , <i>Zonotrichia capensis</i>	Brazil	[11]
pTUMIG03 (KU562788)	R: <i>Turdus rufiventris</i>	<i>Catharus ustulatus</i> , <i>Empidonax hammondi</i> , <i>Hylocichla mustelina</i> , <i>Icterus galbula</i> , <i>Larosterna inca</i> ,	United States	[20], [23], [24], [25], [27], [38], [45], [54],

		<i>Regulus calêndula</i> , <i>Sturnus vulgaris</i> , <i>Toxostoma rufum</i> , <b><i>Turdus migratorius</i></b> , <i>Vermivora celata</i>		
		<i>Spheniscus magellanicus</i> , <i>Turdus albicollis</i> , <i>Turdus amaurochalinus</i> , <i>Turdus flavipes</i> , <i>Turdus subalaris</i> , <i>Turdus rufiventris</i>	Brazil	[7], [11], [15], [16],
		<i>Entomodestes leucotis</i> , <i>Troglodytes aedon</i> , <i>Turdus nigriceps</i> , <i>Turdus serranus</i>	Peru	[7], [41]
		<i>Myioborus miniatus</i> , <i>Turdus albicollis</i> , <i>Turdus flavipes</i> , <i>Turdus fuscater</i> , <i>Turdus grayi</i> , <i>Turdus olivater</i>	Colombia	[49]
		<i>Anairetes fernandezianus</i> <i>Turdus falcklandii</i> , <i>Zonotrichia capensis</i> <i>Turdus rufiventris</i>	Chile Argentina Uruguay	[55] [40], [55] [3], [51]
pVIOLI03 (JX029897)	R: <i>Heliodoxa rubricauda</i> M: <i>Vireo olivaceus</i>	<i>Troglodytes aedon</i> , <b><i>Vireo olivaceus</i></b> <b><i>Vireo olivaceus</i></b> <b><i>Vireo olivaceus</i></b>	Brazil United States Peru	[11] [25] [7]
pPHPAT01 (KY305005)	M: <i>Elaenia albiceps</i>	<i>Ammodramus humeralis</i> , <i>Basileuterus flaveolus</i> <i>Basileuterus leucoblepharus</i> , <i>Coryphospingus pileatus</i> , <i>Paroaria dominicana</i> , <i>Phaeomyias murina</i> , <i>Rynchops niger</i> , <i>Spheniscus magellanicus</i> , <i>Tangara cyanoptera</i>	Brazil	[7], [11], [14], [15], [17], [52]
		<i>Bubo virginianus</i> , <i>Cardinalis cardinalis</i> , <i>Dolichonyx oryzivorus</i> , <i>Geothlypis trichas</i> , <i>Larosterna inca</i> , <i>Quiscalus quiscula</i>	United States	[8], [22], [25], [38], [44]
		<b><i>Phrygilus patagonicus</i></b> <i>Sayornis nigricans</i>	Chile Peru	[40] [53]
hELAALB01 (MK695430)	M: <i>Elaenia albiceps</i>	<i>Anairetes parulus</i> , <b><i>Elaenia albiceps</i></b>	Argentina	[7]
hMYISWA01 (KU562174)	M: <i>Elaenia albiceps</i> M: <i>Myiarchus swainsoni</i> M: <i>Tyrannus melancholicus</i>	<i>Elaenia chiriquensis</i> , <b><i>Myiarchus swainsoni</i></b> <i>Myiodynastes maculatus</i> , <i>Myiopagis viridicata</i> , <i>Phaeomyias murina</i> , <i>Phaethornis malaris</i> , <i>Serpophaga subcristata</i>	Brazil	[7], [16]
hTANDES01 (MT724553)	R: <b><i>Tangara desmaresti</i></b>			
hVIGIL09 (hCOLPLU01) (MH457345)	M: <i>Vireo olivaceus</i>	<i>Euphonia xanthogaster</i> , <i>Microcerculus marginatus</i> <i>Myiodynastes maculatus</i> , <i>Myiornis albiventris</i>	Peru	[7]

		<i>Patagioenas plumbea</i> , <i>Turdus nigriceps</i> , <b><i>Vireo gilvus</i></b> , <i>Willisornis poecilinotus</i>		
		<b><i>Vireo gilvus</i></b> <i>Vireo olivaceus</i>	United States Brazil	[25] [7]
hVIOLI05 (hVIREO02) (KF482350)	<i>M: Vireo olivaceus</i>	<b><i>Vireo olivaceus</i></b>	Peru Colombia Brazil	[53] [28] [7]
hCHIPAR01 (hVIROLI05) (MT724528)	<i>M: Vireo olivaceus</i>	<b><i>Chiroxiphia pareola</i></b>	Peru	[41]
hZOCAP01 (EF153649)	<i>R: Zonotrichia capensis</i>	<b><i>Zonotrichia capensis</i></b>	Chile Peru Colombia Brazil Argentina Ecuador Costa Rica	[39], [40], [39], [53], [56] [28], [39], [57], [7] [7] [39], [57], [39]
IDIUDIU11 (MK947686)	<i>M: Elaenia albiceps</i>	<b><i>Diuca diuca</i></b> <i>Aphrastura spinicauda</i>	Argentina Chile	[7] [58]
IELAALB02 (MK947541)	<i>M: Elaenia albiceps</i>	<b><i>Elaenia albiceps</i></b> <i>Aphrastura spinicauda</i>	Argentina Chile	[7] [58]
IELAALB05 (MK947689)	<i>M: Elaenia albiceps</i>	<b><i>Elaenia albiceps</i></b> <i>Aphrastura spinicauda</i>	Argentina Chile	[7] [58]
IZOLPYR01 (MK947545)	<i>M: Elaenia albiceps</i>	<b><i>Xolmis pyrope</i></b> <i>Aphrastura spinicauda</i>	Argentina Chile	[7] [58]
ITROAED02 (KF767431)	<i>M: Elaenia albiceps</i>	<b><i>Troglodytes aedon</i></b> <i>Anabacerthia striaticollis</i> , <i>Arremon schlegeli</i> , <i>Atlapetes melanocephalus</i> , <i>Basileuterus basilicus</i> , <i>Basileuterus conspicillatus</i> , <i>Buarremon torquatus</i> , <i>Catamenia homochroa</i> , <i>Catharus fuscater</i> , <i>Craniola leuca hellmayri</i> , <i>Diglossa albilatera</i> , <i>Elaenia frantzii</i> , <i>Henicorhina anachoreta</i> , <i>Henicorhina leucophrys</i> , <i>Mecocerculus leucophrys</i> , <i>Mionectes olivaceus</i> , <i>Myiarchus tuberculifer</i> , <i>Myioborus flavivertex</i> , <i>Myioborus miniatus</i> , <i>Myrmotherula schisticolor</i> , <i>Pipra erythrocephala</i> , <i>Pyrrhomyias cinnamomeus</i> , <i>Turdus albicollis</i> , <i>Turdus flavipes</i> , <i>Turdus fuscater</i> , <i>Zonotrichia capensis</i>	Peru Colombia	[41] [49]

## References:

1. Palinauskas, V.; Žiegytė, R.; Ilgūnas, M.; Iezhova, T.A.; Bernotienė, R.; Bolshakov, C.; Valkiūnas, G. Description of the first cryptic avian malaria parasite, *Plasmodium homocircumflexum* n. sp., with experimental data on its virulence and development in avian hosts and mosquitoes. *Int. J. Parasitol.* **2015**, *45*, 51–62. [https://doi.org/10.1016/j.ijpara.2014.08.012] [https://pubmed.ncbi.nlm.nih.gov/25449950/]
2. Ilgūnas, M.; Bukauskaitė, D.; Palinauskas, V. *et al.* Mortality and pathology in birds due to *Plasmodium (Giovannolaia) homocircumflexum* infection, with emphasis on the exoerythrocytic development of avian malaria parasites. *Malar. J.* **2016**, *15*, 256 [https://doi.org/10.1186/s12936-016-1310-x] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc4857288/]
3. Durrant, K.; Beadell, J.; Ishtiaq, F.; Graves, G.; Olson, S.; Gering, E.; . . . Fleischer, R. Avian Hematozoa in South America: A Comparison of Temperate and Tropical Zones. *Ornithol. Monographs.* **2006**, *60*, 98–111. [https://doi.org/10.2307/40166831]
4. Kulma, K.; Low, M.; Besch, S.; Qvarnström, A. Malaria infections reinforce competitive asymmetry between two *Ficedula* flycatchers in a recent contact zone. *Mol Ecol.* **2013**, *22*, 4591–4601. [https://doi.org/10.1111/mec.12409] [https://pubmed.ncbi.nlm.nih.gov/23980765/]
5. Kulma, K.; Low, M.; Besch, S.; Qvarnström, A. Malaria-infected female collared flycatchers (*Ficedula albicollis*) do not pay the cost of late breeding. *PLoS One.* **2014**, *9*, e85822. [https://doi.org/10.1371/journal.pone.0085822] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc3900437/]
6. Garamszegi, L. Z.; Zagalska-Neubauer, M.; Canal, D.; Markó, G.; Szász, E.; Zsebők, S.; Szöllősi, E.; Herczeg, G.; Török, J.; Malaria parasites, immune challenge, MHC variability, and predator avoidance in a passerine bird, *Behav. Ecol.* **2015**, *26*, 1292–1302. [https://doi.org/10.1093/beheco/arv077]
7. Fecchio, A.; Bell, J.A.; Pinheiro, R.B.P.; Cueto, V.R.; Gorosito, C.A.; Lutz, H.; Gaiotti, M.G.; Paiva, L.V.; França, L.F.; Toledo-Lima, G.; Tolentino, M.; Pinho, J.B.; Tkach, V.V.; Fontana, C.S.; Grande, J.M.; Santillán, M.A.; Caparroz, R.; Roos, A.L.; Bessa, R.; Nogueira, W.; Moura, T.; Nolasco, E.C.; Comiche, K.J.M.; Kirchgatter, K.; Guimarães, L.O.; Dispoto, J.H.; Marini, M.Â.; Weckstein, J.D.; Batalha-Filho, H.; Collins, M.D. Avian host composition, local speciation and dispersal drive the regional assembly of avian malaria parasites in South American birds. *Mol. Ecol.* **2019**, *28*, 2681–2693. [https://doi.org/10.1111/mec.15094]
8. Levin, I. I.; Zwiers, P.; Deem, S. L.; Geest, E. A.; Higashiguchi, J. M.; Iezhova, T. A.; Jiménez-Uzcátegui, G.; Kim, D. H.; Morton, J. P.; Perlut, N. G.; Renfrew, R. B.; Sari, E. H.; Valkiunas, G.; Parker, P. G. Multiple lineages of Avian malaria parasites (*Plasmodium*) in the Galapagos Islands and evidence for arrival via migratory birds. *Conserv. Biol.* **2013**, *27*, 1366–1377. [https://doi.org/10.1111/cobi.12127] [https://pubmed.ncbi.nlm.nih.gov/24033638/]
9. Musa, S.; Mackenstedt, U.; Woog, F.; Dinkel, A. Avian malaria on Madagascar: prevalence, biodiversity and specialization of haemosporidian parasites. *Int. J. Parasitol.* **2019**, *49*, 199–210. [https://doi.org/10.1016/j.ijpara.2018.11.001] [https://pubmed.ncbi.nlm.nih.gov/30471288/]
10. Harvey, J.A.; Voelker, G. Avian haemosporidian detection across source materials: prevalence and genetic diversity. *Parasitol. Res.* **2017**, *116*, 3361–3371 [https://doi.org/10.1007/s00436-017-5654-0]
11. Lacorte, G. A.; Félix, G. M.; Pinheiro, R. R.; Chaves, A. V.; Almeida-Neto, G.; Neves, F. S.; Leite, L. O.; Santos, F. R.; Braga, E. M. Exploring the diversity and distribution of neotropical avian malaria parasites—a molecular survey from Southeast Brazil. *PLoS One.* **2013**, *8*, e57770. [https://doi.org/10.1371/journal.pone.0057770] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc3585926/]
12. Marzal, A.; Ricklefs, R.E.; Valkiūnas, G.; Albayrak, T.; Arriero, E.; Bonneaud, C.; Cziráj, G.A.; Ewen, J.; Hellgren, O.; Hořáková, D.; Iezhova, T.A.; Jensen, H.; Križanauskienė, A.; Lima, M.R.; de Lope, F.; Magnussen, E.; Martin, L. B.; Møller, A. P.; Palinauskas, V.; Pap, P. L.; . . . Besch, S. Diversity, loss, and gain of malaria parasites in a globally invasive bird. *PLoS One.* **2011**, *6*, e21905. [https://doi.org/10.1371/journal.pone.0021905] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc3136938/]
13. Chagas, C.R.; Valkiūnas, G.; de Oliveira Guimarães, L.; Monteiro, E.F.; Guida, F.J.; Simões, R.F.; Rodrigues, P.T.; de Albuquerque Luna, E.J.; Kirchgatter, K. Diversity and distribution of avian malaria and related haemosporidian parasites in captive birds from a Brazilian megalopolis. *Malar. J.* **2017**, *16*, 83. [https://doi.org/10.1186/s12936-017-1729-8] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc5316177/]

14. Roos, F.L.; Belo, N.O.; Silveira, P.; Braga, E.M. Prevalence and diversity of avian malaria parasites in migratory Black Skimmers (*Rynchops niger*, Laridae, Charadriiformes) from the Brazilian Amazon Basin. *Parasitol. Res.* **2015**, *114*, 3903–3911. [https://doi.org/10.1007/s00436-015-4622-9] [https://pubmed.ncbi.nlm.nih.gov/26193823/]
15. Vanstreels, R.E.; da Silva-Filho, R.P.; Kolesnikovas, C.K.; Bhering, R.C.; Ruoppolo, V.; Epiphany, S.; Amaku, M.; Ferreira Junior, F.C.; Braga, É.M.; Catão-Dias, J.L. Epidemiology and pathology of avian malaria in penguins undergoing rehabilitation in Brazil. *Vet. Res.* **2015**, *46*, 30. [https://doi.org/10.1186/s13567-015-0160-9] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc4357068/]
16. Fecchio, A.; Svensson-Coelho, M.; Bell, J.; Ellis, V.A.; Medeiros, M.C.; Trisos, C.H.; Blake, J.G.; Loiselle, B.A.; Tobias, J.A.; Fanti, R. et al. Host associations and turnover of haemosporidian parasites in manakins (Aves: Pipridae). *Parasitology*. **2017**, *144*, 984–993. [https://doi.org/10.1017/S0031182017000208] [https://pubmed.ncbi.nlm.nih.gov/28290270/]
17. Ferreira Junior, F. C.; Rodrigues, R. A.; Ellis, V. A.; Leite, L. O.; Borges, M.; Braga, É. M. Habitat modification and seasonality influence avian haemosporidian parasite distributions in southeastern Brazil. *PLoS One*. **2017**, *12*, e0178791. [https://doi.org/10.1371/journal.pone.0178791] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc5456369/]
18. Fecchio, A.; Pinheiro, R.; Felix, G.; Faria, I.P.; Pinho, J.B.; Lacorte, G.A.; Braga, E.M.; Farias, I.P.; Aleixo, A.; Tkach, V.V.; Collins, M.D.; Bell, J.A.; Weckstein, J.D. Host community similarity and geography shape the diversity and distribution of haemosporidian parasites in Amazonian birds. *Ecography*. **2018**, *41*, 505–515. [https://doi.org/10.1111/ecog.03058]
19. De La Torre, G.; Freitas, F.; Fraton, R.; Guaraldo, A. de Angeli Dutra, D.; Braga, E.; Manica, L.; Hemoparasites and their relation to body condition and plumage coloration of the White-necked thrush (*Turdus albicollis*). *Ethol. Ecol. Evol.* **2020**, *32*, 6, 509–526 [https://doi.org/10.1080/03949370.2020.1769739]
20. Ricklefs, R.E.; & Fallon, S.M. Diversification and host switching in avian malaria parasites. *Proc. Biol. Sci.* **2002**, *269*(1494), 885–892. [https://doi.org/10.1098/rspb.2001.1940]
21. Szymanski, M.M.; & Lovette, I.J. High lineage diversity and host sharing of malarial parasites in a local avian assemblage. *J. Parasitol.* **2005**, *91*, 768–774. [https://doi.org/10.1645/GE-417R1.1] [https://pubmed.ncbi.nlm.nih.gov/17089742/]
22. Pagenkopp, K.M.; Klicka, J.; Durrant, K.L.; Garvin, J.C.; Fleischer, R.C. Geographic variation in malarial parasite lineages in the Common Yellowthroat (*Geothlypis trichas*). *Conserv. Genet.* **2008**, *9*, 1577–1588. [https://doi.org/10.1007/s10592-007-9497-6]
23. Oakgrove, K.S.; Harrigan, R.J.; Loiseau, C.; Guers, S.; Seppi, B.; Sehgal, R.N. Distribution, diversity and drivers of blood-borne parasite co-infections in Alaskan bird populations. *Int. J. Parasitol.* **2014**, *44*, 717–727. [https://doi.org/10.1016/j.ijpara.2014.04.011]
24. Walther, E.L.; Carlson, J.S.; Cornel, A. et al. First molecular study of prevalence and diversity of avian haemosporidia in a Central California songbird community. *J. Ornithol.* **2016**, *157*, 549–564 [https://doi.org/10.1007/s10336-015-1301-7]
25. Smith, J.D.; Gill, S.A.; Baker, K.M.; Vonhof, M.J. Prevalence and diversity of avian Haemosporidia infecting songbirds in southwest Michigan. *Parasitol. Res.* **2018**, *117*, 471–489. [https://doi.org/10.1007/s00436-017-5724-3] [https://pubmed.ncbi.nlm.nih.gov/29282527/]
26. Cozzarolo, C.S.; Jenkins, T.; Toews, D.; Brelsford, A.; Christe, P. Prevalence and diversity of haemosporidian parasites in the yellow-rumped warbler hybrid zone. *Ecol. Evol.* **2018**, *8*, 9834–9847. [https://doi.org/10.1002/ece3.4469] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc6202724/]
27. Martinsen, E.S.; Waite, J.L.; Schall, J.J. Morphologically defined subgenera of *Plasmodium* from avian hosts: test of monophyly by phylogenetic analysis of two mitochondrial genes. *Parasitology*. **2007**, *134*, 483–490. [https://doi.org/10.1017/S0031182006001922] [https://pubmed.ncbi.nlm.nih.gov/17147839/]
28. González, A.D.; Lotta, I.A.; García, L.F. Moncada, L.I.; Matta, N.E. Avian haemosporidians from Neotropical highlands: Evidence from morphological and molecular data. *Parasitol. Int.* **2015**, *64*, 48–59. [https://doi.org/10.1016/j.parint.2015.01.007] [https://pubmed.ncbi.nlm.nih.gov/25638289/]
29. Outlaw, D.C.; Ricklefs, R. E. On the phylogenetic relationships of haemosporidian parasites from raptorial birds (Falconiformes and Strigiformes). *J. Parasitol.* **2009**, *95*, 1171–1176. [https://doi.org/10.1645/GE-1982.1]
30. Sarquis-Adamson, Y.; & MacDougall-Shackleton, E.A. Song sparrows *Melospiza melodia* have a home-field advantage in defending against sympatric malarial parasites. *R. Soc. Open Sci.* **2016**, *3*, 160216. [https://doi.org/10.1098/rsos.160216] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc5108946/]
31. Boyd, R.J.; Kelly, T.R.; MacDougall-Shackleton, S.A.; MacDougall-Shackleton, E.A. Alternative reproductive strategies in white-throated sparrows are associated with differences in parasite load following experimental infection. *Biol. Lett.* **2018**, *14*, 20180194. [https://doi.org/10.1098/rsbl.2018.0194] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc6083222/]
32. Mijares, A.; Rosales, R.; Silva-Iturriza, A. Hemosporidian parasites in forest birds from Venezuela: genetic lineage analyses. *Avian Dis.* **2012**, *56*, 583–588. [https://doi.org/10.1637/10058-011312-ResNote.1] [https://pubmed.ncbi.nlm.nih.gov/23050478/]
33. Loiseau, C.; Harrigan, R.J.; Cornel, A.J.; Guers, S.L.; Dodge, M.; Marzec, T.; Carlson, J.S.; Seppi, B.; Sehgal, R.N. First evidence and predictions of *Plasmodium* transmission in Alaskan bird populations. *PLoS One*. **2012**, *7*, e44729. [https://doi.org/10.1371/journal.pone.0044729] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc3446979/]



34. Bosholn, M.; Fecchio, A.; Silveira, P.; Braga, É.M. and Anciães, M. Effects of avian malaria on male behaviour and female visitation in lekking blue-crowned manakins. *J. Avian Biol.* **2016**, *47*, 457–465. [https://doi.org/10.1111/jav.00864]
35. Fecchio, A.; Ribeiro, R.M.; Ferreira, F.C.; de Angeli Dutra, D.; Tolesano-Pascoli, G.; Alquezar, R.D.; Khan, A.U.; Pichorim, M.; Moreira, P.A.; Costa-Nascimento, M.J.; Monteiro, E.F.; Mathias, B.S.; Guimarães, L.O.; Simões, R.F.; Braga, É.M.; Kirchgatter, K.; Dias, R.I. Higher infection probability of haemosporidian parasites in Blue-black Grassquits (*Volatinia jacarina*) inhabiting native vegetation across Brazil. *Parasitol. Int.* **2021**, *80*, 102204. [https://doi.org/10.1016/j.parint.2020.102204]
36. Vanstreels, R.; Dutra, D.A.; Santos, A.P.; Hurtado, R.; Egert, L.; Braga, É.M. First report of avian malaria in a Manx shearwater (*Puffinus puffinus*). *Parasitol. Int.* **2020**, *78*, 102148. [https://doi.org/10.1016/j.parint.2020.102148] [https://pubmed.ncbi.nlm.nih.gov/32464257/]
37. Beadell, J.S.; Ishtiaq, F.; Covas, R.; Melo, M.; Warren, B.H.; Atkinson, C.T.; Bensch, S.; Graves, G.R.; Jhala, Y.V.; Peirce, M.A.; Rahmani, A.R.; Fonseca, D. M., Fleischer, R.C. Global phylogeographic limits of Hawaii's avian malaria. *Proc. Biol. Sci.* **2006**, *273*, 2935–2944. [https://doi.org/10.1098/rspb.2006.3671] [http://www.ncbi.nlm.nih.gov/pmc/articles/pmc1639517/]
38. Spottiswoode, N.; Bartlett, S.L.; Conley, K.J.; Seimon, T.A.; Griffin, D.O.; Sykes, J.M.; Analysis of *Plasmodium* Lineages identified in captive penguins (*Sphenisciformes* spp.), eiders (*Somateria* spp.), and inca terns (*Larosterna Inca*) in a North American zoological collection. *J. Zoo Wildl. Med.* **2020**, *51*, 140–149. [https://doi.org/10.1638/2019-0078] [https://pubmed.ncbi.nlm.nih.gov/32212557/]
39. Doussang, D.; González-Acuña, D.; Torres-Fuentes, L. G.; Loughheed, S. C.; Clemente-Carvalho, R. B.; Greene, K. C.; Vianna, J. A. Spatial distribution, prevalence and diversity of haemosporidians in the rufous-collared sparrow, *Zonotrichia capensis*. *Parasit. Vectors.* **2019**, *12*, 2. [https://doi.org/10.1186/s13071-018-3243-4] [https://pubmed.ncbi.nlm.nih.gov/30606248/]
40. Merino, S.; Moreno, J.; Vásquez, R.A.; Martínez, J.; Sánchez-Monsálvez, I.; Estades, C.F.; Ippi, S.; Sabat, P.; Rozzi, R.; McGehee, S. Haematozoa in forest birds from southern Chile: Latitudinal gradients in prevalence and parasite lineage richness. *Austral. Ecol.* **2008**, *33*, 329–340. [https://doi.org/10.1111/j.1442-9993.2008.01820.x]
41. Galen, S.C.; and Witt, C.C. Diverse avian malaria and other haemosporidian parasites in Andean house wrens: evidence for regional co-diversification by host-switching. *J. Avian Biol.* **2014**, *45*, 374–386. [https://doi.org/10.1111/jav.00375]
42. Beadell, J.S.; and Fleischer, R.C. A restriction enzyme-based assay to distinguish between avian hemospordians. *J. Parasitol.* **2005**, *91*, 683–685. [https://doi.org/10.1645/GE-3412RN] [https://pubmed.ncbi.nlm.nih.gov/16108566/]
43. Kimura, M.; Dhondt, A. A.; Irby J, L. Phylogeographic structuring of *Plasmodium* lineages across the North American range of the house finch (*Carpodacus Mexicanus*). *J. Parasitol.* **2006**, *92*, 1043–1049. [https://doi.org/10.1645/GE-639R.1] [https://pubmed.ncbi.nlm.nih.gov/17152948/]
44. Ishak, H.D.; Dumbacher, J.P.; Anderson, N.L.; Keane, J.J.; Valkiūnas, G.; Haig, S.M.; Tell, L.A.; Sehgal, R.N. Blood parasites in owls with conservation implications for the Spotted Owl (*Strix occidentalis*). *PloS One.* **2008**, *3*, e2304. [https://doi.org/10.1371/journal.pone.0002304]
45. Martinsen, E.S.; Perkins, S.L.; Schall, J.J. A three-genome phylogeny of malaria parasites (*Plasmodium* and closely related genera): evolution of life-history traits and host switches. *Mol. Phylogenet. Evol.* **2008**, *47*, 261–273. [https://doi.org/10.1016/j.ympev.2007.11.012] [https://pubmed.ncbi.nlm.nih.gov/18248741/]
46. Martinsen, E.S.; Sidor, I.F.; Flint, S.; Cooley, J.; Pokras, M.A. Documentation of Malaria Parasite (*Plasmodium* spp.) Infection and Associated Mortality in a Common Loon (*Gavia immer*). *J. Wildl. Dis.* **2017**, *53*, 859–863. [https://doi.org/10.7589/2016-08-195] [https://pubmed.ncbi.nlm.nih.gov/28665230/]
47. Galen, S.C.; Nunes, R.; Sweet, P.R.; et al. Integrating coalescent species delimitation with analysis of host specificity reveals extensive cryptic diversity despite minimal mitochondrial divergence in the malaria parasite genus *Leucocytozoon*. *BMC Evol. Biol.* **2018**, *18*, 128 [https://doi.org/10.1186/s12862-018-1242-x]
48. Turcotte, A.; Bélisle, M.; Pelletier, F.; Garant, D. Environmental determinants of haemosporidian parasite prevalence in a declining population of Tree swallows. *Parasitology.* **2018**, *145*, 961–970. [https://doi.org/10.1017/S0031182017002128] [https://pubmed.ncbi.nlm.nih.gov/29166965/]
49. Pulgarín-R, P.C.; Gómez, C.; Bayly, N.J.; et al. Migratory birds as vehicles for parasite dispersal? Infection by avian haemosporidians over the year and throughout the range of a long-distance migrant. *J. Biogeogr.* **2019**, *46*, 83–96. [https://doi.org/10.1111/jbi.13453]
50. Bueno, M.G.; Lopez, R.P.; de Menezes, R.M.; Costa-Nascimento, M., Lima, G.F.; Araújo, R.A.; Guida, F.J., Kirchgatter, K. Identification of *Plasmodium relictum* causing mortality in penguins (*Spheniscus magellanicus*) from São Paulo Zoo, Brazil. *Vet. Parasitol.* **2010**, *173*, 123–127. [https://doi.org/10.1016/j.vetpar.2010.06.026] [https://pubmed.ncbi.nlm.nih.gov/20638795/]
51. Tostes, R.; Dias, R.; de Oliveira, L.; Senra, M.; Massard, C.L.; & D'Agosto, M. Molecular and Morphological Characterization of a Brazilian Lineage of *Plasmodium* (Novyella) Unalis in *Turdus* Spp. (Passeriformes) of the Atlantic Forest, with Remarks on New Hosts and High Genetic Variation. *J. Parasitol.* **2018**, *104*, 70–78. [https://doi.org/10.1645/16-189] [https://pubmed.ncbi.nlm.nih.gov/28930498/]

52. Vanstreels, R.; Dutra, D.A.; Ferreira-Junior, F.C.; Hurtado, R.; Egert, L.; Mayorga, L.; Bhering, R.; Braga, É.M.; Catão-Dias, J.L. Epidemiology, hematology, and unusual morphological characteristics of *Plasmodium* during an avian malaria outbreak in penguins in Brazil. *Parasitol. Res.* **2019**, *118*, 3497–3508. [<https://doi.org/10.1007/s00436-019-06459-8>] [<https://pubmed.ncbi.nlm.nih.gov/31720833/>]
53. Marzal, A.; García-Longoria, L.; Cárdenas Callirgos, J.M.; Sehgal, R.N.M. Invasive avian malaria as an emerging parasitic disease in native birds of Peru. *Biological Invasions*. **2014**, *17*, 39–45. [<https://doi.org/10.1007/s10530-014-0718-x>]
54. Dodge, M.; Guers, S.L.; Sekercioğlu, Ç. H.; Sehgal, R.N. North American transmission of hemosporidian parasites in the Swainson's thrush (*Catharus ustulatus*), a migratory songbird. *J. Parasitol.* **2013**, *99*, 548–553. [<https://doi.org/10.1645/GE-3134.1>] [<https://pubmed.ncbi.nlm.nih.gov/23030456/>]
55. Martínez, J.; Vásquez, R.; Venegas, C.; Merino, S. Molecular characterisation of haemoparasites in forest birds from Robinson Crusoe Island: Is the Austral Thrush a potential threat to endemic birds? *Bird Conserv. Int.* **2015**, *25*, 139–152. [<https://doi.org/10.1017/S0959270914000227>]
56. Jones, M.R.; Cheviron, Z.A.; Carling, M.D. Spatial patterns of avian malaria prevalence in *Zonotrichia capensis* on the western slope of the Peruvian Andes. *J. Parasitol.* **2013**, *99*, 903–905. [<https://doi.org/10.1645/12-147.1>] [<https://pubmed.ncbi.nlm.nih.gov/23517316/>]
57. Mantilla, J.S.; González, A.D.; Lotta, I.A.; Moens, M.; Pacheco, M.A.; Escalante, A.A.; Valkiūnas, G.; Moncada, L.I.; Pérez-Tris, J.; Matta, N.E. *Haemoproteus erythrogravidus* n. sp. (Haemosporida, Haemoproteidae): Description and molecular characterization of a widespread blood parasite of birds in South America. *Acta Trop.* **2016**, *159*, 83–94. [<https://doi.org/10.1016/j.actatropica.2016.02.025>] [<https://pubmed.ncbi.nlm.nih.gov/26995696/>]
58. Cuevas, E.; Vianna, J.A.; Botero-Delgadillo, E.; Doussang, D.; González-Acuña, D.; Barroso, O.; Rozzi, R.; Vásquez, R.A.; Quirici, V. Latitudinal gradients of haemosporidian parasites: Prevalence, diversity and drivers of infection in the Thorn-tailed Rayadito (*Aphrastura spinicauda*). *Int. J. Parasitol. Parasites Wildl.* **2019**, *11*, 1–11. [<https://doi.org/10.1016/j.ijppaw.2019.11.002>] [<http://www.ncbi.nlm.nih.gov/pmc/articles/pmc6920315/>]